

Floral Scents of a Deceptive Plant Are Hyperdiverse and Under Population-Specific Phenotypic Selection

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Title: Floral scents of a deceptive plant are hyperdiverse and under population-specific phenotypic selection

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ABSTRACT

Floral scent is a key mediator in plant-pollinator interactions; however, little is known to what extent intraspecific scent variation is shaped by phenotypic selection, with no information yet in deceptive plants. We collected inflorescence scent and fruit set of the deceptive fly-pollinated *Arum maculatum* from various populations north vs. south of the Alps. We recorded 291 scent compounds and found that scent and fruit set differed north vs. south of the Alps. Seven and two compounds were under phenotypic selelection in the largest northern and southern population respectively, among them none of the four presently known pollinator attractants of *A. maculatum*. Our study provides the highest number of scent compounds so far reported in a single plant species and provides evidence for the first time that floral scents of a deceptive species are under phenotypic selection. It suggests that regional differences in scent are only partly due to divergent selection.

<u>Key words:</u> *Arum maculatum*, brood-site deception, chemical ecology, geographic variation, hyperdiverse floral scent, phenotypic selection, Psychodidae

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INTRODUCTION

About 88% of angiosperms are cross-pollinated by animals¹ that are attracted to flowers by multifaceted cues². Together with visual cues, the main attractant for pollinators is floral scent^{3,4}. Therefore, scent has strong effects on pollinator visitation and frequency, and hence the plant's reproductive success^{3,5}. With more than 2,000 floral volatile organic compounds (VOCs) described^{4,6}, and an average of 20–60 VOCs per species⁷, floral scent blends can tremendously vary among species in composition and quantity. Consequently, they facilitate discrimination by pollinators among species and contribute to reproductive isolation of (closely related) species^{8–11}.

In addition to interspecific variation, floral scent is also known to vary intraspecifically, both within and among populations^{5,12–16}. Such intraspecific variability might result directly from abiotic (e.g., temperature¹⁷, soil chemistry^{18,19}) and/or biotic factors (e.g., herbivores^{20,21}, microbes²²). Given that scent is heritable^{23–25}, intraspecific differences can also result from varying evolutionary forces, such as natural selection and genetic drift^{4,26,27}.

Although not explicitely demonstrated, genetic drift was suggested to be responsible for strong inter-population differences in floral scents⁵, or to counteract pollinator-mediated selection in two *Yucca* species²⁸). Natural selection on floral scent emission has been shown, both on total scent amount and on individual scent components, by analyses of phenotypic selection by correlating scent phenotypes and fitness measures^{29–36}. Phenotypic selection on floral scent can also vary intraspecifically, potentially leading to variable adaptive responses to spatially different pollinators^{32,36,37}. Until now, studies examining phenotypic selection on floral scent have tested rewarding, but not deceptive species, although the latter also often rely on luring and deceiving their pollinators with scents^{38–40}. Compared to their rewarding relatives, non-rewarding species often display higher variation in scent and other traits attractive to pollinators^{41–43}, and are frequently more pollen-limited^{44–46}. In consequence, they might experience stronger selection on floral scent than rewarding species, as shown for phenotypic floral traits other than scent⁴⁷. An ideal target for studying phenotypic selection on scent is the fly-pollinated and brood-site deceptive *Arum maculatum* L. (Araceae), which attracts their pollinators by olfactory deception.

This perennial herb is widespread in Europe and shows high variation in fruit and seed sets within and among populations^{48,49}. The main pollinators are two psychodid flies (*Psychoda phalaenoides* L. and *P. grisescens* Tonn., Psychodidae) that are attracted by the strong, dunglike inflorescence scent of *A. maculatum* while looking for oviposition sites and/or mating partners^{50–52}. Previous analyses have shown that the scent profile of *A. maculatum* consists of

up to 60 components, also differing among populations 53-58. At least in part, this scent variation appears to reflect variation in the pollinator community of *A. maculatum* across its distribution range 55,58. In Central and much of Western Europe, high abundances of a single *Psychoda* species and sex (females of *P. phalaenoides*) were found 52. In other regions (Mediterranean Europe and Western France), insects occurred in lower abundances but with a higher diversity (mostly both sexes of *P. grisescens* and females of *P. phalaenoides*, plus a few other psychodid species), also with some variation among populations 52. This geographic pollinator pattern is particularly pronounced north vs. south of the Alps 52, D. Laina *et al.*, unpublished and matches a weak genetic (AFLP) subdivision of *A. maculatum* across this geographic barrier 59. Presently, it is unclear whether those regional pollinator and genetic patterns between populations of *A. maculatum* are also reflected in their scent patterns. It is known, however, that the two main pollinating psychodid species have dissimilar floral scent preferences 55,58. Hence, we assume that the dissimilar scent preferences of the two fly species, together with their geographic north-south distribution, could have led to differing selection pressures on scent of *A. maculatum* from north vs. south of the Alps.

In this study, we investigated the floral scent characteristics and fruit set (as an indicator for female fitness) of *A. maculatum* in six populations north of the Alps vs. five populations south of the Alps and tested for phenotypic selection on scent in the two most extensively sampled populations, one per region. Specifically, we asked: (1) Do scent and fruit set differ between populations north vs. south of the Alps, and among populations within regions? (2) Is there phenotypic selection on floral scent? (tested in the most extensively sampled populations north and south of the Alps) And if so, (3) do compounds under selection differ between these populations? Considering the differences in pollinator compositions between regions and also among populations south of the Alps, but not north⁵², we expect to find pronounced differences in scent between the two studied regions but also among southern populations. Considering further regional differences in pollinator spectra and abundance, as well as the different olfactory preferences of pollinator species, we additionally expect lower fruit set and stronger selection on more scent components in the most extensively sampled population south of the Alps, compared to the northern conspecific.

RESULTS

Floral scent

The total absolute amount of scent was highly variable among the 234 sampled individuals (range: 1-2,052 ng inflorescence⁻¹ h⁻¹; Table 1). Across plants from north of the Alps, we overall detected a three-fold lower amount than from those in the South, along with differences among populations within regions (permANOVA: *region*: pseudo- $F_{1,223} = 25.70$, *population* nested within *region*: pseudo- $F_{9,223} = 5.36$, both P < 0.001). From plants of most southern populations, we detected a median of c. 200 ng inflorescence⁻¹ h⁻¹, except for DAO and UDI, where we detected 1.5-fold higher and five-fold lower amounts, respectively. In the North, we detected between 40 and 81 ng inflorescence⁻¹ h⁻¹ in half of the populations, while in the other half amounts were manifold higher (JOS and HOH) or lower (BUR) (Table 1).

Across all scent samples, we detected a total of 291 floral volatiles (283 north vs. 265 south), and 92 of those could be (tentatively) identified (Table **1**, and Supporting Information Table **S2**). A median of 102 compounds per individual was recorded (Fig. **2**), and the number of compounds was independent of the region (permANOVA: pseudo- $F_{1,223} = 1.68$, P = 0.21), but varied among populations within regions (pseudo- $F_{9,223} = 4.47$, P < 0.001). At the population level, between 166 (BUR) and 266 (JOS) compounds were recorded in the North, and between 88 (MON) and 254 (DAO) in the South (Fig. **2**). The two most extensively sampled northern (JOS) vs. southern (DAO) populations covered 92% vs. 87% of their respective regional diversity (Fig. **2**), and together 97% (283/291) of the total number of compounds (Table **1**, and Supporting Information Table **S2**). The five most frequent compounds, found in more than 99% of the samples, were the nitrogen-bearing compound indole, the monoterpenoids 3,7-dimethyloct-1-ene and β -citronellene, the sesquiterpenoid β -caryophyllene, and the unidentified UNK1492 (Table **1**).

The absolute amounts of single compounds significantly differed both between regions (permANOVA: pseudo- $F_{1,223} = 22.52$, P < 0.001) and among populations within regions (pseudo- $F_{9,223} = 6.44$, P < 0.001), but differences were more pronounced between regions (*north* vs. *south* OOB error: 9.9%) than among populations (within *north* OOB error: 28.3%; within *south* OOB error: 25.2%). Only a few abundant compounds dominated the scent bouquet of *A. maculatum*, including indole, β -citronellene, the unknown UNK1415, and 3,7-dimethyloct-2-ene (in both regions), p-cresol (only north), and 2-heptanone (only south) (Table 1). The absolute scent patterns among populations could not be explained by their geographic distances (Mantel's *Rho*: 0.108, P = 0.25).

We also detected differences in the relative amounts of scent components between regions (permANOVA: pseudo- $F_{1,223} = 30.34$, P < 0.001) and among populations (pseudo- $F_{9,223} = 4.87$, P < 0.001; Fig. 3). This scent pattern was also more distinct between regions than among populations within regions (*north* vs. *south* OOB error: 8.2%; among populations within *north* OOB error: 33.0%; among populations within *south* OOB error: 25.2%; see also Supporting Information Figure S1). These relative patterns among populations were also not related to population geographic distances (Mantel's *Rho*: -0.154, P = 0.85).

Based on the *randomForest* analyses, from the 25 compounds each that were most responsible for regional differences in the absolute and relative data, respectively, 20 were common to both datasets (Supporting Information Table S3). Among the latter were 2-heptanone, 2-heptanol, and α - and β -citronellene, all of which were more abundant (in relative and absolute amounts) south of the Alps. Others, such as 1-pentadecanol, the unknown UNK1503, *p*-cresol, and indole occurred in higher amounts north of the Alps (Table 1, and Supporting Information Tables S2, S3). Many of these compounds, but also some non-overlapping compounds (absolute: α -copaene, β -caryophyllene; relative: UNK1409, bicyclogermacrene) generally explained most variation in scent among all the samples (for relative data, see Fig. 3; for absolute data, see Table S4).

Despite the differences in scent between regions and among populations, we also observed considerably high variation in scent within populations, most prominently in the most extensively sampled northern (JOS) and southern (DAO) populations, which harboured almost all of the absolute and relative scent variation of their respective region (for relative data, see Fig. 3).

Fruit set

- Of the 234 individuals surveyed for inflorescence scent, 113 set fruit in summer. Percentages
- of fruit set were significantly higher north (mean \pm sd: $42 \pm \%$, 0–100% Min–Max) than south
- 123 (mean \pm sd: 26 \pm ?%, 0–100% Min–Max) of the Alps (Fig. 4; region: $F_{1,209} = 10.11$, P = 0.002),
- and differed significantly among populations within regions (Fig. 4; population nested within
- 125 region: $F_{8,209} = 2.23$, P = 0.03).

Phenotypic selection on scent

- 128 Of the 19 and three volatile compounds that were tested for β and γ -selection in the two most
- extensively sampled populations north (JOS) vs. south (DAO) of the Alps, respectively, seven
- showed signals of linear β -selection (two of which as a significant interaction), all in the north,

and two of nonlinear γ -selection, all in the south (Fig. 5). Six volatiles correlated positively (positive β -gradient) with relative fruit set (i.e., 2-nonanol, 2-heptanol, α -terpinene, UNK681, and the interaction of UNK1496 with UNK1503), and one (UNK960) negatively, and of the two volatiles with significant γ -gradients one had a positive (sabinene) and the other one (4-terpinenol) a negative gradient (Fig. 5 and Supporting Information Table S5).

Only three compounds that strongly contributed to the absolute (and relative) differences in scent between the regions were under selection (north: 2-heptanol, 2-nonanol, UNK681; Supporting Information Table **S3**), but not others (e.g., 2-heptanone, α - and β -citronellene). Differences in absolute and relative scent traits between the northern JOS and the southern DAO remained significant, regardless of performing the analyses exclusively with the 22 volatiles that correlated with fitness in the pre-selective analyses (absolute: pseudo- $F_{1,109}$ = 12.8, relative: pseudo- $F_{1,109}$ = 21.9, both P < 0.001), or with the 80 compounds that did not (absolute: pseudo- $F_{1,109}$ = 10.0, relative: pseudo- $F_{1,109}$ = 25.2, P < 0.001 each) (see Supporting Information Fig. **S2**).

DISCUSSION

Our study shows that *Arum maculatum* has hyperdiverse inflorescence scents that differ in their composition between populations north vs. south of the Alps. In contrast to our expectations, scent differed not only among populations within the southern but also within the northern region. As expected, individuals from southern populations had lower fruit set than northern ones, yet more volatiles were under phenotypic selection in the most extensively sampled northern population than in the southern one.

Hyperdiversity of floral scent

With 291 floral volatiles recorded, the inflorescence scent diversity of *A. maculatum* is extraordinarily high and, to the best of our knowledge, not matched by any other plant species. In fact, we are not aware of any species from which more than 200 floral compounds are reported, a number that a single *A. maculatum* individual can reach by three quarters (max. = 147 VOCs). This difference in the number of scent compounds between *A. maculatum* and other species cannot just be explained by differences in techniques used for scent analyses, given that scents of a high number of species were analysed using a similar approach as we did (dynamic headspace and thermal desorption of samples). Species closest to the high number in *A. maculatum* include the sapromyiophilous *Sauromatum guttatum* (Araceae, with altogether 196

different VOCs^{60,61}), as well as the insect-pollinated and rewarding *Geonoma macrostachys* (Arecaceae, 176 VOCs⁶²), and *Echinopsis ancistrophora* Cactaceae, 145 VOCs⁶³). Other species for which *c*. 100 VOCs are described likewise include insect-pollinated rewarding species (e.g., *Acleisanthes wrightii*, Nyctaginaceae⁶⁴; *Saraca asoca*, Fabaceae ⁶⁵; *Philodendron bipinnatifidum*, Araceae⁶⁶; Pyrus communis, Rosaceae⁶⁷), but also the sexually deceptive orchid *Ophrys sphegodes*⁶⁸. Thus, high numbers of compounds are found across a wide range of plant families and are apparently not restricted to a specific pollination system.

One explanation for the high diversity of scent compounds in *A. maculatum* is that this species likely imitates various breeding substrates of its pollinators, all potentially differently scented. The two main pollinators, *Psychoda phalaenoides* and *P. grisescens*, breed in rotting manure from cattle and horse, fungi (*P. grisescens*), waste pits, mud-flats, plant litter in drainages and ditches (*P. phalaenoides*), and in the hygropetric zones of river banks and ponds^{69–76}. *Arum maculatum* emits compounds described from quite a number of such substrates, e.g., cattle and horse manure (e.g., indole, *p*-cresol, skatole), fungi (1-octen-3-ol, (*E*)-2-octen-1-ol, 3-octanone), and general degrading and fermenting plant or animal material (e.g., 2,3-heptanedione, acetoin, butanoic acid)^{40,53,77,78}. Highly specialised deceptive plant systems frequently rely on only a few volatiles to attract pollinators – they seem to imitate a more specific model, and thus, release less complex scent blends^{e.g. 40,79–82}.

The number of volatiles detected across the 235 indivduals (11 populations) of A. maculatum (291) is five to ten times higher than previously reported for this species (18–61, and 143 VOCs in total^{53–58,83}). This discrepancy cannot be explained by differences in sample size, as a similar number of individuals were surveyed in previous studies (n = 222 in total, representing 23 populations). Interestingly, we found in some single individuals a similar number of compounds (up to 147 VOCs; median of 102) as overall detected previously. With the exception of two studies^{56,58} who each share one of our sampled populations (JOS and MON, respectively) all previous studies sampled scents in other populations. Thus, some of the differences in the number of compounds detected across studies might reflect region-specific population characteristics (see Fig. 2). More importantly, however, we believe that the discrepancy in the number of compounds recorded largely reflects differences in methodology between the present and previous studies. These are, for example, higher sensitivity of modern GC/MS systems; usage of more selective adsorbent agents (Carbotrap/Tenax-TA vs. solidphase micro-extraction^{55,57} vs. Twister⁵⁸); in situ vs. ex situ sampling^{56,83}; and including all vs. only compounds above a specific threshold in relative amounts^{55,58}. Of the 92 compounds (tentatively) identified in this study, more than half (50) were previously unknown to be

released by *A. maculatum*. Some of these newly described compounds for *A. maculatum* are known from other species of Araceae (e.g., nerol, (E,E)- α -farnesene, 6-methyl-5-heptene-2-ol, γ -terpinene; *Sauromatum guttatum*⁶⁰, *Anthurium* spp. ^{84,85}) or other plant families (e.g., methyl anthranilate, isobutyl butyrate, citronellal^{4,6}). However, this study is, to the best of our knowledge, the first to identify *p*-cresyl butyrate as a floral scent compound.

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Geographic patterns of floral scent

The qualitative, absolute and relative differences in scent detected herein among populations of A. maculatum south of the Alps may be explained by the fact that pollinator spectra are more diverse in abundance, species and sex composition south of the Alps^{52, D. Laina et al., unpublished}. North of the Alps, however, where females of P. phalaenoides are the principal pollinators in all populations^{52, D. Laina et al., unpublished}, even though the other psychodid species also occur in small numbers^{55,86}, the variation in scent among populations is not reflected by variations in pollinator spectra. In our study, all multivariate scent patterns were more pronounced between regions compared to variation among populations within each region. These differences in scent across the Alps are in agreement with strong differences in pollinator spectra 52, D. Laina et al., unpublished and a genetic (AFLP) differentiation of A. maculatum across this geographic barrier⁵⁹, possibly reflecting different colonisation histories during the Pleistocene⁸⁷. Previous studies in A. maculatum also found population effects in scent composition^{55,57,58}; however, our study demonstrates for the first time such population differentiation in scent across the Alps. Interestingly, such regional differences in scent were not recorded in Szenteczki et al.⁵⁸, although the authors analysed inflorescence scents in A. maculatum across Europe, including some populations north and south of the Alps. Intraspecific scent differences between regions and among populations are also known for other plant species 14,15,37,63,88. For some of those species it has been shown that observed scent patterns, as in our study, reflect pollinator and/or genetic patterns^{15,37,88}, but not for others^{14,63}.

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Phenotypic selection on floral scents

In the present study, the absolute and relative scent differences found between the two most extensively sampled populations of their respective region were evident, independent of whether all compounds, only compounds that correlated with fruit set in the pre-selective analyses, or only compounds that did not, were included in the analyses (Supporting Information Fig. S2). Hence, this regional scent pattern appears to be a result of both divergent selection and of other reasons, such as phenotypic plasticity but see58 or genetic drift 89,90. In

support of divergent selection, we detected population-specific signatures of direct selection on scent, potentially as a result of the different olfactory preferences of P. phalaenoides and P. grisescens^{55,58}, the two species most responsible for the regionally different pollinator spectra⁵², D. Laina et al., unpublished. These different selection regimes, however, cannot explain the most obvious differences in scent between A. maculatum from north vs. south of the Alps (see also Supporting Information Fig. S1). This is because most compounds identified as being mainly responsible for those regional differences (e.g., 2-heptanone, 3,7-dimethyloct-1-ene, UNK966; Supporting Information Table S3) did not show signals of direct selection (see Fig. 5). Some other compounds, however, which also differed in their absolute amounts between regions (2heptanol, 2-nonanol, UNK681, sabinene; Supporting Information Table S3, S5) differed in their selection in JOS and DAO (Fig. 5), and some of the differences between regions might thus be due to divergent selection. Interestingly, β -selection on scent was only detected in the population north of the Alps, the region which attracts higher numbers of pollinators and predominantly a single species, i.e. females of P. phalaenoides^{52, D. Laina et al., unpublished}, and has a higher fruit set than south of the Alps (Fig. 4). The preferences by a more consistent single pollinator species might exert strong selection pressures and result in stronger (and thus better detectable) phenotypic selection on specific compounds in the most extensively sampled population north of the Alps, compared to south. There, a more diverse composition of pollinator species^{52, D. Laina et al., unpublished} as it occurs in South of the Alps and their putative different scent preferences^{55,58} might lead to conflicting selection pressures^{33,34,36}.

For the five compounds under phenotypic selection that could be identified by authentic standards (Supporting Information Table **S2**), i.e., 2-nonanol, 2-heptanol, sabinene, 4-terpinenol and α -terpinene, we lack information on their attractiveness to psychodid pollinators of *A. maculatum*. However, the aliphatic components 2-heptanol and 2-nonanol together are found as (sex-)pheromones of female Diptera (Cecidomyiidae⁹¹) and female non-Diptera (Trichoptera⁹²), and as an attractant for bees (Meliponini⁹³). The volatile 2-nonanol alone also attracts kleptoparasitic flies⁹⁴ and is attractive for other Meliponini⁹⁵, where it is used for host-finding and as a pheromone. The monoterpenoids α -terpinene, 4-terpinenol, and sabinene are defence substances of some insects (Coleoptera^{96,97}, Lepidoptera⁹⁸) that repel Coleoptera⁹⁹, but are used by Hymenoptera¹⁰⁰ and Lepidoptera¹⁰¹ for host-finding (beetles) and as an oviposition stimulant. Also, α -terpinene and 4-terpinenol are pheromones in fruit flies¹⁰². It would be very interesting to know whether these five compounds, which are all widespread among floral scents^{4,6}, are attractive to pollinators of *A. maculatum*, especially because they elicit responses in other insects, are known from other sapromyiophilous species^{60,61,77,80,103,104}, and some of

them (α -terpinene and 4-terpinenol) are also known from cattle dung^{105, E. Gfrerer, unpublished}, i.e., one of the oviposition substrates of psychodid flies.

Somewhat unexpectedly, we did not find positive β -selection gradients for the most abundant compounds in the scent of A. maculatum (e.g., β -citronellene, β -caryophyllene, unknown UNK1415), with the expection of 2-heptanol (Fig. 5). Even more surprisingly, we also did not find phenotypic selection for those known to attract P. phalaenoides (i.e. indole, 2heptanone, p-cresol, α -humulene^{50,83}), one of the two main pollinators of this plant species, occurring both north and south of the Alps⁵². This is in contrast to most other studies, where main compounds and/or pollinator attractants showed signals of β -selection and had positive gradients but see 31,34. Possible explanations for these findings are that the main compounds may generally be released in amounts high enough to result in maximum pollinator attractiveness^{see} also 34, that the relationship between the compound and different selection agents is conflicting, i.e., opposing selection by different pollinator species or herbivores^{33,34}, or that different amounts of the compounds have different effects on flower visitors, i.e., the relationship is nonlinear 106–108. Support for the latter hypothesis comes from our nonlinear quadratic analyses, where the amounts of sabinene and 4-terpinenol were correlated with relative fruit set in the southern population DAO (Fig. 5). Although we detected direct nonlinear selection by including quadratic terms in our multivariate models, these quadratic analyses do not uncover all potential nonlinear relationships^{e.g.109}. Hence, we cannot exclude the possibility that such compounds are still under phenotypic selection, which in turn calls for future approaches that allow testing for any kind of nonlinear multivariate relationships.

Deceptive plant species might experience stronger selection than rewarding ones⁴⁷. In deceptive *A. maculatum*, we did not find a higher number of volatiles with signatures of selection (direct: 7% vs. 3–42%), but slightly stronger positive β -selection (-0.3–0.5 Min–Max vs. -0.3–0.4) compared to other, rewarding, species^{30–35}, and a stronger γ -selection (-0.9–9.0 Min–Max vs. -0.5–0.3³⁶, Fig. 5). Future studies on other deceptive plant species that also attract specific pollinators by chemical cues, but have lower levels of fruit set, might reveal even stronger signatures of direct selection.

Conclusions

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Our study on sapromyiophilous *Arum maculatum* reports the highest number of floral volatiles ever found in a single plant species to date. This plant species likely imitates the odours of a multitude of differently scented breeding substrates of its psychodid fly pollinators (e.g., dung, fungi, rotting plant material), which might result in this chemical hyperdiversity. We recorded

pronounced scent differences between populations across the Alps, and this geographic pattern in scent agrees with previously described pollinator and genetic patterns across this geographic barrier. Our results provide for the first time evidence that floral scents of a deceptive species are under phenotypic selection and suggest that populational/regional differences in scent are partly due to divergent selection, but also due to other reasons, such as phenotypic plasticity and genetic drift. In *A. maculatum* and other plants where phenotypic selection on scent was demonstrated^{29–36} the biological role of most compounds under selection is unknown and awaits determination in future studies.

MATERIALS AND METHODS

Study species and populations

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Brood-site deceptive Arum maculatum L. (Araceae) is a rhizomatous perennial woodland herb (2n = 4x = 56) that is widespread throughout Western and Central Europe, including the British Isles, and reaches as far south as Italy, Northern Spain, and the Balkans^{52,110,111}. It exhibitsa sapromyiophilous pollination strategy, is thermogenically active, and emits a strong dung-like scent for attracting fly pollinators during the evening on the first day of anthesis 53,56,112,113. The inflorescence of Arum maculatum consists of a spadix (fleshy spike) and a spathe (bract), is protogynous, and the anthesis lasts less than two days^{51,56,112}. The spathe, which completely encloses the spadix during floral development, partially opens during anthesis to reveal the sterile appendix of the apical part of the spadix. This appendix produces and releases the scent for pollinator attraction^{50,51,53,83}. At the base of the spadix, female (fertile and sterile) flowers are situated lowest, followed upwards by male flowers and staminodes (sterile male flowers). The sequential maturation of female (first) and male flowers (later) prevents selfing¹¹⁴. All flowers remain enveloped by the spathe during anthesis, forming a chamber that is closed by the staminodes throughout the female stage to prevent trapped insects from leaving. Pollinators are attracted in the evening on the first day of anthesis, during the female stage, slip and fall into the floral chamber, and are trapped overnight^{51,52,113,115}. On the next morning, during the male stage, they are dusted with pollen, before being released at around noon when the staminodes wither^{51,52,112}. After pollination in spring, red berry-like fruits develop as an infructescence until summer¹¹⁶.

In 2017–2019, during springtime, we collected scent from randomly chosen A. maculatum individuals of six populations located north of the Alps (n = 107; Northwestern Austria: JOS; Central/Southern Germany: BUR, HOH, MUR, NEC; Northern Switzerland:

RÜM) and from five populations south of the Alps (n = 127; Northern Italy: DAO, LIM, MAH, MON, UDI) (Fig. 1). We kept a minimum distance of one metre between sampled individuals to avoid sampling potential clones, as $Arum\ maculatum$ can reproduce vegetatively. In summer, we harvested fruits of the same plant individuals. At most sites, we surveyed 15 individuals, except for each of the largest population per region (JOS and DAO; n = 70 each), and a northern population (HOH; n = 7) where only a few individuals flowered at the time of sampling (Fig. 1 and Supporting Information Table S1).

Plant volatile collection and analysis

Scent sampling took place on the first day of anthesis during the female stage between 18:00 and 19:30, the period of maximum scent emission⁵⁶, employing a non-invasive dynamic headspace technique. We enclosed each inflorescence *in situ* using an odourless plastic oven bag (c. 30×12 cm; Toppits[®], Melitta, Germany) and immediately collected scent for five minutes at 200 ml min⁻¹ on adsorbent tubes (inner diameter: 2 mm) filled with a mixture of Tenax-TA (mesh 60–80) and Carbotrap B (mesh 20–40; 1.5 mg each; both Supelco, Germany), using a battery-operated vacuum pump (rotary vane pump G12/01 EB, Gardner Denver Austria GmbH, Vienna, Austria)⁵⁶. In the same way, we collected scent samples from leaves and ambient air as negative controls in each population.

The dynamic headspace samples were analysed by thermal desorption-gas chromatography/mass spectrometry (TD-GC/MS)⁵⁶, and obtained data were handled using *GCMSolution* v.4.41 (Shimadzu Corporation, Kyoto, Japan) (for details see Supporting Information Method S1). Components were tentatively identified by comparison of Kováts' retention indices (KRIs¹¹⁷), based on commercially available *n*-alkanes (C₇–C₂₀), and mass spectra to data available in the libraries of Adams¹¹⁸, FFNSC 2, Wiley9, NIST11, and ESSENTIAL OILS (available in *MassFinder 3*, Hochmuth Scientific Consulting, Hamburg, Germany). We established an own library of mass-spectral and KRIs for semi-automatic analysis (Supporting Information Method S1). Whenever possible, components were verified by comparison to authentic reference standards available in the collection of the Plant Ecology Lab of Salzburg University, or to chemically synthesised reference components (Supporting Information Method S2). Of the 267 collected scent samples, 234 yielded a sufficiently informative chromatogram and were included in the analysis (Fig. 1). Ultimately, a compound was only considered if it occurred in more than three scent samples and did not occur in leaf and air controls.

Fruit set

Percentage fruit set (i.e., number of fruits/total number of flowers per individual \times 100) was determined as a measure of female reproductive success. For selection analyses we further estimated relative fruit set (i.e. number of fruits per individual/ mean number of fruits per given population) as a measurement for female reproductive success, standardised per population for the most extensively sampled populations JOS and DAO^{32,35}. Around 50% of the individuals used for scent sampling failed to develop an infructescence, due to unknown reasons such as herbivory, lack of pollination, or other causes (e.g. resource limitation, shallow landslide). In consequence, we only included those plants in subsequent fruit set and selection analyses for which fruit (and scent) data were available ($n_{\text{north}} = 61$, $n_{\text{south}} = 52$). In one southern population (MON), a shallow landslide destroyed all plants, with the exception of one (Fig. 1); hence, this population was excluded from fruit set analyses.

Statistical analyses

Geographic patterns in scent and fruit set data In order to test for geographic differences in floral scent, we performed permutational multivariate analyses of variance (permANOVAs; Anderson, 2001) as implemented in the R package *vegan* v.2.6-6¹¹⁹. We did this on (1) pairwise Bray-Curtis dissimilarities of either absolute or relative scent data (i.e., absolute amount of single compounds or relative amount of single compounds in relation to the total amount of scent in a sample, respectively); and on (2) Euclidean distances of total absolute emission of scent and of total number of floral volatiles per individual. In all these analyses, we used region (north vs. south of the Alps) and population nested within region as explanatory variables (9,999 permutations). To test for geographic patterns (north vs. south Alps) in relation to selection in absolute and relative scent, we focussed on the (reduced) dataset of the two most extensively sampled populations using only compounds that correlated with fruit set in the preselective analyses (see below), or only compounds that did not, with a permANOVA (population as explanatory variable, 9,999 permutations). The Bray-Curtis dissimilarity matrices (based on absolute and relative scent data) were further used to conduct constrained analyses of principal coordinates (CAP¹²⁰) with population as factor, using the capscale function in vegan, in order to visualize similarities and dissimilarities in scent among the samples following 121,122. For each ordination, we also calculated vectors, representing compounds most correlating with the axes (Pearson correlations with *capscale scores*, r > |0.5|, corrected for false-discovery rate¹²³). Additionally, we subjected the absolute and relative data to random forest analyses (Breiman, 2001) by the R package randomForest v.4.6-14¹²⁴ (ntree = 9,999)

bootstrap samples with mtry = 17) to evaluate the distinctness in scent of northern and southern samples (factor region) and among populations within each region (factor population each)¹²⁵. Distinctness was quantified as the average out-of-bag (OOB) error estimate (in %), i.e., the more distinct, the lower the OOB error. From the resulting randomForest objects, we further extracted the importance measurements to determine volatiles crucial for regional distinction.

To test for relationships between the dissimilarity of median absolute and relative scent properties of populations and their geographical distances (in km), we performed Mantel tests with the function *mantel* in *vegan* (9,999 permutations, Spearman's rank correlation). To assess whether absolute amounts of single compounds under selection (see below) differ between the two regions, we performed Mann–Whitney U tests. Differences in percentage of fruit set across regions and among populations within regions were assessed by a generalised linear model (*regions* and *populations* nested within *regions* as factors).

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Analyses of phenotypic selection To estimate the direction and strength of phenotypic selection on individual scent compounds and on total amount of scent, we tested for direct phenotypic selection in the two most extensively sampled populations north (JOS) and south (DAO) of the Alps¹²⁶. These two populations cover a large part of their corresponding regional scent variation (see Fig. 3). As many compounds were quite rare (c. 70% of VOCs in <50% of samples), the scent matrix contained many zeros (non-detects). This zero-inflation can cause severe problems when fitting linear models (see below), as estimates will be biased¹²⁷. That is, the influence of an individual scent compound on fruit set can be either over- or underestimated, leading to potentially wrong conclusions. Additionally, our data has a considerable higher number of factors (VOCs) than samples. Previous studies solved such issues by pre-selecting variables^{30,31,34} to reduce high dimensionality, and/or performed selection analyses only on most abundant compounds^{29,33}, on PC-scores^{29,32,35}, or on physiologically active volatiles³⁴. As we only have very limited knowledge of attractive compounds (four volatiles^{50,83}), and also as minor volatiles can be under selection³⁴, we pre-selected volatiles that correlate with relative fruit set via elastic net, a penalised multivariate linear regression³¹. By adding a penalty to the regression (regularisation), coefficients that are not important for the model are shrunk towards zero (L_1 penalisation). An elastic net additionally selects groups of correlated variables (L_2 penalisation) instead of only one random variable from a multicollinear group of variables, as e.g. LASSO does 128. By combining L_1 with L_2 penalisation, it is well suited for datasets that have a larger number of predictor variables (in our case VOCs) than number of observations, and copes well with multicollinearity 128,129, which is advanteous as volatiles that share the same biosynthetic pathway can be highly multicollinear. Zero-inflation can still influence these analyses; thus, to quantify the impact of non-detects on estimates, we performed a simulation study based on our scent data for JOS and DAO seperateley (see Supplementary Information Notes **S3**). This approach resulted in 93 and 81 scent compounds included in the consecutive elastic net regression analyses for JOS and DAO, respectively. For the simulation and consecutive elastic net analyses 128, we used *glmnet* v.4.1 130 in R.

For calculating the elastic net regression between relative fruit set and the z-transformed variables (individual VOCs and total absolute scent emission) of the reduced data sets, we first cross-validated for α (L_1/L_2 penalty ratio, range: 0–1, 0.1 increment) and λ (penalty strength parameter; range: 100-0.001, -0.1 increment), for JOS and DAO separately. This resulted in an elastic net for the northern population JOS with $\alpha_{(JOS)}$ = 0.9 and for the southern population DAO with $\alpha_{(DAO)}=0.2$, and regression coefficients $\hat{\beta}$ were extracted at $\lambda_{min(JOS)}=0.05$ and $\lambda_{min(DAO)}$ =1.99. Pollinators might respond to different amounts of compounds in a nonlinear manner^{106–108}, potentially resulting in a nonlinear relationship of fitness and scent traits. To account for this, we ran a boruta algorithm (R package Boruta v.7.0.0¹³¹; ntree = 9,999 bootstrap samples with mtry = 9, Bonferroni-adjusted for multiple comparisons) to also identify nonlinear relationships between individual volatiles, total absolute scent emission, and relative fruit set. Boruta is a feature selection method that has shown high stability in variable selection of high-dimensional data sets¹³², and classifies whether a feature (in our case VOCs) is important or not¹³¹. In the northern population JOS, 19 volatiles emerged from the elastic net and four from the boruta analysis; all four of which were already among the 19 linear ones (Fig. 5). In the southern DAO population, no common volatile correlated with fruit set in the elastic net, but three in the *boruta* analysis. No volatile appeared in the analyses of both populations (Fig. 5). Total absolute scent amount did not correlate with fruit set in any of the analyses.

To ultimately determine β -gradients for phenotypic selection on scent (directional selection¹²⁶), we performed with those variables selected by the elastic net model (i.e. $\hat{\beta}$ coefficient > 0) a multivariate linear regression. For volatiles selected by the *boruta* we estimated multivariate quadratic regressions coefficients (γ -gradients, stabilizing and disruptive selection¹²⁶) by squaring the terms and doubling resulting estimates¹⁰⁹, to also test for nonlinear signs of phenotypic selection. For the southern model, we excluded the plant individual "DAO076", as it was determined as an outlier influencing the model (D_{DAO076}=235.4) by Cook's distance¹³³. Although elastic net handles multicollinearity well, resulting individual volatiles might still correlate with another (L_2 penalty, see above). We tested for multicollinearity within the multivariate models by calculating the variance inflation factor

(VIF) (R package car v.3.0.8¹³⁴) for each scent compound in each multivariate model. In the northern model, the VIF value of various compounds was high (> 5) and for the unknowns UNK1496 and UNK1503 even exceeded 10, a threshold that indicates strong multicollinearity¹³⁵. After including these two compounds as an interaction, the VIF of most compounds was below 5 with the exception of 3-octanol and UNK1279 (VIF > 6). Once also the interaction of the latter two volatiles was included in the model, the VIF of all volatiles dropped below 4, and the final northern model had an adjusted R² of 0.71. The southern model had only VIFs below 2, and an adjusted R² of 0.26. All statistical analyses were performed in R v.4.0.2¹³⁶.

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Author contributions

SD, MG, ACH and HPC designed the research; EG and DL conducted the fieldwork; RF executed the scent sample laboratory work; EG and SD built the scent library, and EG analysed all scent and fruit set data; TT identified and synthesised unknown components; MH, WT, RF, SD and EG discussed statistical approaches for selection analyses; MH performed the simulations, and EG all selection analyses; EG wrote the first draft of the manuscript and all authors contributed to the final version.

Data availability statement

The dataset that supports the findings of this study and the R code for the simulation are available at the Dryad digital repository at https://doi.org/10.5061/dryad.pnvx0k6kn

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Supporting Information

Additional Supporting Information may be found online in the Supporting Information section at the end of the article.

Fig. S1 Relative amounts of the most abundant compounds (see Table 1) in the inflorescence scents of *A. maculatum*, and whether or not they are under phenotypic selection.

Fig. S2 Non-metric multidimensional scaling of relative (a, b) and absolute (c, d) *Arum maculatum* scent patterns (a, c) with VOCs correlating with fitness in the pre-selective analyses and (b, d) only with VOCs that did not.

Table S1 Locality information on the sampled populations.

Table S2 Complete inflorescence scent list of *Arum maculatum* (median relative amount per population and area).

Table S3 Volatiles important for regional distinction in relative and absolute scent bouquets.

Table S4 Volatiles most correlating with the *capscale* scores in absolute scent samples.

Table S5 Direct selection gradient β and γ for volatiles of *Arum maculatum*.

Methods S1 Scent analysis (TD-GC/MS), quantification, and set-up of scent library.

Methods S2 Synthesis of reference samples: 2,6-dimethylocta-2,6-diene, 3,7-dimethylocta-2-ene and 2,6-dimethylocta-1,7-diene (α -citronellene)

Methods S3 Simulation to quantify impact of non-detects on selection estimates.

Notes S1 Mass spectrometry of unknown volatiles with significant phenotypic selection gradients.

Figures

Fig 1

Localities of the six populations north (blue) and five populations south (red) of the Alps of *Arum maculatum* sampled for the present study. Numbers in brackets give the number of individuals used for scent and selection analyses. The two most extensively sampled populations are indicated by larger circles. *North*: JOS, Josefiau; BUR, Burg Hohenstein; HOH, Hohendilching; MUR, Murnau; NEC, Horb am Neckar; RÜM, Rümikon; *South*: DAO, Daone; LIM, Limone-Piemonte; MAH, Santa Maria Hoè; MON, Montese; UDI, Udine. The map was prepared using the ETOPO1 Global Relief Model¹³⁷ and ArcGIS v.10.4 (ESRI, Redland, CA).

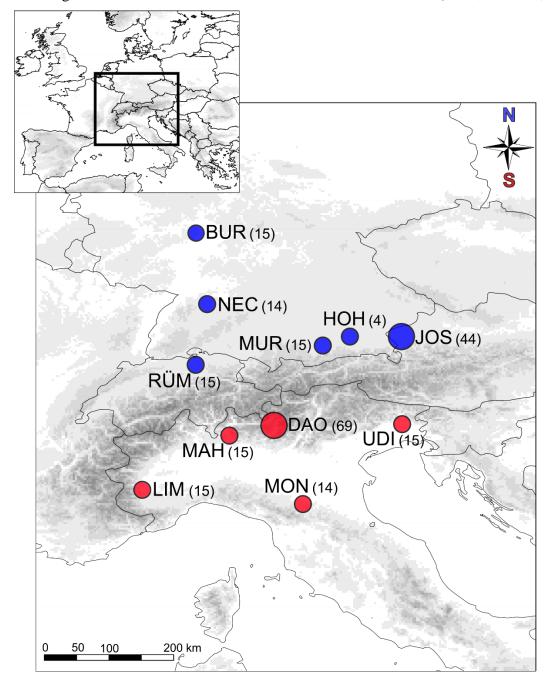
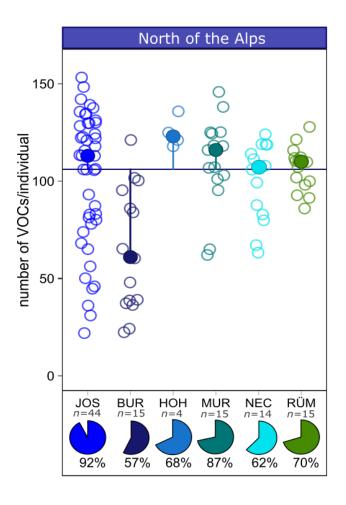


Fig 2

Number of floral scent compounds recorded in individual samples in the six and five populations of *Arum maculatum* from north and south of the Alps, respectively. Filled circles denote the population median of number of volatiles per individual; the vertical lines indicate the distance to the region median (horizontal line); and open circles mark the number of volatiles detected in the individual samples. The sample sizes and the percentage of volatiles detected in a population compared to the number of compounds detected across all samples (291 compounds) are shown as pie charts. See Fig. 1 and Table S1 for identification of population codes.



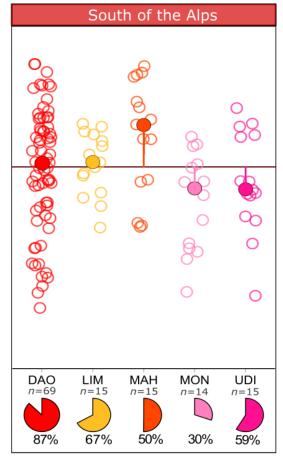
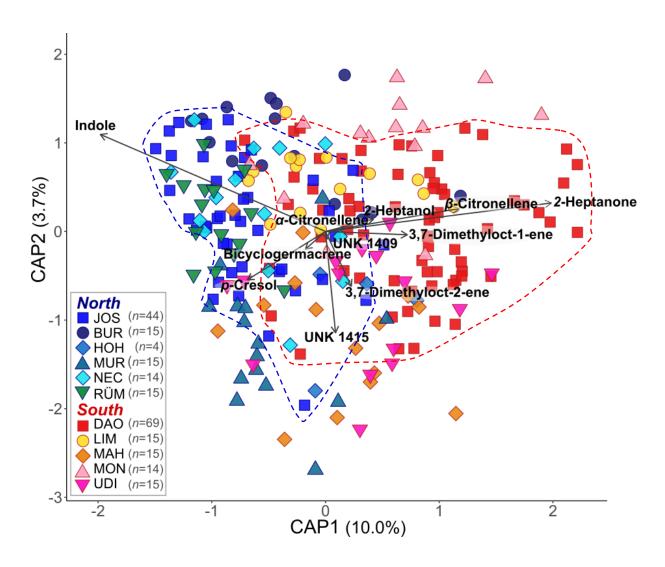


Fig 3
Canonical analysis of principal coordinates (CAP) based on a Bray–Curtis dissimilarity matrix of relative floral scent patterns in *Arum maculatum*. Between four and 69 individual samples were available across the six populations north and five south of the Alps. The vectors depict the volatiles most correlating with the *capscale* scores. See Fig. 1 and Table S1 for identification of population codes.



Fruit set (% female flowers that developed into fruit) of *Arum maculatum* individuals from populations north and south of the Alps, respectively. Filled circles denote the population mean of fruit set; horizontal lines indicate the distance to the region mean (vertical line); and the open circles mark the fruit set in the single individuals. See Fig. 1 and Table S1 for identification of

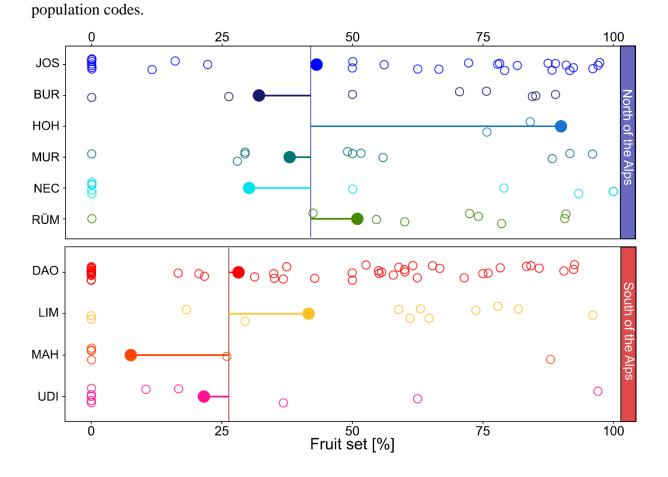
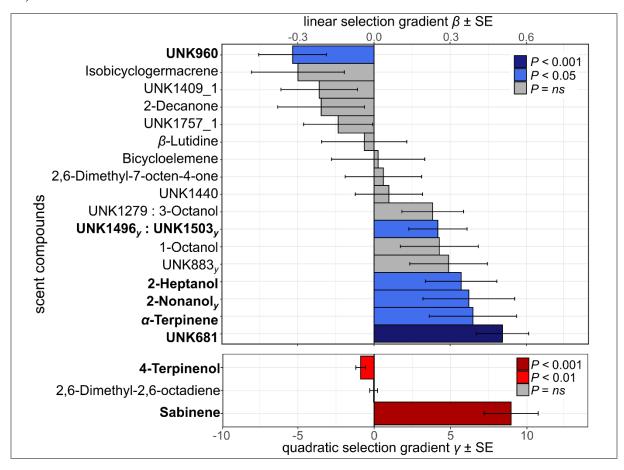


Fig 5

Linear and nonlinear selection gradients β ± SE and γ ± SE for individual floral scent compounds of *Arum maculatum* in the two most extensively sampled populations north (JOS, blue) and south (DAO, red) of the Alps, respectively. Only compounds that resulted from the pre-selective analyses are shown. Scent compounds under significant direct selection (P < 0.05) are in bold and their bars are coloured. Note the different scaling for direct β - and γ -selection. For the northern population, compounds that also correlated in the nonlinear pre-selective analyses are indicated (γ). Individual sample sizes are for JOS (n_{north} = 43) and DAO (n_{south} = 68).



Tables

Table 1 Median amounts of total absolute and relative (contribution of single compounds to total scent) inflorescence scent of *Arum maculatum* surveyed in six and five populations north and south of the Alps, respectively. North and South columns (bold headers) present the average regional median of the corresponding populations (following columns). Volatiles with a median amount of <1% in any population are pooled.

	North	JOS	BUR	НОН	MUR	NEC	RÜM	South	DAO	LIM	MAH	MON	UDI
KRI Compound name	(n=107)	(n=44)	(n=15)	(n=4)	(n=15)	(n=14)	(n=15)	(n=128)	(n=69)	(n=15)	(n=15)	(n=14)	(n=15)
Median total absolute amount of scent trapped (ng inflorescence ⁻¹ h ⁻¹)	67.4	167.2	13.0	565.8	80.7	39.4	41.7	214.7	311.4	203.8	196.9	201.4	42.3
Total number of volatiles	283	269	166	197	204	181	208	265	254	195	146	88	171
Aliphatic components													
893 2-Heptanone*	1.4	1.4	2.4	0.8	1.3	1.1	1.4	6.9	9.3	0.3	2.9	11.9	4.0
902 2-Heptanol*	0.1	0.1	0.3	0.3	0.2	0.2	0.1	1.2	1.9	tr	0.8	2.5	0.5
982 1-Octen-3-ol*	1.9	2.4	tr	2.3	2.0	1.8	1.3	0.3	0.4	tr	6.4	tr	1.3
1096 2-Nonanone*	0.2	0.1	0.1	0.1	0.2	0.1	0.2	0.7	0.9	tr	0.2	1.3	0.4
23 more aliphatic components <1%	0.5	0.5	0.4	2.0	1.7	1.3	0.5	0.7	0.9	0.9	2.0	1.0	0.7
Aromatic components													
1076 p-Cresol*	4.2	1.8	0.1	19.4	11.9	9.2	1.5	0.5	0.5	0.3	0.7	0.6	0.9
4 more aromatic components <1%	tr	tr	tr	0.6	0.3	0.1	0.1	tr	tr	tr	0.1	tr	tr
C5-branched chain components													
4 C5-branched chain components	tr	tr	tr	0.1	tr	0.2	0.1	tr	tr	tr	tr	tr	tr
<1%													
Nitrogen-bearing components													
965 β -Lutidine	0.2	0.1	0.7	0.2	0.4	0.4	0.3	0.1	tr	0.6	0.1	tr	1.3
1310 Indole*	24.2	22.3	20.8	12.6	24.6	33.4	35.6	11.9	11.9	24.8	8.8	12.3	9.5
5 more nitrogen-bearing	0.1	0.1	tr	0.1	tr	tr	0.3	0.1	tr	0.1	0.1	tr	tr
components <1%													
Irregular terpene													
3 irregular terpenes <1%	tr	tr	0.4	0.3	tr	0.6	0.1	0.1	0.1	0.2	tr	0.3	0.1
Monoterpenoids													
914 3,7-Dimethyloct-1-ene*	1.5	1.2	1.1	2.6	2.1	1.8	1.7	4.0	4.3	4.1	2.4	4.6	2.7

935	α-Citronellene*§	0.4	0.3	0.5	0.9	0.5	0.5	0.4	1.3	1.3	1.9	1.1	1.3	1.0
949	β -Citronellene*§	4.2	3.5	8.0	11.6	3.4	7.1	3.5	9.7	10.8	10.1	6.5	9.9	8.2
972	3,7-Dimethyloct-2-ene*	3.1	1.8	2.8	5.1	9.6	2.6	3.3	4.3	4.5	4.4	5.6	2.1	3.2
982	Sabinene*	0.2	tr	1.0	0.2	tr	0.4	0.4	0.4	0.3	1.4	0.3	1.2	tr
1005	2,6-Dimethylocta-2,6-diene*	1.2	0.9	1.0	3.4	4.1	1.0	1.5	1.7	1.8	1.7	1.9	0.5	1.6
1076	Dihydromyrcenol	tr	tr	tr	tr	tr	tr	tr	0.4	0.4	1.0	tr	0.2	0.5
	21 more monoterpenoids <1%	0.3	0.4	tr	2.2	0.9	0.6	0.9	0.6	0.7	1.9	1.2	0.4	1.1
Sesquiterpenoids														
1357	Bicycloelemene	0.4	0.5	0.1	0.5	1.9	0.1	0.5	0.2	0.1	0.2	0.6	0.1	0.9
1399	α-Copaene*	1.0	1.8	1.3	0.5	0.5	0.7	0.8	0.8	0.6	1.0	1.0	1.6	0.6
1434	Isocaryophyllene	0.9	1.3	0.7	0.4	0.6	0.5	1.1	0.9	0.7	1.2	1.3	1.2	0.8
1450	β -Caryophyllene*	3.0	5.5	3.0	2.3	1.4	2.7	2.7	2.9	2.2	3.0	4.0	5.3	2.8
1484	α-Humulene*	2.8	4.7	2.8	1.7	1.2	2.3	2.7	2.3	1.6	2.5	3.0	4.0	2.6
1501	Germacrene D*	0.9	1.3	1.4	0.3	0.3	0.9	0.7	0.5	0.3	0.5	0.7	1.3	0.7
1520	Bicyclogermacrene	0.9	1.0	tr	0.6	2.1	tr	1.3	0.4	0.2	0.2	1.3	tr	1.7
1547	δ -Cadinene	1.2	1.9	1.4	0.6	0.6	1.5	1.1	0.4	0.3	0.5	0.7	0.4	1.0
	10 more sesquiterpenoids <1%	0.4	0.5	0.3	0.4	0.3	0.5	0.6	0.3	0.1	0.3	0.4	0.7	0.3
Unkno	own compounds													
829	UNK 829 <i>m/z</i> : 54,67,110,41,81,39	0.3	0.8	tr	0.2	0.2	0.3	0.1	tr	tr	tr	2.0	tr	0.3
1394	UNK 1394 <i>m/z</i> : 69,55,41,82,95	0.2	0.2	tr	0.1	0.6	0.2	0.2	0.1	0.1	0.1	1.1	0.2	0.1
1409	UNK 1409_1 <i>m/z:</i> 81,55,67,95,41	0.2	0.2	tr	0.4	0.4	0.1	0.1	0.2	0.2	0.1	0.6	0.2	1.1
1415	UNK 1415 <i>m/z</i> : 69, 81,41,95,55	3.7	3.9	1.7	2.3	7.3	3.7	3.4	3.8	3.1	2.8	10.4	2.3	11.3
1492	UNK 1492 <i>m/z</i> : 105,161,91,41,93	1.7	2.7	1.4	0.3	0.5	0.5	1.8	1.2	1.0	1.4	1.6	3.1	0.6
1503	UNK 1503 m/z: 81,107,163	0.8	0.9	0.2	0.4	0.8	0.6	1.0	0.2	0.2	0.1	0.4	0.2	0.3
1524	UNK 1524 <i>m/z</i> : 105,161,204,119,93	0.7	1.0	1.3	0.2	0.2	0.8	0.7	0.4	0.2	0.4	0.5	0.9	0.5
1699	UNK 1699 <i>m/z</i> : 81,163,191,95,123	3.6	4.1	3.0	0.5	1.8	3.2	5.2	1.3	1.1	1.6	2.0	0.8	3.2
	192 more unknowns <1%	2.9	3.9	1.6	4.7	4.9	3.8	4.9	2.7	2.2	4.8	4.6	3.4	3.7
37 1 .11	1 1 1' 4 1 1			T7 //	•	· 1 /T	DI) TII	1	1 6 1					

Volatiles are ordered according to compound class, and within class by Kováts' retention index (KRI). The total number of volatiles is also given.

Abbreviations: * = identification of compound was verified by authentic standards; tr = trace relative amount (<0.05%); m/z = mass-to-charge ratio in decreasing order of abundance. North: JOS, Josefiau; BUR, Burg Hohenstein; HOH, Hohendilching; MUR, Murnau; NEC, Horb am Neckar; RÜM, Rümikon; South: DAO, Daone; LIM, Limone-Piemonte; MAH, Santa Maria Hoè; MON, Montese; UDI, Udine

[§] Synthetic (+)- α - and β -Citronellene coeluted with natural detected α - and β -Citronellene on a chiral column (MEGA-DEX DMT Beta SE, 30m × 0.25mm ID, 0.23μm film) (Gfrerer *et al.* unpublished)