Advances in the synthesis of 1,2-dioxolanes and 1,2-dioxanes

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Recent Advances in the Chemistry of 1,2-Dioxolanes and 1,2-Dioxanes

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1 Introduction

1,2-Dioxolanes and 1,2-dioxanes are particular examples of endoperoxides, which can be found in many natural substances, but have also many perspectives in medicinal chemistry. Indeed, endoperoxides are very famous class of compounds to possess anti-malaric and more generally anti parasitic properties. 1,2,4-Trioxanes or 1,2,4-trioxolanes are for example the main pharmacophore of anti-malarial natural artemisinin and synthetic arterolane (Figure 1).

Figure 1. Structures of 1,2-dioxolane and 1,2-dioxane. Structures of artemisinin, arterolane and diacetone diperoxide, exhibiting structures of 1,2,4-trioxane, 1,2,4-trioxolane and 1,2,4,5-tetroxane.

However, 1,2-dioxolanes and 1,2-dioxanes exhibit also different biological properties, including anti-malarial activities, and by consequence these structures attracted several synthetic chemists for many years in order to develop efficacious methods to prepare these oxygenated heterocycles. Most chemists fear organic

peroxides owing to their reactive and explosive character, such as with diacetone diperoxide (Figure 1). However, 1,2-dioxolanes and 1,2-dioxanes are in general stable compounds under normal conditions due a lower proportion of oxygen atoms involved in peroxide bonds compared to other atoms and a stabilization of the peroxide bond within a favorable 5- or 6-membered ring. Thus, they can generally be easily purified, for example, by silica gel chromatography. Since structures and methods to obtain 1,2-dioxanes and 1,2-dioxolanes are somewhat similar, and given that some families of natural product contain both heterocycles, it seems legitimate to treat both in a single chapter.

Cyclic peroxides were reviewed more than 50 years ago in *Advances in Heterocyclic Chemistry*,¹ therefore, an updated version reviewing most recent works and methods is today required. More recent reviews^{2,3,4} and also book chapters^{5,6} depicted the subject in a more general manner. This chapter encompasses some work presented in these review articles/books, in order to give a general overview of the most successful methods for synthesizing 1,2-dioxolanes and 1,2-dioxanes, but also will focus on the most recent strategies published these 15 last years in the field. A presentation of the natural products containing these structures is a good starting point to understand the importance of developing new methods for the synthesis of 1,2-dioxanes and 1,2-dioxolanes. The existing techniques for preparing these particular heterocycles will be discussed later through the presentation of free radical chain reactions involving oxygen triplet, nucleophilic additions with hydroperoxides, ene reactions or [4+2] cycloadditions involving electrophilic oxygen singlet, [3+2] formal cycloaddition and functionalization of endoperoxyketals involving peroxy-carbenium ion species.

2 Natural products with 1,2-dioxolanes and 1,2-dioxanes scaffolds

Although organic peroxides have a reputation of being reactive and unstable species, it exists a large number of natural products containing 1,2-dioxolane and 1,2-dioxane rings. Their different structures are exhaustively reported in some reviews,^{7,8,9} so this part could be consider as a summary of all reported structures. The different natural substances are going to be presented following their origin and their structural family to which they belong, focusing more on original and important structures.

2.1 Marine metabolites

Marine resources are major sources of endoperoxides, and sponges are in particular the most important pool for this class of compounds. Some cytotoxic or antifungal activities are often reported for these compounds. Thus, primary biological task of these compounds is probably to play a role of chemical defender against certain predators.

2.1.1 1,2-Dioxolane carboxylates

1,2-Dioxolanes are in proportion a minor structure among the peroxy natural products, however sponges of *Plakortis* and *Plakinastrella* species are providing a large number of these scaffolds. Plakinic acids A,¹⁰ C, D,¹¹ E,¹² F,^{13,14} G, H,¹⁵ J,¹⁶ L,¹⁷ N¹⁸ exhibit a fatty acid structure bearing a dioxolane between position 3 and 5. Two methyl substituents are also present at these positions. Their absolute configuration is generally inconsistent and depends on the sponge species studied, leading to one or the other enantiomer. Epimers at C5, with *trans* configuration at position 3 and 5, are called epiplakinic acids, which make it possible to distinguish between *cis* and *trans* diastereomers. Only the isolation work of epiplakinic acids G and H was ambiguous since the authors misused the term "*epi*", driving to some confusion.¹⁵ Indeed, epiplakinic acid H should have been called plakinic acid G to follow the general rule. The absolute stereochemistry was generally determined either by total synthesis such as with plakinic acid A,¹⁹ Mosher ester analysis of the reduced 1,2-dioxolane ring²⁰ or by NMR analysis of the diol with some chiral lanthanum shifting agent,²¹ but recently, circular dichroism also proved to be a reliable method.¹⁶ However attention must be taken when using chiral lanthanum shifting agent, because epiplakinic acid F seemed to have been incorrectly assigned, as demonstrated by its total synthesis, where the optical sign matched with the opposite configuration.²² Indeed optical rotation is strongly driven by the stereochemistry at C3 and C5 in this products family, *cis* and *trans* stereoisomers giving an opposite sign and the fatty chain exerting a lesser influence on it (Figure 2).

Figure 2. Major known 1,2-dioxolanes metabolites isolated from marine sponges *Plakortis* or *Plakinastrella* sp.

The main difference between all plakinic acids relies on the side chain, which provides different chain lengths, unsaturations, and ramifications. Plakinic acids O and P show a more uncommon 1,2-dioxolane placed at position 5 and 7 with a conjugated acid.¹⁸ Some other related compounds were isolated such as epiplakinidoic acid with a carboxylic acid function at the end of the fatty chain,¹⁴ or plakortinic acid A, which exhibits a beautiful bicyclo[4.2.0]octene framework.²³ The structure of plakortisinic acid shows ethyl substitution on both 1,2-dioxolane ring and side chain, and a 4,6-location for the dioxolane, which is quite unusual compared to the plakinic acids series.²⁴ Absolute configuration was deduced from optical rotation value by comparison with a calculated one. Andavadoic acid belongs to plakinic acids since it was isolated from *Plakortis aff simplex* and would have deserved to be differently named.²⁵ Absolute configuration was not reported in the original article but a total synthesis stated later its absolute configuration (Figure 2).²⁶

A bio-guided isolation strategy was mostly at the origin of the discovery of these natural products. Therefore, some moderate antifungal (plakinic acid A, F, J, L) and moderate cytotoxic activity at the micromolar to sub-micromolar range (plakinic

acids C, D, E, F, G, H, epiplakinoic acid, plakortinic acid, plakortisinic acid, andavadoic acid) were observed. Anti-HIV, -parasitic, or -bacterial bioassays were performed for some compounds but their activity in these fields was relatively low (Figure 2).

2.1.2 1,2-Dioxane carboxylates

1,2-Dioxane ring is more representative of the major part of endoperoxides isolated from marine species. The amount of members of this class being quite large, they cannot be all presented in this book chapter. Nonetheless, the major compounds will be shown with some selected examples. Indeed, such as in the plakinic acids family, only small differences exist between some members of a same family.

Most of plakinic acids, isolated from *Plakortis* or *Plakinestrella* sp., exhibit a 1,2-dioxolane ring (paragraph 2.11), but some members show also a 1,2-dioxane entity. Early isolated plakinic acids B,¹⁰ and more recent plakinic acids I,¹⁶ K¹⁷ or M¹⁸ contain this scaffold. Similarly to plakinic acids containing a 1,2-dioxolane ring, these natural products exhibit interesting antifungal activities (Figure 3).

Plakortides, such as plakortisinic acid for 1,2-dioxolanes, were isolated from different *Plakortis* species and exhibit some ethyl substituents in place of classical methyl ones at positions 3 and 4 of the 1,2-dioxane ring as well as on the fatty chain. A selected sample of this large class of compounds are shown herein with plakortides K and Z. This latter one is the simplest compound of the family, therefore it is a good platform for further synthetic studies towards total synthesis of such compounds. This class of natural products exhibits generally some moderate to potent antifungal and/or cytotoxic activities, however an interesting antimalarial activity was also demonstrated for some members (Figure 3).²⁷

Plakortolides are a distinct class of 1,2-dioxane type natural products bearing a carboxylate function, but their originality relies on their bicyclic structure. They were also isolated from different sponge species of *Plakinistrella* or *Plakortis* genera. The stereochemistry of the stereogenic centers can be very different, depending the isolated products, such as plakortolide D¹² or U,²⁸ which generally lead to different alphabetical denominations. A rationalization of this stereochemistry was particularly studied, involving a questioning about the biosynthesis pathway, where a cyclization of a hydroperoxide is more likely than a [4+2] cycloaddition with ¹O₂, which would

lead to a more defined stereochemistry, because of the concerted process (see paragraph 3.4.2).²⁹ An interesting example of a plakortide-like product is plakortoperoxide C, which shows also a 1,2-dioxene scaffold on the side chain. This 1,2-dioxene comes certainly from a [4+2] cycloaddition between $^1\text{O}_2$ and a diene. These compounds exhibited also moderate to good cytotoxic activities, as well as antifungal properties. Some activity against *Toxoplasma gondii* parasite was also reported for some derivatives (Figure 3).³⁰

Stolonoxides are an interesting class of molecules bearing an adjacent tetrahydrofuran ring towards the 1,2-dioxolane ring. These compounds were isolated from tunicate *Stolonica socialis*, and not from a sponge, contrary to most of marine 1,2-dioxane compounds. Such as plakinic acids or plakortides, this class of chemicals contains a carboxylic function with a fatty chain. However, there is no methyl or ethyl substitution on the 1,2-dioxane ring as well as on the side chain. Differences between stolonoxides A to D depend on the stereochemistry on the THF ring and the configuration of the unsaturation at C19-C20.^{31,32} Stolonoxide E and F bear an unprecedented unsaturated cyclooctane ring.³³ Stolonoxides exhibits some potent cytotoxic activities below the micromolar range against some several cell lines (Figure 3).

Another important class of 1,2-dioxanes isolated from marine species is the norterpene cyclic peroxides. These compounds are generally isolated from different sponge species of *Prianos*, *Sigmosceptrella*, *Lacuntrulia*, *Diacarnus* or *Mycale* genera belonging to the demospongiae class. Although most of these compounds are closely related, they bear different names depending the species or the genus of the sponge from which they were isolated. All these molecules bear a propionic acid function adjacent to the 1,2-dioxolane ring at one side and generally a sesquiterpene chain at the other side. Major differences between all these natural products, beside the relative stereochemistry, which can also be different, is the nature of the fatty chain. The farnesyl moiety can stay unchanged, such as in sigmosceptrillin E³⁴ but can also give other types of cyclized structures: monocyclic (muquibilines A and B),^{35,36} bicyclic (trunculin A,³⁷ mycaperoxide A³⁸) or polycyclic structure by means of additional oxidative cyclizations (trunculin X).³⁹ These molecules were reported in general with moderated cytotoxic or antimicrobial activities, but have shown also some antiviral³⁸ and ichthyotoxic activities,⁴⁰ which can explain their role as defensive metabolites for the sponges (Figure 3).

A class of 1,2-dioxanes which can be considered apart, is the peroxyketals, where a methoxy substituent is generally exhibited. The oldest known marine endoperoxide is certainly chondrillin, isolated in 1976 from *Chondrilla sp.* sponge.⁴¹ Some years later were isolated xestins from *Xestospongia sp.*. Xestin B exhibits the same 1,2-dioxene core structure than chondrillin and identical relative configuration; only the side chain being different with a diene framework.⁴² Originally assigned absolute configuration for chondrillin was determined by circular dichroism, but revealed to be wrong, as demonstrated by total synthesis.⁴³ Therefore, chondrillin and xestin B finally share the same absolute configuration, which makes sense since their optical rotation sign is identical. Other 1,2-dioxanes belonging to the same class were also discovered. In one hand, peroxyacarnate A or D were isolated from *Acarnus sp.* sponges,^{44,45} exhibiting an enyne framework on the side chain. On the other hand, manadoperoxide E⁴⁶ and peroxyplakoric acid A1⁴⁷ were isolated from *Plakortis sp.* sponges. These two last substances exhibit a related side chain. Compared to peroxyacarnates, some methyl substituents are present on the two last compounds. Manadoperoxides were reported to show a significant and selective anti-trypanosomal activity (Figure 3).

Figure 3. Illustration of different types of 1,2-dioxane carboxylate marine metabolites, through selected examples.

2.1.3 Other 1,2-dioxane or 1,2-dioxolanes

Other endoperoxides can be found in various marine species, which bring sometimes very original molecules. Thus, sinugibberosides,⁴⁸ isolated from soft coral *Sinularia gibberosa*, provide an unprecedented 1,2-dioxanyl ketal along with an original bicyclo[7.2.0]undecane structure. From the same species was isolated peroxygibberol,⁴⁹ which contains a 1,2-dioxene unit. *Sinularia* genus is also producing sinularioperoxide A and its different congeners,⁵⁰ which exhibit an endoperoxide structure. Denticulatolide⁵¹ was isolated from soft coral *Lobophytum denticulatum* and exhibits an unusual 1,2-dioxane ring within a diterpenoid cembranolide-like macrocyclic structure. Ichthyotoxic activity was reported for this latter compound. Red soft coral *Sarcophyton glaucum* was providing sesquiterpenic dioxosarcoguaiacol,⁵² which possesses an original 1,2-dioxolanyl hemi-ketal framework. Peroxypolasol,⁵³ structure isolated from the Japanese sponge *Epipolasis* sp., is a diterpenic endoperoxide with an original Lingshuiperoxide⁵⁴ is also a sesquiterpene, which was isolated from the Hainan sponge *Dysidea septosa*. The 1,2-dioxene certainly comes from a [4+2] cycloaddition with $^1\text{O}_2$, as well as the butenolide, which would be obtained from the same cycloaddition pathway on a furan ring, followed by a Kornblum-DeLaMare rearrangement. Intricate tricyclic structures gracilioethers A-C were isolated from *Agelas gracilis*,⁵⁵ where gracilioethers A show a 1,2-dioxane ring instead of a cyclic ether. Other gracilioethers E-J were isolated from *Plakinastrella mamillaris*,⁵⁶ which does not belong to demospongiae class, but to the homosclerophorida class. Some significant anti-malarial and anti-leishmanial activities were reported for this family of molecules (Figure 4).

Figure 4. Structures of sinugibberoside B, sinularioperoxide A, peroxygibberol, dioxosarcoguiacol, lingshuiperoxide, peroxyolasol and gracilioether A.

2.2 Terrestrial metabolites

2.2.1 1,2-Dioxolanes and 1,2-dioxanes isolated from plants

Plants are also a major source of 1,2-dioxanes and 1,2-dioxolanes and most of these compounds are terpenoid-like products. Therefore, they can be classified according to their number of isoprene units.

Monoterpenes provide several examples of endoperoxides. In particular ascaridole was probably the first known and isolated natural endoperoxide.⁵⁷ This compound was originally isolated from epazote "*Chenopodium ambrosioides*", which is used traditionally in Mexican food to provide flavor, but also as infusion to improve digestion. The essential oil, which contains about 70% of ascaridole, is reported to be toxic, but is useful for its anthelmintic properties. Acetylsaturejol and isoacetylsaturejol were isolated from *Satureja gillesii*, a Chilean lamiaceae.⁵⁸ The plant is resistant to herbivorous predators and the metabolites that it is producing are found to have significant toxicity toward shrimps *Artemia salina*. 1,2-Dioxolane and 1,2-dioxane structures are both isomeric, and might come from an ene reaction of singlet oxygen (see also paragraph 3.4.1) with pulegone or isopulegone (Figure 5).

Sesquiterpenes are the most representative class of endoperoxides in plants. Bisabolene-like yingzhaosu A⁵⁹ and C⁶⁰ were isolated from *Artabotrys uncinatus*, a traditional chinese herbal medicine for the treatment of malaria, and these compounds were particularly studied. Absolute and relative stereochemistry of yingzhaosu C was notably determined by total synthesis.⁶¹ Arteincultone⁶² and spiroarteincultone⁶³ were isolated from aerial parts of *Artemisia maritima* and/or *Artemisia abrotanum*. They are exhibiting a skeleton related to davanone, a sesquiterpenic tetrahydrofuran. A decahydronaphtalene-like structure can be seen in tirucaladerone,⁶⁴ a caladenequinone derivative recently isolated from *Euphorbia tirucalli*, as well as in schisansphene A,⁶⁵ an eudesmene peroxy-derivative isolated from *Schisandra sphenanthera*. Eudesmanolide derivative tehranolide was isolated from *Artemisia diffusa*,⁶⁶ and exhibits an unprecedented cyclooctane along with an endoperoxy hemiketal. Rugosal A,⁶⁷ was isolated from leaves of *Rosa rugosa* and shows a carotane-like structure. This compound seems to act as an antifungal agent for the plant, which produces this compound when the leaves are damaged, in order

to have a protection against some fungal infections. With a similar structure, isohalpinone is an example of guaiane-like natural endoperoxide, isolated from *Alpinia japonica*.⁶⁸ Another example, now in the guianolide family, is tanaparthin- α -peroxide, isolated from *Achillea setacea*, which exhibits an endoperoxide inside a cyclopentene ring (Figure 5).⁶⁹

Diterpenes are also representatives of different 1,2-dioxanes. Texas broomweed *Amphiachyris amoena* afforded an original double spiro-1,2-dioxane with a labdane-type skeleton, called amoenolide K. It was then proved experimentally that amoenolide A provides this latter compound by a ene reaction with singlet oxygen followed by a further oxo-Michael addition on a butenolide.⁷⁰ African "Tonga croton" *Croton steenkampianus* Gerstner provided some years ago an unprecedented diterpenoic tetracyclic endoperoxide called steenkrotin B.⁷¹ However, a moderate anti-malarial activity was reported for this compound. In a recent total synthesis work trying to access to this natural product, the last cyclizing step, namely addition of hydroperoxide function to the ketone, was unsuccessful.⁷² Yet, the cyclization to the peroxyketal is supposed to be spontaneous, if favored, therefore assignment of the original structure of steenkrotin B can be questioning. Mulinic acid⁷³ is also a diterpenic compound isolated from *Mulinum crassifolium*, a shrub growing in the north of Chile. Its structure was mainly determined by X-ray analysis, which secures its attribution. Internal 1,2-dioxene ring comes obviously from a [4+2] cycloaddition with singlet oxygen (Figure 5).

Some peroxy containing triterpenes were also found. For example, pseudolarolide Q₂ was isolated from the seeds of *Pseudolarix kaempferi*, a chinese tree, and shows a complex 3(4),9(10)-disecocycloartane-type triterpene, occurring a 1,2-dioxolane ring.⁷⁴ Some taraxastane-type triterpenes are also known with an endoperoxide ring, such as 3 β -acetoxy-1 β ,11 α -epidioxy-12-ursene.⁷⁵ This natural product was isolated from the aerial root of *Ficus microcarpa*, a Taiwanese ornamental plant and its elucidation was performed by X-ray crystallography (Figure 5).

Besides terpenes, other class of molecules can be isolated from plants, but there are more scarce examples. Some oxidized fatty acids possessing a 1,2-dioxolane such as thermalic acid A,⁷⁶ could be found from the herbaceous plant *Ophioglossum thermal*, with potent antibacterial activity. Some alkaloids were found to possess endoperoxides such as catharoseumine, an isolated indolomonoterpene from

Catharanthus roseus.⁷⁷ Absolute configuration was determined by ECD and chemical degradation of the endoperoxide, in order to obtain a known natural product. This compound exhibited moderate antimalarial activity. Finally, some flavonoid glycosides, such as apigenosylide B, are showing a spiro-1,2-dioxane.⁷⁸ The main originality in its structure is the endoperoxide attached to the aromatic ring, which is quite unusual. This compound and other apigenosylides were found to display weak antibiotic and cytotoxic activities (Figure 5).

Figure 5. Illustration of different 1,2-dioxanes or dioxenes and 1,2-dioxolanes isolated from terrestrial plants.

2.2.2 1,2-Dioxolanes and 1,2-dioxanes isolated from other kingdoms.

Although plants are a major source of terrestrial endoperoxides, some of these type of compounds can be also isolated from fungus, bacteria or even animals.

Fungi are also a source of numerous endoperoxides, and in particular ergosterol peroxide,⁷⁹ originally isolated from *Aspergillus fumigatus*, is an ubiquitous endoperoxide, obtainable from many sources of mushrooms, but also from some plants, microorganisms and marine species. Some tricyclic sesquiterpenes can also be isolated from different species of fungi, with a structure close to talaperoxide D, isolated from *Talaromyces flavus*.⁸⁰ Some anti-angiogenic activity and moderate cytotoxic activity were reported for this class of compounds.^{80,81} An unprecedented spiroketal peroxide, chloropupukeanolide A, was isolated from *Pestalotia fici*, an endophytic fungus.⁸² This compound displays some significant anti-HIV and cytotoxic activities (Figure 6).

Mycangimycin is an unprecedented polyenic fatty acid bearing a 1,2-dioxolane ring. By some aspects, its structure is related to plakinic acids (paragraph 2.1.1), but the major difference is the absence of methyl substituents on the ring and the side chain. This natural product was isolated from *Streptomyces* strains, which are living in mutualism with an insect, the south pine beetle (*Dendronctonus frontalis*). Mycangimycin is produced to protect the insect against antagonist fungi, therefore this natural substance showed powerful antifungal activities, but also a strong antimalarial one (Figure 6).^{83, 84}

Animals can be also at the origin of the production of important endoperoxides such as prostaglandins H₂ or G₂. These molecules are primary products of radical oxidation of arachidonic acid, and provide from different series of reactions including a Kornblum-DeLaMare rearrangement, all other prostaglandins, but also thromboxanes and prostacyclins.⁸⁵ Prostaglandin H₂ is not only a biosynthetic intermediate, but also displays some specific biological roles (Figure 6).⁸⁶

Figure 6. Examples of different endoperoxide metabolites isolated from fungi (purple), bacteria (green) and animals (red).

3 Methods to synthesize 1,2-dioxolanes, 1,2-dioxanes and 1,2-dioxenes.

3.1 General considerations.

Although it exists many methods to make organic peroxides and endoperoxides, such as 1,2-dioxolanes, 1,2-dioxanes and 1,2-dioxenes, the reagents allowing to make such structures are limited. Indeed, the direct formation of an O-O bond is not really possible, since this bond is weak, alkoxy radicals cannot merge into peroxide bonds (the homolytic cleavage is mostly observed) and since electronegativity of oxygen is also high, unpolung at oxygen atom is nearly impossible; only one example was reported using elemental fluorine, through the formation of a hypofluoride intermediate.⁸⁷ Thus, the only available reagents are: oxygen, which in its stable triplet state promotes mainly radical reactions, while more reactive singlet oxygen is able to provide a source of electrophilic oxygen and promotes cycloaddition reactions, and finally hydrogen peroxide as well as some hydroperoxides derivatives, which can provide a nucleophilic source of organic peroxides. A fourth reagent, which can be used in some specific application is O₃, which procures hydroperoxyketals or acetals from olefins in presence of a protic solvent.

Therefore, the methods to synthesize 1,2-dioxolanes, 1,2-dioxanes and 1,2-dioxenes will be classified depending their method of preparation. Indeed to make an endoperoxide, the elaboration of two C-O bonds are in general needed to introduce the peroxide unit and build the ring. The two bonds can be established either in one step when the two C-O bonds are formed in a chain reaction (particularly with free

radical reactions) or in a concerted fashion (cycloadditions), or in two steps when first the construction of a hydroperoxide or a derivative thereof is first necessary followed by its cyclization (Figure 7).

Figure 7. Different strategies and pathways to build 1,2-dioxanes and 1,2-dioxolanes.

3.2 Radical peroxidations and peroxycyclization.

3.2.1 Radical addition of triplet oxygen and direct formation of an endoperoxide.

3.2.1.1 Cycloaddition of triplet oxygen onto diradical species.

The direct formation of endoperoxide is a possible process through the formation of a diradical species. Thermal decomposition of cyclic azo compounds was early explored in the 80's under oxygen atmosphere and afforded 1,2-dioxanes and/or dioxolanes.⁸⁸ An original approach was conducted some years later and consisted in the decomposition of a bicyclic azo compound under light irradiation with an argon laser at 351.1, 351.4 and 363.8 nm.⁸⁹ The goal was to make some analogs of prostaglandin H₂. The authors started by introducing hydrazide framework **3** by using a [4+2] cycloaddition between azo compound **1** and cyclopentadiene **2**. Further functionalization of the olefin and introduction of a side chain, followed by the elimination of the imide moiety afforded azo compound **4**. At this stage, application of the laser argon in Freon at -16 °C under oxygen afforded expected diastereomers **6a** (*endo*) and **6b** (*exo*) in modest yields after

chromatography at -16°C . Diradical species **5** was reported to be the key intermediate (Scheme 1).

Scheme 1

Latent diradicals are available from some specific dienic systems. An interesting example is illustrated from flash vacuum pyrolysis of furan **7**, which led to the formation of non-aromatic compound **8**. A reorganization of electrons probably took place with recovery of aromaticity as a driving force and formation of diradical species **9**. It is then involved in the formation of 1,2-dioxane **10** with triplet oxygen at low temperature. That is interesting to note this transformation led to the same compounds than those obtained using a [4+2] cycloaddition between a diene and $^1\text{O}_2$ (see paragraph 3.4.2), however it appears that the two transformations are completely different mechanistically speaking (Scheme 2).⁹⁰

Scheme 2

3.2.1.2 Formal cycloaddition of triplet oxygen onto radical cations

Another method to prepare some endoperoxides is the triplet oxygen peroxidation of cation radicals. Original works on this process were using an appropriate one-electron oxidizing reagent, such as dicyanoanthracene with photoactivation. This process was only reserved to electron-rich cyclopropanes, or dienes for example. Otsuji and Mizuno's pioneer work demonstrated that an oxidation through a single electron transfer takes place onto diarylcyclopropane such as product **11** in order to lead to 1,2-dioxolanes.⁹¹ Presumably oxidation occurs on the heteroatom (**12**) and lead by conjugation to the opening of the strain cyclopropyl ring

(**13**). Stabilized radical **13** is then reacting with triplet oxygen (**14**) and further cyclization of the peroxyradical drives to rearomatization to radical cation **15**. Doubtless, an electron transfer from starting molecule **11** takes place in order to propagate the reaction and furnish 1,2-dioxolane **16**. Addition of $\text{Mg}(\text{ClO}_4)_2$ was reported to dissociate dicyanoanthracene radical anion from the radical cation leading to an overall acceleration of the process (Scheme 3).⁹¹

Scheme 3

Research of a more applicable process showed that TiO_2 could be used in place of dicyanoanthracene.⁹² More recent work of Yoon and coworkers showed also that $\text{Ru}(\text{bpz})_3^{2+}$ (**19**) is a more effective catalyst than dicyanoanthracene, mainly by performing the ring expansion with simple visible light apparatus at low temperature and with a wider scope of substrates (Scheme 4).⁹³

Scheme 4

This process was also applicable to the synthesis of 1,2-dioxanes. A pioneer example was reported by the same authors on electron-rich conjugated dienes,

spaced by 3 or 4 atoms. On substrate **20**, photooxidation with dicyanoanthracene leads to the formation of cationic radical **21**, which can cyclize on the other double bond to drive to species **22**. Further reaction with $^3\text{O}_2$ drives to the formation of peroxy radical **23** and cyclization to endoperoxide **24**. Electron transfer from **24** to **20** propagates the reaction furnishing 1,2-dioxane **25**. This transformation can be viewed as a formal [2+2+2] cycloaddition. The presence of two aryl substituents for each olefin appears to be crucial, but limits the scope of the reaction (Scheme 5).⁹⁴

Scheme 5

Fortunately, almost 20 years later was reported an updated protocol by Yoon and coworkers.⁹⁵ They described a similar process on monosubstituted olefins. Particularly, they showed that tris(bipyrazyl)ruthenium(II) complex **19** proved to be a very suitable photosensitizer for such a transformation because of the far superior excited state lifetime. Some years later, Nicewicz and coworkers presented a similar work based on photoexcitation with pyrilium species **26**.⁹⁶ They demonstrated that the presence of an electron-donating group was crucial through substrate screening, and they applied the reaction mainly to 1,1-disubstituted olefins. An example leading to tricyclic structure **30** was particularly interesting. More recently was reported a study using TiO_2 nanoparticles as photosensitizer by Wang and coworkers (Scheme 5).⁹⁷ Although Yoon and coworkers obtained under some conditions some cyclobutanes such as compound **31** as byproducts, it appears that this strained cycle is probably a reversible intermediate formed when oxygen is not reacting, but can nevertheless lead to 1,2-dioxane **32** when placed in photooxygenation conditions. Although previous works mentioned that the photosensitizers used do not produce

any superoxide ion, Wang and coworkers' study showed some evidences of the role of superoxide anion in the formation of dioxanes with TiO_2 as photocatalyst (Scheme 6).

Scheme 6

Using a similar mechanism, some interesting work was developed on cyclopropylamines. Wimalasena and coworkers found that cyclopropylaniline derivatives can react in a single electron oxidation.⁹⁸ Therefore cyclopropylamine **33** could react in presence of air with different reagent to obtain 1,2-dioxolanes **36**. The mechanism is similar to the one depicted in scheme 3, through oxidation and formation of radical cation **34** on the nitrogen, followed by the ring-opening of the cyclopropyl ring by the radical. Further reaction with oxygen lead to 1,2-dioxolane **36**. Because of the instability of these endoperoxides, no yield could be given, even if some NMR spectroscopic evidences were obtained. The instability is probably due to the nitrogen, which possess some basic and nucleophilic properties driving to some degradation such as a Kornblum-DeLaMare rearrangement. Independently, Six and coworkers noticed the formation of an endoperoxide from the reaction mixture of the intramolecular Kulinkovich-DeMeijere reaction of amide **37**.⁹⁹ Instead of isolating expected cyclopropylamine **38**, dioxolane **39** was obtained after purification, due to a spontaneous oxidation with air. Presumably the electron donating aromatic group on nitrogen facilitated this process. Encouraged by this interesting result, they found that anodic oxidation of cyclopropylamines under oxygen drove to 1,2-dioxolanes with a better variety of substrates such as compound **40** converted into endoperoxide **41**.^{100,101} A vinylogous process was also explored from some piperidine derivatives

and proved to be effective.¹⁰² As described by Wimallesena and coworkers, the main difficulty in this study was the instability of the amino-dioxolanes. Only few examples were found to be stable and isolable. Substitution increases stability giving a favorable Thorpe-Ingold effect (Scheme 7).

Scheme 7

A very recent example of Zhou and Lu seemed to use a similar mechanical pathway for the aerobic oxidation of spirovinyl-cyclopropylindoles mediated by some iodine.¹⁰³ Thus, compound **44** was, for example, converted into 1,2-dioxolane **45** in 91% yield with air and 1 mol% of I₂. Some control experiments showed a radical mechanism for this transformation, since photons seem necessary to activate the reaction and the addition of radical inhibitors stops the process. Most of the studied substrates, leading to effective transformations, showed only differences on the substituents on the aromatic ring, which unfortunately limits the scope of this reaction. A plausible mechanism could be firstly the reaction of the nucleophilic amide function in **46** onto iodine to make an I-O bond. Intermediate **46** undergoes then a homolytic cleavage, probably by activation with light irradiation, giving radical cation **47a** in mesomerism with **47b**. The free radical opens then the strained ring by a conjugated process, and radical species **48** reacts then with oxygen to obtain peroxy radical **49**, which can cyclize into 1,2-dioxolane **50**. Electron-transfer from **50** to starting material **44** can then propagate the reaction (Scheme 8).

Scheme 8

A very recent example of cycloaddition of oxygen triplet on annellated 1-aryl-1-carboxyl-cyclopropane derivatives **51a-k**, mediated by visible light in presence of acridine derivative such as [MesAcr] ClO_4 as sensitizer, was also described by Reiser and coworkers to give fused 1,2-dioxolanes.¹⁰⁴ Diverse structure were obtained, mainly some THF derivatives such as **52a-e** in modest yields; pyrrolidine or piperidine derivative **52f** and **52g**; and cyclopentenyl derivatives **52h-i**. The reaction was in contrast ineffective when no aromatic ring was present on the cyclopropyl group and also to prepare tetrahydropyran derivative **52k**. Limitations were also observed when cyclohexenyl rings were used in place of cyclopentenyl rings. (Scheme 9)

Scheme 9

3.2.1.3 Formal cycloaddition of triplet oxygen to cycloalkoxy radicals

Although the spontaneous reaction of 1,2,2-trimethylcyclopropanol with oxygen into 1,2-dioxolane was reported early in 1969,¹⁰⁵ a more general protocol was developed much later in 2001 by Kulinkovich and coworkers, where they showed Manganese (II) abietate (a diterpene carboxylate) in benzene is able to initiate the ring expansion of cyclopropanols **53**.¹⁰⁶ Presumably a superoxomanganese(III) species is involved in the reaction and can abstract an hydrogen from cyclopropanols **53** to form alkoxy radicals **54**, which can freely evolve into an alkyl radical species **55**. This latter can then trap a molecule of oxygen, and a cyclization followed by a propagation process (the two steps probably can work one way or the other) lead peroxy radicals species **56** to the formation of 3-hydroxy-1,2-dioxolanes **58**. These compounds were described to be in equilibrium between the open hydroperoxy-ketone form and endoperoxyde form, leading to thermodynamic mixture of epimers at peroxyacetal function. The usefulness of this heterocycle was neglected at this time (see paragraph 3.5.2), and its transformation into epoxyketones **59** was mainly achieved without characterization of 1,2-dioxolane intermediate **58**. (Scheme 10)

Similarly, a more recent work showed that tertiary cyclopropanols **60** could react in almost quantitative yield in THF with manganese(III) acetylacetonate to

obtain 5,5'-unsubstituted 1,2-dioxolanes **61** in order to prepare chiral epoxyketones **62** with poly-*L*-leucine matrix.¹⁰⁷ (Scheme 10)

Scheme 10

In another example, cyclopropanol **63** could also be readily opened under radical conditions with cobalt(II) acetylacetonate, the mechanism of which is similar to that with manganese. The reaction was originally reported in ethanol with a higher metal charge (5 mol%). Such as previous examples, 1,2-dioxolan-3-ol **64** was not targeted but used as a reactive intermediate for further transformation. Indeed, authors transformed endoperoxide **64** into conjugated ketone **65** by reduction of the peroxide bond with PPh₃ followed by elimination of the hydroxyl group after mesylation. Fourteen examples on poly-functionalized structures were reported with yields given between 69 and 92% over a 3 step sequence.¹⁰⁸ (Scheme 10)

A last example reported by us was the utilization of this radical reaction in the synthesis of saturated analogue of mycangimycin **69** (see Figure 6, paragraph 2.2.2).

The ring expansion was carried out from cyclopropanol **66** with $\text{Co}(\text{acac})_2$ in ethanol and led to 3,5-disubstituted dioxolane **68** after reduction of the hydroxyl group with triethylsilane (see also paragraph 3.5.2).¹⁰⁹ (Scheme 10)

The driving force of these previous transformations is unambiguously the energy strain contained in the cyclopropyl ring, which lead to the formation of an alkyl radical where a peroxidation can take place. Considering cyclobutanes are also strained rings, we recently showed that the radical ring opening of cyclobutanols **70a-i** might drive to the formation of 1,2-dioxan-3-ols **71a-i** through a formal ring expansion process.¹¹⁰ The process is much slower than with cyclopropanols (Scheme 7) and required higher cobalt salt load and higher reaction time to proceed. The reaction is working particularly well with secondary alcohol such as with cyclobutanols **70a-b**, **70e-f**, or **70h**, and is more sluggish when it is substituted by an alkyl group such as with **70b-c**, or **70g**. In contrast the substitution at position 2 improves the stabilization of the reactive radical species, which increased the kinetic of the reaction and led to good yields even at room temperature such as with **70a-b**, **70g**, **70i**. (Scheme 11)

Scheme 11

Although larger ring sizes, such as 5 or 6 member rings, are in general not appropriate to undergo such radical cleavage, some specific examples were reported in the literature about rearrangement of some steroid derivatives in the manner of a Wharton-like fragmentation, but under radical conditions. Thus, Suarez and coworkers discovered a radical fragmentation of steroids containing a cyclopropane ring, mediated by $\text{PhI}(\text{OAc})_2$, I_2 and some light. Following authors' description, decalin ring-opening of compound **72** into dioxolane **73** was generated from the formation of alkoxyradical **74**. Fragmentation of the decalin structure followed by

cyclopropyl ring-opening led to 1,2-dioxolane **77** after reaction with one molecule of oxygen. The remaining alkyl radical reacted with an additional molecule of oxygen, and underwent several transformations, whom a hydrogen atom transfer, to form THF ring present in product **73**. However the authors are evasive concerning this last transformation step, therefore the obtaining of THF ring was not completely understood (Scheme 12).¹¹¹

Scheme 12

3.2.1.4 Formal [2+2+2] cycloaddition of triplet oxygen to enoxy radicals and olefins

Besides the ring opening of cycloalcanols through an alkoxy radical, which can lead to endoperoxides, enols were particularly well studied to promote some formal [2+2+2] cycloadditions with triplet oxygen and a good radical acceptor such as some styrene derivatives.¹¹² Thus in a seminal work, cyclohexanone **78** was reported to react with styrene **79**, oxygen and a combinaison of $\text{Co}(\text{II})(\text{OAc})_2$ & $\text{Mn}(\text{II})(\text{OAc})_2$ at 80°C to give specifically 1,2-dioxan-3-ol **80** (Scheme 13).

The reaction was then more generalized on 1,3 dicarbonyl systems with at least a ketone as function and $\text{Mn}(\text{II})(\text{OAc})_3$ as a radical initiator. Piperidone **81**, for example, gave rise to endoperoxide **83**,¹¹³ whereas piperidinedione **84** afforded dioxanol **86**.¹¹⁴ When a third carbonyl group is used as substituent, the reaction could afford tricyclic structures after a further cyclization. Therefore, spontaneous cyclization and dehydration of endoperoxyketal **88** gave compound **89** when a ketone was involved as a substituent, while an ester such as in structure **92** led to tricyclic compound **93** containing a lactone.^{115,116} The mechanism of the reaction was reported with different hypotheses, particularly in the first mechanistic step of the reaction. It was observed that the limiting step is the formation of enol species **95**. By consequence the easier enol **95** is formed, the faster the reaction is, meaning also

enol species **95** is the reactive species compared to diketone **94**. This also explain the importance of having a 1,3-dicarbonylated substrate to enhance the presence of enol species. Manganese(III) is then inserted on the hydroxyl group and undergoes an hemolytic cleavage to give alkoxy radical **97** at one side and a manganese(II) species at the other side. The formation of a radical cation was also proposed by some authors at this stage.¹¹⁷ The free radical shifts to the more stabilized alkyl position such as with species **98** and then reacts first with radical acceptor **99** (in general a styrene derivative), to give rise to alkyl radical **100**. In a second time, addition of triplet oxygen furnished 1,2-dioxane **102** after H abstraction (Scheme 13).

Scheme 13

This reaction was also applied to some heterocycles exhibiting a 1,3-dicarbonyl framework. Especially, quinolinols are preferentially existing as an enol due to a conjugation with the aromatic ring. Thus, compound **103** reacted with

diphenylethylene **82** to furnish a mixture of dioxane **104** and open-ring **105** (Scheme 11).^{116, 118} This ratio is dependent of the alkyl group involved at position 3. Having no substituent at this position, such as with compound **106**, gave rise to double addition of the olefin. The transformation drove to the formation of compounds **108** and **109**, depending the carbonyl group involved in the tricyclic structure (Scheme 11).¹¹⁹ Other heterocycles such as hydroxybenzoquinone **110** led to similar reaction product **111** (Scheme 11).¹²⁰ Tetronic acid **112**, because of its prominent enol form, was also a very convenient substrate for such a transformation and 1,2-dioxanol **113** was isolated with an excellent yield (Scheme 14).¹²¹

Scheme 14

Intramolecular reaction was also investigated by Gademann and coworkers.¹²² They noticed that compound **114**, an intermediate in the total synthesis of striatin A, was transformed into peroxide **115** by exposure to air and light. Manganese mediated oxidation furnished the same compound in higher yield. This compound exhibited a potent antimalarial activity (Scheme 15).

This formal [2+2+2] cycloaddition was also investigated with other metals on substrates such as compound **116** and cerium(III) chloride in presence of oxygen proved to give similar results compared to manganese (III) acetate (Scheme 15).^{123,124}

Scheme 15

This transformation was also investigated with acetylene derivatives in place of styrene.¹²⁵ Therefore, after some optimizations by using notably a large excess of manganese(III) acetate, Nishino and coworkers found that acetylacetonate **118** and phenyl acetylide **119** drove to the formation of 1,2-dioxolanol **120** with modest yield. Similarly to reaction with styrene derivatives, free radical species from acetylacetonate **118** is probably adding on the triple bond, affording vinyl radical species **121**. Oxygen then reacts with it and the corresponding peroxy radical species **122** is presumably reduced *in situ* by manganese(II) into alkoxy radical **123**, which evolved favorably to alkyl radical **124** by resonance. In turn, this later reacts with another molecule of oxygen to give peroxy radical species **125**, followed by the formation of compound **120** after H abstraction (Scheme 16).

Scheme 16

3.2.1.5 Isayama–Mukaiyama cobalt catalyzed hydroperoxy-silylation

The cobalt catalyzed hydroperoxysilylation was originally developed by Isayama and Mukaiyama,^{126,127} and is probably the most popular method to prepare

hydroperoxides from complex molecules since the reaction is highly chemoselective and regioselective. Indeed, the reaction usually takes place on the most electron rich olefin with formation of a radical species on the most substituted position of the reacting double bond. The mechanism was more recently better studied by Nojima,¹²⁸ where he figured out that cobalt superoxide species, formed from oxygen and Co(II) salts are initiating the reaction with the help of Et₃SiH to form cobalt(III) hydride species. As depicted in Scheme 16, a hydrometallation process takes then place during the hydroperoxysilylation of compound **126**, and surprisingly the metal is reacting predominantly on the most substituted position (Markovnikov's rule). Cobalt(III)-carbon bond in **129** is then easily dissociating, generating alkyl radical species **130** on one side and a cobalt(II) salt on the other side. The regiochemistry appears to be guided by stabilization of this radical species rather than steric hindrance. Alkyl radical species **130** can react then with triplet oxygen to form hydroperoxy radical **131**, which can be reduced with Co(II) species in order to make new superoxo-cobalt species **132**, which can propagate again the process by reaction with Et₃SiH, in order to form a cobalt(III) hydride species (Scheme 17).

Scheme 17

Although the reaction usually does not provide an endoperoxide directly, since the hydroperoxide needs a further cyclization by different methods which would be discussed further (see paragraph 3.2.2.1, 3.3, and 3.5.1), Isayama–Mukaiyama cobalt catalyzed hydroperoxysilylation can provide directly a 1,2-dioxane or a 1,2-dioxolane depending the circumstances and the structure of the substrate. Indeed, some 1,4 or

1,5-diene can lead directly to the formation of these endoperoxides through a radical chain reaction. A first example was described by Nojima and coworkers for the synthesis of analogs of Yingzhaosu A.¹²⁹ A first hydroperoxidation from limonene with some cobalt hydride complex and oxygen was followed by an internal addition of hydroperoxy radical **133** onto the double bond at γ -position, which led to 1,2-dioxane **134**. Further trapping of molecular oxygen, followed by silylation (and in a second step its desilylation with acidic MeOH), drove to dioxane **135**. The transformation was however modest in term of yield and led also to the formation of expected by-products **136** and **137**, coming from individual hydroperoxidation processes. Also, the reaction was applied to other limonene derivatives such as enoate **138**, which gave a similar result. The major comment here is about the *in situ* reduction of the hydroperoxyl function in hydroxyl group with this substrate (Scheme 18).

Scheme 18

The reaction was later more studied by Nojima on acyclic systems.^{130,131} Some specific examples with 1,4-dienes such as with substrate **140** drove to expected 1,2-dioxane ring **141** with 64% yield, while some 1,3-diene systems such as compound **142** could afford 1,2-dioxolane structure **143** under similar reaction conditions. (Scheme 19).

Scheme 19

This reaction was more recently applied as a key step in the total synthesis of many endoperoxides of chamigrane family.¹³² Thus, spirocompound **144** was transformed in a mixture of diastereomers **145a** and **145b** by using the Isayama–Mukaiyama cobalt catalyzed hydroperoxysilylation. The mixture of diastereomers was treated with acidic methanol to cleave the silyl ether, followed by reduction of the hydroperoxide with dimethylsulfide to give steperoxide C (if considering appropriated diastereomer **145b**). Further oxidation of the alcohols in ketone drove to talaperoxide C. Other natural endoperoxides of the chamigrane family have been obtained from these compounds (Scheme 20).

Scheme 20

This radical chain process was also used in a recent work towards the total synthesis of nortribolide. Contrary to the last example (see Scheme 20), the formation of the endoperoxide is just a fulcrum for the synthesis of a specific triol system *via* the peroxide bond reduction with zinc.¹³³ Thus, intermediate **146** was treated under Isayama–Mukaiyama reaction conditions and led to a mixture of compounds **147** and **148** with 10 and 16% yields, respectively, after further optimizations. The yield was not very high but allow to access to a highly functionalized structure with a high chemoselectivity and stereoselectivity. It is also interesting to note that no peroxysilyl ether was isolated, but directly the alcohol function. Such as with compound **138** (Scheme 18), the electron withdrawing group

seems to have an effect on the reducibility of the peroxyradical intermediate. Then, because of the low yield, the authors wanted to test their procedure on other simpler systems such as with eudesmane sesquiterpene **150** in order to give triol **152** through reduction of **151**. Due to the presence of only one remaining olefin in the system, the overall yield after reduction was more satisfying (50% *vs* 16%) (Scheme 21).

We saw that 1,4 and 1,3-dienes could be appropriate substrates to make endoperoxides in the Isayama–Mukaiyama cobalt catalyzed hydroperoxy-silylation, but strained cycles could be also a good alternative to prepare such compounds (see also paragraphs 3.2.1.3 and 3.2.1.5), although examples are more limited. For example, Nojima showed that vinyl cyclopropane **153** could be opened through the formation of radical species **154** at the adjacent position of the strain ring, followed by formation of **155**, which led to trapping of oxygen and ring closing onto the olefin. Further reaction of radical **156** with oxygen followed by silylation drove to 1,2-dioxolane **158** (Scheme 22).¹²⁸

Similarly, Enders used this process as a keystone in the final stage of the total synthesis of gracilioether A. Notably, the ring-opening of the cyclobutane contained in **159** led to the formation of a 1,2-dioxane ring corresponding to gracilioether A, with 39% yield and its epimer with 16% yield. Authors obtained directly the alcohols derivatives rather than the usual triethylsilylperoxy ethers such as with compounds

138 and **146** (Scheme 18 and 21), but unfortunately this phenomenon is not clearly described by the authors (Scheme 23).¹³⁴

Scheme 23

3.2.1.6 Thiyl and selenyl radical-mediated domino reactions

Addition of radical thiyls to olefins under oxygen atmosphere is an old reaction discovered by Kharasch and coworkers in 1950 and called TOCO reaction (Thio-Olefin Co-Oxidation).¹³⁵ This reaction consists, such as in the example depicted in Scheme 23, in the transformation of *n*-propylthiol, styrene and oxygen into sulfoxy-alcohol **161**. Mechanistically, thiyl radical **164** (formed by oxygen or any other radical initiator) is adding onto styrene, giving free radical species **165**, and this step is followed by addition of triplet oxygen and internal oxidation of the sulfide by the peroxy species to give **167** and then **163**. (Scheme 24)

Scheme 24

The application of this reaction many years later by Beckwith and coworkers to dienes¹³⁶ and trienes^{137,138} allowed the preparations of 1,2-dioxolanes when two double bonds were spaced by one methylene segment. Thus, authors could transformed triene **168** into 1,2-dioxolane **169** through the addition of a thiyl radical

to terminal olefin and oxygen trapping of allyl radical **170**, which can then cyclize in turn onto the last olefin in order to perform a free radical chain reaction with another molecule of oxygen. In contrast to the example of Kharasch with a substrate containing only one olefin (Scheme 24), the reaction here does not produce sulfoxide through the reduction of the peroxyradical, keeping the sulfide function. Since the hydroperoxides are more reactive, its selective reduction with PPh_3 guided **169** to alcohol **172** with 49% yield over 2 steps. It is also noteworthy that the reaction was surprisingly diastereoselective since only one isomer was observed (Scheme 25).

Scheme 25

About twenty years later, Bachi and coworkers became interested in the total synthesis of Yingzhaosu A⁵⁹ and several of its synthetic analogues. They found that the application of TOCO reaction on limonene and limonene derivatives would be an elegant pathway to this antimalarial substance, which contains a 1,2-dioxane ring into a bicyclic structure.¹³⁹ Compared to Beckwith and coworkers (Scheme 25), they finally optimized the conditions using a radical initiator such as AIBN or dibenzoyl peroxide in conjunction with light and acetonitrile as solvent.^{140,142} Thus, limonene was for example transformed into a mixture of endoperoxides **174a** and **175b**, corresponding to the formation of four new bonds in only one step! The sulfides were oxidized into sulfoxides **176a** and **177a** and then transformed into aldehyde through a Pummerer rearrangement. This sequence allowed the access to Yingzhaosu A after Mukaiyama aldol reaction and further transformations, but also to many other analogues of this molecule such as **181** or **182**.^{143,144} Moreover, the synthesis of other analogues from limonene derivatives **173b-e** drove to the preparation of many other products such as **178** or compounds **179** or **180** through some Wittig olefination. These two last compounds showed a remarkable anti-malarial activity.¹⁴⁴ (Scheme 26)

Scheme 26

In parallel to Beckwith's work with dienes or trienes (*vide supra*), Feldman and coworkers developed in 1986 a convenient approach for the formation of 1,2-dioxolanes **184** from 1-vinylcyclopropanes **183** bearing phenyl, vinyl or ester substituents at C-2 position of the cyclopropyl ring or alkyl groups at C-1 of the vinyl moiety. Like many radical reactions involving a cyclopropane, this method is based on the β -elimination of cyclopropylmethyl radicals, followed by addition of oxygen triplet.^{145,146} In contrast to the classical TOCO reaction, the newly formed thiyl or selenyl substituent undergoes a final elimination. Indeed, the addition of phenylselenide (or thiyl) radical to alkene **183** drives to cyclopropylmethyl radical **185**, which rearranges into homoallyl radical **186** through a β -fragmentation. This alkyl radical is then trapped by O_2 , providing peroxy radical **187**, which after cyclization and β -elimination of phenylselenyl radical, affords desired dioxolane **184**. Thus, numerous examples of this reaction have been reported giving yields ranging between 41% and 88%. The application of this process to dicyclopropyl compound **189** afforded two contiguous dioxolanes in a mixture of diastereomers **190a** and **190b**, while tricyclopropyl substrate **191** drove to product **192** in 38% yield, with formation of three consecutive 1,2-dioxolane rings, i.e. six new carbon-oxygen bonds in one step! Sulfur showed to be especially effective in this last transformation compared to selenium, which seems to be more suited and selective towards the construction of a single dioxolane ring (Scheme 27).

Scheme 27

Feldman and coworkers were also interested to show that 1,2-dioxolanes could lead to the formation of 1,3-diol. Therefore, they also applied their methodology to the total synthesis of (+/-)-yashabushitriol from vinyl cyclopropane **193**. The thiyl radical gave a better *E* selectivity for the formation of compound **194**, compared to the selenide radical, although this latter gives a slightly higher yield for the transformation into 1,2-dioxolane. The reductive opening of **194** into diol derivative **195** drove to the formation of yashabushitriol after 3 more steps (Scheme 28).¹⁴⁷

Scheme 28

The Feldman's protocol was also applied to the total synthesis of natural products some years later by Wong and coworkers. In a first work, they accomplished the total synthesis of Plakortide E, by applying Feldman's protocol to lactone **197**.¹⁴⁸ Because of the cyclic nature of **197**, only one diastereomer could be obtained. Further reduction of the lactone and the olefin with diimide gave *meso*-diol **199**. A mono protection with a silyl group gave alcohol **200** as a racemate. A kinetic resolution with vinyl acetate and Lipase PS from *Bukhoderia cepaci* in hexane gave unreacted alcohol **(+)-201** or acetate **(-)-202** with >99%ee for both depending the reaction time, respectively 29 h or 3h. Further functionalization of **(+)-201** or **(-)-202** over 11 steps afforded the different enantiomers and diastereomers of Plakortide E. A NMR spectroscopy comparison, as well as a study of the optical rotation sign of the synthetic samples and the natural product confirmed absolute stereochemistry of the 3 stereogenic centers of plakortide E (Scheme 29).

Wong and coworkers were also interested in other natural endoperoxides, such as *epi*-plakinic acid F. For this natural substance, authors were interested to use a diastereoselective Feldman's reaction using a chiral oxazolidinone auxiliary.²² After a screening of different conditions, solvent and chiral auxiliaries, the oxazolidinone derived from valine appeared to be a better chiral inductor, and the use of Sc(OTf)₃, among all other Lewis acids tested as additives, gave the best result in term of diastereoselectivity. Indeed, trans-isomer **205b** was isolated in 72% yield from a mixture of three isomers (90% total yield containing 80% of isomer **205b**). Scandium triflate probably forces the two carbonyl groups to be oriented on the same side due to chelation, such as on intermediate **206**, improving the rigidity of the chelate and by consequence the stereoselectivity of the alkyl radical addition to the

oxygen at the top face of intermediate **206** (80% selectivity). Addition of peroxy radical to the olefin depends more on the conformation of the olefin, but it seems that it adopts the one described in **206** since it drives to the major observed stereochemistry (89%). After 12 steps from **205b** including a carbonyl group homologation and some palladium catalyzed cross coupling reactions to build the fatty chain, the authors could access an enantiopure synthetic sample of *epi*-plakinic acid F (Scheme 30).

Scheme 30

3.2.1.7 Organozinc oxidation through an electrophilic activation of a cyclopropane

Organometallics are known to react with oxygen to produce hydroperoxides or more generally hydroxyl compounds, after an *in situ* reduction. In particular, organoborons¹⁴⁹ and organozinc¹⁵⁰ compounds were reported to produce more chemoselectively hydroperoxides. Although the mechanism of this oxidation is not completely clear, the transformation seems to be a free radical reaction. Recently, López and Vicente reported an original transformation to obtain some furanyl-dioxolanes.¹⁵¹ Indeed, from acetylacetonate **206**, ynal **207**, and olefin **82** in presence of ZnCl_2 (2 mol%) in solvent free conditions, they could obtain acetylfuran **208**, substituted with a cyclopropyl ring. This multi-component reaction works first through the condensation of **206** onto **207** to obtain intermediate **209**. The carbonyl group is then cyclizing onto the alkyne giving **210**, via an activation of the zinc catalyst. Organozinc species is probably giving back a negative charge by conjugation to the oxocarbenium species, driving to furanyl structure **211** containing a zinc carbenoid (proposed intermediate). Taking advantage of this carbenoid, a cyclopropanation takes place with an olefin such as **82** to afford cyclopropane **208**.

Compound **208** could further react with a larger amount of ZnCl_2 under oxygen atmosphere to produce 1,2-dioxolane **212**. Reasonably, ZnCl_2 can activate the cyclopropyl ring, which exhibits a π -character and the oxygen on the furan ring pushes the electron by conjugation leading to a cyclopropyl ring opening, which leads to organozinc species **213**. This step is followed by insertion of oxygen in order to form probably species **214**. A conjugated addition of zinc peroxide affords compounds **212**. The reaction was applied to 6 examples in order to make endoperoxy structures with yields between 62 and 89%, however the method is only limited to diphenylsubstituted cyclopropyl rings for the oxidation step. It is noteworthy that the reaction could be also performed from compounds **206**, **207** and **208** directly in one step if the catalytic charge of ZnCl_2 is increased to 50 mol% directly and if the reaction is performed under oxygen atmosphere (Scheme 31).

3.2.2 Radical Peroxycyclization of Hydroperoxides

3.2.2.1 Synthesis of hydroperoxides: a summary.

We saw previously how possible is the formation of endoperoxides through a free radical chain reaction involving oxygen and allowing the formation of two new carbon-oxygen bonds in a single transformation. Another strategy to prepare 1,2-dioxolanes or 1,2-dioxanes would be to start from a molecule containing a hydroperoxide function to cyclize it. The question would be then how to prepare these hydroperoxides intermediates? Since this book chapter is more focused on the formation of endoperoxides rather than hydroperoxides, and since their reaction

classification would be impossible if we consider both formation of hydroperoxides and their further cyclization (because any combination could be performed), it is preferable to not focus on the description of the synthesis of hydroperoxides.

Nevertheless, it is a mandatory to expose concisely the different methods and strategies, which can be applied for the synthesis of hydroperoxides, in order to give to the reader an overview, although some reviews describe their preparation.^{152,153} Therefore, synthesis of hydroperoxides, as well as synthesis of endoperoxides (*vide supra* Figure 7), can be classified in three main routes:

- Free radical reactions, which involve triplet oxygen, such as the Isayama–Mukaiyama cobalt catalyzed hydroperoxy-silylation of olefin;^{126,127} however oxidations of some organometallics such as boranes¹⁴⁹ or organozinc reagents¹⁵⁰ have also to be considered.

- Nucleophilic additions, which involves hydrogen peroxide and some of its derivatives; any electrophile is generally suitable to some extent: ketones and aldehydes¹⁵⁴ as well as epoxides¹⁵⁵ are suitable substrates under acid catalysis, nucleophilic substitution to halides is also suitable, but generally the reaction is more appropriated on primary halide;¹⁵⁶ however the tertiary halides or alcohols can generate a carbocation which can also react to form hydroperoxides.¹⁵⁷ Addition of hydrogen peroxide to an activated olefin through a peroxymercuration¹⁵⁸ or a peroxyhalogenation¹⁵⁹ provides related compounds.

- Finally electrophilic reactions involving ozone or singlet oxygen; oxygen singlet provides allylic hydroperoxides through an ene-type reaction,¹⁶⁰ whereas ozone, in presence of a protic solvent provides hemiperoxyketals (Figure 8).¹⁶¹

Figure 8. General strategies and pathways to build hydroperoxides.

3.2.2.2 Free radical cyclization of hydroperoxides.

There are few examples of a free radical cyclization of hydroperoxides in the literature. However an interesting example from Carless and coworkers in 1982 showed that a ene-reaction with singlet oxygen (generated *in situ* by photoactivation with tetraphenylporphyrine) on diene **215** afforded a mixture of hydroperoxides **215a-c** in 21, 11, and 13% yield, respectively, through an ene-reaction with singlet oxygen. Further radical mediated cyclization of hydroperoxide **215c** in presence of triplet oxygen and a radical initiator such as di-*tert*-butylperoxyoxalate (DBPO) at room temperature afforded 1,2-dioxolane **216** in 20% yield with the *cis* stereochemistry. The radical initiator allows the production of peroxy radical species, which can then cyclize and trap a new molecule of oxygen (Scheme 31).¹⁶²

Some years later, Boukouvalas and coworkers applied this methodology to the synthesis of Yingzhaosu C (see paragraph 2.2.1).¹⁶³ Thus, tertiary alcohol **218** was treated with an ethereal solution of hydrogen peroxide in presence of amberlyst to accomplish a substitution *via* a carbocation to form hydroperoxide **219**. Treatment

under the conditions that Carless described, with DBPO in oxygen atmosphere, afforded a mixture of 1,2-dioxanes **220a** and **220b** in a 1.7:1 ratio. Addition of *tert*-butyl hydroperoxide improved significantly the yield, up to 89%. Chemoselective reduction of the hydroperoxide with PPh₃ afforded racemic Yingzhaosu C and its epimer in few steps. This work allowed them to clearly reassign the relative configuration of Yingzhaosu C as *cis*, whereas natural product was initially reported as *trans* (Scheme 32).⁶⁰

Scheme 32

3.3 Cyclization through nucleophilic addition or substitution

In contrast to free radical chain reactions, which can mostly provide 1,2-dioxolane or 1,2-dioxane units directly from oxygen in one step, cyclizations using a nucleophilic addition or substitution are predominantly prepared from hydroperoxides. However, these methods are generally stereospecific and allow more easily the preparation of endoperoxides with controlled and defined stereochemistry. Few examples, presenting a direct formation of an endoperoxide from hydrogen peroxide or one of its derivatives, are in general a two-step sequence in a one-pot process. Synthesis of hydroperoxides was discussed in paragraph 3.2.2.1, but reactions presented in this section will be disclosed following their cyclization step.

3.3.1 Nucleophilic substitution

3.3.1.1 Nucleophilic substitution (S_N2) on halides or sulfonates

Nucleophilic substitution by some hydrogen peroxide derivatives in alkylation-type reactions is complicated due to the low reactivity of peroxyanion in this transformation and the propensity of organic peroxides to degrade under basic conditions. Nevertheless, double alkylation from sulfonates was early studied by Corey and Nicolaou in 1975. They reported a first example of a direct formation of 1,2-dioxolane by using potassium superoxide and 18-crown-6 ether to improve its solubility in DMSO.¹⁶⁴ This reaction relies first on the formation of a peroxyradical derivative, which is reduced *in situ* in a peroxyanion that reacts further in an intramolecular fashion to displace the second leaving group. Thus, treatment of dimesylate **219** under these conditions at 25 °C for 3 minutes afforded *cis*-1,2-dioxolane **220** in moderate yield (Scheme 33).

Following a similar strategy, several endoperoxides have been prepared from ditriflates and bis-(tributyltin)peroxide by Salomon and Salomon.¹⁶⁵ Therefore, dioxolanes **224a-b** and 1,2-dioxane **224c** could be synthesized from compound **223a-b** and **223c** respectively, in acceptable yields. These two examples showed the utilization of more reactive sulfonates towards alkylation with oxygen derivatives and were applied to few substituted substrates (Scheme 33).

Scheme 33

Use of hydroperoxides for further cyclization by alkylation was more widely applied because it circumvents the problem of the poorly effective intermolecular alkylation reaction, thus the hydroperoxide function could be introduced by a more convenient method. Furthermore, alkyl halides need generally to be activated by a silver salt to be effective alkylating agents toward the hydroperoxy function. Therefore, in a pioneer example, Bloodworth and coworkers managed to cyclize product **225** into 1,2-dioxolane **226a** with *endo* configuration and inversion of configuration by using Ag₂O for 63 h, and 1,2-dioxolane **226b** with *exo* configuration

and retention of configuration by using silver trifluoroacetate for 1.5 h.¹⁶⁶ No yield was given for these transformations, however retention of configuration observed for the reaction with silver trifluoroacetate was discussed and authors proposed that formation of bromonium ion **227** might be probably at the origin of this unexpected process (Scheme 34).

Similarly, Kishi and coworkers used the same transformation on a cyclopentyl system in their studies towards prostaglandin endoperoxide (PGH₂). An oxidative hydroboration was performed on cyclopentene **228** to give hydroperoxide **229**. Intramolecular alkylation of the latter with Ag₂O afforded endoperoxide **230** in 58% yield (Scheme 34).^{167,168}

Scheme 34

More recent work by Dussault and coworkers showed that a double cyclization could be performed from hydroperoxyketals.¹⁶⁹ Notably, dimesilate **231** was transformed in ketone **232**, but not in hydroperoxyketal as expected. Further reaction with H₂O₂ and H₂SO₄ afforded directly spiro-compound **233**. Homologated substrate **233**, in contrast, could be converted directly by ozonolysis in hydroperoxyketal **235**. However no spontaneous cyclization was observed in this case. Treatment with *t*-BuOK and 18-crown-6 ether, however, could lead to expected structure **236** (Scheme 35).

Application of these last conditions to the total synthesis of peroxyplakoric acids, and in particular A₃ derivative, allowed the construction of the 1,2-dioxane unit.¹⁷⁰ Indeed, intermediate **237** underwent an ozonolysis in presence of methanol to furnish hemiperoxyketal **238**. Cyclization with *t*-BuOK and 18-crown-6 ether afforded in good yield 1,2-dioxane **239**. Further functionalizations including a hydrozirconation, a Negishi cross-coupling and a deprotection afforded ultimately

compound **240**. Unfortunately authors could not complete their final objective (Scheme 35).

Scheme 35

Lastly was reported the total synthesis of dicarnoxide C by Seifert, and their key step for the construction of the 1,2-dioxolane unit was also an intramolecular alkylation.¹⁷¹ Authors performed first an Isayama–Mukaiyama cobalt catalyzed hydroperoxy-silylation on compound **241** followed by a fluoride treatment to remove the silyl group and install the hydroperoxide function. Then the cyclization of **242** was achieved with DBU as base at 60 °C. Functionalization of **243** by introducing the fatty sesquiterpene part, using a Julia-Kochi olefination and further functional modifications, afforded Diacarnoxide C as a mixture of diastereomers. Indeed, introduction of the hydroperoxyl function could not be controlled and authors were unable to separate the different diastereomers at any stage of the synthesis. However most of the analytical data matched with those of the natural product (Scheme 36).

3.3.1.2 Nucleophilic substitution (S_N1) *via* a carbocation.

Preparation of 1,2-dioxolane can be also achieved through a S_N1 pathway. So, in this example, allylic alcohol **245** was transformed into allylic hydroperoxides **246a** and **246b** in a 10:90 ratio by using an ene-reaction with singlet oxygen. The mixture was then treated with a mild Lewis acid such as In(OTf)₃ to drive to **247** through carbocation **248** (Scheme 37).¹⁷²

This mechanism takes also part in the total synthesis of propindilactone G.¹⁷³ Thus, hydroperoxysilane **250** could be obtained *via* an Isayama-Mukaiyama hydroperoxysilylation of **249** and cyclization on hydroxyl group was then performed using BF₃•OEt₂. A complete inversion of the stereochemistry in **251** was observed due to the orientation of the peroxy group in **250**. Eleven steps were then necessary to obtain propindilactone G. It is also noteworthy that intermediate **251** is closely related to pseudolarolides as in Figure 5 (*vide supra*). (Scheme 38)

Scheme 38

3.3.1.3 Nucleophilic substitution on strained heterocycles

Epoxides and oxetanes are convenient intermediates, which can easily undergo a nucleophilic substitution with hydrogen peroxides or any hydroperoxide after activation of the heterocycle with a Lewis or Bronsted acid. The advantages of this method are the possibility to also introduce a hydroxyl group in α or β position of the peroxide function with acids conditions, compared to the basic conditions, and the availability of numerous methods to obtain enantiopure epoxides and in less extend oxetanes, which makes it possible to obtain endoperoxides with a good control of the stereochemistry.

Xu and coworkers applied early this method to assign both relative and absolute configuration of Yingzaosu C. Indeed, since chiral epoxides are readily available in the two enantiomeric forms by using titanium catalyzed asymmetric Sharpless epoxidation, authors used this method for preparing the four possible stereoisomers of Yingzhaosu C.^{61,174} Thus, compound **252** was transformed into mixture of diastereomers **253a-b** and **253c-d** after a Sharpless epoxidation with one or the other enantiomer of diisopropyl tartrate (DIPT), acetylation of the primary alcohol, Isayama-mukaiyama hydroperoxydation and desilylation. It is noteworthy that the hydroperoxidation brings a mixture of diastereomers and that authors first tried to cyclize directly the hydroperoxides through formation of a peroxy anion upon desilylation, but no epoxide ring opening was observed under these conditions, with KF. However acid-mediated conditions allowed cleanly cyclization, affording 1,2-dioxanes **254a-d**. At this step, the different diastereomers could be separated by chromatography and after three more steps performed individually on each products,

the four different possible stereoisomers of yingzhaosu C were thus obtained. Optical rotation study allowed authors to conclude that natural yingzhaosu C is a mixture of enantiomers with the (+)-one being predominant. However they could not conclude themselves about the relative stereochemistry of yingzhaosu C because they could not determine which synthetic product was the *cis* or *trans* isomer, but slightly later Boukouvalas and coworkers¹⁶³ (see Scheme 31) could in contrast conclude that the *cis* isomer is the natural product (stereoisomers a and d). (Scheme 39)

Scheme 39

Gemma, Campiani and coworkers developed also strategy based on a hydroperoxidation of olefin **255** followed by an epoxide ring-opening for the total synthesis of dihydroplakortin and 6-*epi*-dihydroplakortin.¹⁷⁵ After a synthetic study,¹⁷⁶ they were able to access natural products by cyclizing the mixture of diastereoisomer **256** into compounds (6*S*)-**257** and (6*R*)-**257**, which were both separated by chromatography at this step. A 5 steps sequence, *i.e.* an oxidative cleavage of diol, homologation through Wittig reaction, oxidation and esterification, furnished both of desired compounds: dihydroplakortin and its 6-epimer. (Scheme 40)

Scheme 40

The strategy based on epoxide ring-opening was also applied by Vatèle and coworkers in two different works towards the total synthesis of endoperoxides. Firstly, epoxy-alcohol **253** underwent a hydroperoxidation to give compound **254** (as a mixture of diastereomers) and an acid catalysis allowed the formation of 1,2-dioxane **255**, a hypothetical precursor in their work towards total synthesis of Plakortolide E.¹⁷⁷ In a second study towards total synthesis of andavadoic acid, the same strategy was exactly applied through hydroperoxydation of **256**.²⁶ However the conditions used and the regioselectivity of the oxirane ring opening were different to obtain **258a-b**. Indeed, a 5-*exo*-tet cyclization was observed in contrast to a 6-*endo*-tet in the case of the transformation of **254** into **255**, and some basic conditions were also preferred (Scheme 41).

Scheme 41

Epoxides were also used in a cyclization cascade towards the total synthesis of stolonoxides A-F. Deng and coworkers were first interested by applying their enantioselective catalytic peroxidation of enal from **266** with aminoquinidine organocatalyst **268** and hydroperoxide **269**.¹⁷⁸ After transformation of the aldehyde into a methyl ester, peroxyketal **267** was obtained with 90% of the desired diastereomer. Protecting group switch of the peroxyether function followed by Shi epoxidation afforded intermediate **270**. Cleavage of the protecting group followed by a base catalyzed ring opening cascade of the two epoxides, afforded endoperoxide **272** in 52% yield. The direct use of the peroxyketal present in **267** did not allow authors to continue the synthesis because of some side-reactions during the deprotection step and an acid-catalyzed ring opening of the epoxides drove to a poor yield for **272** (15%) along with some bis-THF byproducts, which correspond to a cascade cyclization from a reduced version of hydroperoxide **270**. (Scheme 42)

Scheme 42

Cyclization of hydroperoxides could be also conducted with oxetanes. Dussault and coworkers showed this process is applicable to any oxetane such as compounds **273a-d** to give rise to 1,2-dioxolones **274a-d**.¹⁷⁹ The reaction takes place on the more substituted position with generally an inversion of the configuration. When the reaction was conducted with homologated compounds **273e-f**, 1,2-dioxanes **274e-f** were obtained. However, conditions had to be optimized and reaction time was also extended from 1-2 h to 2 days, meaning 6-*exo-tet* cyclization is much slower. An erosion of the stereoselectivity was also observed for compound **274f**, since partial formation of a carbocation could have arisen in this case (Scheme 43).

Cyclization was also accomplished from hemiperoxyketals. Thus, olefins **275a-c** underwent an ozonolysis to generate this function. Surprisingly, authors isolated directly 1,2-dioxolanes **276a-c** without the help of any acid catalysis. However, this transformation was difficult to apply to the synthesis of the corresponding 1,2-dioxanes.¹⁷⁹ (Scheme 43).

Scheme 43

3.3.2 Addition to carbon-carbon double bonds and related functions

3.3.2.1 Peroxymercuration

Mercury is metal well-known for efficiently promoting nucleophilic addition on olefins or alkynes, because of its high carbophilicity, which means this metal can give an electrophilic character to any π -bond system. The process was also applicable to hydrogen peroxide or hydroperoxide through a peroxymercuration. Pioneer work from Porter and coworkers showed the interest of this transformation for the synthesis of 1,2-dioxanes or 1,2-dioxolanes from hydroperoxides containing an olefin in γ or δ -position.¹⁸⁰ Mercurations are mechanistically related to halogenations, and the metal stays incorporated in final product unless any cleavage of the carbon-mercury bond is undertaken. Moreover, mercury and its salts are known to be highly toxic and due to stoichiometric amount involved in the process, no recent application was reported for any peroxy cyclization.

However, the most noteworthy applications were probably achieved by Bloodworth and coworkers in the 80's where a double peroxymercuration from hydrogen peroxide and diene **277a-b** could be carried out, giving 1,2-dioxolane **278a** or 1,2-dioxane **278b**.^{181,182,183} Carbon-mercury bonds could be then cleaved either by

reduction with NaBH_4 and soda to give products **279a-b**, while oxidation with bromine afforded products **280a-b** (Scheme 44).

An application to the synthesis of some synthetic analogues of plakinic acids was reported from conjugated esters **281a-d**. Thus, first peroxymercuration was performed through formation of **281a-d** followed by a second one giving **273a-d**. A reductive demercuration delivered finally 1,2-dioxolanes **284a-d** and after a final saponification, plakinic acid derivatives **285a-d** were obtained (Scheme 44).¹⁸⁴

Scheme 44

Besides olefins, cyclopropanes possess also a π -bond character. Thus, a peroxycyclization from a hydroperoxide was reported on such substrates. Indeed hydroperoxides **287** underwent a cyclization in presence of perchloric acid and mercury(II) salts to give, in about 80% yield, organomercury compounds **288** as intermediates, isolated with a 60:40 *cis:trans* ratio. Reduction of the organomercury function with alkaline sodium borohydride afforded corresponding dioxolanes **289** in 52-60% yields (Scheme 45).¹⁸⁵

Scheme 45

3.3.2.2 Peroxyhalogenation

Like mercury cations, halides have the ability to activate an olefin through the formation of a halonium ion. Bloodworth and coworkers particularly studied this transformation and they found out that two mechanisms are in competition.¹⁸⁶ For instance, reaction of hydroperoxide **290** with *N*-iodosuccinimide (NIS) furnished **291a** as a 1:1 mixture of diastereomers, suggesting a radical mechanism through the formation of peroxyradical **292a**. In contrast reaction with *N*-bromosuccinimide (NBS) afforded 1,2-dioxolane **291b** in a 80:20 mixture of diastereomer, meaning the mechanism is mainly ionic, *via* bromonium **293b**, due to the stereospecificity of the ionic pathway. However, **291b** being in a 80:20 mixture of diastereomers might come from a minor radical process such as for NIS. *N*-Chlorosuccinimide (NCS) was unreactive for such transformation, and optimized conditions with *t*-butyl hypochlorite and pyridine overcame the difficulties initially encountered, affording **291c** (Scheme 46).¹⁸⁷

Peroxyhalogenation was later used in many syntheses, such as cyclization of **294** to 1,2-dioxolane **295** with Br₂; the stereospecificity of the reaction being total for this example.¹⁸⁸ A cycloperoxyiodination was also described from hydroperoxide **297**, synthesized from meldrum's acid derivative **296** by a copper mediated peroxidation. Authors used 1,3-diiodo-5,5-dimethylhydantoin (DIH) as iodonium source, which gave a good regioselectivity and diastereoselectivity for the formation of 1,2-dioxolane **298** (Scheme 46).¹⁸⁹

A more recent example was reported by Das and coworkers on the haloperoxycyclization of oxazoles.¹⁹⁰ Thus, derivatives **299** were transformed into hydroperoxides **300** by a S_N1 type reaction and further peroxycyclizations were achieved using PIDA and zinc halide. PIDA is oxidizing *in situ* the bromide or chloride into corresponding chlorine or bromine cation source, making further cycloperoxyhalogenation possible to yield, for example, **301a** or **302b**. Among other halogen oxidant these conditions proved to be the most high yielding. However zinc iodide was unable to furnish the desired 1,2-dioxanes. Therefore, up to fifteen spiro-1,2-dioxanes with different substituents on the oxazole were obtained (Scheme 46).

Scheme 46

3.3.2.3 Metal catalyzed peroxycyclization

Some recent methods are involving metal catalyzed peroxycyclization to produce 1,2-dioxanes or 1,2-dioxolanes. Palladium mediated reactions were particularly studied. Thus, Woerpel and coworkers presented a Wacker-type reaction to transform hydroperoxyalkenes **302a-f** into 1,2-dioxanes **303a-f**.¹⁹¹ The reaction involves palladium(II) acetate with some benzoquinone to reoxidize the metal species. Indeed, such as in the Wacker oxidation, substrates **302a-f** are coordinated to Pd(II) species in order to activate the olefin and perform an intramolecular addition from the hydroperoxide. Endoperoxides **303a-f** were then obtained from intermediates **305**, which undergo a β -elimination of the metal. Palladium(0) is then reoxidized in palladium(II) by the benzoquinone. The yield of this reaction is sometimes moderate, due to the hydroperoxide reduction by-products. A divergent protocol was also studied, using benzoquinone 10 mol% and AgCO₃ as an oxidant, however the yield could not really be improved, although it suppressed the reduction process, but gives instead other oxidation by-products (Scheme 47).

Scheme 47

Wong and coworkers, in their total synthesis of plakortide E, studied a synthetic methodology towards the palladium catalyzed oxidation of divinylcyclopropanes **306a-d** into 1,2-dioxolanes **307a-d**.¹⁴⁸ After using molecular oxygen, which gave unsatisfactory results, they optimized the conditions using $\text{Pd}(\text{PPh}_3)_4$ and urea peroxide to obtain such endoperoxides. However the diastereoselectivity was poor and authors could not use this transformation in the total synthesis of plakortide E (see also Chapter 3.2.1.6, Scheme 28). Although the transformation is original, it can be postulated that cyclopropanes are opened by Pd(0) species to give bis- π -allyl palladium species **309a-d**, which can eventually be oxidized with oxygen peroxide into Pd(IV) species. A double Tsuji-Trost like process can then take place with hydrogen peroxide to furnish 1,2-dioxolane **307a-d**. (Scheme 48)

Scheme 48

Such as palladium, gold is known to behave an excellent carbophilicity. Thus, use of this metal was reported as a small application of a peroxycyclization.¹⁹⁰ Meldrum's acid derivatives **312a-b** were transformed into endoperoxyketals **313a-b** in presence of $\text{PPh}_3\text{AuNTf}_2$ and PPTS in MeOH. The first important step is the *endo*-dig peroxycyclization giving **315a-b**. After a proto-demetalation, enol ether function in **316a-b** was acetalized with methanol and PPTS. (Scheme 49)

Scheme 49

3.3.2.4 Conjugated addition

Conjugate olefins with electron withdrawing groups are also potential electrophiles for conjugate addition of hydroperoxides, due to an electron density deficiency at the β -position. Addition of hydrogen peroxide to conjugate systems is a well-known old process, but generally results in the formation of epoxides, the reaction being known as Weitz-Scheffer epoxidation.¹⁹² This epoxidation is driven by the formation of an enolate just after the addition of the hydroperoxy anion, which

can in turn react onto the electrophilic peroxide intramolecularly to give an oxirane. Thus, buffered or acidic conditions might preferentially keep the peroxide bond free.

Kobayashi M. and coworkers especially developed a methodology for the synthesis of 1,2-dioxanes such as **320** by using first $\text{Sc}(\text{OTf})_3$ mediated peroxyhemiacetalization of ketone **318** followed in second by peroxycyclization of **319** with diethylamine in trifluoroethanol.^{193,194,195} These conditions were particularly optimized for this transformation, the major by-products being the oxirane through Weitz-Scheffer epoxidation¹⁹² or a ketone through a Kornblum-DeLaMare rearrangement.¹⁹⁶ Indeed, a base seems essential to induce a good nucleophilicity to the hydroperoxide for the addition onto the conjugated ester, but it also promotes these side reactions. Trifluoroethanol, because of its acidic character, slows them down, notably by buffering the reaction medium. (Scheme 50)

These reaction conditions were also applied in different syntheses of endoperoxides. Therefore, Dussault and coworkers used this peroxycyclization in the total synthesis of peroxycarboates A&D through the obtention of compound **321**;¹⁹⁷ Wu and coworkers obtained many different spiro-1,2-dioxanes such as **322** by using the same method;^{198,199,200} Ortega and coworkers also used this protocol for synthesis of 1,2-dioxolane **323** or 1,2-dioxane **324**;²⁰¹ Vatèle and coworkers used the Kobayashi's procedure in the final step of the synthesis of plakortolide E and (-)-*ent*-plakortolide E;²⁰² and finally Harwood and coworkers also applied the Et_2NH catalyzed peroxycyclization method in the final step of the total synthesis of mycaperoxide B.²⁰³ It can be noted that all yields are from poor to modest in this transformation, due to the above mentioned side-reactions and many protocols needed to be reoptimized on certain substrates to afford desired endoperoxides (Scheme 50).

Peroxy cyclization was also applied to other electron-withdrawing groups than esters. In these cases, Et_2NH catalyzed peroxy cyclization in trifluoroethanol was not always necessary. Indeed, after peroxyhemiacetalization of compounds **325a-b**, 1,2-dioxanes **326a-b** were obtained spontaneously but in poor yields.¹⁹⁵ Interestingly, conjugated nitro substrate **328** drove also spontaneously to 1,2-dioxane **330** in modest yield after a $\text{Sc}(\text{OTf})_3$ mediated peroxyhemiacetalization.^{199,200} These two last results suggest that peroxy cyclization was promoted by Lewis acid catalysis with these two different types of substrates (Scheme 51).

A very significant work about conjugate addition was accomplished by List and coworkers. Indeed, they succeeded in preparing enantioselectively, among nineteen examples, various 1,2-dioxolan-3-ols of type **332** from enones of type **331**, and more

selectively with respect to the Weitz-Scheffer peroxidation driving to oxiranes of type **333**.^{204,205} Organocatalysis is mediated by aminoquinine **268**, in presence of trichloroacetic acid. Addition of hydrogen peroxide to the enone passes in fact through the formation of conjugated iminium ion **334**, which is more reactive towards nucleophilic addition. Adduct **335** evolves back to ketone function, which drove then to dioxolane **332**. Some representative examples such as **332a-d** show that this reaction makes it possible to introduce different functionalities with excellent enantioselectivities, which is currently not so frequent in the synthesis of endoperoxides. However, the reaction seems limited to poorly functionalized ketones (for R²), because groups other than methyl give 1,2-dioxolanes with poor yields and a significant amount of oxiranes **333**. We also personally experienced no conversion with more functionalized ketones (Scheme 52).

Scheme 52

3.3.3 Addition to the carbonyl group

Hydrogen peroxide is a very good nucleophile towards carbonyl group, and by consequence, its addition onto 1,3- and 1,4 diketones system was early studied in the 60's.^{206,207} Depending the stoichiometry of H_2O_2 , adducts **336a-c** and **337a-c** could be obtained from acetylacetone or acetonylacetone. Notably, the use of two equivalents lead to formation of dimeric or polymeric products (Scheme 53).

This method was also applied to the synthesis of tricycle **338**, for the research of new anti-malarial drugs.²⁰⁸ Dussault and coworkers were able to cyclize compounds **340a-b** under acid catalysis, which provided spiro-products **341a** and **341b** after *in situ* silyl deprotection, (Scheme 53).¹⁶⁹

Scheme 53

Terent'ev developed an original methodology from 1,3-diketones, by using some iodine and H_2O_2 to promote an iodoperoxidation from an internal olefin.²⁰⁹ Thus compounds **342a-e** were transformed into 1,2-dioxolanes **343a-e**, whereas diene such as **342f** was transformed into original endoperoxide **343f** through a double iodo-cyclization. The mechanism can evolve between two pathways: in path A, an iodocyclization from ketone group takes place, which produces oxycarbenium species **344a-e** and finally **343a-e** after H_2O_2 addition, while path B describes an inversion of the steps. A mechanical study would be necessary to determine the most plausible route, in particular by using a sequential addition of the reagents in one direction or the other and by analyzing the possible differences in the relative stereochemistry (Scheme 54).

Scheme 54

An interesting study by Woerpel and coworkers concerns the addition of hydrogen peroxide to 1,3-keto-epoxides **348**, which afforded *trans*-**349** and *cis*-**349** in a ratio of about 70:30.²¹⁰ Aromatic substituents generally gave modest yields, whereas alkyl type substituents conducted the best conversions. Mechanistically, the first reacting function is difficult to determine between ketone and epoxide, but the most interesting feature in this transformation is probably the fact that the reaction first leads to the formation of the kinetic products **349**, while a prolonged reaction time mainly leads to the formation of dioxolanes *cis/trans*-**350**, the thermodynamic products. The formation of peroxy-carbenium species **351** is crucial for the formation of both kinetic and thermodynamic products, so addition of hydroperoxide to the carbonyl group is obviously the critical step in this reaction to afford 1,2-dioxolanes **350**. (Scheme 55)

Scheme 55

Gemma, Campiani and coworkers transposed their study toward the ring opening of epoxides with hydroperoxide,^{175,176} to carbonyl derivatives. Therefore, after Isayama–Mukaiyama hydroperoxidation of olefins **352a-r**, lactone **353a-r** were reduced with DIBAL and resulting lactols **354a-r** underwent a cyclization onto oxycarbenium species **355a-r** to furnish 18 examples of 1,2-dioxanes, fused with a THF ring.²¹¹ Some amino derivatives were also recently prepared from **356f**, using reductive amination, but the yields were low presumably to some competitive Kornblum-DeLaMare rearrangement.²¹² (Scheme 56)

The process was further applied to tetrahydropyranyl compounds. Thus, after hydroperoxidation of olefin **357a-c**, hydroperoxycyclization was performed under the same conditions to obtain intricate dioxanes **360a-c**.²¹³ (Scheme 56)

Application of the two above strategies to ketone derivatives **361a-ac** and **363a-c** afforded in presence of various alcohols, more than 30 ketals derivatives such as **362a-c** or **364a**.²¹⁴ (Scheme 57)

All these products prepared by the team of Gemma and Campiani were tested on *Plasmodium f.* parasites. However, the antimalarial activity was in general modest compared to drugs currently used to treat this disease (Schemes 56&57).

Scheme 56

Scheme 57

3.4 Peroxycyclization involving Electrophilic Singlet Oxygen

Singlet oxygen is an unstable and very reactive oxygen species with a short lifetime, prepared *in situ* from triplet oxygen with a photo-sensitizer under light activation. Peroxides and endoperoxides can be obtained with this reagent from olefins, generally through two sort of processes: an ene-reaction or a [4+2] cycloaddition. Whereas ene-reaction can be only applied to mono-olefins, [4+2] cycloaddition are specifically applicable to dienes or higher conjugated systems. However, in this last scenario, the two processes can be in competition, depending the substrate, which leads to the formation of a hydroperoxide instead of a 1,2-dioxene.

3.4.1 Ene reaction

Ene reaction with oxygen singlet consists generally in the preparation of hydroperoxides; however, in the presence of conjugated ketones, this reaction rises to the formation of hemi-endoperoxiketals. Specifically, Snider and coworkers studied this reaction for the synthesis of some natural products. Therefore, racemic chondrillin and plakorin were synthesized from conjugated ketone **365** by reaction with singlet oxygen, generated with the help of a photo-sensitizer under light irradiation, in a DCM:MeOH mixture.²¹⁵ Transketalization of **366** with MeOH drove to a separable mixture of chondrillin and plakorin. Light irradiation presumably enolizes the conjugated ketone which can then react with singlet oxygen to form after

elimination hydroperoxide **369**. Further UV-mediated isomerization leads **369** finally to **370**, which prefers to adopt an endoperoxide form. If the enolization of **365** seems obvious, Addition of singlet oxygen could react from species **367** through an ene reaction or a [4+2] cycloaddition process. However, since hydroperoxide **369** has been isolated and identified as a by-product in this transformation, the ene reaction is probably the most plausible mechanistic pathway. A very similar strategy was adopted by Dussault in an enantioselective synthesis of chondrillin and plakorin. In particular, they used a light mediated isomerization a double bond from an intermediate such as **369** (Scheme 58).²¹⁶

Ene reaction with singlet oxygen was also applied by Snider and coworkers to the total synthesis of acetylsaturejone and isoacetylsaturejone.²¹⁷ Photooxygenation of intermediate **371** with rose Bengal afforded acetylsaturejone through a direct ene reaction with oxygen triplet. In contrast photooxygenation with CuSO₄ as a sensitizer afforded isoacetylsaturejone through formation of enol **372**. Differences observed in these two conditions are not clearly understood. However, the ene reaction is probably the biomimetic pathway towards the production of these last natural products (Scheme 58).

Scheme 58

The ene reaction was also utilized more recently in the synthesis of the diterpene (+)-Jungermatrobrunin A by Lei, X. and coworkers, where the final step was the peroxidation of **373** with singlet oxygen.²¹⁸ Extensive screening of reaction conditions appointed that rose Bengal as sensitizer in MeCN/pyridine (40:1) using a 24W white LED during 36h gave the best results for this transformation, giving (+)-Jungermatrobrunin A in 57% yield (65% conversion). Light wavelength was very important since UV promoted the rearrangement to product **374** (Scheme 59).

Scheme 59

In a very specific example towards photooxygenation of some allylstannes, Dussault and coworkers demonstrated that singlet oxygen is able to induce directly the formation of 3,5-disubstituted dioxolane rings from functionalized allyl stannanes through a rearrangement including a 1,2-migration of the SnBu_3 group.²¹⁹ Thus allylstannanes **376a-c** were transformed in expected hydroperoxides **377a-c** through a classical ene-reaction or in dioxolanes **378a-c** via a migration of SnBu_3 onto reactional intermediate **379a-c**, evolving into species **380a-c**, which lead to the endoperoxide. The scope of the reaction is limited to few examples. Only **378b** was obtained as a sole reaction product, while **378a** or **378c** were obtained with less selectivity towards hydroperoxides **377a** or **377c**. The transformation seems stereospecific, but is pretty difficult to predict. Nevertheless, the nature of the “X” group, the stereochemistry of the double bond and the solvent seem crucial to the selectivity between **377** and **378** (Scheme 60).

Scheme 60

3.4.2 [4+2] Cycloaddition

The [4+2] cycloaddition of singlet oxygen with dienic systems is probably the most famous transformation for synthesizing endoperoxides, with approximately 1500 occurrences in the literature *via* a search with Scifinder®. The peroxidation of ergosterol was early described by Windaus and Brunken in the 20's by using similar conditions what we use today, *i.e.* an oxygen source, a filament lamp to produce photons and a photosensitizer such as eosin to convert triplet oxygen in singlet oxygen.²²⁰ Because of the high number of occurrences, a selection of research reports will be here presented, focusing on most recent and/or most noteworthy examples.

As explained in the introduction of paragraph 3.4, [4+2] cycloaddition with singlet oxygen can in competition with ene-reaction. The outcome mainly depends on the structure of the substrate, thus Harwood and coworkers principally studied this phenomenon towards the total synthesis of mycaperoxide B.²²¹ Indeed, they hypothesized that mycaperoxide B is probably produced biosynthetically through [4+2] cycloaddition followed by reduction of the olefin contained in the 1,2-dioxene. However, their study could not validate their proposed biomimetic pathway, since diene **381a** underwent a ene-reaction giving **382a** (for both *Z:E* or *E:E* isomers), in contrast to substrate **381b** (non-methylated olefin), which underwent a [4+2] cycloaddition giving 1,2-dioxene **383b**. (Scheme 61)

Scheme 61

The [4+2] cycloaddition was nevertheless used successfully in the total synthesis of some natural endoperoxides. Steliou and Yao presented a study of total syntheses of unnamed natural products belonging to the plakortide family (see paragraph 2.1.2), where the final key step was a [4+2] cycloaddition with singlet oxygen. Thus, diene **384** underwent a cycloaddition to produce 1,2-dioxenes **385a** and **385b** as a mixture of diastereomers (Scheme 62).²²²

Jung and coworkers also used a [4+2] cycloaddition during the synthesis of 6-*epi*-plakortolide E. A cycloaddition reaction from diene **386** afforded dioxenes **387a** and **387b** in the same ratio than the stereoisomeric purity of the starting material. Diastereomer **387a** was then transformed in 4 steps into 6-*epi*-plakortolide E, notably by applying an iodolactonisation followed by a radical reduction of the remaining iodide (Scheme 62).²²³

Whereas 1,2-dioxenes can be kept as a final objective because the target needs the unsaturation inside the endoperoxide, there are some noteworthy examples where the 1,2-dioxene could be converted into a 1,2-dioxane. Because metal catalyzed hydrogenation often lead to homolytic reduction of peroxide bond, the utilization of diimide seems obviously the reagent of choice for this transformation. Thus, guianolide derivative **388** underwent a cycloaddition reaction to make thaperoxide, a synthetic anti-malarial agent, and a diimide reduction gave access to 1,2-dioxane **389**.²²⁴ Many endoperoxide analogs of **389** were then prepared (Scheme 63).

Another example of a selective 1,2-dioxene reduction is represented by the synthesis of gracilioether F by Wong and coworkers.²²⁵ Application of a [4+2] cycloaddition on **390** with singlet oxygen, followed by a diimide reduction gave rise to dioxane **391**. The endoperoxide was then reduced and the primary alcohol oxidized to give the natural product (Scheme 63).

Scheme 63

Such as with the past example depicted in Scheme 57, 1,2-dioxenes are more often used as intermediates to prepare other specific functionalities. Nicolaou and coworkers used the 1,2-dioxenes present in **393** to perform a double Kornblum-DeLaMare rearrangement, producing conjugated 1,4-hydroxyketones, which were further hydrogenated to give Epicoccin (Scheme 64).^{226,227}

In a study towards mulinane diterpenoids, Xie and coworkers prepared mulinic acid by [4+2] cycloaddition from diene **394**. Reaction of mulinic acid with a ruthenium complex gave rise to the formation of isomulinic acid (Scheme 64).²²⁸

Another interesting example transformed 1,2-dioxenes into furans under Appel reaction conditions. Thus, diene **395** underwent first a cycloaddition with singlet oxygen to form 1,2-dioxene **396** and further reaction with PPh_3 and CBr_4 led to **397** in good yield.²²⁹ The tribromomethylanion degrades the peroxide into a hydroxyketone through a Kornblum-deLaMare rearrangement and a dehydration by reaction of the ketal with the bromotriphenylphosphonium cation leads to the furan. Further functionalization through an olefin cross-metathesis reaction gave F_5 methyl ester (Scheme 64).

Scheme 64

Another noteworthy example concerns a work towards the total synthesis of Okundoperoxide, where the 1,2-dioxene was converted into another endoperoxide.²³⁰ Diene **399** underwent first a [4+2] cycloaddition with oxygen singlet giving dioxene **400**. A transformation of the diol in allylic epoxide **401** allowed then to perform the last key step, involving the silyl deprotection triggering with the endoperoxide ring opening and the final cyclization onto the olefin through a Brønsted acid activation of the oxirane (Scheme 65).

Cabrera and coworkers performed a similar transformation onto tetracyclic phenol **403** to produce endoperoxide **408**.²³¹ First, a [4+2] cycloaddition reaction with singlet oxygen takes place, and hydroperoxy group is then released upon elimination by the geminal hydroxyl group. Intermediate **405** is then subjected to a supplementary formal peroxycyclization with singlet oxygen affording endoperoxide **408**. During this last sequence, the reaction seems to not pass exactly through a [4+2] cycloaddition, following author's hypotheses. (Scheme 65)

Scheme 65

3.5 Peroxycyclization and Functionalization of Endoperoxyketals through the Generation of Peroxycarbenium Species.

We saw in paragraph 3.3.3 that carbonyl groups are ideal substrates to prepare some peroxyketals through the creation of a new C-O bond. Peroxyketals, through the formation of reactive peroxycarbenium species are interesting precursors to build new endoperoxides *via* the formation of a new C-C bond. In particular, two main processes will be described in this section: 1) Formal [3+2] cycloaddition between peroxycarbenium species and olefins, allowing the formation of 1,2-dioxolanes. 2) Functionalization of endoperoxyketals, providing an excellent route to many functionalized 1,2-dioxanes or 1,2-dioxolanes.

3.5.1 Formal [3+2] cycloadditions

Seminal work of Nojima and coworkers in the 80's demonstrated for the first time that some 1,2-dioxolanes could be obtained from ozonides (namely 1,2,4-

trioxolanes) by reaction with some 1,1-disubstituted olefins in presence of $\text{BF}_3 \cdot \text{OEt}_2$.^{232,233} Thus, ozonide **409**, was for example, transformed into 1,2-dioxolane **410** by reaction with methylstyrene **91**, releasing some benzophenone. The mechanism of the reaction is somehow similar to all the reactions that will be following in this section. An activation of the peroxyketal by the Lewis acid such as in **411** induces the fragmentation of the ozonide in acetophenone and peroxy-carbenium **412**. The ene-reaction of **91** onto peroxy-carbenium species **412** takes then place to afford in a first step carbonium species **413** and in a second step 1,2-dioxolane **410**. Because formation of **410** is sequential through species **413**, the reaction cannot be considered like a concerted [3+2] cycloaddition but more like a formal process, which influences stereochemistry outcome of the reaction by giving a mixture of diastereomers (Scheme 66).

Scheme 66

Since ozonides possess an axial symmetry, Lewis acid mediated fragmentation of these heterocycles can provide in general two different peroxy-carbenium species. Therefore, Dussault and coworkers were able some years later to apply this formal [3+2] cycloaddition on disubstituted ozonides such as with **414**.^{234,235} In this case, a mixture of products **415** and **416** was obtained from the formation of peroxy-carbenium species **417** and **418**. Regioselectivity in the fragmentation of ozonides cannot always be controlled, which is a drawback with these substrates (Scheme 67).

Scheme 67

A major improvement, in the formal [3+2] cycloaddition between olefins and peroxy-carbenium species, was the utilization of acyclic hemiperoxyketals or hemiperoxyacetals as precursors to perform this transformation.^{236,237} Allylation of these species such as with allyltrimethylsilane, afforded directly 1,2-dioxanes. No regioselectivity issue is observed with the peroxyketals compared to ozonides, but some low yields in this reaction can be attributed to competition between alkoxy and hydroperoxyl group as nucleofuge. The reaction is working pretty well on ketal derivatives but the reaction is more difficult with peroxyacetals such as **419c**. Using a methoxyethylether rather than a methoxy group as nucleofuge improves the yield because of its ability to chelate titanium or tin chloride. Thus compounds **420a-d** were prepared from alkoxyhydroperoxy ketals **419a-d**, however no diastereoselective ratio was reported for these compounds meaning they were probably prepared as 1:1 mixture of diastereomers (Scheme 68).

Scheme 68

Reaction was further studied and exemplified by Woerpel and coworkers on different peroxyketals.^{238,239} Their major input was firstly to use protected

peroxyketals as silylether to promote the reaction. Triethylsilyl group is particularly suitable because protection of the hydroperoxide greatly improves the process during activation of the peroxyketal with Lewis acid, and this silyl group is sufficiently labile under reaction conditions to perform a spontaneous cyclization, rather than allowing an elimination to occur as in any Sakurai or ene-reaction. In a first study, authors used a disilylperoxyketals **421-423** to generate peroxycarbenium species (method A),²³⁸ while in a second study they used silyloxyperoxyketals **424-426** (method B).²³⁹ The use of method A or B is dependent of the substrates and both methods are complementary. Various 1,2-dioxolanes were then prepared (**427-431**) giving yields in the same range, whatever the starting material, prepared from both methods. (Scheme 69)

Scheme 69

Whereas the scope of the reaction was mainly limited to allylsilanes and 1,1-disubstituted alkenes, the formal [3+2] cycloaddition with other type of olefins fails. Indeed, Wu and coworkers observed that expected reaction products rearranged into some side products presenting some alkyl group migration, instead of the expected

1,2-dioxolane.²⁴⁰ This phenomenon was already observed by Nojima²³² and later by us under strong Lewis acid conditions.²⁴¹ Nevertheless, Wu and coworkers discovered that vinyl ether and vinylamine derivatives are convenient reagents for the [3+2] cycloaddition reactions with peroxy-carbenium species.²⁴⁰ Thus, many examples with 2,3-dihydrofuran were performed giving several fused bicyclic compounds. The reaction worked also with *N*-Boc-2,3-dihydropyrol and with acyclic derivatives to give for example **432e** or **432f**. This method allows more generally the introduction of an alkyl substituent in position 4 of the 1,2-dioxolane (Scheme 70).

Scheme 70

The [3+2] cycloaddition was also experienced in an intramolecular fashion, like from compound **441**, which was converted into a mixture of diastereomers **442a** and **442b**, respectively in 22 and 64% yield.²⁴² (Scheme 71)

Scheme 71

The formal [3+2] cycloaddition, using the conditions developed by Woerpel and coworkers, was then used in many application towards the search of new bioactives 1,2-dioxolanes. Thus, Vennerstrom and coworkers developed a library of new anti-malarial candidates exhibiting different endoperoxides, inspired from the synthetic antiparasmodal agent arterolane. They particularly replaced the 1,2,4-trioxolane present in this bioactive compound by a 1,2-dioxolane such as in product **443**.^{243,244,245} However the results were disappointing in term of antimalarial activity, compound **443** being more than 100-fold less active than arterolane against different strains of *Plasmodium falciparum*. Woerpel and Clardy developed and tested also many new 1,2-dioxolanes from the derivatives presented in scheme 69.²⁴⁶ For instance, compound **444** is a chimera between a dioxolane and chloroquine, and which shows potent anti-malarial activity on different strains of the parasite; the beneficial effect of the endoperoxide being significant compared to chloroquine alone. Woerpel also found that compound **427b**, called FINO₂ is a very promising anticancer agent, acting in the ferroptosis through GPX4 activation (Figure 9).^{247, 248}

Figure 9. Different bioactive 1,2-dioxolanes prepared by a [3+2] formal cycloaddition.

3.5.2 Functionalization of Endoperoxyketals and acetals

The creation of peroxycarbenium species is not only useful for the preparation of 1,2-dioxolanes through a formal [3+2] cycloaddition, but can be also reactive intermediates for the functionalization of endoperoxyketals. Indeed, because the peroxide is already substituted, no [3+2] formal cycloaddition can occur on such species, just a substitution being observed.

Dussault and coworkers were the first ones to investigate such a transformation. The method was particularly attractive since it would give easy access to many natural products belonging to the family of plakinic acids.²⁴⁹ 1,2-Dioxolan-3-ol derivatives by themselves are not ideal substrates because of the weak leaving group ability of the hydroxyl group under many Lewis acid conditions. Therefore, a Brønsted acid catalyzed transketalization brought to the synthesis of more potential substrates such as **445a-c** giving after Sakurai or Mukaiyama aldol reaction products **449-455**, dioxolane **446c** affording **455**, **447c** yielding **457**, and finally dioxene **448a** leading to compound **458**.²³⁷ The nature of the nucleofuge contributes to the effectiveness of the reaction, thus, **445c** exhibits the most effective leaving group ability in these series. Indeed, yields are in general superior with this substituent, due to the ability of methoxyethyl ether to form a chelate with Ti(IV) or Sn(IV). The reaction was working with allylsilanes (Sakurai reaction) but also with silyl enol ether or *S*-silyl ketene acetals (Mukaiyama aldol reaction). The stereoselectivity of the reaction is still difficult to control, thus attempts to produce enantiopure aldol products **454** or **455** with some chiral auxiliary failed. However the Oppolzer auxiliary allowed the separation of diastereomers.²⁵⁰ The method was also applied to the synthesis of plakinic acid A (*vide supra* Figure 2) *via* the obtention of thioester **457**.¹⁹ The preparation of all possible enantiomers and diastereomers of plakinic acid A in combination with the optical rotation study toward the sign and its absolute value allowed the proposition of an absolute configuration of plakinic acid A. Dussault also were interested in applying a Sakurai reaction on dioxene **448a**. The reaction was working for the first time on an endoperoxyacetal derivative to give **458**, but the reaction was probably working due to an improved stabilization of the peroxycarbenium species with the unsaturation.²³⁷ Indeed, the reaction was reported to be ineffective with a saturated derivative (Scheme 72).

Scheme 72

The peroxyendoketals can also be reduced by silanes in acidic conditions. List and coworkers presented some applications of the 1,2-dioxolan-3-ols obtained by an enantioselective conjugate addition of hydrogen peroxide to enones (see paragraph 3.3.2.4, Scheme 52). Thus, by using first a transketalization with MeOH, followed by a treatment with Et_3SiH and TiCl_4 , 3,5-disubstituted 1,2-dioxolane **460** could be obtained from **459** over 2 steps. **460** could also be obtained directly by using triflic acid from hydroxyl derivative **459**, albeit in lower yield compared to the two-step sequence (Scheme 73).²⁰⁵

We also reported this reduction in a chemical reaction towards the synthesis of a saturated analogue of mycangimycin using List's conditions with triflic acid. This work was presented more precisely in paragraph 3.2.1.3, Scheme 10 (Scheme 73).¹⁰⁹

Scheme 73

The two previous examples in Scheme 73 showed how preparing 3,5-disubstituted 1,2-dioxolanes by reduction of the peroxyketal, leading to major *cis*-diastereomers. However the direct preparation of 3,5-disubstituted 1,2-dioxolanes or 1,2-dioxanes from endoperoxyacetals is still unsolved since most of the previous examples were reported from peroxyketals, excluding substrate **448a**, which drove to the formation of **458**, because of the enhanced stabilization of peroxycarbenium ion with the unsaturation (see Scheme 72). Our own experience on 3-methoxy-1,2-dioxolanes led us to the same conclusion about their poor reactivity. To overcome these difficulties, acetate was found to be a more appropriate nucleofuge.²⁴¹ However, the protocol of acetylation was crucial, since classical conditions (Ac₂O, pyridine or DMAP) were unsuccessful due to the reactivity of peroxides towards amines. Lewis acid catalyzed acetylation was in contrast very effective and could provide these corresponding reactive substrates. Addition of different nucleophiles was performed using SnCl₄ or TiCl₄ as Lewis acids (0.9 equiv). The reaction was allowed with very diverse nucleophiles, from allylsilanes (**461-469**) to silyl enol ethers of ketone derivatives (**472-473**), to silanes (**470**, **471**, **474**). The results were different by using TiCl₄ or SnCl₄, the first one giving high *trans* selectivities but low yields, whereas the second one delivered 1,2-dioxolanes in significant higher yields but with lower diastereoselectivities. It was found that 3,5-disubstituted 1,2-dioxolanes underwent a more pronounced but selective degradation of the *cis*-diastereomer with TiCl₄, rationalizing these results.²⁴¹ (Scheme 74)

Scheme 74

Slightly later, we reported the same transformation using improved experimental conditions. Utilization of mild Lewis acids such as $\text{Sc}(\text{OTf})_3$ or $\text{InCl}_3/\text{TMSCl}$ proved to be very convenient catalysts for such a reaction.²⁵¹ Besides the catalytic amounts of Lewis acids used (5 mol%), the room temperature conditions instead of -40°C facilitate the experimental protocol. No significant degradation was observed with these catalysts compared to the previous study.²⁴¹ Moreover, it was found that these conditions expanded the scope of the reaction to other nucleophiles such as *O*- or *S*-silyl ketene acetals (**476** and **477-478**, Scheme 75). However, deactivated allylsilanes such as bromo- or chloro-allylsilanes were ineffective with these catalysts (**464** & **467**, Scheme 75). This concludes, finally, that the two protocols, *i.e.* stoichiometric amount of a strong Lewis acid or catalytic amount of a mild Lewis acid, are complementary to each other (Scheme 75).

Scheme 75

This last method was also applied to the synthesis of functionalized 1,2-dioxanes. A general approach for the synthesis of 1,2-dioxan-3-ols was developed by us using an oxidative ring expansion of cyclobutanols (see paragraph 3.2.1.3, scheme 11), therefore the access to substituted 1,2-dioxanes could be possible.¹¹⁰ Indeed, the reaction between various silylated nucleophiles and 1,2-dioxan-3-yl acetates in presence of Sc(OTf)₃ afforded mostly the desired products in good yields. The selectivity, compared to the reaction with 1,2-dioxolan-3-yl acetates,^{241,251} was in general higher, giving the 3,6-*cis*-isomer as major product. Thus, allylated product **479** was isolated as a sole diastereomer whereas other nucleophiles gave **480** and **481** with more moderated selectivities. 6,6-Disubstituted 1,2-dioxanyl-3-acetates are also viable substrates, and the selectivity towards the *cis*-diastereomer depends of the nature of the substituents: *tert*-butyl group discriminating strongly the addition over methyl group to one side (**482**, **486**); the selectivity is more moderate with other substituents (**489**, **490**). Aromatic substituent gave poor yield for the substitution (**491**), because it facilitates side reactions such as eliminations (Kornblum-DeLaMare rearrangement). Different nucleophiles could be used from ketones (**483**) or substituted allylsilanes (**484**) or siloxyfurans (**485**). (Scheme 76)

Scheme 76

4 Conclusion

This book chapter reviews the most advanced and popular methods to access to 1,2-dioxolanes and 1,2-dioxanes. The most studied methods are probably the one using radical processes involving oxygen triplet. Indeed, they allow to conveniently and directly obtain the endoperoxides, through a radical chain reaction with a free reagent; however they lack to give the desired product in a diastereoselective or enantioselective manner. Nucleophilic addition of hydroperoxides needs, in contrast, to have generally the hydroperoxy function already present in the molecule to lead to endoperoxides, which consists in a final cyclization; however these process are more prone to give stereoselective and stereospecific reactions. Oxygen singlet is also a popular reagent, especially to performed [4+2] cycloadditions, but is limited to few substrates (ene-reactions) or to the construction of some 1,2-dioxenes or 1,2-dioxanes by extension. The use of peroxy-carbenium ion is a more recent method developed from the 90's, which is an interesting tool to access to new structure, in particular 1,2-dioxolanes through a formal [3+2] cycloaddition; however an effort will have to be furnished in the future to develop asymmetric versions of this reaction. Many natural products exhibiting a 1,2-dioxane or a 1,2-dioxolane have been reported with

antibacterial, antifungal, antimalarial, antiviral or cytotoxic activities, a high proportion of which was isolated from marine sponges. However, few total syntheses of these compounds were studied compared to the large number of natural substances reported. This means that still some efforts are needed to access these structures. Efforts to find and develop new methods in a stereoselective fashion have to be conducted in the future to overcome present limitations.

5 References

1. Schulz, M.; Kirschke, K. Cyclic Peroxides. In *Advances in Heterocyclic Chemistry*; Elsevier, 1967; Vol. 8, pp 165–217. [https://doi.org/10.1016/S0065-2725\(08\)60607-8](https://doi.org/10.1016/S0065-2725(08)60607-8).
2. Terent'ev, A. O.; Borisov, D. A.; Vil', V. A.; Dembitsky, V. M. *Beilstein J. Org. Chem.* **2014**, 10 (1), 34–114. <https://doi.org/10.3762/bjoc.10.6>.
3. McCullough, K. J. and Nojima, M. *Curr. Org. Chem.* **2001**, 5, 601–636. <https://doi.org/10.2174/1385272013375346>
4. Pinet, A.; Nguyen, L. T.; Figadère, B.; Ferrié, L. *Eur. J. Org. Chem.*, early view. <https://doi.org/10.1002/ejoc.202000980>.
5. Korshin, E. E.; Bachi, M. D. Synthesis of Cyclic Peroxides. In *PATAI'S Chemistry of Functional Groups*; American Cancer Society, 2009. <https://doi.org/10.1002/9780470682531.pat0351>.
6. *Category 5, Compounds with One Saturated Carbon Heteroatom Bond: Peroxides*, 1st ed.; Berkessel, Ed.; Georg Thieme Verlag: Stuttgart, 2009. <https://doi.org/10.1055/b-003-121800>.
7. Liu, D.-Z.; Liu, J.-K. *Nat. Prod. Bioprospect.* **2013**, 3 (5), 161–206. <https://doi.org/10.1007/s13659-013-0042-7>.
8. Casteel, D. A. *Nat. Prod. Rep.* **1999**, 16 (1), 55–73. <https://doi.org/10.1039/A705725C>.
9. Casteel, D. A. *Nat. Prod. Rep.* **1992**, 9 (4), 289–312. <https://doi.org/10.1039/NP9920900289>.
10. Phillipson, D. W.; Rinehart, K. L. *J. Am. Chem. Soc.* **1983**, 105 (26), 7735–7736. <https://doi.org/10.1021/ja00364a045>.
11. Davidson, B. S. *J. Org. Chem.* **1991**, 56 (23), 6722–6724. <https://doi.org/10.1021/jo00023a048>.
12. Horton, P. A.; Longley, R. E.; Kelly-Borges, M.; McConnell, O. J.; Ballas, L. M. *J. Nat. Prod.* **1994**, 57 (10), 1374–1381. <https://doi.org/10.1021/np50112a006>.
13. Chen, Y.; Killday, K. B.; McCarthy, P. J.; Schimoler, R.; Chilson, K.; Selitrennikoff, C.; Pomponi, S. A.; Wright, A. E. *J. Nat. Prod.* **2001**, 64 (2), 262–264. <https://doi.org/10.1021/np000368+>.
14. Jiménez-Romero, C.; Ortiz, I.; Vicente, J.; Vera, B.; Rodríguez, A. D.; Nam, S.; Jove, R. *J. Nat. Prod.* **2010**, 73 (10), 1694–1700. <https://doi.org/10.1021/np100461t>.
15. Sandler, J. S.; Colin, P. L.; Hooper, J. N. A.; Faulkner, D. J. *J. Nat. Prod.* **2002**, 65 (9), 1258–1261. <https://doi.org/10.1021/np020228v>.
16. Dalisay, D. S.; Quach, T.; Nicholas, G. N.; Molinski, T. F. *Angew. Chem. Int. Ed.* **2009**, 48 (24), 4367–4371. <https://doi.org/10.1002/anie.200900888>.
17. Dalisay, D. S.; Quach, T.; Molinski, T. F. *Org. Lett.* **2010**, 12 (7), 1524–1527. <https://doi.org/10.1021/ol100249v>.
18. Jamison, M. T.; Dalisay, D. S.; Molinski, T. F. *J. Nat. Prod.* **2016**, 79 (3), 555–563. <https://doi.org/10.1021/acs.jnatprod.5b00951>.
19. Dai, P.; Trullinger, T. K.; Liu, X.; Dussault, P. H. *J. Org. Chem.* **2006**, 71 (6), 2283–2292. <https://doi.org/10.1021/jo0522254>.

20. Cafieri, F.; Fattorusso, E.; Taglialatela-Scafati, O.; Ianaro, A. *Tetrahedron* **1999**, 55 (22), 7045–7056. [https://doi.org/10.1016/S0040-4020\(99\)00332-4](https://doi.org/10.1016/S0040-4020(99)00332-4).
21. Ghosh, I.; Zeng, H.; Kishi, Y. *Org. Lett.* **2004**, 6 (25), 4715–4718. <https://doi.org/10.1021/ol048061f>.
22. Tian, X.-Y.; Han, J.-W.; Zhao, Q.; Wong, H. N. C. *Org. Biomol. Chem.* **2014**, 12 (22), 3686–3700. <https://doi.org/10.1039/C4OB00448E>.
23. Jiménez-Romero, C.; Rodríguez, A. D.; Nam, S. *Org. Lett.* **2017**, 19 (6), 1486–1489. <https://doi.org/10.1021/acs.orglett.7b00547>.
24. Mohammed, R.; Peng, J.; Kelly, M.; Yousaf, M.; Winn, E.; Odde, S.; Bie, Z.; Xie, A.; Doerksen, R. J.; Hamann, M. T. *Aust. J. Chem.* **2010**, 63 (6), 877–885. <https://doi.org/10.1071/CH09665>.
25. Rudi, A.; Afanii, R.; Gravalos, L. G.; Akin, M.; Gaydou, E.; Vacelet, J.; Kashman, Y. *J. Nat. Prod.* **2003**, 66 (5), 682–685. <https://doi.org/10.1021/np020589a>.
26. Barnych, B.; Fenet, B.; Vatile, J.-M. *Tetrahedron* **2013**, 69 (1), 334–340. <https://doi.org/10.1016/j.tet.2012.10.022>.
27. Yong, K. W. L.; Barnych, B.; De Voss, J. J.; Vatile, J.-M.; Garson, M. J. *J. Nat. Prod.* **2012**, 75 (10), 1792–1797. <https://doi.org/10.1021/np3005634>.
28. Yong, K. W. L.; Lambert, L. K.; Hayes, P. Y.; De Voss, J. J.; Garson, M. J. *J. Nat. Prod.* **2012**, 75 (3), 351–360. <https://doi.org/10.1021/np200619q>.
29. Yong, K. W. L.; De Voss, J. J.; Hooper, J. N. A.; Garson, M. J. *J. Nat. Prod.* **2011**, 74 (2), 194–207. <https://doi.org/10.1021/np100620x>.
30. Perry, T. L.; Dickerson, A.; Khan, A. A.; Kondru, R. K.; Beratan, D. N.; Wipf, P.; Kelly, M.; Hamann, M. T. *Tetrahedron* **2001**, 57 (8), 1483–1487. [https://doi.org/10.1016/S0040-4020\(00\)01134-0](https://doi.org/10.1016/S0040-4020(00)01134-0).
31. Fontana, A.; González, M. C.; Gavagnin, M.; Templado, J.; Cimino, G. *Tetrahedron Lett.* **2000**, 41 (3), 429–432. [https://doi.org/10.1016/S0040-4039\(99\)02077-8](https://doi.org/10.1016/S0040-4039(99)02077-8).
32. Durán, R.; Zubía, E.; Ortega, M. J.; Naranjo, S.; Salvá, J. *Tetrahedron* **2000**, 56 (33), 6031–6037. [https://doi.org/10.1016/S0040-4020\(00\)00554-8](https://doi.org/10.1016/S0040-4020(00)00554-8).
33. Reyes, F.; Rodríguez-Acebes, R.; Fernández, R.; Bueno, S.; Francesch, A.; Cuevas, C. *J. Nat. Prod.* **2010**, 73 (1), 83–85. <https://doi.org/10.1021/np900700h>.
34. Ovenden, S. P. B.; Capon, R. J. *J. Nat. Prod.* **1999**, 62 (2), 214–218. <https://doi.org/10.1021/np980223r>.
35. Kashman, Y.; Rotem, M. *Tetrahedron Lett.* **1979**, 20 (19), 1707–1708. [https://doi.org/10.1016/S0040-4039\(01\)93630-5](https://doi.org/10.1016/S0040-4039(01)93630-5).
36. Sperry, S.; Valeriote, F. A.; Corbett, T. H.; Crews, P. *J. Nat. Prod.* **1998**, 61 (2), 241–247. <https://doi.org/10.1021/np970467w>.
37. Capon, R. J.; MacLeod, J. K.; Willis, A. C. *J. Org. Chem.* **1987**, 52 (3), 339–342. <https://doi.org/10.1021/jo00379a004>.
38. Tanaka, J.; Higa, T.; Suwanborirux, K.; Kokpol, U.; Bernardinelli, G.; Jefford, C. W. *J. Org. Chem.* **1993**, 58 (11), 2999–3002. <https://doi.org/10.1021/jo00063a016>.
39. Hirade, H.; de Voogd, N. J.; Suzuka, T.; Tanaka, J. *Tetrahedron* **2019**, 75 (33), 4620–4625. <https://doi.org/10.1016/j.tet.2019.07.005>.
40. Albericci, M.; Braekman, J. C.; Daloze, D.; Tursch, B. *Tetrahedron* **1982**, 38 (13), 1881–1890. [https://doi.org/10.1016/0040-4020\(82\)80037-9](https://doi.org/10.1016/0040-4020(82)80037-9).
41. Wells, R. J. *Tetrahedron Lett.* **1976**, 17 (30), 2637–2638. [https://doi.org/10.1016/S0040-4039\(00\)91755-6](https://doi.org/10.1016/S0040-4039(00)91755-6).
42. Quinoa, E.; Kho, E.; Manes, L. V.; Crews, P.; Bakus, G. J. *J. Org. Chem.* **1986**, 51 (22), 4260–4264. <https://doi.org/10.1021/jo00372a029>.
43. Dussault, P. H.; Woller, K. R. *J. Am. Chem. Soc.* **1997**, 119 (16), 3824–3825. <https://doi.org/10.1021/ja970174p>.

44. Yosief, T.; Rudi, A.; Wolde-ab, Y.; Kashman, Y. *J. Nat. Prod.* **1998**, 61 (4), 491–493. <https://doi.org/10.1021/np970407q>.
45. Fontana, A.; d'Ippolito, G.; D'Souza, L.; Mollo, E.; *J. Nat. Prod.* **2001**, 64 (1), 131–133. <https://doi.org/10.1021/np0002435>.
46. Chianese, G.; Fattorusso, E.; Scala, F.; Teta, R.; Calcinai, B.; Bavestrello, G.; Dien, H. A.; Kaiser, M.; Tasdemir, D.; Taglialatela-Scafati, O. *Org. Biomol. Chem.* **2012**, 10 (35), 7197–7207. <https://doi.org/10.1039/C2OB26124C>.
47. Kobayashi, M.; Kondo, K.; Kitagawa, I. *Chem. Pharm. Bull.* **1993**, 41 (7), 1324–1326. <https://doi.org/10.1248/cpb.41.1324>.
48. Chen, S.-P.; Ahmed, A. F.; Dai, C.-F.; Lu, C.-K.; Hu, W.-P.; Wang, J.-J.; Sheu, J.-H. *Tetrahedron* **2006**, 62 (29), 6802–6807. <https://doi.org/10.1016/j.tet.2006.04.088>.
49. Ahmed, A. F.; Kuo, Y.-H.; Dai, C.-F.; Sheu, J.-H. *J. Nat. Prod.* **2005**, 68 (8), 1208–1212. <https://doi.org/10.1021/np050114u>.
50. Chao, C.-H.; Hsieh, C.-H.; Chen, S.-P.; Lu, C.-K.; Dai, C.-F.; Wu, Y.-C.; Sheu, J.-H. *Tetrahedron Lett.* **2006**, 47 (13), 2175–2178. <https://doi.org/10.1016/j.tetlet.2006.01.107>.
51. Uchio, Y.; Eguchi, S.; Kuramoto, J.; Nakayama, M.; Hase, T. *Tetrahedron Lett.* **1985**, 26 (37), 4487–4490. [https://doi.org/10.1016/S0040-4039\(00\)88937-6](https://doi.org/10.1016/S0040-4039(00)88937-6).
52. Sawant, S. S.; Youssef, D. T. A.; Sylvester, P. W.; Wali, V.; Sayed, K. A. E. *Nat. Prod. Commun.* **2019**. <https://doi.org/10.1177/1934578X0700200201>.
53. Umeyama, A.; Machida, M.; Nozaki, M.; Arihara, S. *J. Nat. Prod.* **1998**, 61 (11), 1435–1436. <https://doi.org/10.1021/np980210d>.
54. Huang, X.-C.; Li, J.; Li, Z.-Y.; Shi, L.; Guo, Y.-W. *J. Nat. Prod.* **2008**, 71 (8), 1399–1403. <https://doi.org/10.1021/np8002035>.
55. Ueoka, R.; Nakao, Y.; Kawatsu, S.; Yaegashi, J.; Matsumoto, Y.; Matsunaga, S.; Furihata, K.; van Soest, R. W. M.; Fusetani, N. *J. Org. Chem.* **2009**, 74 (11), 4203–4207. <https://doi.org/10.1021/jo900380f>.
56. Festa, C.; De Marino, S.; D'Auria, M. V.; Deharo, E.; Gonzalez, G.; Deyssard, C.; Petek, S.; Bifulco, G.; Zampella, A. *Tetrahedron* **2012**, 68 (49), 10157–10163. <https://doi.org/10.1016/j.tet.2012.09.106>.
57. Wallach, O. *Liebigs Ann.* **1912**, 392 (1), 49–75. <https://doi.org/10.1002/jlac.19123920104>.
58. Labbe, C.; Castillo, M.; Connolly, J. D. *Phytochemistry* **1993**, 34 (2), 441–444. [https://doi.org/10.1016/0031-9422\(93\)80026-O](https://doi.org/10.1016/0031-9422(93)80026-O).
59. Liang, X. T.; Yu, D. Q.; Wu, W. L.; Rang, H. C., *Acta Chim. Sinica*, **1979**, 37, 215. http://sioc-journal.cn/Jwk_hxb/EN/Y1979/V37/I3/215
60. Zhang, L.; Zhou, W.-S.; Xu, X.-X. *J. Chem. Soc., Chem. Commun.* **1988**, 8, 523–524. <https://doi.org/10.1039/C39880000523>.
61. Xu, X.-X.; Dong, H.-Q. *Tetrahedron Lett.* **1994**, 35 (50), 9429–9432. [https://doi.org/10.1016/S0040-4039\(00\)78561-3](https://doi.org/10.1016/S0040-4039(00)78561-3).
62. Rücker, G.; Breitmaier, E.; Mayer, R.; Manns, D. *Arch. Pharm.* **1987**, 320 (5), 437–441. <https://doi.org/10.1002/ardp.19873200510>.
63. Rücker, G.; Manns, D.; Wilbert, S. *Arch. Pharm.* **1993**, 326 (8), 457–460. <https://doi.org/10.1002/ardp.19933260806>.
64. Duong, T.-H.; Beniddir, M. A.; Genta-Jouve, G.; Nguyen, H.-H.; Nguyen, D.-P.; Nguyen, T.-A.-T.; Mac, D.-H.; Boustie, J.; Nguyen, K.-P.-P.; Chavasiri, W.; Le Pogam, P. *Fitoterapia* **2019**, 135, 44–51. <https://doi.org/10.1016/j.fitote.2019.04.001>.
65. Ma, W.-H.; Tan, C.-M.; He, J.-C.; Duan, P.-S.; Qin, L.-P. *Chem. Nat. Compd.* **2011**, 47 (5), 713–715. <https://doi.org/10.1007/s10600-011-0042-y>.
66. Rustaiyan, A.; Sigari, H.; Jakupovic, J.; Grenz, M. *Phytochemistry* **1989**, 28 (10), 2723–2725. [https://doi.org/10.1016/S0031-9422\(00\)98075-0](https://doi.org/10.1016/S0031-9422(00)98075-0).

67. Hashidoko, Y.; Tahara, S.; Mizutani, J. *Phytochemistry* **1989**, 28 (2), 425–430. [https://doi.org/10.1016/0031-9422\(89\)80026-3](https://doi.org/10.1016/0031-9422(89)80026-3).
68. Itokawa, H.; Morita, H.; Osawa, K.; Watanabe, K.; Itaka, Y. *Chem. Pharm. Bull.* **1987**, 35 (7), 2849–2859. <https://doi.org/10.1248/cpb.35.2849>.
69. Todorova, M.; Vogler, B.; Tsankova, E. **2000**, 55 (9–10), 840–842. <https://doi.org/10.1515/znc-2000-9-1028>.
70. O'Mathúna, D. P.; Doskotch, R. W. *J. Nat. Prod.* **1995**, 58 (9), 1407–1418. <https://doi.org/10.1021/np50123a011>.
71. Adelekan, A. M.; Prozesky, E. A.; Hussein, A. A.; Ureña, L. D.; van Rooyen, P. H.; Liles, D. C.; Meyer, J. J. M.; Rodríguez, B. *J. Nat. Prod.* **2008**, 71 (11), 1919–1922. <https://doi.org/10.1021/np800333r>.
72. Xuan, J.; Zhu, A.; Ma, B.; Ding, H. *Org. Lett.* **2018**, 20 (13), 4153–4156. <https://doi.org/10.1021/acs.orglett.8b01875>.
73. Loyola, L. A.; Morales, G.; Rodríguez, B.; Jiménez-Barbero, J.; Torre, M. C. de la; Perales, A.; Torres, M. R. *Tetrahedron* **1990**, 46 (15), 5413–5420. [https://doi.org/10.1016/S0040-4020\(01\)87848-0](https://doi.org/10.1016/S0040-4020(01)87848-0).
74. Tan, J.-M.; Qiu, Y.-H.; Tan, X.-Q.; Tan, C.-H. *Helv. Chim. Acta* **2011**, 94 (9), 1697–1702. <https://doi.org/10.1002/hlca.201100166>.
75. Chiang, Y.-M.; Kuo, Y.-H. *J. Nat. Prod.* **2001**, 64 (4), 436–439. <https://doi.org/10.1021/np0004808>.
76. Dong, J.-W.; Cai, L.; Li, X.-J.; Peng, L.; Xing, Y.; Mei, R.-F.; Wang, J.-P.; Ding, Z.-T. *Fitoterapia* **2016**, 109, 212–216. <https://doi.org/10.1016/j.fitote.2015.12.019>.
77. Wang, L.; He, H.-P.; Di, Y.-T.; Zhang, Y.; Hao, X.-J. *Tetrahedron Lett.* **2012**, 53 (13), 1576–1578. <https://doi.org/10.1016/j.tetlet.2012.01.060>.
78. Lee, S.-S.; Lin, Y.-S.; Chen, C.-K. *J. Nat. Prod.* **2009**, 72 (7), 1249–1252. <https://doi.org/10.1021/np9000653>.
79. Wieland, P.; Prelog, V. *Helv. Chim. Acta* **1947**, 30 (4), 1028–1030. <https://doi.org/10.1002/hlca.19470300414>.
80. Li, H.; Huang, H.; Shao, C.; Huang, H.; Jiang, J.; Zhu, X.; Liu, Y.; Liu, L.; Lu, Y.; Li, M.; Lin, Y.; She, Z. *J. Nat. Prod.* **2011**, 74 (5), 1230–1235. <https://doi.org/10.1021/np200164k>.
81. Chokpaiboon, S.; Sommit, D.; Teerawatananond, T.; Muangsin, N.; Bunyapaiboonsri, T.; Pudhom, K. *J. Nat. Prod.* **2010**, 73 (5), 1005–1007. <https://doi.org/10.1021/np100103j>.
82. Liu, L.; Niu, S.; Lu, X.; Chen, X.; Zhang, H.; Guo, L.; Che, Y. *Chem. Commun.* **2010**, 46 (3), 460–462. <https://doi.org/10.1039/B918330B>.
83. Scott, J. J.; Oh, D.-C.; Yuceer, M. C.; Klepzig, K. D.; Clardy, J.; Currie, C. R. *Science* **2008**, 322 (5898), 63–63. <https://doi.org/10.1126/science.1160423>.
84. Oh, D.-C.; Scott, J. J.; Currie, C. R.; Clardy, J. *Org. Lett.* **2009**, 11 (3), 633–636. <https://doi.org/10.1021/ol802709x>.
85. Hamberg, M.; Samuelsson, B. *Proc. Natl. Acad. Sci. USA* **1973**, 70 (3), 899–903. <https://doi.org/10.1073/pnas.70.3.899>.
86. Moncada, S.; Vane, J. R. *Pharmacol. Rev.* **1978**, 30 (3), 293–331. <http://pharmrev.aspetjournals.org/content/30/3/293>.
87. Nissen, J. H.; Drews, T.; Schröder, B.; Beckers, H.; Steinhauer, S.; Riedel, S. *Chem. Eur. J.* **2019**, 25 (64), 14721–14727. <https://doi.org/10.1002/chem.201903620>.
88. Daniel Little, R.; Losiski-Dang, L.; Venegas, M. G.; Merlic, C. *Tetrahedron Lett.* **1983**, 24 (42), 4499–4502. [https://doi.org/10.1016/S0040-4039\(00\)85938-9](https://doi.org/10.1016/S0040-4039(00)85938-9).
89. Wilson, R. M.; Schnapp, K. A.; Merwin, R. K.; Ranganathan, R.; Moats, D. L.; Conrad, T. T. *J. Org. Chem.* **1986**, 51 (21), 4028–4035. <https://doi.org/10.1021/jo00371a021>.

90. Chou, C.-H.; Trahanovsky, W. S. *J. Org. Chem.* **1995**, 60 (17), 5449–5451.
<https://doi.org/10.1021/jo00122a024>.
91. (1) Mizuno, K.; Kamiyama, N.; Ichinose, N.; Otsuji, Y. *Tetrahedron* **1985**, 41 (11), 2207–2214.
[https://doi.org/10.1016/S0040-4020\(01\)96594-9](https://doi.org/10.1016/S0040-4020(01)96594-9).
92. (1) Maeda, H.; Nakagawa, H.; Mizuno, K. *Photochem. Photobiol. Sci.* **2003**, 2 (11), 1056–1058.
<https://doi.org/10.1039/B306186H>.
93. (1) Lu, Z.; Parrish, J. D.; Yoon, T. P. *Tetrahedron* **2014**, 70 (27–28), 4270–4278.
<https://doi.org/10.1016/j.tet.2014.02.045>.
94. (1) Mizuno, K.; Tamai, T.; Hashida, I.; Otsuji, Y.; Kuriyama, Y.; Tokumaru, K. *J. Org. Chem.* **1994**, 59 (24), 7329–7334. <https://doi.org/10.1021/jo00103a025>.
95. (1) Parrish, J. D.; Ischay, M. A.; Lu, Z.; Guo, S.; Peters, N. R.; Yoon, T. P. *Org. Lett.* **2012**, 14 (6), 1640–1643. <https://doi.org/10.1021/ol300428q>.
96. (1) Gesmundo, N. J.; Nicewicz, D. A. *Beilstein J. Org. Chem.* **2014**, 10 (1), 1272–1281.
<https://doi.org/10.3762/bjoc.10.128>.
97. (1) Qiao, X.; Biswas, S.; Wu, W.; Zhu, F.; Tung, C.-H.; Wang, Y. *Tetrahedron* **2018**, 74 (20), 2421–2427. <https://doi.org/10.1016/j.tet.2018.03.040>.
98. (1) Wimalasena, K.; Wickman, H. B.; Mahindaratne, M. P. D. *Eur. J. Org. Chem.* **2001**, 2001 (20), 3811–3817. [https://doi.org/10.1002/1099-0690\(200110\)2001:20<3811::AID-EJOC3811>3.0.CO;2-6](https://doi.org/10.1002/1099-0690(200110)2001:20<3811::AID-EJOC3811>3.0.CO;2-6).
99. Ouhamou, N.; Six, Y. *Org. Biomol. Chem.* **2003**, 1 (17), 3007–3009.
<https://doi.org/10.1039/B306719J>.
100. Madelaine, C.; Six, Y.; Buriez, O. *Angew. Chem.* **2007**, 119 (42), 8192–8195.
<https://doi.org/10.1002/ange.200702903>.
101. Madelaine, C.; Buriez, O.; Crousse, B.; Florent, I.; Grellier, P.; Retaillieu, P.; Six, Y. *Org. Biomol. Chem.* **2010**, 8 (24), 5591–5601. <https://doi.org/10.1039/C0OB00308E>.
102. Nuter, F.; Dimé, A. K. D.; Chen, C.; Bounaadja, L.; Mouray, E.; Florent, I.; Six, Y.; Buriez, O.; Marinetti, A.; Voituriez, A. *Chem. Eur. J.* **2015**, 21 (14), 5584–5593.
<https://doi.org/10.1002/chem.201406138>.
103. Xiong, C.; Cheng, K.; Wang, J.; Yang, F.; Lu, J.; Zhou, Q. *J. Org. Chem.* **2020**, 85 (14), 9386–9395. <https://doi.org/10.1021/acs.joc.0c00652>.
104. Budde, S.; Goerdeler, F.; Floß, J.; Kreitmeier, P.; Hicks, E. F.; Moscovitz, O.; Seeberger, P. H.; Davies, H. M. L.; Reiser, O. *Org. Chem. Front.* **2020**, 7 (14), 1789–1795.
<https://doi.org/10.1039/D0QO00168F>.
105. Gibson, D. H.; DePuy, C. H. *Tetrahedron Lett.* **1969**, 10 (27), 2203–2206.
[https://doi.org/10.1016/S0040-4039\(01\)88122-3](https://doi.org/10.1016/S0040-4039(01)88122-3).
106. Kulinkovich, O. G.; Astashko, D. A.; Tyvorskii, V. I.; Ilyina, N. A. *Synthesis* **2001**, 2001 (10), 1453–1455. <https://doi.org/10.1055/s-2001-16089>.
107. Elek, G. Z.; Borovkov, V.; Lopp, M.; Kananovich, D. G. *Org. Lett.* **2017**, 19 (13), 3544–3547.
<https://doi.org/10.1021/acs.orglett.7b01519>.
108. Han, W.-B.; Li, S.-G.; Lu, X.-W.; Wu, Y. *Eur. J. Org. Chem.* **2014**, 2014 (18), 3841–3846.
<https://doi.org/10.1002/ejoc.201402175>.
109. Nguyen, T. L.; Ferrié, L.; Figadère, B. *Tetrahedron Lett.* **2016**, 57 (47), 5286–5289.
<https://doi.org/10.1016/j.tetlet.2016.10.051>.
110. M. Martín López; Nicolas, J.; Pinet, Alexis; Figadère, Bruno; Ferrié, L. *ChemRxiv* **2020**, Preprint.
<https://doi.org/10.26434/chemrxiv.13386917.v1>.
111. Boto, A.; Betancor, C.; Suárez, E. *Tetrahedron Lett.* **1994**, 35 (30), 5509–5512.
[https://doi.org/10.1016/S0040-4039\(00\)73537-4](https://doi.org/10.1016/S0040-4039(00)73537-4).
112. Iwahama, T.; Sakaguchi, S.; Ishii, Y. *Chem. Commun.* **2000**, No. 23, 2317–2318.
<https://doi.org/10.1039/B007182J>.

113. Kumabe, R.; Nishino, H.; Yasutake, M.; Nguyen, V.-H.; Kurosawa, K. *Tetrahedron Lett.* **2001**, 42 (1), 69–72. [https://doi.org/10.1016/S0040-4039\(00\)01884-0](https://doi.org/10.1016/S0040-4039(00)01884-0).
114. Asahi, K.; Nishino, H. *Tetrahedron* **2005**, 61 (47), 11107–11124. <https://doi.org/10.1016/j.tet.2005.09.029>.
115. Asahi, K.; Nishino, H. *Tetrahedron* **2008**, 64 (8), 1620–1634. <https://doi.org/10.1016/j.tet.2007.12.017>.
116. Asahi, K.; Nishino, H. *Eur. J. Org. Chem.* **2008**, 2008 (14), 2404–2416. <https://doi.org/10.1002/ejoc.200701232>.
117. Linker, T. *J. Organomet. Chem.* **2002**, 661 (1), 159–167. [https://doi.org/10.1016/S0022-328X\(02\)01824-7](https://doi.org/10.1016/S0022-328X(02)01824-7).
118. Kumabe, R.; Nishino, H. *Tetrahedron Lett.* **2004**, 45 (4), 703–706. <https://doi.org/10.1016/j.tetlet.2003.11.054>.
119. Nishino, H.; Kumabe, R.; Hamada, R.; Yakut, M. *Tetrahedron* **2014**, 70 (7), 1437–1450. <https://doi.org/10.1016/j.tet.2014.01.013>.
120. Biyogo, A. M.; Curti, C.; El-Kashef, H.; Khoumeri, O.; Terme, T.; Vanelle, P. *RSC Adv.* **2016**, 7 (1), 106–111. <https://doi.org/10.1039/C6RA25138B>.
121. Nishino, H.; Haque, A. *Heterocycles*, **2011**, 83 (8), 1783. <https://doi.org/10.3987/COM-11-12241>.
122. Daepfen, C.; Kaiser, M.; Neuburger, M.; Gademann, K. *Org. Lett.* **2015**, 17 (21), 5420–5423. <https://doi.org/10.1021/acs.orglett.5b02773>.
123. Christoffers, J.; Werner, T.; Frey, W.; Baro, A. *Eur. J. Org. Chem.* **2003**, 2003 (24), 4879–4886. <https://doi.org/10.1002/ejoc.200300439>.
124. Rössle, M.; Werner, T.; Frey, W.; Christoffers, J. *Eur. J. Org. Chem.* **2005**, 2005 (23), 5031–5038. <https://doi.org/10.1002/ejoc.200500487>.
125. Tsubusaki, T.; Nishino, H. *Tetrahedron* **2009**, 65 (18), 3745–3752. <https://doi.org/10.1016/j.tet.2009.02.045>.
126. IsayamaShigeru; MukaiyamaTeruaki. *Chem. Lett.* **2006**. <https://doi.org/10.1246/cl.1989.573>.
127. Isayama, S. *Bull. Chem. Soc. Jpn.* **1990**, 63 (5), 1305–1310. <https://doi.org/10.1246/bcsj.63.1305>.
128. Tokuyasu, T.; Kunikawa, S.; Masuyama, A.; Nojima, M. *Org. Lett.* **2002**, 4 (21), 3595–3598. <https://doi.org/10.1021/ol0201299>.
129. Tokuyasu, T.; Kunikawa, S.; Abe, M.; Masuyama, A.; Nojima, M.; Kim, H.-S.; Begum, K.; Wataya, Y. *J. Org. Chem.* **2003**, 68 (19), 7361–7367. <https://doi.org/10.1021/jo030107f>.
130. Tokuyasu, T.; Kunikawa, S.; McCullough, K. J.; Masuyama, A.; Nojima, M. *J. Org. Chem.* **2005**, 70 (1), 251–260. <https://doi.org/10.1021/jo048359j>.
131. Wu, J.-M.; Kunikawa, S.; Tokuyasu, T.; Masuyama, A.; Nojima, M.; Kim, H.-S.; Wataya, Y. *Tetrahedron* **2005**, 61 (42), 9961–9968. <https://doi.org/10.1016/j.tet.2005.08.025>.
132. Chen, H.-J.; Wu, Y. *Org. Lett.* **2015**, 17 (3), 592–595. <https://doi.org/10.1021/ol503603t>.
133. Hu, X.; Musacchio, A. J.; Shen, X.; Tao, Y.; Maimone, T. J. *J. Am. Chem. Soc.* **2019**, 141 (37), 14904–14915. <https://doi.org/10.1021/jacs.9b08001>.
134. Li, Q.; Zhao, K.; Peuronen, A.; Rissanen, K.; Enders, D.; Tang, Y. *J. Am. Chem. Soc.* **2018**, 140 (5), 1937–1944. <https://doi.org/10.1021/jacs.7b12903>.
135. Kharasch, M. S.; Nudenberg, W.; Mantell, G. J. *J. Org. Chem.* **1951**, 16 (4), 524–532. <https://doi.org/10.1021/jo01144a005>.
136. Beckwith, A. L. J.; Wagner, R. D. *J. Am. Chem. Soc.* **1979**, 101 (23), 7099–7100. <https://doi.org/10.1021/ja00517a069>.
137. Barker, P. J.; Beckwith, A. L. J.; Fung, Y. *Tetrahedron Lett.* **1983**, 24 (1), 97–100. [https://doi.org/10.1016/S0040-4039\(00\)81337-4](https://doi.org/10.1016/S0040-4039(00)81337-4).

138. Beckwith, A. L. J.; Wagner, R. D. *J. Chem. Soc., Chem. Commun.* **1980**, No. 11, 485–486. <https://doi.org/10.1039/C39800000485>.
139. Bachi, M. D.; Korshin, E. E. *Synlett* **1998**, 1998 (2), 122–124. <https://doi.org/10.1055/s-1998-1587>.
140. Korshin, E. E.; Hoos, R.; Szpilman, A. M.; Konstantinovski, L.; Posner, G. H.; Bachi, M. D. *Tetrahedron* **2002**, 58 (12), 2449–2469. [https://doi.org/10.1016/S0040-4020\(02\)00126-6](https://doi.org/10.1016/S0040-4020(02)00126-6).
141. Szpilman, A. M.; Korshin, E. E.; Rozenberg, H.; Bachi, M. D. *J. Org. Chem.* **2005**, 70 (9), 3618–3632. <https://doi.org/10.1021/jo050074z>.
142. O'Neill, P. M.; Verissimo, E.; Ward, S. A.; Davies, J.; Korshin, E. E.; Araujo, N.; Pugh, M. D.; Cristiano, M. L. S.; Stocks, P. A.; Bachi, M. D. *Bioorganic & Medicinal Chem. Lett.* **2006**, 16 (11), 2991–2995. <https://doi.org/10.1016/j.bmcl.2006.02.059>.
143. Bachi, M. D.; Korshin, E. E.; Hoos, R.; Szpilman, A. M.; Ploypradith, P.; Xie, S.; Shapiro, T. A.; Posner, G. H. A. *J. Med. Chem.* **2003**, 46 (12), 2516–2533. <https://doi.org/10.1021/jm020584a>.
144. O'Neill, P. M.; Stocks, P. A.; Pugh, M. D.; Araujo, N. C.; Korshin, E. E.; Bickley, J. F.; Ward, S. A.; Bray, P. G.; Pasini, E.; Davies, J.; Verissimo, E.; Bachi, M. D. *Angew. Chem. Int. Ed.* **2004**, 43 (32), 4193–4197. <https://doi.org/10.1002/anie.200453859>.
145. Feldman, K. S.; Parvez, M. *J. Am. Chem. Soc.* **1986**, 108 (6), 1328–1330. <https://doi.org/10.1021/ja00266a051>.
146. Feldman, K. S.; Simpson, R. E. *J. Am. Chem. Soc.* **1989**, 111 (13), 4878–4886. <https://doi.org/10.1021/ja00195a047>.
147. Feldman, K. S.; Simpson, R. E. *Tetrahedron Lett.* **1989**, 30 (50), 6985–6988. [https://doi.org/10.1016/S0040-4039\(01\)93404-5](https://doi.org/10.1016/S0040-4039(01)93404-5).
148. Sun, X.-Y.; Tian, X.-Y.; Li, Z.-W.; Peng, X.-S.; Wong, H. N. C. *Chem. Eur. J.* **2011**, 17 (21), 5874–5880. <https://doi.org/10.1002/chem.201003309>.
149. Brown, H. C.; Midland, M. M. *J. Am. Chem. Soc.* **1971**, 93 (16), 4078–4080. <https://doi.org/10.1021/ja00745a060>.
150. Klement, I.; Lütjens, H.; Knochel, P. *Tetrahedron* **1997**, 53 (27), 9135–9144. [https://doi.org/10.1016/S0040-4020\(97\)00603-0](https://doi.org/10.1016/S0040-4020(97)00603-0).
151. Mata, S.; González, J.; Vicente, R.; López, L. A. *Eur. J. Org. Chem.* **2016**, 2016 (15), 2681–2687. <https://doi.org/10.1002/ejoc.201600393>.
152. Berkessel, A.; Vogl, N. Synthetic Uses of Peroxides. In *PATAI'S Chemistry of Functional Groups*; American Cancer Society, 2009. <https://doi.org/10.1002/9780470682531.pat0352>.
153. Gandhi, H.; O'Reilly, K.; Gupta, M. K.; Horgan, C.; O'Leary, E. M.; O'Sullivan, T. P. Advances in the Synthesis of Acyclic Peroxides. *RSC Adv.* **2017**, 7 (32), 19506–19556. <https://doi.org/10.1039/C6RA28489B>.
154. An, X.; Zha, Q.; Wu, Y. *Org. Lett.* **2019**, 21 (5), 1542–1546. <https://doi.org/10.1021/acs.orglett.9b00425>.
155. Li, Y.; Hao, H.-D.; Wu, Y. *Org. Lett.* **2009**, 11 (12), 2691–2694. <https://doi.org/10.1021/ol900811m>.
156. Groendyke, B. J.; Modak, A.; Cook, S. P. *J. Org. Chem.* **2019**, 84 (20), 13073–13091. <https://doi.org/10.1021/acs.joc.9b01979>.
157. Sakurai, S.; Kato, T.; Sakamoto, R.; Maruoka, K. *Tetrahedron* **2019**, 75 (2), 172–179. <https://doi.org/10.1016/j.tet.2018.11.048>.
158. Bloodworth, A. J.; Spencer, M. D. *J. Organomet. Chem.* **1990**, 386 (3), 299–304. [https://doi.org/10.1016/0022-328X\(90\)80002-H](https://doi.org/10.1016/0022-328X(90)80002-H).
159. Terent'ev, A. O.; Krylov, I. B.; Borisov, D. A.; Nikishin, G. I. *Synthesis* **2007**, 2007 (19), 2979–2986. <https://doi.org/10.1055/s-2007-990776>.
160. Stephenson, L. M. *Tetrahedron Lett.* **1980**, 21 (11), 1005–1008. [https://doi.org/10.1016/S0040-4039\(00\)78824-1](https://doi.org/10.1016/S0040-4039(00)78824-1).

161. Velluz, L.; Muller, G.; Mathieu, J.; Poittevin, A. *Tetrahedron* **1960**, 9 (3), 145–148.
[https://doi.org/10.1016/0040-4020\(60\)80002-6](https://doi.org/10.1016/0040-4020(60)80002-6).
162. Carless, H. A. J.; Batten, R. J. *Tetrahedron Lett.* **1982**, 23 (45), 4735–4738.
[https://doi.org/10.1016/S0040-4039\(00\)85700-7](https://doi.org/10.1016/S0040-4039(00)85700-7).
163. Boukouvalas, J.; Pouliot, R.; Fréchette, Y. *Tetrahedron Lett.* **1995**, 36 (24), 4167–4170.
[https://doi.org/10.1016/0040-4039\(95\)00714-N](https://doi.org/10.1016/0040-4039(95)00714-N).
164. Corey, E. J.; Nicolaou, K. C.; Shibasaki, M.; Machida, Y.; Shiner, C. S. *Tetrahedron Lett.* **1975**, 16 (37), 3183–3186. [https://doi.org/10.1016/S0040-4039\(00\)91450-3](https://doi.org/10.1016/S0040-4039(00)91450-3).
165. Salomon, M. F.; Salomon, R. G. *J. Am. Chem. Soc.* **1977**, 99 (10), 3500–3501.
<https://doi.org/10.1021/ja00452a051>.
166. Bloodworth, A. J.; Eggelte, H. J. *Tetrahedron Lett.* **1981**, 22 (2), 169–172.
[https://doi.org/10.1016/0040-4039\(81\)80179-7](https://doi.org/10.1016/0040-4039(81)80179-7).
167. Takahashi, K.; Kishi, M. *J. Chem. Soc., Chem. Commun.* **1987**, 10, 722–724.
<https://doi.org/10.1039/C39870000722>.
168. Takahashi, K.; Kishi, M. *Tetrahedron* **1988**, 44 (15), 4737–4746. [https://doi.org/10.1016/S0040-4020\(01\)86176-7](https://doi.org/10.1016/S0040-4020(01)86176-7).
169. Ghorai, P.; Dussault, P. H.; Hu, C. *Org. Lett.* **2008**, 10 (12), 2401–2404.
<https://doi.org/10.1021/ol800657m>.
170. Xu, C.; Schwartz, C.; Raible, J.; Dussault, P. H. *Tetrahedron* **2009**, 65 (47), 9680–9685.
<https://doi.org/10.1016/j.tet.2009.09.068>.
171. Schneider, M.-A.; Seifert, K. *Eur. J. Org. Chem.* **2017**, 2017 (45), 6739–6746.
<https://doi.org/10.1002/ejoc.201700922>.
172. Griesbeck, A. G.; Goldfuss, B.; Jäger, C.; Brüllingen, E.; Lippold, T.; Kleczka, M. *ChemPhotoChem* **2017**, 1 (5), 213–221. <https://doi.org/10.1002/cptc.201600041>.
173. Wang, Y.; Chen, B.; He, X.; Gui, J. *J. Am. Chem. Soc.* **2020**, 142 (11), 5007–5012.
<https://doi.org/10.1021/jacs.0c00363>.
174. Xu, X.-X.; Dong, H.-Q. *J. Org. Chem.* **1995**, 60 (10), 3039–3044.
<https://doi.org/10.1021/jo00115a019>.
175. Gemma, S.; Gabellieri, E.; Sanna Coccone, S.; Martí, F.; Taglialatela-Scafati, O.; Novellino, E.; Campiani, G.; Butini, S. *J. Org. Chem.* **2010**, 75 (7), 2333–2340.
<https://doi.org/10.1021/jo1001559>.
176. Gemma, S.; Martí, F.; Gabellieri, E.; Campiani, G.; Novellino, E.; Butini, S. *Tetrahedron Letters* **2009**, 50 (41), 5719–5722. <https://doi.org/10.1016/j.tetlet.2009.07.137>.
177. Barnych, B.; Vatéle, J.-M. *Tetrahedron* **2012**, 68 (19), 3717–3724.
<https://doi.org/10.1016/j.tet.2012.03.024>.
178. Hu, L.; Lu, X.; Deng, L. *J. Am. Chem. Soc.* **2015**, 137 (26), 8400–8403.
<https://doi.org/10.1021/jacs.5b05345>.
179. Dai, P.; Dussault, P. H. *Org. Lett.* **2005**, 7 (20), 4333–4335. <https://doi.org/10.1021/ol051407h>.
180. Nixon, J. R.; Cudd, M. A.; Porter, N. A. *J. Org. Chem.* **1978**, 43 (21), 4048–4052.
<https://doi.org/10.1021/jo00415a014>.
181. Bloodworth, A. J.; Loveitt, M. E. *J. Chem. Soc., Chem. Commun.* **1976**, No. 3, 94–95.
<https://doi.org/10.1039/C39760000094>.
182. Bloodworth, A. J.; Khan, J. A. *J. Chem. Soc., Perkin Trans. 1* **1980**, 0 (0), 2450–2457.
<https://doi.org/10.1039/P19800002450>.
183. Bloodworth, A. J.; Loveitt, M. E. *J. Chem. Soc., Perkin Trans. 1* **1978**, 0 (6), 522–530.
<https://doi.org/10.1039/P19780000522>.
184. Bloodworth, A. J.; Bothwell, B. D.; Collins, A. N.; Maidwell, N. L. *Tetrahedron Lett.* **1996**, 37 (11), 1885–1888. [https://doi.org/10.1016/0040-4039\(96\)00143-8](https://doi.org/10.1016/0040-4039(96)00143-8).

185. Bloodworth, A. J.; Korkodilos, D. *Tetrahedron Lett.* **1991**, 32 (47), 6953–6956.
[https://doi.org/10.1016/0040-4039\(91\)80453-D](https://doi.org/10.1016/0040-4039(91)80453-D).
186. Bloodworth, A. J.; Curtis, R. J. *J. Chem. Soc., Chem. Commun.* **1989**, No. 3, 173–175.
<https://doi.org/10.1039/C39890000173>.
187. Bloodworth, A. J.; Tallant, N. A. *Tetrahedron Lett.* **1990**, 31 (48), 7077–7080.
[https://doi.org/10.1016/S0040-4039\(00\)97247-2](https://doi.org/10.1016/S0040-4039(00)97247-2).
188. Kishali, N. (Horasan); Sahin, E.; Kara, Y. *Org. Lett.* **2006**, 8 (9), 1791–1793.
<https://doi.org/10.1021/ol060272s>.
189. Krabbe, S. W.; Do, D. T.; Johnson, J. S. *Org. Lett.* **2012**, 14 (23), 5932–5935.
<https://doi.org/10.1021/ol302848m>.
190. Das, P.; Hasan, M. H.; Mitra, D.; Bollavarapu, R.; Valente, E. J.; Tandon, R.; Raucher, D.; Hamme, A. T. *J. Org. Chem.* **2019**, 84 (11), 6992–7006. <https://doi.org/10.1021/acs.joc.9b00746>.
191. Harris, J. R.; Waetzig, S. R.; Woerpel, K. A. *Org. Lett.* **2009**, 11 (15), 3290–3293.
<https://doi.org/10.1021/ol901046z>.
192. Weitz, E.; Scheffer, A. *Chem. Ber.* **1921**, 54 (9), 2327–2344.
<https://doi.org/10.1002/cber.19210540922>.
193. Murakami, N.; Kawanishi, M.; Itagaki, S.; Horii, T.; Kobayashi, M. *Tetrahedron Lett.* **2001**, 42 (41), 7281–7285. [https://doi.org/10.1016/S0040-4039\(01\)01493-9](https://doi.org/10.1016/S0040-4039(01)01493-9).
194. Murakami, N.; Kawanishi, M.; Itagaki, S.; Horii, T.; Kobayashi, M. *Bioorg. Med. Chem. Lett.* **2002**, 12 (1), 69–72. [https://doi.org/10.1016/S0960-894X\(01\)00673-4](https://doi.org/10.1016/S0960-894X(01)00673-4).
195. Kawanishi, M.; Kotoku, N.; Itagaki, S.; Horii, T.; Kobayashi, M. *Bioorg. Med. Chem. Lett.* **2004**, 12 (20), 5297–5307. <https://doi.org/10.1016/j.bmc.2004.04.051>.
196. Yaremenko, I. A.; Vil', V. A.; Demchuk, D. V.; Terent'ev, A. O. *Beilstein J. Org. Chem.* **2016**, 12 (1), 1647–1748. <https://doi.org/10.3762/bjoc.12.162>.
197. Xu, C.; Raible, J. M.; Dussault, P. H. *Org. Lett.* **2005**, 7 (12), 2509–2511.
<https://doi.org/10.1021/ol050291m>.
198. Jin, H.-X.; Liu, H.-H.; Zhang, Q.; Wu, Y. *Tetrahedron Lett.* **2005**, 46 (34), 5767–5769.
<https://doi.org/10.1016/j.tetlet.2005.06.110>.
199. Jin, H.-X.; Zhang, Q.; Kim, H.-S.; Wataya, Y.; Liu, H.-H.; Wu, Y. *Tetrahedron* **2006**, 62 (33), 7699–7711. <https://doi.org/10.1016/j.tet.2006.05.065>.
200. Li, Y.; Zhang, Q.; Wittlin, S.; Jin, H.-X.; Wu, Y. *Tetrahedron* **2009**, 65 (34), 6972–6985.
<https://doi.org/10.1016/j.tet.2009.06.050>.
201. Guerra, F. M.; Zubía, E.; Ortega, M. J.; Moreno-Dorado, F. J.; Massanet, G. M. *Tetrahedron* **2010**, 66 (1), 157–163. <https://doi.org/10.1016/j.tet.2009.11.026>.
202. Barnych, B.; Vatéle, J.-M. *Org. Lett.* **2012**, 14 (2), 564–567. <https://doi.org/10.1021/ol203185f>.
203. Silva, E. M. P.; Pye, R. J.; Brown, G. D.; Harwood, L. M. *Eur. J. Org. Chem.* **2012**, 2012 (6), 1209–1216. <https://doi.org/10.1002/ejoc.201101477>.
204. Reisinger, C. M.; Wang, X.; List, B. *Angew. Chem. Int. Ed.* **2008**, 47 (42), 8112–8115.
<https://doi.org/10.1002/anie.200803238>.
205. Lifchits, O.; Mahlau, M.; Reisinger, C. M.; Lee, A.; Farès, C.; Polyak, I.; Gopakumar, G.; Thiel, W.; List, B. *J. Am. Chem. Soc.* **2013**, 135 (17), 6677–6693. <https://doi.org/10.1021/ja402058v>.
206. Milas, N. A.; Mageli, O. L.; Golubovic, Aleksandar.; Arndt, R. W.; Ho, J. C. J. *J. Am. Chem. Soc.* **1963**, 85 (2), 222–226. <https://doi.org/10.1021/ja00885a022>.
207. Rieche, A.; Bischoff, C.; Pulz, M. *Chem. Ber.* **1962**, 95 (8), 2005–2008.
<https://doi.org/10.1002/cber.19620950822>.
208. Howarth, J.; Wilson, D. *Bioorg. Med. Chem. Lett.* **2003**, 13 (12), 2013–2015.
[https://doi.org/10.1016/S0960-894X\(03\)00326-3](https://doi.org/10.1016/S0960-894X(03)00326-3).

209. Zdvizhkov, A. T.; Terent'ev, A. O.; Radulov, P. S.; Novikov, R. A.; Tafeenko, V. A.; Chernyshev, V. V.; Ilovaisky, A. I.; Levitsky, D. O.; Fleury, F.; Nikishin, G. I. *Tetrahedron Lett.* **2016**, 57 (8), 949–952. <https://doi.org/10.1016/j.tetlet.2016.01.061>.
210. Kandur, W. V.; Richert, K. J.; Rieder, C. J.; Thomas, A. M.; Hu, C.; Ziller, J. W.; Woerpel, K. A. *Org. Lett.* **2014**, 16 (10), 2650–2653. <https://doi.org/10.1021/ol500835f>.
211. Gemma, S.; Kunjir, S.; Sanna Coccone, S.; Brindisi, M.; Moretti, V.; Brogi, S.; Novellino, E.; Basilico, N.; Parapini, S.; Taramelli, D.; Campiani, G.; Butini, S. *J. Med. Chem.* **2011**, 54 (16), 5949–5953. <https://doi.org/10.1021/jm200686d>.
212. Gemma, S.; di Cerbo, L.; Relitti, N.; Vallone, A.; Brindisi, M.; Brogi, S.; Chemi, G.; Novellino, E.; Campiani, G.; Butini, S. *Tetrahedron Letters* **2018**, 59 (49), 4330–4333. <https://doi.org/10.1016/j.tetlet.2018.10.059>.
213. Gemma, S.; Kunjir, S.; Brindisi, M.; Novellino, E.; Campiani, G.; Butini, S. *Tetrahedron Letters* **2013**, 54 (10), 1233–1235. <https://doi.org/10.1016/j.tetlet.2012.12.075>.
214. D'Alessandro, S.; Alfano, G.; Di Cerbo, L.; Brogi, S.; Chemi, G.; Relitti, N.; Brindisi, M.; Lamponi, S.; Novellino, E.; Campiani, G.; Gemma, S.; Basilico, N.; Taramelli, D.; Baratto, M. C.; Pogni, R.; Butini, S. *Bioorganic Chemistry* **2019**, 89, 103020. <https://doi.org/10.1016/j.bioorg.2019.103020>.
215. Snider, B. B.; Shi, Z. *J. Am. Chem. Soc.* **1992**, 114 (5), 1790–1800. <https://doi.org/10.1021/ja00031a038>.
216. Dussault, P. H.; Eary, C. T.; Woller, K. R. *J. Org. Chem.* **1999**, 64 (6), 1789–1797. <https://doi.org/10.1021/jo981128q>.
217. Snider, B. B.; O'Neil, S. V. *Synth. Commun.* **1995**, 25 (7), 1085–1091. <https://doi.org/10.1080/00397919508012671>.
218. Wu, J.; Kadonaga, Y.; Hong, B.; Wang, J.; Lei, X. *Angew. Chem. Int. Ed.* **2019**, 58 (32), 10879–10883. <https://doi.org/10.1002/anie.201903682>.
219. Dussault, P. H.; Zope, U. R. *Tetrahedron Lett.* **1995**, 36 (13), 2187–2190. [https://doi.org/10.1016/0040-4039\(95\)00231-Z](https://doi.org/10.1016/0040-4039(95)00231-Z).
220. Windaus, A.; Brunken, J. *Liebigs. Ann.* **1928**, 460 (1), 225–235. <https://doi.org/10.1002/jlac.19284600113>.
221. Harwood, L. M.; Robertson, J.; Swallow, S. *Synlett* **1999**, 1999 (09), 1359–1362. <https://doi.org/10.1055/s-1999-2851>.
222. Yao, G.; Steliou, K. *Org. Lett.* **2002**, 4 (4), 485–488. <https://doi.org/10.1021/ol016943y>.
223. Jung, M.; Ham, J.; Song, J. *Org. Lett.* **2002**, 4 (16), 2763–2765. <https://doi.org/10.1021/ol026285x>.
224. Sun, L.; Shah, F.; Helal, M. A.; Wu, Y.; Pedduri, Y.; Chittiboyina, A. G.; Gut, J.; Rosenthal, P. J.; Avery, M. A. *J. Med. Chem.* **2010**, 53 (21), 7864–7868. <https://doi.org/10.1021/jm1006462>.
225. Shen, X.-Y.; Peng, X.-S.; Wong, H. N. C. *Org. Lett.* **2016**, 18 (5), 1032–1035. <https://doi.org/10.1021/acs.orglett.6b00161>.
226. Nicolaou, K. C.; Lu, M.; Totokotsopoulos, S.; Heretsch, P.; Giguère, D.; Sun, Y.-P.; Sarlah, D.; Nguyen, T. H.; Wolf, I. C.; Smee, D. F.; Day, C. W.; Bopp, S.; Winzeler, E. A. *J. Am. Chem. Soc.* **2012**, 134 (41), 17320–17332. <https://doi.org/10.1021/ja308429f>.
227. Nicolaou, K. C.; Totokotsopoulos, S.; Giguère, D.; Sun, Y.-P.; Sarlah, D. *J. Am. Chem. Soc.* **2011**, 133 (21), 8150–8153. <https://doi.org/10.1021/ja2032635>.
228. Liu, Y.-T.; Li, L.-P.; Xie, J.-H.; Zhou, Q.-L. *Angew. Chem. Int. Ed.* **2017**, 56 (41), 12708–12711. <https://doi.org/10.1002/anie.201706994>.
229. Lee, R. J.; Lindley, M. R.; Pritchard, G. J.; Kimber, M. C. *Chem. Commun.* **2017**, 53 (47), 6327–6330. <https://doi.org/10.1039/C7CC03229C>.
230. Mori, N.; Sakoda, D.; Watanabe, H. *Tetrahedron Lett.* **2017**, 58 (40), 3884–3886. <https://doi.org/10.1016/j.tetlet.2017.08.064>.

231. Cabrera-Afonso, M. J.; Lucena, S. R.; Juarranz, Á.; Urbano, A.; Carreño, M. C. *Org. Lett.* **2018**, 20 (19), 6094–6098. <https://doi.org/10.1021/acs.orglett.8b02515>.
232. Yoshida, M.; Miura, M.; Nojima, M.; Kusabayashi, S. *J. Am. Chem. Soc.* **1983**, 105 (20), 6279–6285. <https://doi.org/10.1021/ja00358a016>.
233. Miura, M.; Yoshida, M.; Nojima, M.; Kusabayashi, S. *J. Chem. Soc., Chem. Commun.* **1982**, No. 7, 397–398. <https://doi.org/10.1039/C39820000397>.
234. Dussault, P. H.; Lee, H.-J.; Liu, X. *J. Chem. Soc., Perkin Trans. 1* **2000**, 0 (17), 3006–3013. <https://doi.org/10.1039/B001391I>.
235. Dussault, P. H.; Liu, X. *Tetrahedron Lett.* **1999**, 40 (36), 6553–6556. [https://doi.org/10.1016/S0040-4039\(99\)01399-4](https://doi.org/10.1016/S0040-4039(99)01399-4).
236. Dussault, P. H.; Zope, U. *Tetrahedron Lett.* **1995**, 36 (21), 3655–3658. [https://doi.org/10.1016/0040-4039\(95\)00622-J](https://doi.org/10.1016/0040-4039(95)00622-J).
237. Dussault, P. H.; Lee, I. Q.; Lee, H.-J.; Niu, Q. J.; Schultz, J. A.; Zope, U. R. *J. Org. Chem.* **2000**, 65 (25), 8407–8414. <https://doi.org/10.1021/jo991714z>.
238. Ramirez, A.; Woerpel, K. A. *Org. Lett.* **2005**, 7 (21), 4617–4620. <https://doi.org/10.1021/ol051703u>.
239. Hurlocker, B.; Miner, M. R.; Woerpel, K. A. *Org. Lett.* **2014**, 16 (16), 4280–4283. <https://doi.org/10.1021/ol5020015>.
240. Xu, Z.-J.; Wittlin, S.; Wu, Y. *Chem. Eur. J.* **2017**, 23 (9), 2031–2034. <https://doi.org/10.1002/chem.201605871>.
241. Pinet, A.; Nguyen, T. L.; Bernadat, G.; Figadère, B.; Ferrié, L. *Org. Lett.* **2019**, 21 (12), 4729–4733. <https://doi.org/10.1021/acs.orglett.9b01616>.
242. Dussault, P. H.; Lee, H.-J.; Niu, Q. J. *J. Org. Chem.* **1995**, 60 (4), 784–785. <https://doi.org/10.1021/jo00109a001>.
243. Wang, X.; Dong, Y.; Wittlin, S.; Charman, S. A.; Chiu, F. C. K.; Chollet, J.; Katneni, K.; Mannila, J.; Morizzi, J.; Ryan, E.; Scheurer, C.; Steuten, J.; Santo Tomas, J.; Snyder, C.; Vennerstrom, J. L. *J. Med. Chem.* **2013**, 56 (6), 2547–2555. <https://doi.org/10.1021/jm400004u>.
244. Zhao, Q.; Vargas, M.; Dong, Y.; Zhou, L.; Wang, X.; Sriraghavan, K.; Keiser, J.; Vennerstrom, J. L. *J. Med. Chem.* **2010**, 53 (10), 4223–4233. <https://doi.org/10.1021/jm100226t>.
245. Wang, X.; Dong, Y.; Wittlin, S.; Creek, D.; Chollet, J.; Charman, S. A.; Santo Tomas, J.; Scheurer, C.; Snyder, C.; Vennerstrom, J. L. *J. Med. Chem.* **2007**, 50 (23), 5840–5847. <https://doi.org/10.1021/jm0707673>.
246. Martyn, D. C.; Ramirez, A. P.; Beattie, M. J.; Cortese, J. F.; Patel, V.; Rush, M. A.; Woerpel, K. A.; Clardy, J. *Bioorganic & Medicinal Chem. Lett.* **2008**, 18 (24), 6521–6524. <https://doi.org/10.1016/j.bmcl.2008.10.083>.
247. Gaschler, M. M.; Andia, A. A.; Liu, H.; Csuka, J. M.; Hurlocker, B.; Vaiana, C. A.; Heindel, D. W.; Zuckerman, D. S.; Bos, P. H.; Reznik, E.; Ye, L. F.; Tyurina, Y. Y.; Lin, A. J.; Shchepinov, M. S.; Chan, A. Y.; Peguero-Pereira, E.; Fomich, M. A.; Daniels, J. D.; Bekish, A. V.; Shmanai, V. V.; Kagan, V. E.; Mahal, L. K.; Woerpel, K. A.; Stockwell, B. R. *Nat. Chem. Biol.* **2018**, 14 (5), 507–515. <https://doi.org/10.1038/s41589-018-0031-6>.
248. Abrams, R. P.; Carroll, W. L.; Woerpel, K. A. *ACS Chem. Biol.* **2016**, 11 (5), 1305–1312. <https://doi.org/10.1021/acschembio.5b00900>.
249. Dussault, P. H.; Liu, X. *Org. Lett.* **1999**, 1 (9), 1391–1393. <https://doi.org/10.1021/ol990954y>.
250. Dussault, P. H.; K. Trullinger, T.; Cho-Shultz, S. *Tetrahedron* **2000**, 56 (47), 9213–9220. [https://doi.org/10.1016/S0040-4020\(00\)00894-2](https://doi.org/10.1016/S0040-4020(00)00894-2).
251. Pinet, A.; Figadère, B.; Ferrié, L. *Adv. Synth. Cat.* **2020**, 362 (5), 1190–1194. <https://doi.org/10.1002/adsc.201901145>.