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► **To cite this version:**

Thomas Lesaffre. Population-level consequences of inheritable somatic mutations and the evolution of mutation rates in plants. Proceedings of the Royal Society B: Biological Sciences, Royal Society, The, 2021. hal-03404994

**HAL Id: hal-03404994**

**<https://hal.archives-ouvertes.fr/hal-03404994>**

Submitted on 26 Oct 2021

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POPULATION-LEVEL CONSEQUENCES OF INHERITABLE SOMATIC  
MUTATIONS AND THE EVOLUTION OF MUTATION RATES IN PLANTS

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**ABSTRACT.** Inbreeding depression, that is the decrease in fitness of inbred relative to outbred individuals, was shown to increase strongly as life expectancy increases in plants. Because plants are thought to not have a separated germline, it was proposed that this pattern could be generated by somatic mutations accumulating during growth, since larger and more long-lived species have more opportunities for mutations to accumulate. A key determinant of the role of somatic mutations is the rate at which they occur, which likely differs between species because mutation rates may evolve differently in species with contrasting life-histories. In this paper, I study the evolution of the mutation rates in plants, and consider the population-level consequences of inheritable somatic mutations given this evolution. I show that despite substantially lower somatic and meiotic mutation rates, more long-lived species still tend to accumulate larger amounts of deleterious mutations because of the increased number of opportunities they have to acquire mutations during growth, leading to higher levels of inbreeding depression in these species. However, the magnitude of this increase depends strongly on how mutagenic meiosis is relative to growth, to the point of being close to nonexistent in some situations.

# 1 Introduction

1 Plant growth is fueled by cell divisions occurring in meristems. Each shoot is produced by  
2 an apical meristem and may bear axillary meristems, which are typically situated in the  
3 axils of leaves and grow out to become the apical meristem of a new shoot upon activation  
4 (Burian et al., 2016). As meristematic cells generate all the tissues constituting the shoot,  
5 any mutation occurring in a meristematic cell will be borne by all the cells it gave rise to,  
6 leading to genetic mosaicism within individual plants (Schoen and Schultz, 2019). Fur-  
7 thermore, because meristems also give rise to reproductive tissues, mutations occurring  
8 during growth before the differentiation of the germline, that is somatic mutations, may  
9 be present in the gametes and hence be inherited (although how frequently the inheritance  
10 of somatic mutations occurs is currently unknown, Lanfear, 2018). All else being equal, it  
11 follows that the larger and the older a given plant grows, the more cell divisions it under-  
12 goes and the more somatic mutations it should accumulate and transmit to its offspring,  
13 potentially leading to a higher mutation load in more long-lived and larger species since  
14 it is thought that most mutations are deleterious (Eyre-Walker and Keightley, 2007).

15 Inbreeding depression, that is the decrease in fitness of inbred relative to outbred  
16 individuals (Charlesworth and Charlesworth, 1987), is thought to be mostly generated  
17 by recessive deleterious mutations maintained at mutation-selection balance in popula-  
18 tions (Charlesworth and Willis, 2009). Hence, Scofield and Schultz (2006) proposed that  
19 somatic mutations accumulation could lead to higher inbreeding depression in larger and  
20 more long-lived species. Consistent with this view, inbreeding depression was indeed shown  
21 to increase strongly as life expectancy increases across plant species (Duminil et al., 2009;  
22 Angeloni et al., 2011). Furthermore, Bobiwash et al. (2013) showed that substantial in-

23 breeding depression was generated by somatic mutations in a study performed at the  
24 phenotypic level in old *Vaccinium angustifolium* clones. This is, however, the only em-  
25 pirical test of Scofield and Schultz (2006)'s idea. Besides, recent theoretical investigations  
26 have shown that variations in inbreeding depression can in principle be generated by dif-  
27 ferences in the fitness effect of mutations between species with contrasting life-histories  
28 (Lesaffre and Billiard, 2020), so that somatic mutations accumulation may not always be  
29 needed to explain variations in the magnitude of inbreeding depression across plant species.  
30 Moreover, theoretical investigations of the population-level consequences of somatic mu-  
31 tations accumulation are lacking, so that their role in the maintenance of high inbreeding  
32 depression in long-lived species remains poorly understood. Indeed, theoretical studies  
33 regarding somatic mutations in plants either focused on the case of favorable mutations,  
34 conferring resistance against herbivores (e.g. Antolin and Strobeck, 1985), or studied the  
35 fate of deleterious mutations subject to intra-organismal selection (Otto and Orive, 1995;  
36 Pineda-Krch and Lehtilä, 2002), but never considered the population-level consequences  
37 of recessive deleterious mutations (Schoen and Schultz, 2019). In summary, deleterious  
38 somatic mutations accumulation has been proposed as a mechanism to explain the rarity  
39 of selfing species among long-lived plants (Scofield and Schultz, 2006), consistent with  
40 empirical measures of inbreeding depression, but theoretical support for this idea remains  
41 scarce.

42 An important determinant of the consequences of somatic mutations accumulation is  
43 the rate at which said mutations accumulate during growth, that is the somatic mutation  
44 rate, which is defined here as the number of mutations occurring per unit of vegetative  
45 growth. This rate is likely influenced by evolutionary mechanisms similar to those affect-

46 ing mutation rates in general. For example, [Kimura \(1967\)](#) showed that mutation rates  
47 should be shaped by the opposition between the increase in the number of deleterious  
48 mutations borne by individuals with higher mutation rates on the one hand, which causes  
49 indirect selection against genetic variants increasing mutation rates to increase, and the  
50 direct fitness cost there is to increasing the fidelity of DNA replication on the other hand.  
51 Besides, [Lynch \(2011\)](#) proposed that selection to decrease the mutation rate should be-  
52 come weaker than genetic drift at some point in finite populations, thereby favoring the  
53 persistence of non-zero mutation rates. Nevertheless, the inheritability of somatic muta-  
54 tions in plants and their intrinsic link with growth and life expectancy likely contribute  
55 to shape the evolution of mutation rates in a specific manner which was never tackled  
56 theoretically. Great interest was however taken in empirically detecting somatic muta-  
57 tions and comparing mutations rates in a variety of plants species ranging from the very  
58 short-lived *Arabidopsis thaliana* to ancient, centuries old trees. In an analysis performed  
59 across many plant families, [Lanfear et al. \(2013\)](#) showed that taller species among pairs of  
60 sister species have significantly lower rates of molecular evolution, measured as the number  
61 of substitutions per site per  $10^6$  years. They argued that contrary to animals, this pattern  
62 is not a mere reflection of differences in generation time, which would reflect different rates  
63 of genome copying per unit of time, because somatic genome copying events contribute  
64 to the inheritable genetic variation in plants. Instead, they proposed that this pattern  
65 may be due to slower growth in taller species, which results in a lower number of mitosis  
66 (and therefore mutations) per unit of time. Consistent with this view, it was shown at the  
67 cellular level that axillary meristems cells are set aside early during the growth of a shoot  
68 ([Burian et al., 2016](#)), resulting in a number of cell divisions increasing linearly with the

69 number of branching events in trees although the number of terminal branches increases  
70 exponentially. Furthermore, multiple studies showed that somatic mutation rates tend  
71 to be considerably lower in taller, more long-lived species (Schmid-Siegert et al., 2017;  
72 Plomion et al., 2018; Hofmeister et al., 2019; Orr et al., 2020; Wang et al., 2019; Hanlon  
73 et al., 2019). For instance, Orr et al. (2020) found the somatic mutation rate per gener-  
74 ation to be only ten times higher in *Eucalyptus melliodora* than in *Arabidopsis*, despite  
75 being  $> 100$  times larger in size.

76 Thus, empirical evidence indicates that more long-lived species have acquired mecha-  
77 nisms to reduce the amount of mutations accumulated during growth on the one hand, but  
78 still present high levels of inbreeding depression on the other hand, which suggests that  
79 more long-lived species still accumulate more mutations despite above mentioned limiting  
80 mechanisms. The aim of the present study is to disentangle the relationship between these  
81 two observations. I first study the evolution of the mutation rate in plants, and then con-  
82 sider the number of mutations and the magnitude of inbreeding depression maintained at  
83 mutation-selection balance, given the evolutionarily stable mutation rate reached by the  
84 population. To do so, I extend the work of previous authors (Kimura, 1967; Gervais and  
85 Roze, 2017) to the case of a perennial population in which individuals grow as they age and  
86 accumulate mutations in doing so. I obtain analytical predictions which are then tested  
87 against the output of individual-centered simulations. I show that the evolutionarily stable  
88 mutation rate should decrease in plants as life expectancy increases, because deleterious  
89 mutations have more time to accumulate in more long-lived species. Furthermore, I show  
90 that despite substantially lower per year mutation rates, more long-lived species still tend  
91 to accumulate larger amounts of deleterious mutations because of higher per generation,

92 leading to higher levels of inbreeding depression in these species. However, the magnitude  
93 of this increase depends strongly on how mutagenic meiosis is relative to growth.

## 2 Methods

94 **Model outline.** I consider a large population of hermaphroditic diploids. Individuals  
95 survive between mating events with a constant probability  $S$ . Juveniles may only settle in  
96 replacement of deceased individuals, so that population size is kept constant. Individuals  
97 are assumed to be made of a trunk, which grows by one section between each flowering  
98 event (FIG. 1). This growth model is neither intended to depict a particular kind of plants  
99 nor to be a realistic model of plant growth. It was chosen because it is the simplest growth  
100 model incorporating within-individual genetic mosaicism. Besides, as long as mutations  
101 do not interfere with the growth process, as it is the case here (see below), more compli-  
102 cated growth models would only alter the age distribution of sections within individuals,  
103 which should not qualitatively alter the results presented in this study provided that older  
104 individuals are still made of older sections on average.

105 Mutations at the selected loci occur both during meiosis and somatic growth. The  
106 meiotic mutation rate of a given individual ( $u$ ), which includes both mutations occurring  
107 during meiosis and during the development of disposable reproductive parts, is determined  
108 by its genotype at a single modifier locus. At this locus, I consider the fate of a rare mutant  
109 ( $m$ ) with a weak effect ( $\varepsilon$ ) competing with a resident allele ( $M$ ). This mutant allele is  
110 codominant with the resident, so that an individual's meiotic mutation rate is given by  
111  $u_{MM} = u_0$ ,  $u_{Mm} = u_0 + \varepsilon$ , or  $u_{mm} = u_0 + 2\varepsilon$ , depending on its genotype at the modifier.

112 Mutations occur due to the unrepaired misincorporation of nucleotides during DNA

113 replications, or due to DNA lesions occurring between replications which are not repaired  
114 in time before the next replication event (*i.e.* cell division), so that they end up being  
115 incorporated in the daughter cells' genome (Gao et al., 2016). Because there is, to my  
116 knowledge, no reason to expect DNA repair mechanisms to fundamentally differ between  
117 meiotic and somatic cell divisions, I hypothesized that meiotic and somatic mutation rates  
118 should evolve jointly to some extent. Importantly however, these two rates differ in at  
119 least two ways. First, they are not defined on the same scale. Indeed, while the somatic  
120 mutation rate is usually defined as a number of mutations per unit of growth, as it is the  
121 case in the present model, meiotic mutation rates are defined at the scale of a reproductive  
122 event. Thus, they may each cover a very different number of cell divisions, especially  
123 since recent empirical evidence has shown that the number of cell divisions separating  
124 axillary buds stem cells from those of the apical meristem they emerged from may be  
125 much lower than previously thought due to strong quiescence mechanisms (Burian et al.,  
126 2016). Second, meiotic cell divisions necessarily include recombination, causing additional  
127 double-strand DNA breaks and therefore giving the opportunity for more mutations to  
128 occur during meiosis than during mitosis (Magni and Von Borstel, 1962). Hence, the  
129 relationship between these two mutation rates is not straightforward, because different  
130 genetic events may happen and different numbers of cell divisions may occur over the course  
131 of a growth season and during a reproductive event. In the absence of a more mechanistic  
132 model, it is hard to give a biologically well-motivated shape to this relationship. Thus, in  
133 an effort to keep the model as simple as possible, I will assume that somatic mutations  
134 accumulate at rate  $\gamma u$  per unit of growth (that is, per section, FIG. 1), where  $\gamma$  is a  
135 positive real number which allows one to tune the intensity of somatic mutation relative



136 to meiotic mutation. In other words, I assume there is a linear relationship between the  
 137 two rates.

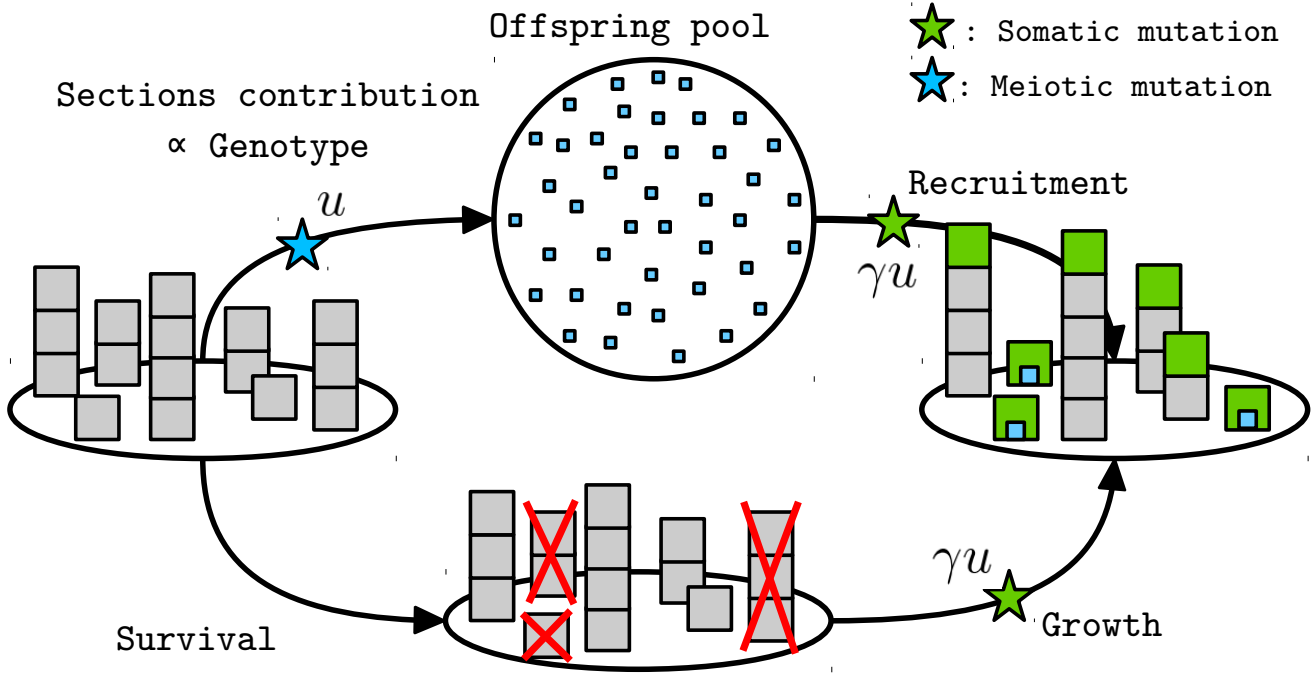


FIGURE 1: Life cycle of the modeled population. Small blue squares depict seeds. Green squares depict the sections grown during the last growing season. Juveniles go through one growing season before reproducing, and are therefore made of a single section as depicted by the green squares wrapping the small blue ones. Stars show the steps at which meiotic (blue) and somatic (green) mutation occurs. The rate at which mutation occurs is indicated beside each star.

138 I assume that any section can contribute to reproduction (FIG. 1). Self-fertilisation  
 139 occurs at rate  $\alpha$ , a fraction  $\sigma$  of which imperatively occurs within the same section. The  
 140 remaining fraction  $1 - \sigma$  can occur between sections within the individual. Introducing  
 141  $\sigma$  into the model enables one to study the effect of within versus between sections selfing  
 142 more easily.

143 A section's fecundity is determined by its genotype at a very large number of biallelic  
 144 loci acting multiplicatively. At these loci, allele 0 is an healthy allele, while allele 1 is a

145 mutated allele which diminishes the section's fecundity by a proportion  $s$ . In heterozygotes,  
 146 allele 1 expresses proportionally to its dominance coefficient  $h$ . Following previous authors  
 147 (Gervais and Roze, 2017), I also introduce a DNA replication fidelity cost function,  $f$ ,  
 148 which is an increasing function of the meiotic mutation rate  $u$ . Gervais and Roze (2017)  
 149 considered a variety of cost functions and came to qualitatively similar conclusions in every  
 150 case. Yet, most of their results were obtained using the cost function given in Equation  
 151 (1),

$$f(u) = e^{-\frac{c}{u}}, \quad (1)$$

152 where  $c$  is the cost of replication fidelity, which is also used in this study. Thus, the  
 153 fecundity of a section is given by

$$W = f(u) \times (1 - s)^{n_{\text{hom}}} (1 - sh)^{n_{\text{het}}}, \quad (2)$$

154 where  $n_{\text{hom}}$  and  $n_{\text{het}}$  are the number of mutations borne in the homozygous and heterozy-  
 155 gous states, respectively.

156 **Analytical methods.** To study the model, I use the theoretical framework described  
 157 in Kirkpatrick et al. (2002), which relies on indicator variables to describe individuals'  
 158 multilocus genotypes. In the analytical work, the effect of the proportion of obligate  
 159 within-section selfing ( $\sigma$ ) is neglected since it is difficult to incorporate and will prove  
 160 to have very little impact on the results. For the sake of brevity, derivations of the  
 161 results presented in the following sections are detailed in Appendices I.1 and I.2 for results  
 162 regarding the evolution of mutation rate and the mutation-selection equilibrium properties

163 of the population given the evolutionarily stable mutation rate, respectively.

164 **Individual-centered simulations.** Individual-centered simulations were run to test  
165 the validity of analytical approximations. The simulation program was coded in C++11, is  
166 available from GitHub and has been given a DOI using Zenodo (10.5281/zenodo.5166952).  
167 In this program, individuals are represented by two chromosomes of length  $\lambda$  (expressed in  
168 cM) with the modifier situated at the center and along which mutations can occur at any  
169 position, so that infinitely many selected loci are effectively modeled (Roze and Michod,  
170 2010).

171 **Modeled loci.** Following the work of Gervais and Roze (2017), it is assumed that  
172 infinitely many alleles exist, coding for any value of  $u \in [0, +\infty[$  exist at the modifier.  
173 Mutations at the modifier occur at rate  $u_m = 10^{-3}$ , and the value coded by the new  
174 allele is sampled from a Gaussian distribution centered on the former allele value with  
175 standard deviation  $\sigma_m = 10^{-2}$ , which is truncated at zero to prevent the modifier from  
176 going out of range. At selected loci, the number of mutations occurring on a chromosome  
177 during a given mutation event is sampled from a Poisson distribution with mean  $u$  ( $\gamma u$  for  
178 somatic growth), and their position is sampled from a uniform distribution. Recombination  
179 is modeled by exchanging segments between homologous chromosomes. The number of  
180 crossing-overs is sampled in a Poisson distribution with mean  $\lambda$  and their positions are  
181 sampled from a uniform distribution along chromosomes. Every time a mutation occurs,  
182 the age of the section at which it occurred along the individual is stored, so that the  
183 genotype of any section within an individual can be reconstructed at any time from the  
184 individual genome. This method allows one to gain substantial computation time because

185 mutations are stored only once per individual instead of being copied once for each new  
186 section.

187 **Sequence of events.** The population is kept of constant size,  $N$ . Between each  
188 mating event, individuals have a constant survival probability  $S$ . If they survive, they  
189 grow by one section, and mutations occur at rate  $\gamma u$  in this section. If they die, they are  
190 replaced by an offspring produced by the population. Any section within any individual  
191 can be chosen as a parent, with a probability proportional to its fecundity (Equation  
192 2). The offspring is produced by self-fertilisation with probability  $\alpha$ , in which case the  
193 chosen section mates with itself with probability  $\sigma$ , and with any section within the same  
194 individual with probability  $1 - \sigma$ . When selfing occurs between sections, a second parental  
195 section is selected within the individual. When the offspring is not produced by self-  
196 fertilisation, which occurs at rate  $1 - \alpha$ , it is produced by random mating and a second  
197 parent is selected from the whole population. Mutation occurs at rate  $u$  during meiosis.

198 **Measurements.** Once the equilibrium was reached, that is when both the muta-  
199 tion rate and the average number of mutations per chromosome were at equilibrium, the  
200 average number of mutations per chromosome in seeds, the average mutation rate and  
201 inbreeding depression were measured. Although individuals are chimeric in the model, I  
202 stuck with measuring inbreeding depression at the individual level to be in line with its  
203 formal definition. To do so, I counted how many times each individual was chosen as  
204 a parent before it died (*i.e.* I measured its lifetime reproductive success) and used this  
205 quantity as a measure of lifetime fitness. Individuals were marked as being produced by  
206 outcrossing (0), selfing within the same section (1), and selfing between sections within

207 the same individual (2), so that I was able to measure fitness differences between these  
208 various categories of individuals. Namely, I measured inbreeding depression, that is the  
209 decrease in fitness of selfed individuals relative to the outcrossed ( $\delta_{01}$ ), and autogamy de-  
210 pression (Schultz and Scofield, 2009; Bobiwash et al., 2013), that is the decrease in fitness  
211 of within-section selfed individuals relative to between-sections ones ( $\delta_{12}$ ). Ten replicates  
212 were run for each parameter set. Simulations were kept running for  $10^6$  and  $2 \times 10^5$  re-  
213 productive seasons for life expectancies lower and higher than 200 reproductive seasons,  
214 respectively. Results were averaged over the last  $10^5$  reproductive cycles and  $2 \times 10^4$  for  
215 life expectancies lower and higher than 200 reproductive seasons, respectively, and the  
216 95% confidence interval around the mean was also recorded.

### 3 Results

217 In what follows, life expectancy ( $E$ ) will be used to discuss results instead of survival  
218 probability ( $S$ ) for the sake of clarity and biological relevance. Given survival probability  
219  $S$ , life expectancy can be computed as

$$E = \frac{1}{1 - S}. \quad (3)$$

#### 3.1 Evolutionarily stable mutation rate

220 Let us first study the evolution of the mutation rate. It is shown in Appendix I.1 that the  
221 evolution of the mutation rate is the result of the opposition between the direct cost of  
222 DNA replication fidelity, which is higher when the mutation rate is lower, and the indirect  
223 selection caused by deleterious alleles which tend to be more frequently linked with modifier

224 alleles increasing the mutation rate (Equation A23). The resulting evolutionarily stable  
225 mutation rate is given by

$$u^* = \sqrt{-\frac{c}{\hat{s}_{ind}}}, \quad (4)$$

226 where  $\hat{s}_{ind}$  encapsulates the intensity of indirect selection acting on the modifier. Its  
227 expression is derived in Appendix I.1.5. FIGURE 2 shows the evolutionarily stable mutation  
228 rate as a function of life expectancy (top row), along with the intensity of indirect selection  
229 (bottom row), for cases where  $\gamma = 1$ ,  $\gamma = 0.1$  and  $\gamma = 0.01$ . I chose to focus on cases  
230 where  $\gamma \leq 1$ , that is on cases where more mutations are produced during meiosis (plus the  
231 production of disposable reproductive parts) than during the development of a new section,  
232 on the basis of three lines of evidence. First, direct observations of plant development at  
233 the cellular level indicate that cells destined to form axillary meristems undergo much fewer  
234 divisions than other cells from the moment they are produced in the apical meristem, which  
235 suggests that the number of cell divisions per branching event, and therefore the number of  
236 opportunities for mutations to accumulate, may be lower than previously thought (Burian  
237 et al., 2016). Second, estimates of somatic mutation rates per unit of growth tend to  
238 be low (Orr et al., 2020). Third, to my knowledge, the only experiment comparing the  
239 mutagenicity of meiosis and mitosis was performed by Magni and Von Borstel (1962)  
240 in yeast. They found meiosis to be 6 to 20 times more mutagenic than mitosis, which  
241 further suggests that  $\gamma$  may tend to be lower than 1. Besides, performing simulations with  
242  $\gamma > 1$  proved to be very challenging since the number of mutations accumulated in the  
243 population quickly became very high, causing simulations to run very slowly and consume  
244 a lot of resources.

Equilibrium mutation rate and indirect selection as a function of life expectancy ( $\sigma = 0.5$ )

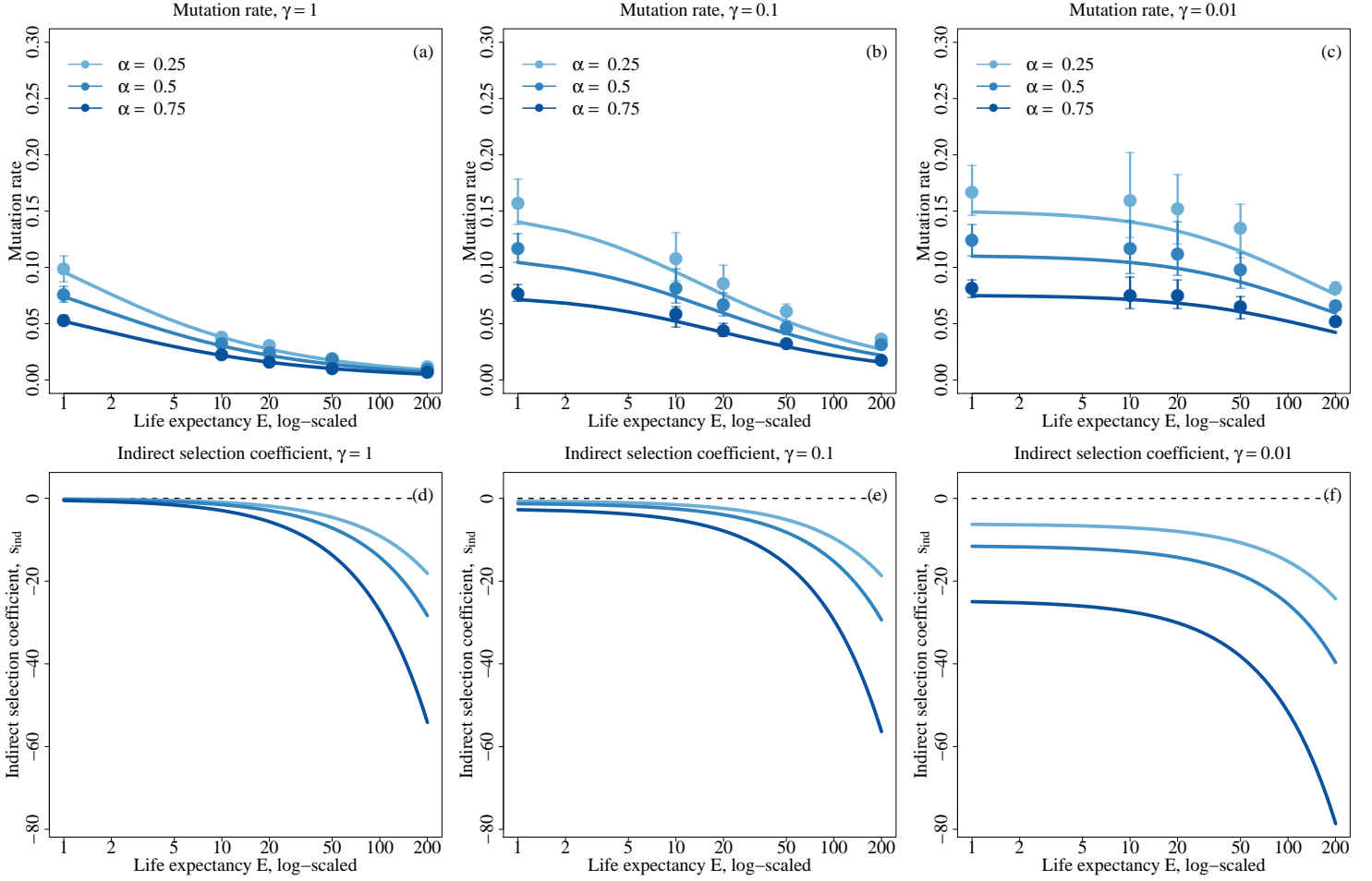


FIGURE 2: Evolutionarily stable mutation rate (top) and intensity of indirect selection (bottom) as a function of life expectancy (log-scaled) for various selfing rates (colors) and for  $\gamma = 1$  (left),  $\gamma = 0.1$  (middle) and  $\gamma = 0.01$  (right). Other parameters values are  $s = 0.05$ ,  $h = 0.3$ ,  $c = 0.0014$ ,  $\lambda = 20$ , and  $\sigma = 0.5$ . Dots depict simulation results and error bars depict the 95% confidence intervals. Lines depict analytical predictions.

245 The evolutionarily stable mutation rate decreases with life expectancy for all  $\gamma$  values  
 246 (FIG. 2a-c). In both cases, this is due to the greater number of opportunities to accumulate  
 247 deleterious mutations in more long-lived species because they go through more growth  
 248 events, which in turn causes indirect selection to increase against alleles increasing the  
 249 mutation rate because deleterious mutations become more numerous (FIG. 2d-f).

250 The mutation rate also decreases as the selfing rate ( $\alpha$ ) increases, which may seem

251 counter-intuitive since selfing tends to reduce the number of deleterious mutations seg-  
252 regating in the population through purging [Roze \(2015\)](#). However, self-fertilisation also  
253 causes genetic associations between selected loci and the modifier to increase, thereby in-  
254 creasing indirect selection and resulting in a decrease of the evolutionarily stable mutation  
255 rate when the selfing rate increases as shown by [Gervais and Roze \(2017\)](#). The results pre-  
256 sented in [FIG. 2](#) were obtained assuming half of selfing events occurred imperatively within  
257 the same section ( $\sigma = 0.5$ ). Cases with  $\sigma = 0$  and  $\sigma = 1$  were also investigated and yielded  
258 very similar results, which are presented in [FIG. S3](#) and [S4](#), respectively, in [Appendix II](#).  
259 The very small effect of  $\sigma$  on the results is due to the relatively low evolutionarily stable  
260 mutation rate, which causes few somatic mutations to occur during growth, and to the  
261 fact that weak selection was assumed so that mutations have little effect on their bearer's  
262 fitness.



Mutations / haplotype and inbreeding depression as a function of life expectancy ( $\sigma = 0.5$ )

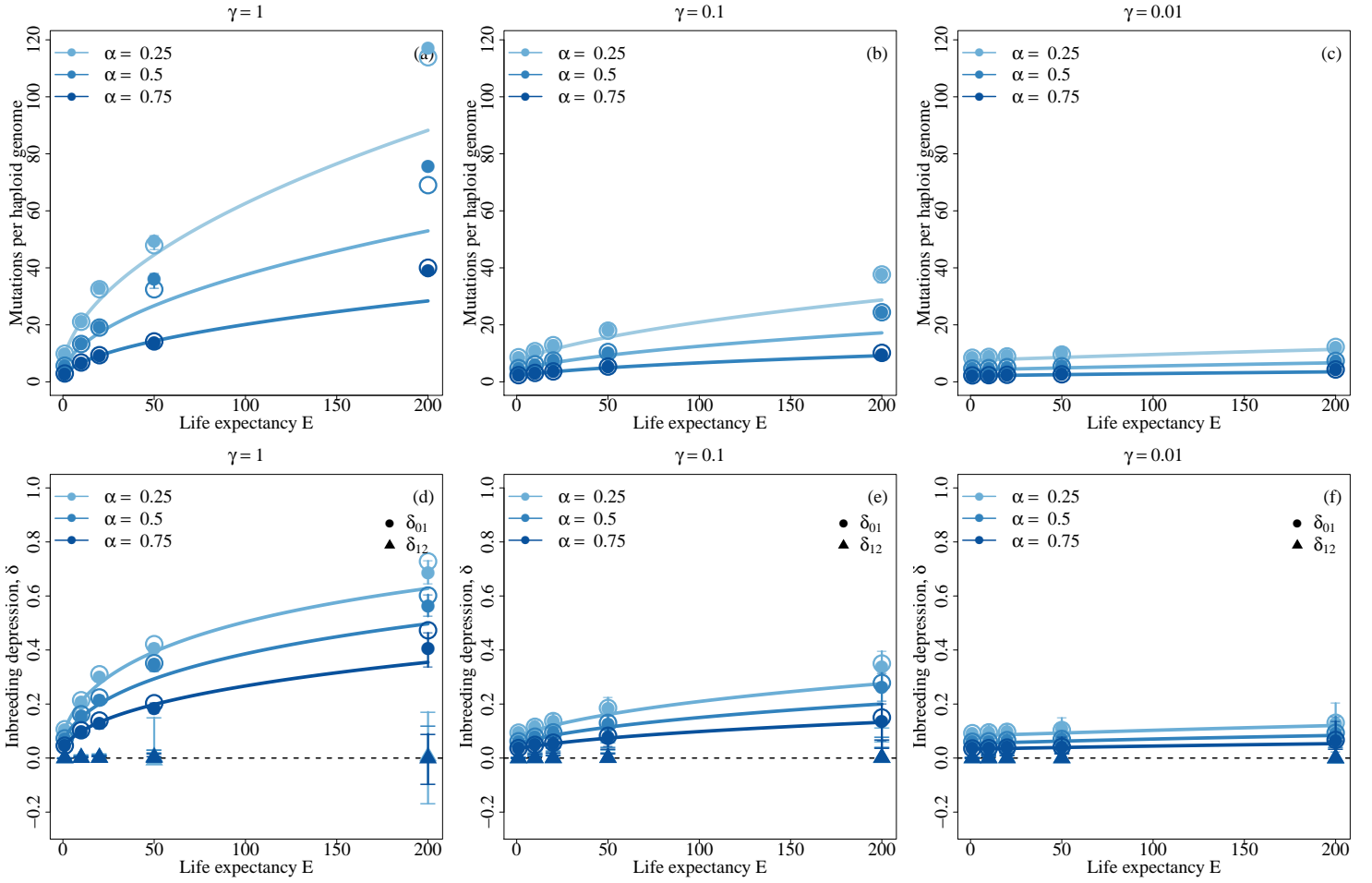


FIGURE 3: Average number of mutations per haploid genome (top) and inbreeding depression (bottom) as a function of life expectancy (log-scaled) for various selfing rates (colors) and  $\gamma = 1$  (left),  $\gamma = 0.1$  (middle) and  $\gamma = 0.01$  (right). Other parameters values are  $s = 0.05$ ,  $h = 0.3$ ,  $c = 0.0014$ ,  $\lambda = 20$ , and  $\sigma = 0.5$ . Filled dots depict simulation results and error bars depict the 95% confidence intervals. Lines depict analytical predictions. Open circles depict the value predicted by our analytical model when the equilibrium mutation rate from simulations is used instead of Equation 4. On the bottom row, dots indicate inbreeding depression ( $\delta_{01}$ ), while triangles indicate autogamy depression ( $\delta_{12}$ ).

### 3.2 Mutation-selection balance

263 Once the mutation rate has reached an equilibrium and the population is at mutation-  
 264 selection balance, I show in Appendix I.2.1 that a leading order approximation of the

265 average number of mutations per haploid genome in juveniles ( $n$ ) is given by

$$n \approx \frac{\hat{u}^*}{s[h + F(1 - h)]} - \gamma u^* \frac{S}{1 - S}, \quad (5)$$

266 where  $u^* = \sqrt{-\frac{c}{\hat{s}_{ind}}}$ , and  $\hat{u}^* = \left(1 + \frac{\gamma}{1 - S}\right) u^*$  depicts the total mutation rate of the

267 population over the course of one timestep, including both meiotic and somatic mutations.

268 As for inbreeding depression calculated between outcrossed and selfed individuals ( $\delta_{01}$ ), it

269 is given by

$$\delta_{01} = 1 - \exp \left[ -s(1 - 2h) \frac{1 + F}{2} \left( \frac{\hat{u}^*}{s[h + F(1 - h)]} - \gamma u^* \frac{S}{1 - S} \right) \right], \quad (6)$$

270 where  $F = \frac{\alpha}{2 - \alpha}$ , to leading order in  $s$ . Again, I do not consider the impact of the

271 proportion of selfing occurring within or between sections ( $\sigma$ ) in the analytical model since

272 it is negligible. FIGURE 3 shows the number of mutations per haploid genome among

273 juveniles ( $n$ , top row), and inbreeding and autogamy depression ( $\delta_{01}$  and  $\delta_{12}$ , bottom

274 row) at mutation-selection balance. Deviations between analytical predictions (lines) and

275 simulations results (dots) are observed. They can be explained by the slight differences

276 between the predicted evolutionarily stable mutation rate and the equilibrium mutation

277 rate reached by simulations, which build up large differences in  $n$  when life expectancy

278 becomes high. Indeed, when the equilibrium mutation rate from the simulations is used

279 to predict  $n$  instead of Equation (4), the agreement between predictions (open circles) and

280 simulation results (dots) is restored.

281 The number of mutations maintained  $n$  increases with life expectancy in every case

282 due to the greater amount of opportunities for mutations to accumulate in more long-

283 lived species. Indeed, the denominator of the first term in Equation (5) shows that the  
284 intensity of selection is independent of life expectancy, while the total mutation rate  $\hat{u}^*$  is  
285 an increasing function of life expectancy in all investigated cases despite the fact that the  
286 equilibrium mutation rate *per mutagenic event* ( $u^*$ ) decreases in more long-lived species  
287 (FIG. S1 in Appendix II shows the total mutation rate as a function of life expectancy in  
288 said cases). The increase of  $n$  with life expectancy becomes much lower when  $\gamma$  decreases  
289 to the point of being barely noticeable with  $\gamma = 0.01$ , despite the fact that the equilibrium  
290 meiotic mutation rate is slightly higher in that case. This result is generated by the joint  
291 effect of  $\gamma$ , which reduces the contribution of somatic mutations as it decreases, and of  
292 the evolution of mutation rate which is lower at the evolutionary equilibrium in more  
293 long-lived species (as an additional illustration, FIG. S2 in Appendix II compares the  
294 obtained  $\hat{u}^*$  with the one expected if the evolutionarily stable mutation rate for annuals,  
295 that is  $E = 1$ , is assumed for all life expectancies for various  $\gamma$  values). As a result of these  
296 effects, inbreeding depression gets lower as  $\gamma$  decreases and increases as life expectancy  
297 increases, but this increase becomes less and less marked for smaller  $\gamma$  values. Furthermore,  
298 consistent with the negligible effect  $\sigma$  had on the evolution of the mutation rate, almost  
299 no autogamy depression is generated (triangles in FIG. 2, bottom row).

## 4 Discussion

300 In this paper, I studied the evolution of the mutation rate when mutations accumulating  
301 during growth are assumed to be inheritable, and considered the consequences of such  
302 mutation accumulation for mutation load and inbreeding depression in species with varying  
303 degrees of perenniality.

## 4.1 Evolution of the mutation rate

304 I showed that the evolutionarily stable mutation rate decreases as life expectancy increases  
305 because of the greater number of opportunities to accumulate mutations during growth  
306 in more long-lived species, which makes indirect selection against alleles increasing the  
307 mutation rate stronger. However, although the mutation rate per mutagenic event ( $u$ ),  
308 that is per growth season or per meiosis in the present model, decreased in more long-lived  
309 species, the total mutation rate ( $\hat{u}$ ), that is the rate at which mutations entered the pop-  
310 ulation through both somatic growth and meiosis, increased. Hence, results indicate that  
311 while we should expect more efficient mechanisms reducing the accumulation of deleteri-  
312 ous mutations during growth to evolve in more long-lived species, so that their per unit of  
313 growth and per year mutation rate should be lower, their per generation mutation rates  
314 should still be higher. These predictions are in line with empirical evidence, which suggest  
315 that mutation rates per generation tend to be higher in more long-lived species although  
316 the mutation rates per unit of growth tend to be lower (Hofmeister et al., 2019; Hanlon  
317 et al., 2019; Orr et al., 2020).

318 I modeled the evolution of the mutation rate following the work of Kimura (1967), by  
319 assuming there is a direct fitness cost to DNA replication fidelity opposing the indirect  
320 selection generated by deleterious mutations linked to the modifier, so that the mutation  
321 rate was maintained greater than zero in response to a trade-off. An alternative mecha-  
322 nism, which is not mutually exclusive with the trade-off described above, was put forward  
323 by Lynch (2011). They proposed that selection should always act to reduce the mutation  
324 rate, down until it becomes so low that the selective advantage brought by any further  
325 reduction should be overwhelmed by genetic drift, thus maintaining non-zero mutation

326 rates because alleles further decreasing the mutation rate should at some point become  
327 effectively neutral, and thereby creating a lower bound for the evolution of the mutation  
328 rate (Lynch, 2011). This lower bound is inevitably influenced by effective population size,  
329 as it plays on the relative strength of selection and genetic drift. In the present model,  
330 I overlooked Lynch (2011)'s lower bound by assuming a large and fixed population size.  
331 Yet, effective population sizes are expected to be higher in more long-lived species in  
332 which generations overlap (Felsenstein, 1971; Charlesworth, 1980; Petit and Hampe, 2006;  
333 Duminil et al., 2009), which implies the lower bound described by Lynch (2011) should  
334 be met for lower mutation rates in said species. Hence, we should expect the decrease  
335 in the evolutionarily stable mutation rate described in this study to become sharper in  
336 conditions where Lynch (2011)'s lower bound is expected to matter for the evolution of  
337 the mutation rate.

## 4.2 Inbreeding depression

338 The larger total mutation rate in more long-lived species led to the maintenance of more  
339 mutations in the population at mutation-selection balance, and therefore to higher in-  
340 breeding depression in these species, consistent with results from meta-analyses which  
341 found inbreeding depression to increase in larger-statured, more long-lived species (Du-  
342 minil et al., 2009; Angeloni et al., 2011). Importantly however, the magnitude of the  
343 increase in the total mutation rate, and therefore in inbreeding depression with life ex-  
344 pectancy depended strongly on the relative mutagenicity of meiosis and growth, which  
345 was controlled by the  $\gamma$  parameter in this model. Indeed, while the increase in inbreeding  
346 depression was strong when  $\gamma$  was close to 1, that is when the same amount of mutation

347 was produced during meiosis and during growth between two flowering seasons, it became  
348 smaller as  $\gamma$  decreased, to the point of being barely noticeable for  $\gamma = 0.01$ . This was due  
349 to both the decrease of the evolutionarily stable total mutation rate ( $\hat{u}^*$ ) and to the de-  
350 crease of  $\gamma$ , which made the contribution of somatic mutations to the mutation load more  
351 and more negligible compared with meiotic mutations. Hence, according to the results  
352 presented in this paper, for somatic mutations to be the main driver of the empirically  
353 observed increase in inbreeding depression in more long-lived species, roughly the same  
354 amount of mutations should be produced during growth between two flowering seasons  
355 and during reproduction.

### 4.3 Mating system evolution

356 Inbreeding depression is thought to be one of the main factors preventing the evolution of  
357 self-fertilisation (Lande and Schemske, 1985; Barrett and Harder, 2017). In Angiosperms,  
358 consistent with the observed increase in inbreeding depression in more long-lived species,  
359 there exists a strong correlation between mating systems and life-histories. Indeed, many  
360 self-fertilising species are annuals whereas most long-lived species are strictly outcrossing  
361 (Barrett and Harder, 1996; Munoz et al., 2016). Thus, somatic mutations accumulation  
362 was proposed as an explanation for this correlation (Scofield and Schultz, 2006). While the  
363 results presented in this study indicate that inbreeding depression increases with respect  
364 to life expectancy due to somatic mutations accumulation, particularly when  $\gamma$  is large,  
365 this increase is tempered by the decrease of the evolutionarily stable mutation rate with  
366 life expectancy. Furthermore, in agreement with results obtained by Gervais and Roze  
367 (2017), I showed that the evolutionarily stable mutation rate decreases as the selfing rate

368 increases because the modifier becomes more strongly associated with selected loci. These  
369 decreases of the mutation rate with respect to mating system and life expectancy, together  
370 with the purging effect of self-fertilisation (Roze, 2015), result in a substantial drop in the  
371 magnitude of inbreeding depression as the selfing rate increases in more long-lived species,  
372 potentially opening the way for the evolution of self-fertilisation. Hence, whether somatic  
373 mutations accumulation is sufficient to explain the correlation between life-history and  
374 mating system in Angiosperms when the mutation rate is allowed to evolve jointly with  
375 the mating system remains an open question.

#### 4.4 Autogamy depression

376 In order to empirically estimate the contribution of somatic mutations accumulation to  
377 inbreeding depression using phenotypic data, a method was developed by Schultz and  
378 Scofield (2009). This method, called the autogamy depression test, relies on the compar-  
379 ison of the fitnesses of individuals produced by selfing within an inflorescence with those  
380 of individuals produced by selfing between distant inflorescences on the plant's crown  
381 (Schultz and Scofield, 2009; Bobiwash et al., 2013). In this paper, I performed such test  
382 by measuring autogamy depression ( $\delta_{12}$ ). Contrary to inbreeding depression, I found auto-  
383 gamy depression to be almost null in every case, even in situations where the contribution  
384 of somatic mutations accumulation to inbreeding depression was high. This result can be  
385 explained by the low evolutionarily stable mutation rates, and by the fact that we only  
386 considered mutations with a weak fitness effect. It suggests that the autogamy depression  
387 test should only be able to detect mutations with a large fitness effect in large individ-  
388 uals, where mutations have had time to accumulate. Thus, it implies that detecting no

389 autogamy depression in a given population cannot be taken as evidence of a negligible  
390 contribution of somatic mutations accumulation to the population's mutation load.

## 4.5 Germline segregation and relative mutagenicity of growth and meiosis

391 The results presented above suggest that valuable insights into the evolutionary relevance  
392 of somatic mutations and the evolution of the mutation rate in plants could be gained by  
393 further investigating the  $\gamma$  parameter in this model, which depicts the relative mutagenicity  
394 of meiosis and growth between two flowering seasons, and is likely influenced by at least  
395 three important factors that were either overlooked or only partially accounted for in this  
396 study.

### 4.5.1 Relative mutagenicity of meiosis and mitosis

397 First, it is necessarily influenced by how mutagenic meiotic divisions are in comparison with  
398 mitotic divisions, about which little is known although one may expect meiotic divisions to  
399 generate more mutations, as they generate many more double strand DNA breaks which  
400 are required for recombination and are known to be particularly mutagenic events (Magni  
401 and Von Borstel, 1962; Arbel-Eden and Simchen, 2019).

### 4.5.2 Number of cell divisions separating meristems

402 Second, it is influenced by the number of mitoses occurring between flowering buds. This  
403 number depends on the growth habit of the considered species, because fast growing species  
404 undergo more mitoses per unit of time than slow growing species, and because the rate  
405 at which mitoses occur, and thus the growth rate, may interact with the evolution of the



406 mutation rate. For instance, investing in a higher fidelity of DNA replication may tend to  
407 slow down individual growth.

408 The number of mitoses separating two meristems also depends on patterns of meris-  
409 tematic stem cell divisions that were recently brought to light (Burian, 2021). Indeed,  
410 although it has long been thought that the germline remains unsegregated up until a  
411 meristem switches to the floral state in plants, Burian et al. (2016) showed that within  
412 the apical meristem, the stem cells give rise to a specific cell lineage which will serve as  
413 the axillary meristems' stem cells and spend most of their time in a quiescent, almost  
414 non-dividing state, contrary to surrounding cell lineages which divide vigorously to effect  
415 plant growth. Thanks to this mechanism, the number of mitoses separating two meristems  
416 is greatly reduced and so is the number of cell divisions separating the seed from the ga-  
417 metes, as the germline directly emerges from these stem cells. Hence, although it is clear  
418 that the plant germline remains undifferentiated up until reproduction is triggered, it may  
419 be considered segregated prior to differentiation, because the cells giving rise to it do not  
420 suffer the same fate as surrounding somatic cell lineages, thus behaving as a functional  
421 germline (Romberger et al., 1993; Burian, 2021). The exact timing of such segregation  
422 during plant development is, however, not known (Lanfear, 2018). Therefore, it is impor-  
423 tant to point out that the results presented in this study not only hold if the germline  
424 segregates late in development, but that they would also hold if the germline was actually  
425 segregated as early as the first embryonic cell division (as it is the case in animals) and  
426 remained sheltered within meristems. Indeed, such segregated germline would still have  
427 to go through a non-zero number of mitotic cell divisions to be passed from one meristem  
428 to the next due to developmental constraints (Burian, 2021), so that the number of cell

429 divisions it goes through before reproduction would still be affected by individual growth  
430 and be higher in more long-lived, larger species. In summary, the validity of the results  
431 presented in this paper does not depend on the degree to which the germline is actually  
432 segregated in plants, but the existence of a functional germline as described above, ir-  
433 respective of when germline segregation occurs, supports the idea that plants acquired  
434 physiological mechanisms favoring lower values of  $\gamma$ .

### 4.5.3 Intra-organismal selection

435 Finally, apart from mechanisms reducing the amount of mutations produced during growth,  
436 deleterious mutations may also be affected by intra-organismal selection, which may not  
437 only reduce the growth rate by eliminating mutated cells, but also efficiently purge dele-  
438 rious mutations from the organism, so that little to no somatic mutation may be present  
439 in the gamete, which could make  $\gamma$  smaller among the mutations effectively transmit-  
440 ted to offspring. This could in turn affect the evolution of the mutation rate. Little is  
441 known, however, about the actual efficacy of intra-organismal selection in removing dele-  
442 terious mutations since it was seldom investigated theoretical (Otto and Orive, 1995), and  
443 mostly empirically demonstrated to occur in the case of strongly beneficial mutations (e.g.  
444 Edwards et al., 1990; Simberloff and Leppanen, 2019).

445 The various elements discussed above show that  $\gamma$  is an emerging property of the  
446 interaction between a variety of physiological mechanisms rather than a fixed quantity,  
447 which advocates for the development of theoretical models treating it as such rather than  
448 as a fixed parameter, by incorporating growth, mutation and selection at the cellular level.

## Acknowledgements

I would like to thank Diala Abu Awad, Sylvain Billiard, Ludovic Maisonneuve, Denis Roze and Roman Stetsenko for taking the time to discuss this work, and Sylvain Billiard and Ludovic Maisonneuve for their comments on the manuscript. This work was funded by the European Research Council (NOVEL project, grant #648321). I also thank the Région Hauts-de-France, and the Ministère de l'Enseignement Supérieur et de la Recherche (CPER Climibio), and the European Fund for Regional Economic Development for their financial support.

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