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Elie Ayoub, Perrine Cologon, Marie Marsiglia, Jean-Baptiste Salmon,
Christine Dalmazzone

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1 Development of online separation and surfactant
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4 AUTHOR NAMES

5 Elie Ayoub^a, Perrine Cologon^{*a}, Marie Marsiglia^a, Jean-Baptiste Salmon^b, Christine
6 Dalmazzone^a

7

8 AUTHOR ADDRESS

9 ^a IFP Energies nouvelles, 1 et 4 avenue de Bois-Préau, 92852 Rueil-Malmaison, France

10 ^b Laboratoire du futur – UMR5258, 178 avenue du Dr. Schweitzer, 33600 Pessac, France

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12

13

14 1. INTRODUCTION

15 Both the growth of global oil consumption and the decline of new conventional oil fields
16 discoveries are observed (Bera and Mandal, 2015; Hirasaki et al., 2011; Thomas, 2008; Sheng,
17 2014). To match the current oil demand, reservoir recovery rate must be increased. Conventional
18 oil production methods, namely first and secondary recovery, lead together to the recovery of
19 around 20 to 40% of the original oil in place (Bera and Mandal, 2015; Hirasaki et al., 2011;
20 Thomas, 2008; Sheng, 2011; Sheng, 2014). Then, the flow rate of produced oil tends towards zero,
21 partly due to the action of capillary forces that traps the residual oil in the pores of the reservoir
22 (Reed and Healy, 1977). The dimensionless capillary number Ca expresses the ratio of viscous
23 drag forces to capillary forces. It has been experimentally demonstrated that the residual oil
24 saturation in the reservoir decreases as this capillary number increases (Hakiki et al., 2017; Lake,
25 1989; Oughanem et al., 2015). Consequently, the action of capillary forces can be counterbalanced
26 in several ways into the pores of the reservoir. It is possible to increase the injection velocity or
27 the viscosity of the moving water phase by using solutions of polymers. However, these solutions
28 are quickly limited by practical aspects related to the injection power of the pumps and the risks
29 of rock fracking. Thus, the best way to significantly increase the capillary number is the reduction
30 of the interfacial tension between oil and water (IFT) by several decades. This drastic reduction is
31 obtained by using specific chemical surfactants within a tertiary recovery technique called
32 surfactant chemical EOR (cEOR) or “surfactant flooding” (Sheng, 2011). This process consists in
33 injecting a surfactant formulation which lowers the IFT between water and oil and thus facilitates
34 the oil mobilization towards the producing wells (Bourel and Schechter, 1988; Jing et al. 2021;
35 Hirasaki et al. 2011; Sheng, 2010). In the case of sandstone reservoirs, anionic surfactants are
36 essentially employed, due to their low tendency to adsorb on negatively charged surfaces (Chen

37 and Schechter, 2021). The main chemical structures are sulfates, like alkyl sulfates, alcohol ethoxy
38 or propoxy sulfates and sulfonates, like alkyl aryl sulfonates, linear or branched alkyl benzene
39 sulfonates, alpha-olefin or internal olefin sulfonates (Negin et al., 2017).

40 The formulation of the surfactants, which is the key parameter of surfactant flooding, is
41 specifically designed to achieve ultra-low IFT (lower than 10^{-2} mN/m) and create a Winsor III
42 microemulsion (WIII) with the oil at reservoir temperature (Haegel et al., 2008; Hakiki et al., 2015;
43 Salager et al., 2013; Thomas et al., 1999). Winsor III system can be achieved by balancing the
44 respective affinities of the hydrophobic and hydrophilic parts of the surfactants with regard to the
45 aqueous and oily phases. The formulation design is therefore performed through salinity–scan
46 tests, which are generally used to investigate the phase diagrams of anionic surfactant formulations
47 (Hirasaki et al., 2011). From a practical point of view, salinity modifies the distribution of the
48 anionic surfactant between the oil, brine and microemulsion phases (Fukumoto et al., 2016;
49 Fukumoto et al., 2018; Salager et al., 2013). Below the optimal salinity, the surfactant is preferably
50 soluble in brine, giving a Winsor I system (WI), consisting of an oil-in-water microemulsion in
51 equilibrium with an excess of oil phase. At a salinity higher than the optimum, it preferably
52 becomes soluble in the oily phase, giving a Winsor II system (WII), consisting of a water-in-oil
53 microemulsion in equilibrium with an excess of aqueous phase. At the optimal salinity S^* , the
54 surfactant is mainly concentrated in the intermediate microemulsion phase, giving a Winsor III
55 system consisting of a bicontinuous microemulsion in equilibrium with the excess aqueous and
56 oily phases. It is noteworthy that the IFT is the lowest at this optimal salinity.

57 Most of the time, the optimal formulation can be first developed through robotic salinity-scan tests
58 (AlSofi et al., 2021; Morvan et al., 2008). This is essentially done on model oil phases (alkanes)
59 representative of the Equivalent Alkane Carbon Number (EACN) of the crude oil under

60 investigation (Queste et al., 2007). In a final step, the formulation is optimized with the real crude
61 oil (Oukhemanou et al., 2014).

62

63 The efficiency of the designed formulation is then evaluated using a test called "coreflood
64 experiment". This refers to a specific setup which operates on a rock sample under pressure and
65 temperature conditions very close to those of the reservoir (Flaaten et al., 2009; Hirasaki et al.,
66 2011; Levitt et al., 2009; Mohammadi et al., 2009). The measurement of the quantity of oil
67 recovered and of surfactants retained on the rock makes it possible to determine the performance
68 of the surfactant flooding process, hence the efficiency of the formulation, the environmental
69 footprint and the profitability of the process. Evaluating the economics of the process and
70 environmental footprint are of paramount importance. For these two parameters, it is worthwhile
71 to recover the highest volume of oil by injecting the lowest amount of surfactants, meaning that
72 most of surfactants must be adsorbed on the water-oil interface and only a negligible part on the
73 rock. Therefore, the volume of oil produced by surfactant flooding and the quantity of surfactant
74 adsorbed on the rock must be determined for each coreflood experiment. The effluents resulting
75 from a coreflood test are complex mixtures of oil, brine and surfactants. They are generally
76 collected in graduated cylinders or tubes as the experiment progresses. The analyses are then
77 performed a posteriori, offline. The quantity of oil recovered is calculated from volumetric
78 measurements (estimation of the water-to-oil volume ratio in each tube) or by a NMR analysis of
79 the rock sample at the end of the test (Fleury and Deflandre, 2003). The presence of quite stable
80 emulsions in the tubes induces a lower accuracy of the volumetric measurement. The concentration
81 of surfactants is measured in the aqueous phase using various techniques such as potentiometric
82 Hyamine titration (Liu et al., 2010), high performance liquid chromatography (HPLC), or even

83 UV-visible spectroscopy, though offline (Liang et al., 2021). It is noteworthy that carrying out
84 these analyses is a tremendous challenge, especially for the determination of the surfactant
85 concentration. A main issue concerns the presence of emulsions in the effluents that can sometimes
86 be very stable (Dalmazzone et al., 2012). The type of these emulsions is generally oil-in-water and
87 the presence of micronic oil droplets may interfere with the Hyamine dosage or foul the liquid
88 chromatography column. Another trouble comes from the distribution of surfactants between the
89 oily and aqueous phases. Depending on the experimental conditions, it is possible that the
90 surfactants are partly trapped in the oil phase. These surfactants are not quantified in the mass
91 balance, which may lead to an overestimation of the quantity of surfactants adsorbed or retained
92 on the rock and consequently to the rejection of a formulation for economic and environmental
93 reasons.

94 To overcome these difficulties, a micro/millifluidic device capable of treating and separating
95 coreflood effluents online has been designed. This setup allows a relevant quantification of the
96 surfactant concentration, online or offline. The overall approach consists in (1) performing an
97 online dilution of the effluent with deionized water to decrease the salinity and thus promote
98 surfactant transfer to the aqueous phase, (2) separating online the oil and the aqueous phase using
99 a membrane, and (3) measuring the surfactant concentration.

100

101 In this paper, the designed micro/millifluidic setup is detailed. The validation of the methodology
102 was first performed on a model system of surfactants and oil. Then, the whole micro/millifluidic
103 assembly was tested at the outlet of a real coreflood experiment to evaluate the potential of the
104 developed methodology in realistic and representative conditions.

105

106 2. MATERIALS AND METHODS

107 **2.1 Chemicals**

108 Sodium dodecylbenzenesulfonate (SDBS) (technical grade) was purchased from Sigma-Aldrich.
109 Isobutanol (C₄H₁₀O) with >99% purity and n-decane (C₁₀H₂₂) with 99%_{wt} purity were provided
110 by Alfa Aesar. Sodium chloride (NaCl) with 99.9% purity was purchased from VWR chemicals.
111 Organol Red was obtained from Prolabo. All compounds were used without further purification.
112 Aqueous solutions were prepared with Milli-Q water (resistivity higher than 18.2 MΩ/cm). The
113 industrial surfactants, Alkylglyceryl Ether Sulfonate (AGES – 33wt.% active matter) and Internal
114 Olefin Sulfonate (IOS – 33wt.% active matter), were provided by Solvay. Arabian light crude oil,
115 filtered over 5 μm, was chosen in this study because of its low paraffin content, avoiding clogging
116 of the tubings at room temperature. Details of the Arabian light composition and characteristics
117 are given in Table 1.

118

119 **Table 1.** Density and viscosity at 20°C and SARA composition of the Arabian light crude oil

Density at 20°C (g/cm³)	0.88
Viscosity at 20°C (mPa.s)	12
Saturates (wt%)	53.8
Aromatics (wt%)	32.6
Resins (wt%)	10.6
Asphaltenes (wt%)	2.7

120

121 **2.2 Phase diagrams**

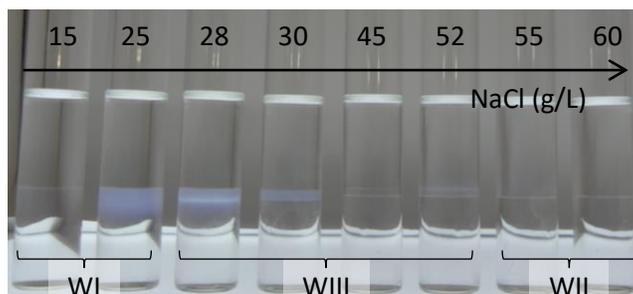
122 Phase diagrams are of interest in order both to optimize EOR formulations through the
123 identification of the optimal salinity and to treat the effluent with the appropriate methodology.
124 While WIII microemulsions are the aim of EOR formulation design, WI microemulsions are
125 preferred to treat the effluents as the surfactant is mainly contained in the aqueous phase for such
126 systems.

127 Model fluid mixtures were prepared with decane and an aqueous phase composed of Milli-Q water,
128 SDBS, NaCl brine and isobutanol. SDBS is a sulfonate anionic surfactant. It was selected because
129 it is a representative surrogate for EOR surfactants due to its sulfonate polar moiety and its carbon
130 chain length, and also because its aromatic ring makes UV-visible absorption measurement
131 possible. Fluids representative of real systems were prepared with filtered Arabian light crude oil
132 and an aqueous phase composed of Milli-Q water, an industrial IOS-AGES blend and NaCl brine.
133 For both systems, salinity scans were done to determine the different Winsor zones.
134 Microemulsions were prepared at room temperature in glass tubes using the following procedure.

- 135 • First, solutions of SDBS (16 g/L) and NaCl (200 g/L) were prepared in separate flasks. The
136 mass of SDBS and NaCl was measured with an electronic weighing scale (AE200, Mettler
137 Toledo) with a precision of ± 0.1 mg.
- 138 • Then, 3.5 mL of the SDBS solution was added in a glass tube. Also, 0.40 mL of isobutanol
139 (5.83 %vol. of aqueous phase) was added to ensure the dissolution of the surfactants.
- 140 • Next, the volume of the aqueous phase was adjusted to 7 mL with Milli-Q water and NaCl
141 solutions whose respective volumes enable to achieve the set salinity (ranging from 15 to
142 80 g/L). At that point, the solution was carefully mixed.

- 143
- Finally, 7 mL of decane was poured along the glass tube walls, resulting in a Water to Oil Ratio (WOR) of one. The interface was gently stretched by turning the tubes upside down approximately 6 times without creating an emulsion.
- 144
- 145
- Then, the samples were left for one month at room temperature to ensure complete thermodynamic equilibrium.
- 146
- 147

148 Middle phases corresponding to Winsor III systems were observed for salinities ranging from
149 28 to 52 g/L (Figure 1). The phase transitions can be described as follows: Winsor I type
150 microemulsion (oil-in-water) from 0 to 25 g/L NaCl, Winsor III (bicontinuous) from 28 to 52
151 g/L NaCl, and Winsor II (water-in-oil) from 55 to 80 g/L NaCl, in agreement with previous
152 studies (Fukumoto et al., 2016).



154 **Figure 1.** Photograph of formulated microemulsions with salt increments. Formulations are
155 composed of water, NaCl, SDBS, isobutanol and decane. WOR is unity. WI, WII and WIII are the
156 abbreviations of Winsor I, II and III respectively.

157

158 The same protocol was followed for the microemulsions formulated with crude oil and industrial
159 surfactants. Decane was replaced by Arabian Light crude oil and a 4 g/L solution of IOS-AGES
160 mixture was used instead of SDBS. Note that isobutanol was not used in this case and the WOR
161 was set to 0.7. The samples were kept at 40 °C during four months in order to reach equilibrium.
162 A middle phase was observed for salinities ranging from 70 to 75 g/L. Phase transitions in terms

163 of microemulsions can be described as follows: Winsor I (oil-in-water) from 0 to 65 g/L NaCl,
164 Winsor III (bicontinuous) from 70 to 75 g/L NaCl, and Winsor II (water-in-oil) from 80 to 130 g/L
165 NaCl.

166

167 **2.3 Water-in-oil titration**

168 Mass fraction of water in the oil phase or the microemulsion phase was measured by Karl Fischer
169 titration. The titration was conducted with 787-KF Titrino from Metrohm with a reagent of
170 Hydranal-Composite 5 purchased from Fluka. A weighed sample (about 10 mg) was injected into
171 a measuring cell, and the amount of water in the sample was obtained from the volume of reagent
172 needed to reach a significant shift in potential. All the samples were analyzed at least three times.

173

174 **2.4 Surfactant quantification**

175 Different techniques were used to measure the concentration of surfactant in the aqueous phase.
176 Conventional offline analyses included Hyamine titration and High-performance liquid
177 chromatography (HPLC).

178 For Hyamine titration, 1 mL of the sample was introduced in the measuring cell of the titrator
179 (862-compact Titrosampler, Metrohm). Incremental volumes (30 μ L) of titrant (Hyamine 1622-
180 solution, Merck Millipore) were added to the sample until a shift in potential was reached.

181 HPLC analysis was also employed to confirm the results of Hyamine titration. Briefly, samples
182 were injected into an Agilent 1260 Infinity system coupled to an Evaporative Light Scattering
183 Detector (ELSD G4260B). A mixture of ammonium acetate buffer (pH = 5) and acetonitrile was
184 used as mobile phase at a flow rate of 1 mL/min. Once separated, the surfactants were detected in

185 the ELSD (nebulization temperature: 40°C, evaporator temperature: 70°C). Both analytical
186 techniques were calibrated with standard solutions.

187 Additionally, online measurements were specifically developed: UV-visible absorption spectra
188 were acquired on an AvaSpec-ULS2048 spectrophotometer (Avantes) equipped with a deuterium-
189 halogen light source offering a wavelength range of 200 nm to 1100 nm. A micro-flow cell (18 μ L
190 volume) with a 10 mm Z-design optical path was used for online measurements (Avantes).

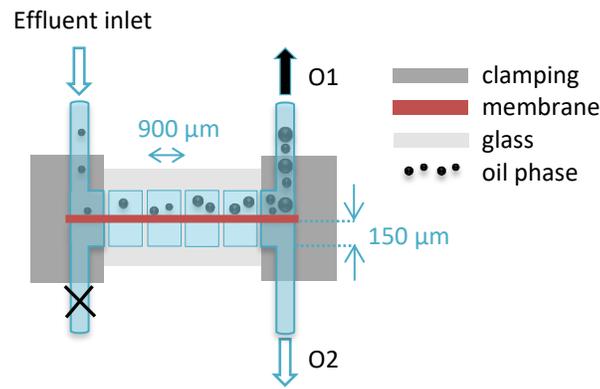
191

192 **2.5 Micro/Millifluidic experimental setup**

193 Low pressure syringe pumps neMESYS 290N (Cetoni) were used to pump the fluids. Teflon
194 tubings from Cluzeau Info Labo (1/16" external diameter and 762 μ m internal diameter) were
195 used to connect all parts of the setup. Oil-in-water emulsions were created with two coaxial
196 capillaries. The chip was manufactured by inserting a circular capillary with a 200 μ m internal
197 diameter in a square capillary of 400 μ m internal width. This enables to create oil-in-water
198 emulsions at the tip of the inner capillary in the continuous aqueous phase (Utada et al., 2005).
199 Decane was colored with organol red for visualization purposes.

200 The membrane separation device is sold by Dolomite under the name "Membrane Chip Interface"
201 and is specifically designed for online liquid-liquid separation. It consists of two microfluidic glass
202 chips engraved on one side, a hydrophilic treated PTFE membrane (Part No. 3200347, Dolomite)
203 and a metal structure which seals the whole system. Each glass chip consists of a serpentine
204 channel with a length of 21.2 cm, width of 900 μ m and height of 150 μ m. The hydrophilic
205 membrane (25 mm in diameter and 0.2 μ m nominal pore size) is sandwiched between the two
206 chips. It contains four inlets, two on each side of the membrane, to ensure the insertion of the tubes
207 allowing the connections. In our case, one of the bottom inlets was closed in order to have one

208 inlet on the upper side and two outlets on both sides of the membrane. Outlets are referred to as
209 O1 and O2 for the upper and lower sides respectively. The whole system is schematically presented
210 in Figure 2.

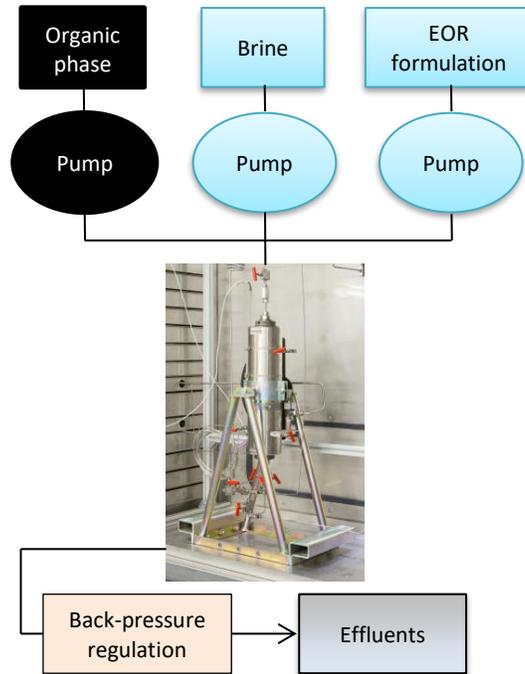


211
212 **Figure 2.** Schematic sectional view of the membrane separation device. The membrane stands in
213 red and both outlets for the separated effluent are identified (O1 and O2). The black circles
214 correspond to droplets of oil transported in the flowing aqueous phase (light blue).

215
216 Visualization of the fluids circulating in the system is partly possible thanks to a small opening in
217 the metal structure.

218 219 **2.6 Coreflood experiment**

220 A coreflood experiment setup, designed at IFPEN and schematically presented in Figure 3, was
221 used with a "Bentheimer" type core (38.9 mm long and 34.98 mm in diameter). This sandstone
222 outcrop sample is often used to mimic reservoir behavior for EOR studies. The core is placed in a
223 sleeve in a coreholder Hassler cell. An overburden pressure is then applied on the sleeve to ensure
224 that no leak occurs between the core and the sleeve. All the experiments were carried out at a pore
225 pressure of 10 bar and a confining pressure of 40 bar. Temperature was kept at 40°C throughout
226 the experiment.



227

228 **Figure 3.** Schematic view of a coreflood experiment.

229

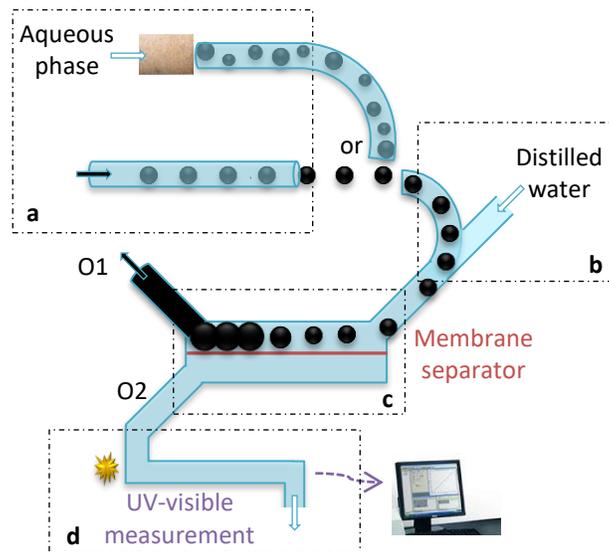
230 General EOR workflow has been described in details elsewhere (Moreau et al., 2010). Briefly, the
 231 following steps were applied: first of all, a forced imbibition step was performed with brine to
 232 determine the pore volume of the core (28.1 mL). In a second step, the rock was saturated with a
 233 filtered Arabian Light crude oil. Then, water injection was carried out to mimic a "waterflooding"
 234 step in order to determine the residual oil saturation (56%, i.e. 15.7 mL of oil). Finally, the
 235 formulation was injected at a flow rate of 5 mL/h for 52 hours, followed by a brine injection at 70
 236 g/L NaCl for 50 hours.

237

238 3. RESULTS AND DISCUSSION

239 The purpose of this study is to develop a method leading to accurate measurement of surfactant
240 concentrations in EOR effluents. Therefore, an efficient oil/water separation is needed. A dilution
241 with water is also performed as a first step to decrease the salinity and switch the aqueous phase
242 of the effluent to a Winsor I microemulsion because surfactants are mostly in the aqueous phase
243 in this case.

244 Figure 4 displays a schematic view of the experimental setup developed to achieve these steps
245 online. The whole assembly includes an effluent collection or generation with coaxial capillaries
246 for model experiments to optimize the following steps (Figure 4.a), a dilution step (Figure 4.b), a
247 membrane separation device (Figure 4.c) and an online UV-visible absorption measurement
248 (Figure 4.d). Each step has been individually optimized, before assembling and characterizing the
249 whole setup.



250

251 **Figure 4.** Schematic view of the experimental setup developed in this study. It includes: a. effluent
252 collection or generation with coaxial capillaries, b. dilution with distilled water, c. membrane
253 separation and d. UV-visible absorption measurement. The black circles show schematically oil
254 droplets flowing in the aqueous phase (light blue).
255

256 3.1 Oil-in-water emulsion creation

257 Surrogate oil-in-water emulsions were obtained with coaxial capillaries (Figure 4.a). Water flow
258 rate ranged from 1 to 10 mL/h which is representative of flooding conditions. Droplet size could
259 easily be tuned by modifying the oil flow rate relatively to the one of water. For this study, the
260 flow rate of the aqueous continuous phase was always twice the one of the organic phase. This
261 resulted in oil droplets having a mean diameter of about 400 μm .

262

263 **3.2 Dilution step**

264 The effluent, either created by coaxial capillaries or collected at the outlet of the coreflow, was
265 then diluted with Milli-Q water (Figure 4.b) using a syringe pump and a T-junction. The dilution
266 water flow rate was equivalent to the initial flow rate of the water phase in the emulsion. This step
267 enabled to reduce the salinity by a factor of two and thus highly increased the IFT between the
268 aqueous phase and oil, promoting phase separation. In a first attempt, a microfluidic herringbone
269 design was used to maximize the mixing efficiency as described in the work of Stroock et al.
270 (2002). However, in this configuration, oil droplets were sheared into smaller droplets which
271 limited the oil/water separation likely due to the fouling of the membrane. A simple T-junction
272 appeared to be sufficient to dilute the effluent.

273

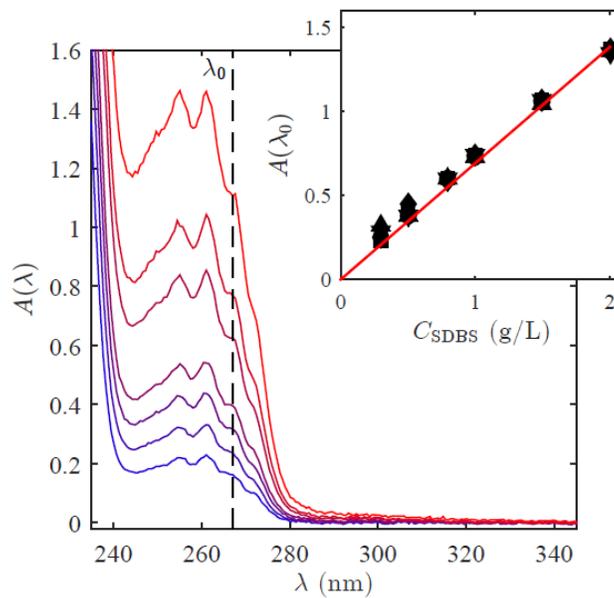
274 **3.3 Membrane separation device**

275 The oil/water separation step is represented schematically in Figure 4.c and detailed previously in
276 Figure 2. Due to the hydrophilic nature of the membrane, the fluid in outlet O2 is expected to be
277 mostly aqueous, whereas the fluid in outlet O1 is expected to be composed of a mixture of organic
278 and aqueous phases.

279

280 **3.4 Surfactant quantification**

281 Online UV-visible absorption spectroscopy was developed in the framework of this study as a
282 quantification technique of the surfactants in aqueous samples (Figure 4.d). UV-visible spectra
283 obtained for different concentrations of SDBS are shown in Figure 5. The absorbance at 267 nm
284 (λ_0) exhibited a linear relationship with SDBS concentrations (see the insert of Figure 5) according
285 to the Beer-Lambert law, whatever the salinity (0 to 10 g/L NaCl). The choice of this wavelength
286 to monitor SDBS is explained in the next sections.



287

288 **Figure 5.** Absorbance ($A(\lambda)$) for different concentrations of SDBS in Milli-Q water at room
289 temperature; Insert : Absorbance $A(\lambda_0)$ at the wavelength $\lambda_0 = 267$ nm chosen to monitor SDBS
290 concentration.

291

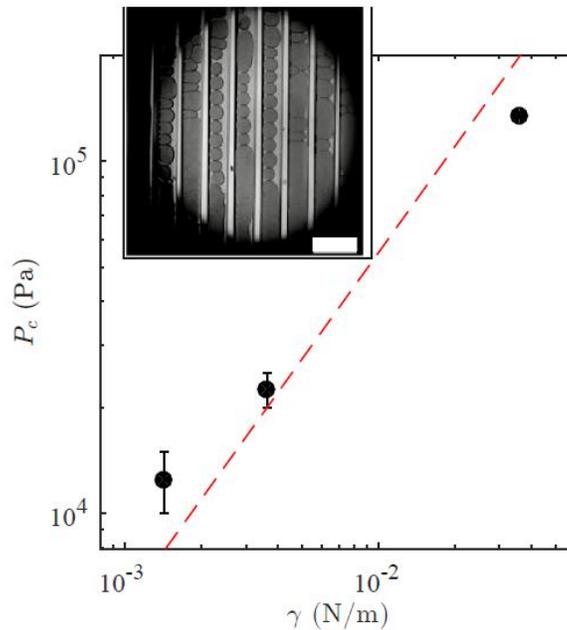
292 **3.5 Characterization of the membrane separation device**

293 First, the impact of the interfacial tension between the aqueous phase and decane on the membrane
294 separation efficiency has been studied. Therefore, different Winsor I emulsions, corresponding to

295 different interfacial tensions, were formed and injected directly in the separation device using the
296 coaxial capillary device. Then, the injection was stopped to immobilize the fluids in the device,
297 and O1 was closed. In this configuration the fluid can only flow through the membrane and the
298 objective of this experiment was to determine the pressure at which the decane breakthrough
299 occurs. Thus, different pressures were applied at the inlet of the device (pressure controller MFCS-
300 EZ, Fluigent) while observing the membrane area under a stereomicroscope using the opening of
301 the Dolomite membrane device (see insert in Figure 6 for a typical image using bright-field
302 imaging).

303 At low imposed pressures, decane droplets are getting closer and some of them coalesce. At a well-
304 defined critical transmembrane pressure drop P_c , one clearly sees decane droplets starting to flow
305 through the membrane.

306 The interfacial tensions were measured with a Krüss SDT spinning drop tensiometer. These
307 experiments showed that for an emulsion with an interfacial tension of 35.7 ± 0.2 mN/m, the
308 breakthrough pressure is about ~ 135 kPa. For an emulsion with an IFT of 3.6 ± 0.4 mN/m, the
309 breakthrough pressure decreases down to about ~ 22.5 kPa. And this pressure decreases further
310 down to about ~ 12.5 kPa for an IFT of 1.4 ± 0.1 mN/m. These data are plotted in Figure 6.



311

312 **Figure 6.** Breakthrough pressure (P_c) of the decane through the hydrophilic membrane versus
 313 interfacial tension (γ). The red dashed line is the linear law $P_c = 2\gamma/r$ with $r = 0.4\mu\text{m}$. Insert:
 314 bright-field image of the membrane separation device taken with a stereomicroscope before
 315 applying pressure at the inlet.

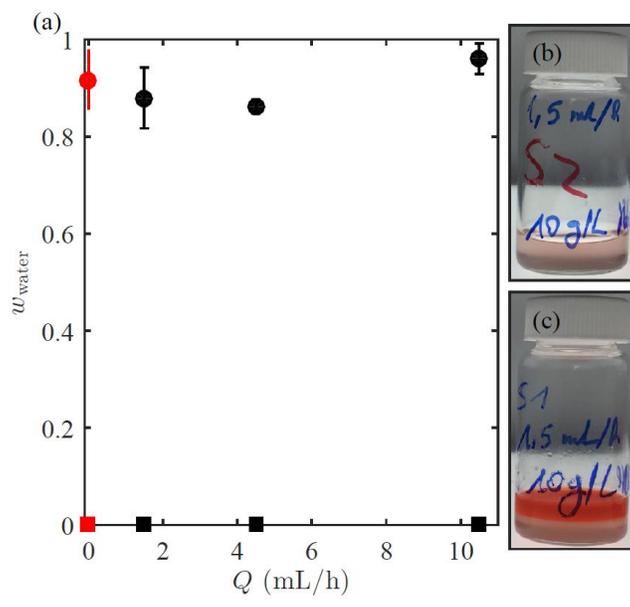
316

317 Capillary pressure is related to the interfacial tension of the fluids according to the application of
 318 Young-Laplace equation in a narrow tube of circular cross-section with a zero contact angle giving
 319 the capillary pressure $2\gamma/r$, with γ the interfacial tension (N/m) and r the pore radius of the
 320 membrane (m). So, for a constant pore size, as a first approximation, the breakthrough pressure P_c
 321 is proportional to the IFT. Figure 6 shows the experimental data, as well as the best linear fit of
 322 the breakthrough pressure according to $P_c = 2\gamma/r$. While this relationship does not perfectly fit
 323 our data, this crude estimate yields a pore size of $0.4\mu\text{m}$ in agreement with the value given by the
 324 manufacturer ($0.2\mu\text{m}$) considering the complex geometry of the pores. Without being predictive,
 325 these data allow us to determine the range of pressure for which we can be sure that the oil droplets
 326 do not pass through the membrane for a given IFT.

327

328 The second characterization of the membrane separation device was to check its ability to separate
329 an emulsion with flow rates ranging between 1 and 10.5 mL/h, in the same order of magnitude as
330 the flow rates usually used for coreflood experiments. For this purpose, a Winsor I emulsion was
331 created with the model system (with decane being colored with organol red for the ease of
332 visualization) and injected in the membrane separation device at different flow rates. The fluids
333 recovered at the two outlets were collected in flasks. Two phases were recovered at Outlet 1: a red
334 phase and a pink one (Figure 7.b); but only one phase in Outlet 2, a pink one (Figure 7.c). The red
335 phase of O1 is decane and the pink phase of O1 and O2 is mainly water. Its pink coloration is due
336 to the presence of SDBS at a concentration higher than the critical micellar concentration, which
337 allows the dispersion of organol red, insoluble in water without SDBS. Similar behaviors were
338 observed for three different flow rates (1.5, 4.5, and 10.5 mL/h) at various salinities (0, 5, 10, and
339 15 g/L). Note that all the tested salinities lead to Winsor I microemulsions at thermodynamic
340 equilibrium.

341 Figure 7.a shows the water percentage measured by Karl-Fisher titration, versus the emulsion flow
342 rate at the entrance of the separation device. Water content was measured in the upper phase (red
343 oily phase) recovered in O1 and in the aqueous phase recovered in O2. Initial fluids were also
344 analyzed, giving the results in red ($Q = 0$ mL/h). These values ($92\% \pm 6\%$ for aqueous phase –
345 below the quantification limit for the organic phase) were in agreement with the expected
346 concentrations.



347

348 **Figure 7.** a. Water content (w_{water}) measured by Karl Fischer titration in the supernatant oily
 349 phase at Outlet 1 (black square) and in the aqueous phase at Outlet 2 (black circle) after separation
 350 with the membrane separation device. The water content of the initial phases is specified in red
 351 (red square for decane and red circle for the aqueous solution of SDBS). Pictures of b. Outlet 2
 352 flask and c. Outlet 1 flask.

353

354 In O2 (black circles) water contents comprised between 88 and 96 %_{vol} were obtained, depending
 355 on the injection flow rate. Concerning the upper phase of O1, Karl Fisher titration gives the same
 356 result as the initial decane, which allows us to claim that the red phase is pure decane. Therefore,
 357 the membrane separation device leads to a >90% pure aqueous phase, which is considered
 358 satisfactory regarding the final application.

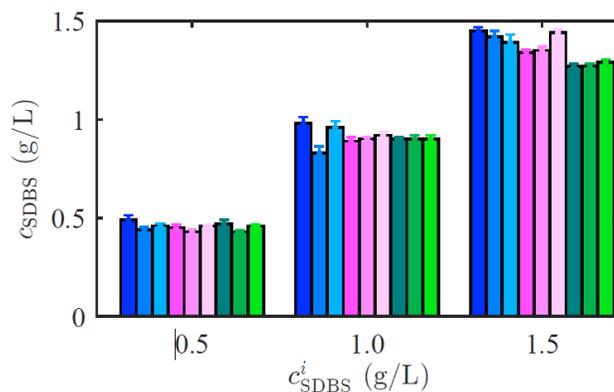
359

360 3.6 Characterization of the full experimental setup

361 This section aims at evaluating the potential of the proposed methodology for separating the
 362 organic and aqueous phases and measuring the surfactant concentration in the aqueous phase. So
 363 the complete setup was assessed, including dilution, oil/water separation and surfactant

364 quantification. Results obtained with either model fluids or real systems are presented in this
365 section.

366 First, experiments were performed with SDBS concentrations of 0.5, 1 and 1.5 g/L, and salinities
367 equal to 5, 10, and 15 g/L; at these salinities, Winsor I microemulsions are obtained at equilibrium.
368 Figure 8 shows the concentrations of SDBS measured after oil/water separation at three different
369 flow rates (1.5, 4.5, and 10.5 mL/h) for 9 formulations containing initially various amounts of
370 NaCl and SDBS. SDBS concentrations presented in Figure 8 are the average of 8 measurements
371 performed online with the UV-visible spectrometer at 267 nm during an hour. The measured
372 concentrations are around 10 % lower than the expected concentrations. This difference could be
373 due to the presence of small oil droplets that can pass through the membrane because the tested
374 salinities lead to a Winsor I microemulsion (cf. next paragraph), leading to a different
375 spectroscopic response. To confirm the SDBS concentration, the separated aqueous phase was also
376 measured by Hyamine titration. The results show that the initial concentrations of 0.5 ± 0.05 ,
377 1 ± 0.05 , and 1.5 ± 0.04 g/L were obtained.



378

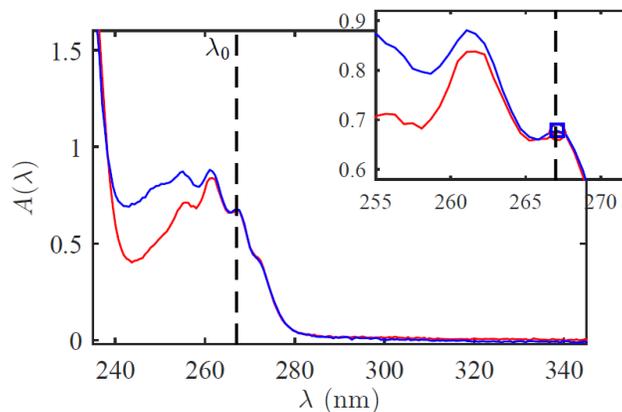
379 **Figure 8.** SDBS concentrations (c_{SDBS}) in the phase obtained in Outlet 2 measured with UV-
380 visible spectroscopy, after separation with the membrane separation device for different flow rates
381 (1.5, 4.5, and 10.5 mL/h from dark to light color), different salinities (5 g/L in blue, 10 g/L in pink,
382 15 g/L in green) and different initial SDBS concentrations (c_{SDBS}^i).

383

384 Both UV-visible measurements and Hyamine titration results show that in the case of the model
385 systems, a reliable online surfactant measurement at $\pm 10\%$ can be obtained.

386 Furthermore, experiments were carried out to assess potential interactions between the aqueous
387 phase containing surfactants and decane possibly resulting in changes in the UV-visible spectra.

388 Comparisons were made between a raw 1 g/L SDBS solution in 5 g/L NaCl (blue spectrum in
389 Figure 9) and the same solution contacted with decane for seven days at room temperature (red
390 spectrum in Figure 9).



391

392 **Figure 9.** Absorbance $A(\lambda)$ of a 1 g/L SDBS solution in 5 g/L NaCl before (blue) and after (red)
393 contacting decane at room temperature.

394

395 A significant decrease in absorbance was observed in the 240-260 nm range. This might be
396 explained by the formation of micelles in the aqueous phase, which bias the measurement. These

397 objects have been quantified by Dynamic Light Scattering (DLS) showing that their size is around
398 20 nm in diameter. This observation led us to select a wavelength providing a constant signal for

399 the SDBS quantification (267 nm, Figure 9).

400 Secondly, the performance of the whole experimental setup was assessed with real systems:
 401 filtered Arabian light crude oil and an aqueous phase composed of an industrial blend of surfactants
 402 at concentrations of 1, 2, and 4 g/L at a salinity of 35 g/L. At this salinity, a Winsor I microemulsion
 403 is obtained. Table 2 shows the concentrations measured with Hyamine titration at three different
 404 flow rates (1.5, 4.5, and 10.5 mL/h). The obtained concentrations are in good agreement with the
 405 expected ones.

406 **Table 2.** Industrial surfactant concentrations in the phase obtained in Outlet 2 measured with
 407 Hyamine titration, after separation with the membrane separation device for different flow rates
 408 (1.5, 4.5 and 10.5 mL/h) at a salinity of 35 g/L.

409

Expected industrial surfactant concentration (g/L)	Measured industrial surfactant concentration		
	1.5 mL/h (g/L)	4.5 mL/h (g/L)	10.5 mL/h (g/L)
1	1.1±0.1	1.1±0.1	1.1±0.1
2	2.1±0.2	2.1±0.2	2.1±0.2
4	3.7±0.4	3.7±0.4	3.9±0.4

410

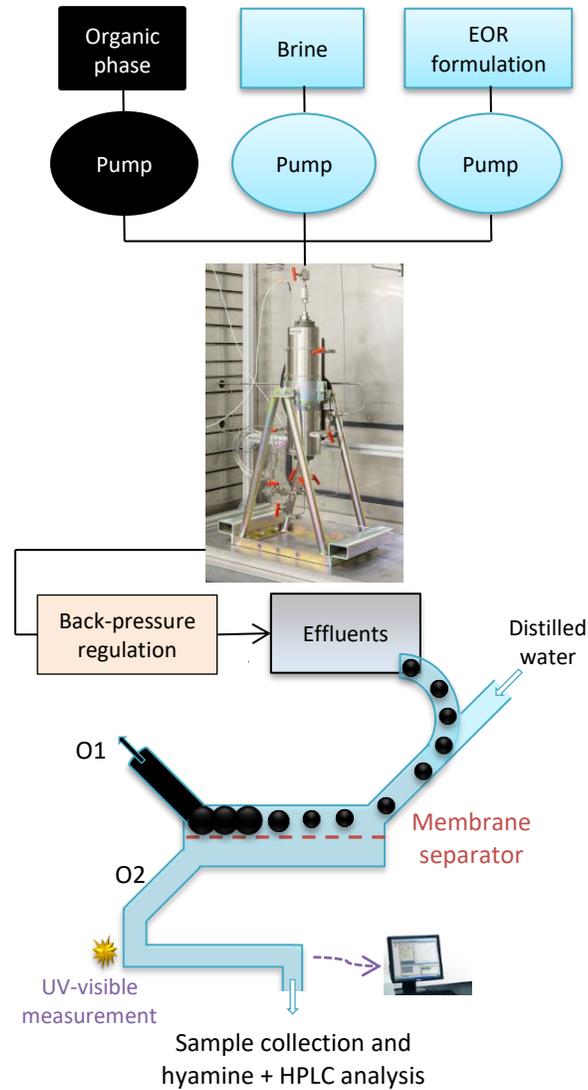
411 These results confirm that the experimental setup is efficient to separate the effluents (oil-in-water
 412 emulsions), enabling accurate quantification of surfactants in the aqueous phase with offline
 413 measurements, here Hyamine titration.

414 Note that no online UV-visible measurement could be exploited during these experiments because
 415 the contact between crude oil and water phase containing surfactants resulted in the dispersion of
 416 compounds from the oil. Spectra were so altered in this case that no reliable quantification could
 417 be considered.

418

419 **3.7 Coreflood Application**

420 The implementation of our experimental setup on a coreflood experiment is presented in Figure
421 10.



422
423 **Figure 10.** Schematic view of a coreflood experiment and downstream implementation of our
424 experimental setup.

425
426 The formulation for surfactant flooding was optimized through a salinity scan with NaCl from a
427 mixture of surfactants composed of 4 g/L of AGES and 4 g/L of IOS, both provided by Solvay.
428 The optimal NaCl salinity leading to WIII microemulsions is about 70 g/L at 40°C.

429 While the formulation was injected, the effluents of the coreflood experiment were diverted into
430 the microfluidic setup, located after the back-pressure regulation (Figure 10). After dilution with
431 water and separation, the aqueous phase was pumped into to the UV-visible spectroscopy flow cell
432 and an autosampler which collected the fluids at a frequency of 1 tube per hour for offline
433 surfactant measurements.

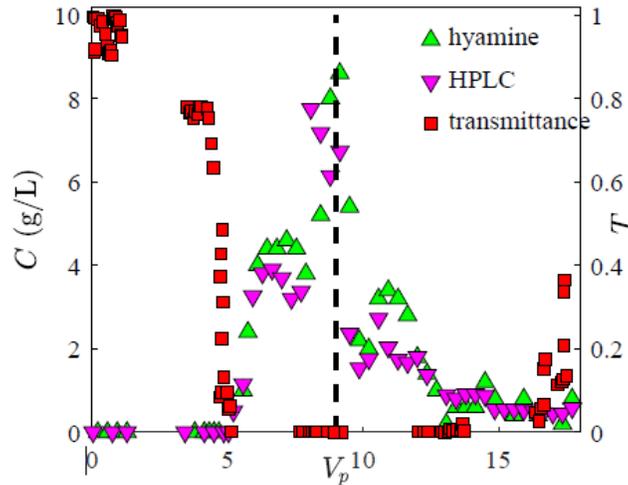
434 Only one phase was recovered at the aqueous outlet O2 of the membrane separation device (Figure
435 11.b), while a mixture of oil and water was collected in the oily outlet O1. No free oil phase was
436 seen, nor any phase separation, in the tubes of O2, even one month after the end of the experiment.
437 Evolution of the color of the effluents collected at Outlet 2 was noticed throughout the experiment.
438 The samples changed from transparent in the beginning to a pale yellow color that gets darker, and
439 at the end samples gradually became transparent again. This color change could be related to the
440 formation of an oil-in-water microemulsion with very tiny droplets (a few tens of nm droplets as
441 shown before) that can flow through the pores of the separation membrane.



442
443 **Figure 11.** Aspect of the coreflood effluents once collected in tubes: a. without using our
444 experimental setup, b. after using our experimental setup in the aqueous outlet (Outlet 2). A clear
445 improvement of water quality is noticed thanks to the present setup.

446
447 This water quality is compatible with an offline determination of the surfactants by Hyamine
448 titration and by HPLC under completely satisfactory conditions without further purification steps.

449 Figure 12 shows the evolution of the surfactant concentrations measured offline in the separated
450 water phase, as a function of injected porous volume (V_p). Results of both analyses (Hyamine –
451 green triangles, and HPLC – purple triangles) are quite consistent.



452

453 **Figure 12.** Analysis of the separated aqueous effluents from the coreflood: Evolution of the
454 concentration of industrial surfactants (C) measured by Hyamine titration (green triangle) and
455 HPLC (purple triangle) and evolution of the transmittance at 255 nm (red square) vs. injected
456 porous volume (V_p). The dotted line stands for the end of the surfactant injection step.

457

458 It is important to note that online measurement by UV-visible spectroscopy cannot be used to
459 quantitatively monitor surfactant concentration due to the presence of other interfering substances,
460 as previously mentioned above. However, it can advantageously be used to follow the
461 breakthrough of surfactants and identify their presence in specific samples generated throughout
462 the coreflood experiment. In this case it is necessary to use the information obtained in the
463 transmission mode. The evolution of the transmittance measured online at a wavelength of 255 nm
464 is superimposed in Figure 12. A clear breakthrough of surfactants corresponds to the point where
465 transmittance falls to zero. As long as the effluents contain surfactant, the transmittance value
466 remains close to zero, and then begins to increase again as the surfactant concentration itself

467 becomes negligible. This information can advantageously be used to select the relevant tubes for
468 further offline analysis.

469 This experiment led us to the conclusion that the experimental setup makes it possible to obtain a
470 "clean" aqueous phase (i.e. containing no free oil) from the effluents for the quantification of
471 surfactants. The quality of this aqueous phase enables to provide accurate results using offline
472 techniques (Hyamine titration, HPLC) and avoids the time-consuming manual oil/water separation
473 step. Moreover, the use of online UV-visible spectroscopy analysis allows an online monitoring
474 of the surfactant presence which is a significant improvement over the current protocols where this
475 information is obtained retrospectively.

476 4. CONCLUSIONS

477 In this work, a micro/millifluidic setup was developed with the aim of quantifying online the
478 surfactant concentration at the outlet of a coreflood experiment. The experimental setup includes:
479 a dilution chip to transfer the surfactants in the water phase, a membrane-based separation device,
480 and an online UV-visible absorption spectrometer for surfactant monitoring.

481 The performance of the membrane separation device was first evaluated. The range of pressure for
482 which an oily phase does not flow through the membrane was determined depending on the
483 interfacial tension. In this pressure range, the performance of the membrane separation device was
484 assessed with various flow rates (1 to 10 mL/h) and salinities (0 to 15 g/L NaCl). The fluid
485 collected downstream the membrane contains around 90% water, confirming the efficiency of the
486 device over a wide range of operating conditions.

487

488 The whole experimental setup was then successfully tested on model fluid mixtures (decane, NaCl
 489 brine and SDBS) and real systems (Arabian Light oil, NaCl brine and a mixture of IOS-AGES).
 490 The membrane separation device allowed a very efficient separation of oil-in-water emulsions and
 491 it was possible to accurately measure the concentration of surfactants in the separated aqueous
 492 phase.

493 The setup was finally implemented downstream a coreflood experiment to test its efficiency under
 494 industrial conditions. The oil/water separation proved to be efficient, providing an aqueous phase
 495 compatible with further offline analyses like potentiometric Hyamine titration or HPLC. The direct
 496 online surfactant measurement with UV-visible spectroscopy was unfortunately prevented by the
 497 presence of interferences due to the contact with crude oil. However, the transmittance signal could
 498 advantageously be exploited to continuously follow the breakthrough of surfactants and target the
 499 relevant samples for offline analyses, which is clearly an improvement compared to classical
 500 protocols. Table 3 summarizes the upgrading reached thanks to the experimental setup developed
 501 in the present study.

502 **Table 3.** Improvements obtained in the present study, compared to classical monitoring
 503 approaches

	Classical methods	Improvements obtained in the present study
Oil/water separation	Offline gravity separation	Online membrane separation
Surfactant analysis	Offline	Online/Offline
Surfactant analysis technique	Potentiometric titration	UV-visible (online) or HPLC (offline)
Measurement accuracy	Low: (1) poor oil/water separation leading to high uncertainties, (2) part of the surfactant remains trapped in the oil phase and is therefore not quantified.	Greatly improved: (1) very low residual oil amounts, (2) the dilution step enables to transfer all surfactant in the aqueous phase.

504

505

506 AUTHOR INFORMATION

507 **Author Contributions**

508 The manuscript was written through contributions of all authors. All authors have given approval
509 to the final version of the manuscript.

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