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Review

Toxicity and remediation of pharmaceuticals and pesticides using metal oxides and carbon nanomaterials

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Abbreviations: CIP, chlorophenol; CMs, carbamates; CNTs, carbon nanotubes; CPF, chlorpyrifos; CYP450, Cytochrome P450; 2D, two-dimensional; 2,4-D, 2,4-dichlorophenoxyacetic acid; DC, desphenyl-chloridazon; DCIP, dichlorophenol; DFB, decafluorobiphenyl; DFC, diclofenac sodium; DFT, density functional theory; 2,4-DP, 2-(2,4-dichlorophenoxy)propionic acid; E, sorption energy; E_a, activation energy; EC50, maximal effective concentration; ECOSAR, ecological structure-activity relationship; EDA, electron donor-acceptor; Eg, energy gap; EG, ethylene glycol; F-g-CN, functionalized graphite carbon nitride; G, graphene; GAC, granular activated carbon; g-C₃N₄, graphitic carbon nitride; GO, graphene oxide; GO-IL, graphene oxide modified with imidazolium-based ionic liquid; GOS, graphene oxide sheet; QODs, graphene quantum dots; HA, humic acid; h-BN, hexagonal boron nitride; HO-MWCNT, hydroxylated multi-walled carbon nanotube; HSP, heat shock protein; IARC, international agency for research on cancer; IL, ionic liquid; IOMs, iron oxide minerals; IONPs, iron oxide nanoparticles; LEV, levofloxacin; MCPA, 4-chloro-2-methylphenoxyacetic acid; MCPP, methylchlorophenoxypropionic acid; MDC, methyl-desphenylchloridazon; MeNa, sodium methanolate; MWCNTs, multi-walled carbon nanotubes; NAC, NH₄Cl-modified activated carbon; NADPH, nicotinamide adenine dinucleotide phosphate; N-CNT, nitrogen-doped carbon nanotube; NOR, norfloxacin; NSGO, nanosheet graphene oxide; OD, outer diameter; OFLX, ofloxacin; O-MWCNT, oxidized multi-walled carbon nanotube; OPAC, orange peel activated carbon; PAC, powdered activated carbon; PEC, predicted environmental concentration; PEF, pefloxacin; PEG, polyethylene glycol; PGO, porous graphene oxide; PNEC, predicted no-effect concentration; PNT, phenacetin; POPs, persistent organic pollutants; PPCPs, personal care products; PRO, propranolol; PZC, point of zero charge; q_{max}, maximum adsorption capacity; RRIP, recycled rusted iron particles; ROS, reactive oxygen species; SAC, standard activated carbon; S_{BET}, specific surface area; SPIONP, superparamagnetic iron oxide nanoparticle; SWCNTs, single-walled carbon nanotubes; T, temperature; 2,4,5-T, 2,4,5-trichlorophenoxyacetic acid; TCIP, trichlorophenol; TDPs, trace drugs and pesticides; TSAC, tangerine seed activated carbon; WHO, world health organization; WSAC, walnut shell activated carbon.

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- Water contamination by drugs/pesticides led to environmental issues.
- Metal oxides/carbon-based sorbents used for drugs/pesticides removal.
- Residual drugs in the human body cause serious issues e.g., genetic disorders, etc.
- Presence of pesticides in the body leads to Parkinson's disease and breast cancer.
- Chemisorption and physisorption are main mechanisms for drugs and pesticides removal.

A B S T R A C T

The worldwide development of agriculture and industry has resulted in contamination of water bodies by pharmaceuticals, pesticides and other xenobiotics. Even at trace levels of few micrograms per liter in waters, these contaminants induce public health and environmental issues, thus calling for efficient removal methods such as adsorption. Recent adsorption techniques for wastewater treatment involve metal oxide compounds, e.g. Fe_2O_3 , ZnO , Al_2O_3 and ZnO-MgO , and carbon-based materials such as graphene oxide, activated carbon, carbon nanotubes, and carbon/graphene quantum dots. Here, the small size of metal oxides and the presence various functional groups has allowed higher adsorption efficiencies. Moreover, carbon-based adsorbents exhibit unique properties such as high surface area, high porosity, easy functionalization, low price, and high surface reactivity. Here we review the cytotoxic effects of pharmaceutical drugs and pesticides in terms of human risk and ecotoxicology. We also present remediation techniques involving adsorption on metal oxides and carbon-based materials.

Keywords:

Human risk
Pharmaceutical
Pesticides
Metal oxides
Carbon nanostructures

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1. Introduction

Population growth, overexploitation of finite water supplies, and contamination by human, agricultural and industrial practices

will lead to a future water shortage if advanced techniques are not rapidly designed to clean water (Márquez et al., 2014). By 2050, water and food demand are predicted to rise by 50% and 70%,

respectively, from current levels (Márquez et al., 2014). The shortage of high-quality water supplies has already resulted in the expansion of wastewater treatment and recycling systems to ensure that treated water is healthy for reuse (Kidak and Doğan, 2018). Over the last decade, the concentration of organic pollutants such as drugs and pesticides has considerably increased, from nanograms to micrograms per liter in urban wastewater, freshwater, and drinking. Contaminants often originate primarily from municipal, hospital, veterinary, and farm wastewater. The growing water pollution is thus of great concern for human health (Kidak and Doğan, 2018; Márquez et al., 2014; Nikolaou et al., 2007).

According to the World Health Organization (WHO) report at 2012, the concentrations of pharmaceuticals in surface water, groundwater, and partially treated water are typically less than $0.1 \mu\text{g L}^{-1}$, and concentrations in the treated water are generally below $0.05 \mu\text{g L}^{-1}$ (Organization, 2012). The widespread exposure to medications and pesticides has raised concerns about their adverse effects on the environment and human health. Residues re-enter humans and animals' bodies from various water resources and cause adverse effects such as drug resistance and metabolism disruption in the body (Zhang, 2003). Therefore, removing pharmaceuticals from wastewater resources is a pressing issue to safeguard the environment.

Traces of drugs and pesticides in various water resources indicate the low efficiency of traditional systems of water treatment such as filtration, reverse osmosis, and direct contact membrane distillation. Moreover, some pollutants such as diclofenac, paraben, carbamazepine, and diazepam are not fully biologically removable by fungi, bacteria and activated sludge (Dehaghi et al., 2014). Various techniques such as adsorption (Ahmadijokani et al., 2020a, 2020b, 2020c, ndAhmadijokani et al., 2020a, n.d.; Ahmed et al., 2017), reverse osmosis (Rodriguez-Mozaz et al., 2015), air stripping (Zareei and Ghoreyshi, 2011), and biological methods (Ahmed et al., 2017; Carboneras et al., 2018; Goswami et al., 2018; Marican and Duran-Lara, 2018; Saleh et al., 2020) have been deployed widely to remove pollutants, yet adsorption appears promising for several reasons (Ahmad et al., 2010). For instance, adsorption surpasses other water treatment strategies because of the lower initial price, higher efficiency, simplicity of layout, and lower maintenance. Furthermore, adsorption does not contribute to the creation of harmful compounds (Ahmad et al., 2010).

Enhanced adsorption can be accomplished using metal oxides, which as crystalline materials that possess a metal cation and an oxide anion, e.g. MgO , Al_2O_3 , Fe_2O_3 , Fe_3O_4 , CuO , and ZnO (Gupta et al., 2007; Srivastava et al., 2020). Indeed, metal oxide-based adsorbents bear functional groups, e.g., $-\text{OH}$, $-\text{C}=\text{O}$, and $-\text{COOH}$, and active sites, which can adsorb a wide range of contaminants (Batzill, 2011; Mahmoud et al., 2015; Wu et al., 2016; Bagbi et al., 2017; Li et al., 2016). Carbon-based materials as adsorbents are also under active research because these materials have large surface area, high porosity and surface reactivity, and they can be easily functionalized (Y. Wang et al., 2019; Nazal, 2020). Carbon-based materials such as graphene oxide (Umbreen et al., 2018), activated carbon (Alves et al., 2018), carbon nanotubes (Wang et al., 2019a,b,c), quantum carbon dots (Koe et al., 2020), and graphene (Ali et al., 2019b), are considered the most effective adsorbents for the removal of pollutants, e.g., dyes and pesticides.

The high adsorption capacity of carbonaceous materials has been applied to treat organic pollutants such as drugs, dyes, pesticides and odors (I. Ali et al., 2019b; Umbreen et al., 2018) and for chemical transformations (Varma, 2019). Their unusual physical and chemical properties include exceptionally high tensile strength, high young modulus, good chemical and thermal stability, and excellent thermal and electrical conductivity, thus rendering

carbonaceous materials ideal candidates for adsorption applications (I. Ali et al., 2019b; Umbreen et al., 2018). Dissolved organic pollutants are removed from water by water-insoluble carbon-based adsorbents. The adsorption capacity of carbon-based adsorbents depends on porosity, chemical properties and adsorbent durability (Rafati et al., 2018; Shaker and Yakout, 2016; Srivastava et al., 2020). Here we review cytotoxicity effects of drugs and pesticides on humans, and then present the use of metal oxide and carbon nanostructure as efficient adsorbents to remove pesticides and drugs from aqueous media.

2. Cytotoxic effects of drugs and pesticides in water

The presence of pesticides and drugs is one of the factors affecting the health of aquatic environments. The major sources, environmental pathways, and toxicity of active pharmaceutical ingredients and pesticides are shown in Fig. 1S (Supplementary file), (Seo et al., 2005). These contaminants change the quality of water resources via their transit from the land to adjacent rivers or water bodies. They are then absorbed by aquatic plants and reach human bodies through the food chain. This pollution transfer cycle causes antagonistic or synergistic effects with acute and long-term results (Bianchi et al., 2017; Hartz et al., 2019; Wolfram et al., 2019). Due to the importance of serious problems caused by the excessive use of these materials, the following sections describe the main impacts of drugs and pesticides on human health.

2.1. Ecotoxicity of drugs and pesticides

In recent years, the use of drugs and pesticides has annually increased, and their abuse has become more rampant. These conditions cause drugs and pesticides or their metabolites to persist in the environment as they are relatively stable in the environment and are difficult to control using conventional methods.

2.1.1. Drugs

To date, most studies on the residual effects of drugs on human life are focused on drugs used in large amounts, like non-steroidal anti-inflammatory, β -blockers, fat regulators, and to some extent, psychiatric drugs (Touraud et al., 2011). These lingering drugs illustrate the toxic effects that cause genetic disorderliness, sickness, and cell death. Unhealthy lifestyle behaviors and curative medicines are factors that negatively affect prenatal development and increase the children's susceptibility to diseases. The toxicological mechanisms of chemicals and lifestyle factors include the generation of reactive oxygen species and oxidative cell injury (Al-Gubory, 2014; Gray et al., 2013). In eco-toxicological terms, dissolved amounts of the drug can be assessed by human blood plasma analysis as an elementary valuation of the eco-toxicological authority of a substance. Then, these results can be compared with blood plasma levels of mammalian in aquatic creatures. Comparative data may reveal whether chronic exposure is required for evaluation in specific long-term trials or not (Länge and Dietrich, 2002). Following the European Directive EC 93/67/EEC (European Commission, 1993), chemical classification is performed based on their EC_{50} values in three categories; $\text{EC}_{50} < 1 \text{ mg L}^{-1}$, $1\text{--}10 \text{ mg L}^{-1}$, and $10\text{--}100 \text{ mg L}^{-1}$ are considered very toxic, toxic, and harmful, respectively, to aquatic organisms (Grabarczyk et al., 2020).

Environmental drug contamination is also found in various states, such as sewage sludge in the ground, leaching from landfills, or irrigating farmable land with treated/untreated effluent in the ground (Kümmerer, 2008). The production and formulation of drugs and their use by patients have caused their improper disposal in the aquatic environment. They mainly enter the aquatic milieu in

low ($< \text{ng L}^{-1}$) or high concentrations (mg L^{-1}) through human excretion, industrial effluents, and recipient streams with remarkable adverse effects on wildlife and human life (aus der Beek et al., 2016; Boxall et al., 2012; Larsson, 2014). The limited data on measured environmental concentration for bromazepam coincide with a confined eco-toxicological profile. The chronic data on fishes, daphnids, and green microalgae were investigated using the ecological structure-activity relationships-class program (ECOSAR), and the values obtained were 174, 6458, and $2995 \mu\text{g L}^{-1}$, respectively. For eco-toxicological assays of bromazepam, the behavior of the fish *Danio rerio* was evaluated and demonstrated the reduction of shoal cohesion. Citalopram with a concentration of $4000 \mu\text{g L}^{-1}$ presented chronic toxicity for *Ceriodaphnia dubia* and *Daphnia similis* during an 8-day experiment (Cunha et al., 2019).

To undertake the environmental risk assessment, the drug concentration in the natural world and the lowest drug concentration with side effects are often examined. Prediction of effects of chemicals parameter on flora and fauna can be ascertained by quantitative structure-activity relationships or common ecotoxicity experiments (Johnson et al., 2008). Consequences for the children include cranial and facial abnormalities and digitalis, along with growth retardation. The required experiments for conducting a robust environmental hazard evaluation of drugs are established by subjecting fishes to genotoxic xenobiotics; effects can increase cancer occurrence, DNA damage, and nuclear abnormalities (Johnson et al., 2008; Priyan et al., 2020).

Anti-cancer drugs mostly induce cytotoxic/genotoxic effects and may damage DNA synthesis and alter the mechanism of endocrine glands regulating tumor growth. Hence, they are considered a threat. There are currently no specific guidelines on how to assess the residual drug risk for human health. However, to evaluate the risk of indirect exposure to drugs, data on the toxicity of drug components are applied to calculate the acceptable daily intake. These acceptable daily intake values are thought to have no pharmacological or toxicological predictive figures but are applied for the predicted no-effect concentrations of human/environmental vulnerability. The predicted no-effect concentrations values can be compared with the predicted environmental concentrations using evaluation patterns (Touraud et al., 2011). The risk ratio of predicted environmental concentrations to predicted no-effect concentrations for human health is presented in Table 1S (Supplementary file), which includes the therapeutic and generic names of some drugs and the acceptable daily intake for drinkable water or fish-eating in humans, especially children whose acceptable daily intake are lower than adults. Antibiotics for bacteria control in humans and animals can potentially be hazardous when entering the terrestrial environment. Antibiotic-rich fertilizers used as a source of nutrients for plants raise concerns about the effects of antibiotics on plant growth, soil enzyme/fauna activities, and nutrient cycles. These antibiotics can directly affect the animals and plants in the soil via the ensuing toxicity or indirectly affect the availability of nutrients by altering the microfauna and microflora (Kumar et al., 2005).

One of the discrete pharmacokinetic processes is drug metabolism. In biological drug transfer, lipophilic centers convert to hydrophilic centers to help assist in expelling drugs from the body. Biotransmission is the metabolic degradation of xenobiotics through a specific enzyme process, e.g., the CYP450 family. This process is vital because the drugs with a lipophilic class stay longer in the body. The prolonged shelf life of the drug may lead to poisoning. The well-known pathways of drug metabolism are shown in Fig. 2S (Supplementary file) (De Groot, 2006; Jaladanki et al., 2020).

CYP450 manages the metabolic reactions of drugs, wherein the reactions impact some special atoms of the molecules depending

on the electronic factors. Of note, CYP450 controls the entry/exit channel and the catalytic site. The catalytic region of cytochromes comprises three sections of the base, wall, and dome. The drug situations inside the cavity are principally defined based on the chemical characteristics of the dome section (Fig. 3a–S) (Supplementary file); the cavity walls of CYP450 are hydrophobic. The related walls conduct the hydrophobic functions of drugs toward the cavity of Fe-porphyrin. Moreover, this location of drugs is assisted by the amino acids existing in the arch section. Generally, the CYP450 enzyme participates in a catalytic cycle of eight stages (Fig. 3b–S) (Supplementary file):

- Stage 1: replacing the CYP450 enzyme with a water molecule for binding to the substrate.
- Stage 2: reducing Fe(III) to Fe(II) to provide an electron by redox proteins (NADPH-CYP450 reductase).
- Stage 3: binding of O_2 with Fe(II) to electron transfer from Fe(II) to O_2 .
- Stage 4–5: enzyme-substrate complexation for taking another electron from redox proteins (cytochrome b5).
- Stage 6–7: addition of H^+ to complex. This complex leads to the failure of O–O bond and escape of H_2O to create Cpd I. Subsequently, Cpd I withdrew H^+ from the substrate to set up a radical substrate.
- Stage 8: The oxygen and hydrogen are transferred to the radical substrate and finally release heme iron as Fe(III). The general catalytic cycle is shown in (Fig. 3b–S) (Supplementary file):

2.1.2. Pesticides

Pesticides are biologically converted to more water-soluble metabolites upon absorption by the body. The biological transfer rate can be fast for organophosphate insecticides (hours to days), or very gradual for organochlorine insecticides, from decades to years. The purpose of biotransformation is essentially detoxification and elimination. The latter event caused the bio-agglomeration of these lipophilic compounds in fatty contexture. Several mechanisms, such as oxidative stress and receptor-mediated mechanisms, are presumed to induce carcinogenicity by pesticides. For oxidative stress, the cytochrome P450 (CYP) catalyzes the normal oxidation of pesticides and produce hydroxylated metabolites (Mechanism 1). CYP450s lead to the oxidative metabolism of organochlorines and polychlorinated biphenyls; hence, the generation of superoxide (Mechanism 2). Other mechanisms including inflammatory and aberrant epigenetic, are in the development stage of the study. Epigenetic rectifications of tumor suppressor genes and their on-cogenes altering their tumor expression can also be introduced as molecular stimuli of pathogenesis during cancer progression and progression stages (Banerjee et al., 2001). To evaluate the toxicity and operational mechanisms of pesticide rotenone, tebufenpyrad, and fenazaquin, dose-dependent cell death was studied in the period of exposure 24–48 h. Exposure of cells with various doses of pesticides showed a decrease in ATP and superoxide generation. In other mechanisms, redox-active pesticide metabolites, viz. quinones, or bipyridinium compounds, generate superoxide. Moreover, the electron transport cascades in mitochondria were impaired, producing an excess of superoxide flux. Nicotinamide adenine dinucleotide phosphate (NADPH) oxidase can be activated by pesticides that liberate superoxides (Banerjee et al., 2001; Green et al., 2008; Sherer et al., 2007). The oxidative equations pertaining to CYP and its inefficacious cycling for superoxide production are depicted in Fig. 4S (Supplementary file).

Pesticides may boost the incidence of Parkinson's disease through disruption of mitochondrial function (Sherer et al., 2007). 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine induces Parkinson's

disease) with symptoms such as dopaminergic degeneration, α -synuclein aggregation, high oxidative damage, and α -synuclein aggregation by inhibition of complex I. Dose-dependent cell death of human neuroblastoma cell line (SK-N-MC) neuroblastoma cells, exposed to pesticide treatment, exhibited a rank order toxicity: pyridaben > rotenone > fenpyroximate > fenazaquin > tebufenpyrad. The most toxic compound of pyridaben illustrated the significant toxicity at a concentration of 10 pmol L⁻¹ (picomoles per liter), while the toxicity of tebufenpyrad and fenazaquin was attained at a concentration of 1 M (Sherer et al., 2007). Also, pesticides can cause stinging eyes, rashes, blisters, blindness, nausea, dizziness, diarrhea and death. Examples of known chronic effects are cancers, birth defects, reproductive harm, neurological and developmental toxicity, immunotoxicity, and disruption of the endocrine system (Hallenbeck and Cunningham-Burns, 2012).

Although pesticides are not genotoxic by themselves, their binding efficiency to steroid and xenobiotic receptors may lead to creating alterations in gene expression programs. Pesticide-induced cancers appear to be caused by effects on biomarkers, genetic sensitivity, oxidative stress, and DNA lesion (Gray et al., 2013). Based on the available epidemiological evidence, there is a link between pesticide exposure and cancer. Alteration by pesticides may damage DNA and contribute to cancer mutagenic mechanisms. Many xenochemicals can block or mimic the hormone function by attaching the endogenous ligands to the steroid nuclear receptor family and displacing them, thereby activating the function of the receptor, leading to a change in gene expression (Kretschmer and Baldwin, 2005; Schug et al., 2011). Moreover, insecticides can stress midgut cells, and heat shock protein 70 (HSP70) can eliminate this damage. Major damage of morphological alterations was precluded through the amplified expression of this stress protein. The localization of HSP70 may be related to areas where detoxification is performed (de Souza et al., 2019). Leading pesticide compounds and their action mechanism on body organs are summarized in Table 2S (Supplementary file).

There is evidence that persistent organic pollutants are related to type 2 diabetes. Inflammation in fatty organs, lipotoxicity in living organs such as pancreas, liver, muscle, and mitochondrial abnormality or impairment leads to the obesity and type 2 diabetes epidemic's initial mechanism. Persistent organic pollutants contaminate the food materials and our environments; hence, they can increase human's adipose tissue after accumulation in the tissue. Persistent organic pollutants with continuous release into circulation can reach critical organs and induce chemical-specific or physiological responses. The low doses of persistent organic pollutants induce three phases (viz., phase I, phase II, and phase III xenobiotic metabolism pathways) that enhance the excretion of persistent organic pollutants. Phase II leads to chronically decreasing inconsumption of mitochondrial function. This decrease in glutathione wreck the mitochondrial function and the reduction is acutely dependent on the inflammation and lipid accumulation of extrauterine. On the other hand, an increased amount of persistent organic pollutants mixtures can enhance proteins' synthesis with cytoprotective and restorative functions. This activation may ameliorate mitochondrial function and theoretically reduce the risk of type 2 diabetes. Therefore, U-shaped inverted associations can be expected. In other words, persistent organic pollutants can induce a dual effect on adipose (i.e., alteration in hyperplasia and hypertrophy) by various endocrine disrupting mechanisms depending on the type or dose of POPs (Han et al., 2020; Lee et al., 2014; Mansouri and Reggabi, 2020). Fig. 5S (Supplementary file) presents the overview of correlations of persistent organic pollutants and type 2 diabetes and their conceivable mechanisms.

3. Environmental and human risks of metal oxides and carbon-based nanoparticles

The success of nanotechnology is due to the unique properties of nanoparticles. Although nanoparticles have numerous advantages in various arena of the scientific world and in the daily life of human communities (Makvandi et al., 2020; C.Y. Wang et al., 2020), they may also have toxic effects on the plant, animals, and humans. A large quantity of nanoparticles has recently entered the environment because of the growing applications and development. Humans and animals are exposed to the nanoparticles through inhalation, swallow, and skin uptake, and after nanoparticles entry, they can damage the cells and ultimately the organs by complicated mechanisms (Sengul and Asmatulu, 2020). However, toxicity of metal oxide nanoparticles and carbon-based nanomaterials is not well known and is still being assayed for potential ecological and human health hazards. According to the obtained results from *in-vivo* and *in-vitro* assays, the cytotoxicity and gene toxicity of metal oxide nanoparticles on animals is related to the production of reactive oxygen species (ROS) and oxidative stress that can lead to lipid peroxidation, inflammation induction, animals' organs damage, the nervous system interference, DNA damage, and affect offspring (Srivastava et al., 2020; Zhu et al., 2019). There are few reports on the toxicity of metal oxide nanoparticles to human's body (Djurišić et al., 2015; Schneider et al., 2017), especially the influences of metal oxide nanoparticles on oral cells, mucosal cells, so these toxic influences on human life should not be ignored, and more study in this field is required in the future.

The industrial usage and widespread application of carbon-based nanomaterials have caused a rapid increase in their production, so the anxiety about their harmful effects on the environment and human is raised. Carbon-based nanomaterials have possible long-term ecological consequences, while the ecological and human health dangers of these nanomaterials have not been systematically investigated. Cytotoxicity towards animal, algae, and bacteria cells has been observed for graphene oxide, reduced graphene oxide (Shareena et al., 2018), graphene quantum dots (Hashemi et al., 2020), and carbon nanotubes chiefly multi-walled carbon nanotube.

The graphene oxide cytotoxic effect to microbial communities was confirmed at 50–300 mg L⁻¹, and there was no substantial decrease in viability of human cells after exposure to the non-cytotoxic dosage of 15 µg mL⁻¹ for graphene oxide (100 nm) and graphene quantum dots (50 nm) (Hashemi et al., 2020). The considerable alterations in the expression level of Bax, Bcl2, miR-21, miR-29a, and PTEN genes, and changes in mitochondrial activity at cellular level were observed after treatment with non-cytotoxic dosage. The basal level of genes and mitochondrial membrane potential were more influenced by the graphene oxide in compared to graphene quantum dots. The miRNA alteration can alter the fate of cells, and all these genes are involved in the development and metastasis of breast tumor. Thus, the secondary toxic influence of graphene oxide-100 and graphene quantum dots-50 at a non-toxic dosage was confirmed on both, the cellular and molecular level.

The carcinogenic risks of carbon nanotubes were published by the International Agency for Research on Cancer (IARC) in 2014 (Kobayashi et al., 2017). The carcinogenicity effect of single wall carbon nanotube is less than multi wall carbon nanotube for humans (Heller et al., 2020), and the suspected carcinogenic effects of single wall carbon nanotube (>55%; diameter < 2 nm and length = 5–15 µm) was reported by both IARC and chemical secretariat monographs. Exposure to multi wall carbon nanotubes stimulate local and systemic inflammation, contributing to formation of the pre-metastatic and metastatic niches, along with

damaging the central nervous system (Falahian et al., 2018) and promoting the metastatic cascade of breast cancer (Lu et al., 2019). The detrimental health influences in animal studies comprise cardiac inflammation, acute and chronic respiratory damage, and cancer such as mesothelioma (Ellenbecker et al., 2018). The long-term sustainability of carbon nanotubes for water treatment is questionable without the nano-safety guidelines, and so the current risk measurements of carbon nanotubes are overshadowed by uncertainties (Das et al., 2018).

Despite the intrinsic cytotoxicity of metal oxides and carbon-based nanomaterials as described above, abundant outstanding features of these nano-adsorbents make their application in the water treatment field extremely attractive. As a result, the topic need to address how to make the useful appliance of nanoparticles in terms of benefits while decreasing their toxic impact on the environment and human life.

4. Adsorption process and the adsorbents

The term adsorption, initially introduced by Kayser in 1881 (Dąbrowski, 2001), is a physicochemical process that typically happens in the solid-liquid form. Although adsorption is fully efficient in both liquid-liquid and gas-liquid phases, it is primarily used to treat water in solid-liquid phases. Adsorption is the mechanism by which molecules of a liquid or gaseous material are bound to a solid adsorbent (Dąbrowski, 2001; del Mar Orta et al., 2019; Derylo-Marczewska et al., 2017) and this process requires the aggregation of materials at the interface between the two phases. Essentially, the adsorption of molecules by internal or exterior surfaces of a solid could be termed as adsorption. The substance taken out of the liquid phase is called the adsorbate, and the material providing the adsorption surface is called the adsorbent. The adsorption process can be classified into three groups: physisorption, chemisorption, and ion exchange. Van der Waals and covalent forces are the main forces that control physical and chemical adsorption, respectively (related to the bonding between the adsorbent and the adsorbent) (Crini et al., 2018). Several factors like the pH of the aqueous solution, the sorbent dosage, the contact time, the concentration of the main substances, and the temperature hugely influence the adsorption of pollutants. The optimization of these factors plays a significant role in the adsorption of contaminants (Ali et al., 2019a), which is invariably accompanied by the release of energy. The heat released from the physical and chemical adsorption is between 5–10 and 10–100 kcal mol⁻¹, respectively (Ali et al., 2019a; Crini et al., 2018).

Adsorbents play an important role in the elimination of water contaminants and their efficiency depends on the structural features. The important properties of good adsorbent comprise biocompatibility, non-toxicity, readily available, simple renewability, and biodegradability (Nasrollahzadeh et al., 2021a). They must have great adsorption capacity due to the high surface area, along with good thermal stability and high abrasion resistance. Adsorbent compounds can be organic, mineral, or biological, and they are usually employed in the form of spherical, rod, wire, tube, or particle with sizes ranging from nanometer to millimeter (Singh et al., 2018). The adsorbents are generally categorized into three classes: (I) polar and hydrophilic oxygen-containing materials such as metal oxides (Gusain et al., 2019), (II) non-polar and hydrophobic carbon-based materials such as activated carbon, graphite, graphitic carbon nitride, graphene, carbon nanotubes, carbon nanofibers, fullerenes, and nanodiamonds (Nasrollahzadeh et al., 2021b), and (III) polar or non-polar polymer-based materials (Ismail et al., 2020). A great deal of effort has been expanded over the last few years on the usage of various adsorbents for water/wastewater purification (Srivastava et al., 2020; Zare et al., 2018).

4.1. Metal oxides

Metal oxide nanoparticles exhibit significant potential for wastewater remediation due to their low toxicity, thermal stability, large surface area, porous structures, easy recovery, and the presence of Lewis acid-base sites in their structures (Nagpal and Kakkar, 2019).

4.1.1. Iron oxides

Iron oxide nanoparticles, in the pure, doped, and composite forms, have been widely used for the adsorption of organic pollutants (D'Cruz et al., 2020; Shrivastava et al., 2019). Magnetic nanoparticles, including hematite (α -Fe₂O₃), maghemite (γ -Fe₂O₃), and magnetite (Fe₃O₄), were commonly deployed for the elimination of contaminants from aqueous media (Bhateria and Singh, 2019). The reason for the popularity of magnetic nanoparticles as an adsorbent is their easy separation via an external magnetic field and the reduced energy consumption and costs in the recycling process; the use of magnetic nanoparticles in the adsorption of organic pollutants has been highlighted (Bhateria and Singh, 2019). For example, superparamagnetic iron oxide nanoparticles (SPIONP, Fe₃O₄) have been synthesized via the co-precipitation procedure and used for the sorption of levofloxacin (LEV) drug from aqueous solutions (Al-Jabari et al., 2019). The maximum removal percentage of levofloxacin by superparamagnetic Fe₃O₄ was achieved at pH ~6.5, which indicated that the affinity of neutral/zwitterion form of levofloxacin onto the adsorbent is greater than their cationic and anionic counterparts. Based on these data, a plausible mechanism was proposed, which is schematically depicted in Fig. 1.

Modified iron oxide nanoparticles have good potential for the wastewater treatment process as has been exemplified in the rapid green synthesis of magnetic nano-sorbents by using extracts such as *Nelumbo nucifera* stalks, *Colocasia esculenta* corms, and *Piper betle* leaves; they provide an efficient, facile, and environmentally friendly route for the modification of nanoparticles (Misra et al., 2018), and as illustrated, in the good adsorption efficiency for carbamazepine.

Also, the green synthesized iron oxide nanoparticles (IONPs; Fe₃O₄) using an extract of *Excoecaria cochinchinensis* leaves (Lin et al., 2018) were applied for the elimination of rifampicin (RIF) from aqueous media (Fig. 2) (Cai et al., 2019); isotherm and kinetic results demonstrated the monolayer chemisorption of rifampicin onto IONPs with a q_{max} of 84.8 mg g⁻¹. The adsorption mechanism of rifampicin onto Fe₃O₄ nanoparticles at optimum pH 5.5 comprised electrostatic attraction and chemisorption between active sites of Fe₃O₄ (–COOH, –COH, and C π electron) with functional groups of the rifampicin molecules.

A greener protocol for the preparation of crystalline hexagonal hematite nanoparticles has been reported using the supernatant culture of a newly isolated bacterium, *Bacillus cereus* SVK1 (Rajendran et al., 2015). The applicability of eco-friendly bio-synthesized hematite nanoparticles was explored for the adsorptive removal of carbamazepine from water (Rajendran and Sen, 2018). The main reason for high removal percent (>90%) is believed to be related to the hydrogen bonding between amide groups of carbamazepine and the functional groups on hematite.

Modified nanoparticles with functional groups increase the surface functionality along with the surface wettability and stability (Gao and Yan, 2012). Indeed, the hydrophobic, electrostatic, van der Waals, π - π interactions and hydrogen bonding of adsorbent functional groups cause the adsorption of organic contaminants onto the modified metal oxides. For example, the magnetic sorbent (103-HC-DS/Fe-5) was synthesized via modification of hydrocalumite-iron oxide with dodecyl sulfate to remove agricultural chemicals, atrazine (q_{max} = 4.5 mg g⁻¹) and chlorpyrifos

Fig. 1. (a) pH-dependent chemical structures of levofloxacin (LEV), (b) effect of pH on levofloxacin adsorption, (c) effect of initial concentration on levofloxacin adsorption, and (d) plausible mechanism for levofloxacin adsorption onto superparamagnetic iron oxide nanoparticles at optimal conditions (pH = 6.5, levofloxacin concentration = 20 ppm, time = 240 min, and temperature = 298 K). Reproduced with modification with permission from (Al-Jabari et al., (2019)).

Fig. 2. Green preparation of iron oxide nanoparticles (IONPs), the adsorption of rifampicin (RIF) onto IONPs at optimal conditions (pH = 5.5, RIF concentration = 20 mol L⁻¹, time = 240 min, and temperature = 298 K), and representation of adsorption mechanism. Reproduced with modification with permission from (Cai et al., (2019)). (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)

($q_{\max} = 72.9 \text{ mg g}^{-1}$), from aqueous solutions (Milagres et al., 2020). Further, Milagres et al. reported that the 103-HC-DS/Fe-5 sorbent could be successfully utilized for the elimination of acetamiprid, atrazine, chlorpyrifos, and thiamethoxam agrichemicals from river water (Milagres et al., 2020).

The Fe_3O_4 -red mud nanoparticles have been prepared and used for the removal of ciprofloxacin from wastewater via a batch technique (Aydin et al., 2019); their adsorption capacity for ciprofloxacin was determined to be 111.11 mg g^{-1} and the removal percent for ciprofloxacin being over 90% from real wastewater. The combination of Fe_3O_4 and red mud has overcome the disadvantages such as iron leaching, agglomeration of nanoparticles, and separation difficulties from aqueous solutions relating to pristine materials. The low price and ready availability of the red mud as processes waste renders the Fe_3O_4 -red mud nanoparticles a suitable alternative to commercial adsorbents.

Facile and one-pot hydrothermal procedure have been documented for the preparation of pure hematite ($\alpha\text{-Fe}_2\text{O}_3$) nanoparticles (Nassar et al., 2018) with high adsorption capacity (70 mg g^{-1}) for the elimination of cephalexin antibiotic from aqueous media at optimum conditions ($\alpha\text{-Fe}_2\text{O}_3$ dosage = 0.05 g, cephalexin solution volume = 25 mL, pH = 7.5, cephalexin initial concentration = 150 mg L^{-1} , time = 180 min, and temperature = 25°C). The strong electrostatic attractions between the CO_2^- groups of negatively charged deprotonated-cephalexin molecules ($\text{pK}_{\text{a}2} < 7.5$) and the positively charged $\alpha\text{-Fe}_2\text{O}_3$ nanoparticles ($\text{pH}_{\text{PZC}} \text{ of } \alpha\text{-Fe}_2\text{O}_3 > 7.5$) were responsible for the obtained maximum adsorption capacity at pH = 7.5.

Some studies have been reported on the adsorption of oxolinic acid, levofloxacin, diclofenac, ketoprofen, and naproxen by goethite adsorbent, and have been demonstrated the good adsorption efficiency of goethite (Cheng et al., 2019; Qin et al., 2018; Yu et al., 2019). Mechanistic insight into interactions between two iron oxide minerals (IOMs; ferrihydrite and goethite) with different crystal structures and tetracycline was presented by Wu et al. (Fig. 3) (Wu

et al., 2019). It was indicated that ferrihydrite, an amorphous iron oxide with a larger specific surface area and smaller pore size, exhibited a better adsorption capability than goethite, a crystalline iron oxide, to remove of tetracycline from aqueous media; both IOMs formed stable inner-sphere complexation and had specific tetracycline sorption. Although both IOMs systems had electrostatic interactions with tetracycline sorbate, the significant effect for tetracycline adsorption onto goethite was only discernible. After tetracycline adsorption on ferrihydrite, Fe^{2+} ion was released in solution, indicating a redox chemical reaction. Na^+ and Ca^{2+} ions had no significant influence on tetracycline sorption, whereas Cu^{2+} ions formed a complex with tetracycline and its size became larger. Therefore, the Cu tetracycline complex reduced the tetracycline sorption onto ferrihydrite with a smaller pore size but improved tetracycline sorption onto goethite. The ensuing results from the ionic force influence provided proof about iron oxide mineral effect on the mobility of tetracycline.

An innovative approach for the adsorption and persulfate-based advanced oxidation of ibuprofen has been reported by Yin et al. using recycled rusted iron particles (RRIP) from heat pads (Fig. 4) (Yin et al., 2018); RRIP core-shell structure comprises zero-valent iron (Fe^0) in the core, and amorphous FeOOH and $\alpha\text{-FeOOH}$ on the shell. The $q_{\max}$ of RRIP for ibuprofen sorption was 3.47 mg g^{-1} , which was 4.8 times greater than the synthetic $\alpha\text{-FeOOH}$. The larger pore volume/size and higher specific surface area of RRIP compared with $\alpha\text{-FeOOH}$ was the reason for the enhanced ibuprofen adsorption. Also, the iron (hydr)oxides created on the rusted iron particles surface generate a large number of sites for the adsorption of ibuprofen. The RRIP activated persulfate can degrade the adsorbed and dissolved ibuprofen by generating both the hydroxyl ($\text{OH}\cdot$) and sulfate ($\text{SO}_4\cdot^-$) radicals.

4.1.2. Zinc oxide

Other metal oxides or their composites have been investigated either theoretically or experimentally for the removal of drug and

Fig. 3. Pore size-distribution map of (a) ferrihydrite, (b) goethite, and (c) the mechanistic insight for tetracycline adsorption onto ferrihydrite and goethite. Reprinted with permission from (Wu et al., (2019)).

Fig. 4. Recycling of rusted iron particles (RRIP) with core-shell Fe–FeOOH structure from used heat pads (PDS), the application of RRIP as an adsorbent for removal of ibuprofen, the radical formation and ibuprofen degradation in the RRIP activated persulfate via persulfate-based advanced oxidation and presentation of adsorption and degradation mechanism. Reprinted with modification with permission from (Yin et al., (2018)).

pesticide residues. The $\text{Zn}_{12}\text{O}_{12}$ nanocluster containing 6 tetragonal and 8 hexagonal rings with 36 Zn–O bonds was synthesized by Onori et al., and its adsorption capability and electronic sensitivity to the anticancer drug platinum, was explored via density functional theory (DFT) calculations in aqueous solution (Onori and Alipour, 2018). Platinum simultaneously interacted with Zn and O atoms of Zn–O bond via the Cl, Pt, and H atoms and lay on a ZnO nanocluster hexagonal ring with an adsorption energy of $-31.7 \text{ kcal mol}^{-1}$ (298 K). After drug adsorption, the ZnO nanocluster valence level shifted to higher energies, and the high destabilization of the HOMO level decreased the HOMO-LUMO energy gap (Eg) of the ZnO nanocluster. Thus, Eg decrease and conduction electrons increase of adsorbent largely enhanced the electrical conductivity. The enhancement in ZnO nanocluster electrical conductivity after platinum adsorption was converted to an electrical signal, thus rendering ZnO nanocluster a promising candidate for platinum detection.

Besides theoretical investigations, the ZnO nanoparticles that were prepared via the precipitation method were evaluated for the elimination of ciprofloxacin hydrochloride from aqueous media via batch experiments (Dhiman and Sharma, 2019a). The highest adsorption of 85.4% at pH 4 and assessment of isotherm and kinetic equations for ciprofloxacin antibiotic adsorption onto ZnO nanoparticles illustrated that favorable monolayer sorption with uniform distribution of sorbate onto sorbent surface occurred via physical interactions.

In another report, competitive removal of [ofloxacin hydrochloride + ciprofloxacin hydrochloride] and [paracetamol + diclofenac sodium] from binary solutions was studied using ZnO nanoparticles (Dhiman and Sharma, 2019b). The ZnO nanoparticles displayed good adsorption efficiency for the studied drugs in primary solutions and pH played an important role in drug sorption onto ZnO nanoparticles. The optimal pH was chosen at pH = 8; the pH below ZnO point zero charge ($\text{pH}_{\text{PZC}} = 8.7$), $\text{pK}_{\text{a}1} < \text{pH} < \text{pK}_{\text{a}2}$ for ofloxacin and ciprofloxacin, pK_{a}

diclofenac sodium $< \text{pH}$, and pK_{a} paracetamol $> \text{pH}$. Thus, there were strong electrostatic attractions between the positively charged surface of ZnO with the negatively charged carboxyl group of zwitterionic ofloxacin and ciprofloxacin, as well as with anionic diclofenac but the hydrophobic and electrostatic interactions did not play a role in the paracetamol adsorption. In binary solutions, the presence of the second drug had a negative effect on the removal of the first drug, and vice versa. So, the adsorption of each drug was significantly reduced in the presence of other drugs.

4.1.3. Cerium oxide

The preparation of CeO_2 nanofibers from Ce(1,3,5-benzene tricarboxylate)(H_2O)₆ (Ce-BTC) metal-organic frameworks and their application was investigated for the pesticide adsorption from aqueous media (Abdelillah Ali Elhoussein et al., 2018). The Ce-BTC metal-organic frameworks were synthesized via the hydrothermal method and CeO_2 nanofibers were created from Ce-BTC nanoparticles by calcination method at 650°C for 3 h. The CeO_2 nanofibers were used for 2,4-dichlorophenoxyacetic acid (2,4-D) adsorption from the water by a batch system with the q_{max} value to be 95.78 mg g^{-1} at 308 K. Based on the kinetic and isotherm studies, the 2,4-D sorption onto CeO_2 was performed through boundary layer diffusion and intra-particle diffusion.

4.1.4. Cobalt oxide and copper oxide

Other research works for drug adsorption onto metal oxides were exemplified by doxycycline hyclate sorption onto cobalt oxide (Co_3O_4) (Abbas et al., 2019) and ciprofloxacin hydrochloride adsorption onto copper oxide (CuO) (Sharma and Dhiman, 2017) in aqueous solutions.

4.1.5. Metal oxides composites

Despite the good performance of metal oxide nanoparticles, metal oxide composites had a better performance for removing organic pollutants due to the synergistic effects of metal oxides

participating in their structures. Magnetically retrievable mesoporous magnesium ferrite (MgFe_2O_4) has been explored for chlorpyrifos adsorption from real pesticide wastewater (Sharma and Kakkar, 2018). The mesoporous MgFe_2O_4 with high surface area ($170 \text{ m}^2 \text{ g}^{-1}$) was prepared using benign starting materials and urea as a dual-purpose mediator in the one-step solvothermal method (Fig. 5a). Batch adsorption experiments demonstrated that the effective adsorption of chlorpyrifos occurred onto MgFe_2O_4 adsorbent at $\text{pH} > 9$ (Fig. 5b); the proposed chlorpyrifos adsorption mechanism is depicted in Fig. 5c. Accordingly, a chemisorption process occurred via the interaction of hydroxylated MgFe_2O_4 surface with the electronegative atoms (chlorine, oxygen, and sulfur) and the chlorpyrifos aromatic ring followed by degradation into small organic species. This investigation revealed that the mesoporous MgFe_2O_4 with high adsorption efficiency can be an interesting selection for wastewater treatment.

The adsorption capability of ZnO – MgO nanocomposite and MgO nanoparticles were evaluated and compared for the elimination of antibiotics from aqueous media via response surface methodology (Fakhri and Behrouz, 2015); fast and efficient linezolid adsorption occurred onto MgO nanoparticles and ZnO – MgO nanocomposite via a simple procedure. The maximum adsorption capacity of ZnO – MgO nanocomposite (140.28 mg g^{-1}) was better than MgO nanoparticles (123.45 mg g^{-1}) because of the higher specific surface area of the nanocomposite.

The composite of iron and copper oxides (Fe/Cu oxides) has been synthesized using the extract of pine needle and was applied

as an efficient adsorbent for ofloxacin and norfloxacin removal from aqueous media (Fig. 6a) (Ma et al., 2020). The maximum adsorption capacity of Fe/Cu oxide for ofloxacin and norfloxacin were calculated 1.26 mmol g^{-1} and 1.64 mmol g^{-1} , respectively. The thermodynamic parameters established the endothermic and spontaneous nature of adsorption, and kinetic results revealed that the diffusion had an important role in the adsorption rate-limiting step. The ofloxacin and norfloxacin adsorption mechanism involved a combination of electrostatic attraction ($\text{pH} < \text{pH}_{\text{pZC}} < \text{pK}_{\text{a1}}$), hydrogen bonds formation, π – π stacking, hydrophobic interaction, and surface complexation (Fig. 6b).

Similar studies were performed in the field of water purification by metal oxides nanocomposites for the removal of anticonvulsant drug oxcarbazepine onto 3D porous NiFe_2O_4 (Parashar et al., 2020) and adsorption of riboflavin onto silver-deposited iron oxide magnetic ($\text{Ag/Fe}_3\text{O}_4$) nanoparticles (Akhond et al., 2016). The excellent adsorption efficiencies demonstrated by the most used metal oxides and their composites for various drugs/pesticides removal are presented in Table 1.

4.2. Carbon nanostructures

Carbon is among the most plentiful and versatile elements on the earth with a wide-ranging application (Benzigar et al., 2018). Carbon-based nano-compounds exhibit special and extraordinary characteristics, including tremendous thermal and mechanical stability, corrosion resistance, hardness, and low density (Z. M. Li

Fig. 5. (a) Preparation of mesoporous magnesium ferrite (MgFe_2O_4); the MgFe_2O_4 was produced in 3 steps: I) the formation of the homogeneous content from magnesium acetate, ferric nitrate, and sodium acetate with polyethylene glycol (PEG) in ethylene glycol (EG), II) the Mg – Fe alkoxide/glycolate precursor creation and III) the crystalline MgFe_2O_4 preparation via calcination, (b) the effect of MgFe_2O_4 dosage on the adsorption capacity and removal percentage, and (c) the plausible mechanism for the chlorpyrifos adsorption onto the MgFe_2O_4 surface. Reprinted with modification with permission from (Sharma and Kakkar, (2018)).

Fig. 6. (a) A plausible mechanism for synthesis of Fe/Cu oxides composite particles using pine needles extracts, (b) pH effects on the norfloxacin and ofloxacin removal, and the norfloxacin and ofloxacin adsorption mechanism onto Fe/Cu oxides composite particles. Reproduced with modification with permission from (Ma et al., (2020)).

et al., 2019a,b). These unique properties have led to the deployment of carbon-based nanomaterials in diverse fields comprising environmental remediation (Zhang et al., 2019), biotechnology (Molaei, 2019), drug delivery (Panwar et al., 2019), coating (Yi et al., 2019), packaging (Sobhan et al., 2019), and energy storage (Hu et al., 2019), among others (Bi et al., 2019). The carbon-based nanomaterials synthesis for targeted applications is interesting because of their unique properties, including flexibility, superior directionality, high surface area, and outstanding optical properties, which made them special and more challenging than bulk materials for specialized uses (Gan et al., 2019; Nekouieian et al., 2019; M. Yang et al., 2019). The carbon-based nanomaterials cover a range of nano-structural materials and their common forms encompass activated carbon, graphene, graphene oxide, reduced graphene oxide, graphitic carbon nitride ($\text{g-C}_3\text{N}_4$), carbon nanotubes, and carbon/graphene quantum dots. Several methods pertaining to the preparation and

incorporation of carbon-based nanomaterials for the elimination of contaminants from aqueous media are discussed in this review.

4.2.1. Activated carbon

The activated carbon typical elemental composition consists of around 80% carbon with the rest material comprising hydrogen, nitrogen, oxygen, and sulfur, alongside main functional groups such as phenols, carbonyl, carboxyl, quinones, lactones, and others. Two types of often used activated carbon are powdered activated carbon and granular activated carbon. The pretreatment, carbonization, and activation are the three essential steps during the production of activated carbon. The activated carbon is characterized as carbon-rich material with a well-built internal porous structure that is usually created via the pyrolysis and chemical treatment of natural sources, including wood (Danish and Ahmad, 2018), coal (Shokry et al., 2019), rice husks (Menya et al., 2018), sugarcane bagasse

Table 1

Data on the drugs or pesticides removal by metal oxides and their composites.

Adsorbents	Pollutants	pH	Temperature (K)	Time (min)	$q_{\max}$ (mg g ⁻¹)	Reference
Fe ₃ O ₄	Carbamazepine	7	300	180	0.715	Misra et al. (2018)
Fe ₃ O ₄	Levofloxacin	6.5	298	240	6.094	Al-Jabari et al. (2019)
Fe ₃ O ₄	Rifampicin	5.5	303	60	84.8	Cai et al. (2019)
α -Fe ₂ O ₃	Cephalexin	7.5	298	180	52.28	Nassar et al. (2018)
Fe ₂ O ₃	Carbamazepine	—	298	150	2.89	Rajendran and Sen (2018)
α -FeOOH	Diclofenac	5.23	298	480	0.025	Zhao et al. (2017)
α -FeOOH	Tetracycline	5.5	298	5760	1.726	Wu et al. (2019)
Ferrihydrite	Tetracycline	5.5	298	5760	3.822	Wu et al. (2019)
Fe ₃ O ₄ /red mud	Ciprofloxacin	6	298	180	110.15	Aydin et al. (2019)
Ag/Fe ₃ O ₄	Riboflavin	6	298	2	18.14	Akhond et al. (2016)
ZnO	Ciprofloxacin	4	298	150	8.88	Dhiman and Sharma (2019a)
MgO	Linezolid	10	308	—	123.45	Fakhri and Behrouz (2015)
ZnO–MgO	Linezolid	10	308	—	140.28	Fakhri and Behrouz (2015)
CuO	Ciprofloxacin	4	298	135	8.15	Sharma and Dhiman (2017)
Cobalt oxide	Doxycycline hyclate	6	313	90	4.484	Abbas et al. (2019)
NiFe ₂ O ₄	Oxcarbapazine	2.5	303	20	0.103	Parashar et al. (2020)
Fe/Cu oxides	Norfloxacin	8.12	293	180	523.7	Ma et al. (2020)
Fe/Cu oxides	Ofloxacin	8.87	293	180	455.3	Ma et al. (2020)
Fe ₃ O ₄	Methoxychlor	2	298	55	163.9	Nayak et al. (2018)
Fe ₃ O ₄	Atrazine	2	298	55	77.5	Nayak et al. (2018)
MgFe ₂ O ₄	Chlorpyrifos	10	295	360	4461	Sharma and Kakkar (2018)
ZnO	Naphthalene	4	298	40	66.80	Kaur et al. (2017)
CTAB–ZnO ^a	Naphthalene	4	298	40	89.96	Kaur et al. (2017)
BMTF–IL–ZnO ^b	Naphthalene	4	298	40	148.3	Kaur et al. (2017)
Hr–MgO ^c	Chlorpyrifos	7	295	60	3974	Sharma and Kakkar (2017)
CeO ₂	2,4-Dichloro phenoxy acetic acid	—	308	100	95.78	Abdelillah Ali Elhussein et al. (2018)

^a **CTAB–ZnO**: Cetyltrimethylammonium bromide functionalized Zinc oxide.^b **BMTF–IL–ZnO**: 1-Butyl-3-methylimidazolium tetrafluoroborate functionalized Zinc oxide.^c **Hr–MgO**: Hierarchical magnesium oxide.

(Guo et al., 2020), orange peel (Tovar et al., 2019), bamboo (Negara et al., 2019), date stones (Chowdhury et al., 2019), and other organic wastes (A. Rahman et al., 2019; B. Yang et al., 2019; Yek et al., 2019).

The activated carbons with a broad variety of functional groups, well-organized micro, meso, and macro-pores, and high surface area have been applied as the oldest, most popular, and most common sorbent for the adsorption of organic contaminants in water and wastewater remediation (Mansour et al., 2018; Völker et al., 2019). The main emphasis has been on improving the activation procedures in an optimum manner with proper precursors for different pollutants because the activation method greatly influences pore structure and surface functional groups of activated carbons (Bhatnagar et al., 2013). The activation procedure is typically performed by acidic/basic-, microwave-, ozone-, and plasma treatment wherein the activated carbon surface modification is carried out by physical, chemical, and physiochemical treatment for higher sorption of specific contaminants (Ahmed, 2017). Of note, physical and chemical activation are the two major types of treatments. The physical activation occurs in the presence of inert gases, O₂ absence or activation by oxidizing agents such as steam, air, CO₂ and their combination, while the chemical activation occurs via carbonization, impregnation with base (KOH, NaOH and Na₂CO₃), alkali metal salt (AlCl₃ and ZnCl₂) or acid (H₃PO₄ and H₂SO₄), and strongly dehydration (Shejale et al., 2020). The activation by using chemical oxidants is performed under mild conditions as compared to physical activation and is created by the chemical-treated activated carbons with higher micro-porosity and larger surface area (Ahmed, 2017). Thus, the source material, activation method, and any additional modifications determine the surface area (up to 2500 m² g⁻¹) and the accurate internal pore composition of activated carbon.

4.2.1.1. Drugs adsorption onto activated carbons. Many studies have been reported for the adsorption of drug from aqueous solution onto activated carbons, e.g., diclofenac (de Franco et al., 2018) and

triclosan (Bernal et al., 2020a) onto the activated carbons, atenolol (Haro et al., 2017), and phenol, salicylic acid, and methylparaben (Bernal et al., 2020b) onto commercial granular activated carbons, ampicillin (Del Vecchio et al., 2019), and acetaminophen and salicylic acid (Bernal et al., 2020c) onto granular activated carbon, ranitidine onto activated carbon prepared from lemon peel (Bhattacharyya et al., 2019), metformin hydrochloride onto *Zea mays* tassel activated carbon (Kalumpha et al., 2020), metronidazole and sulfamethoxazole onto walnut shell-based activated carbon (Teixeira et al., 2019), etodolac onto microporous activated carbon derived from apricot, peach stones, and almond shell mixture biomasses (Erdem et al., 2020), acetaminophen onto cashew nut shell biomass-derived activated carbons (Geczo et al., 2020), acetylsalicylic acid and sodium diclofenac onto lovegrass (Poaceae) derived activated carbon (Cimirro et al., 2020), and amoxicillin onto activated carbon obtained from the sugar and alcohol industry byproducts (Schultz et al., 2020).

The activated carbon textural properties have been tailored to improve its potential for diclofenac adsorption from aqueous media (Moral-Rodríguez et al., 2019). The activated carbons were prepared as activated carbon 0 ($S_{\text{BET}} = 799 \text{ m}^2 \text{ g}^{-1}$, $V_{\text{Micro}} = 0.31 \text{ cm}^3 \text{ g}^{-1}$), activated carbon 12 ($S_{\text{BET}} = 1146 \text{ m}^2 \text{ g}^{-1}$, $V_{\text{Micro}} = 0.45 \text{ cm}^3 \text{ g}^{-1}$), activated carbon 24 ($S_{\text{BET}} = 1279 \text{ m}^2 \text{ g}^{-1}$, $V_{\text{Micro}} = 0.50 \text{ cm}^3 \text{ g}^{-1}$) and activated carbon 40 ($S_{\text{BET}} = 1801 \text{ m}^2 \text{ g}^{-1}$, $V_{\text{Micro}} = 0.66 \text{ cm}^3 \text{ g}^{-1}$) by modifying a commercial activated carbon via physical treatment with CO₂ for 0, 12, 24 and 40 h during the activation process. The $q_{\max}$ of activated carbons was mainly attributed to the activated carbons porous structure or activated carbons textural properties and was considerably improved by increasing the activation time. Hence, the activated carbon-potential for the removal of emerging pollutants can be significantly improved by tailoring the activated carbons textural properties.

The influence of hydrophobicity/lipophilicity of pharmaceuticals and personal care products (PPCPs), caffeine ($\log K_{\text{ow}} = -0.07$),

Table 2

The recent drugs and pesticide adsorption onto activated carbons.

Activation Method	Modification conditions	Carbon source	Adsorbate	Adsorption conditions	Adsorption isotherm	Adsorption kinetic	$q_{\max}$ (mg g ⁻¹)	Reference
Chemical activation	KOH, 4 h	Prosopis juliflora	Tetracycline	pH = 7.0 $C_0 = 1 \text{ mg L}^{-1}$ $V = 100 \text{ mL}$, $m = 1 \text{ g L}^{-1}$ $T = 303 \text{ K}$, $t = 120 \text{ min}$	Langmuir	Pseudo-second-order	28.81	Manjunath et al. (2020)
Chemical activation	KOH, 4 h	Prosopis juliflora	Metronidazole	pH = 7.0 $C_0 = 1 \text{ mg L}^{-1}$ $V = 100 \text{ mL}$, $m = 1 \text{ g L}^{-1}$ $T = 303 \text{ K}$, $t = 120 \text{ min}$	Langmuir	Pseudo-second-order	25.06	Manjunath et al. (2020)
Chemical activation	KOH, 4 h	Prosopis juliflora	Sulfadiazine	pH = 7.0 $C_0 = 1 \text{ mg L}^{-1}$ $V = 100 \text{ mL}$, $m = 1 \text{ g L}^{-1}$ $T = 303 \text{ K}$, $t = 120 \text{ min}$	Langmuir	Pseudo-second-order	18.48	Manjunath et al. (2020)
Chemical activation	I) Carbonization (500 °C, 2 h) II) NaOH (700 °C, 90 min)	Date press cake	Cefixime	pH = 4.0 $C_0 = 500 \text{ mg L}^{-1}$ $V = 25 \text{ mL}$, $m = 1 \text{ g L}^{-1}$ $T = 298 \text{ K}$, $t = 240 \text{ min}$	Redlich-Peterson	Elovich Pseudo-second-order	557.9	Hasanzadeh et al. (2020)
Chemical activation	I) Carbonization (500 °C, 2 h) II) KOH (700 °C, 90 min)	Date press cake	Cefixime	pH = 4.0 $C_0 = 500 \text{ mg L}^{-1}$ $V = 25 \text{ mL}$, $m = 1 \text{ g L}^{-1}$ $T = 298 \text{ K}$, $t = 240 \text{ min}$	Redlich-Peterson	Elovich Pseudo-second-order	571.5	Hasanzadeh et al. (2020)
Chemical activation	I) Steam-pyrolysis (900 °C, 3 h) II) $\text{HNO}_3 + \text{H}_2\text{O}_2$ (200 °C, 1 h)	Sucrose Polystyrene	Tetracycline	pH = 7.0 $C_0 = 60 \text{ mg L}^{-1}$ $V = 25 \text{ mL}$, $m = 0.1 \text{ g}$ $T = 298 \text{ K}$, $t = 120 \text{ min}$	Langmuir	Pseudo-second-order	909.2	Hubetska et al. (2020)
Chemical activation	I) Carbonization (300 °C) II) Impregnation (H_3PO_4 , 24 h) III) Chemical activation (400 °C)	Zea mays tassel	Metformin hydrochloride	pH = 9.5 $C_0 = 152.5 \text{ mg L}^{-1}$ $V = 20 \text{ mL}$, $m = 0.575 \text{ g}$ $T = 293 \text{ K}$, $t = 67.5 \text{ min}$	Langmuir	Pseudo-second-order	44.84	Kalumpha et al. (2020)
Chemical activation	I) Impregnation (FeSO_4 , 700 °C) II) Pyrolysis (700 °C, 2 h)	Apricot and peach stones + Almond shell	Etodolac	pH = 4.0 $C_0 = 20 \text{ mg L}^{-1}$ $V = 100 \text{ mL}$, $m = 5 \text{ g L}^{-1}$ $T = 298 \text{ K}$, $t = 150 \text{ min}$	Freundlich	Pseudo-second-order	19.74	Erdem et al. (2020)
Physical activation	Physically activated with CO_2	Coconut shell	Methylparaben	pH = 7.42 $C_0 = 6.58 \text{ mmol L}^{-1}$ $V = 25 \text{ mL}$, $m = 100 \text{ mg}$ $T = 293 \text{ K}$, $t = 7 \text{ days}$	Langmuir	—	1.58 mmol g ⁻¹	Bernal et al. (2020b)
Chemical activation	I) H_3PO_4 (85 °C, 2 h) II) Carbonization (400–700 °C, 2h)	Cashew nut shell	Acetaminophen	pH = — $C_0 = 50 \text{ mg L}^{-1}$ $V = 25 \text{ mL}$, $m = 12.5 \text{ mg}$ $T = - \text{K}$, $t = 24 \text{ h}$	Langmuir	Pseudo-second-order	146	Geczo et al. (2020)
Chemical activation	I) Impregnation (ZnCl_2 , 80 °C, 2 h) II) Pyrolysis (700 °C, 1 h)	Lovegrass (Poaceae)	Sodium diclofenac	pH = 4.0 $C_0 = 200 \text{ mg L}^{-1}$ $V = 20 \text{ mL}$, $m = 60 \text{ mg}$ $T = 298 \text{ K}$, $t = 60 \text{ min}$	Liu	Pseudo-second-order	312.4	Cimirro et al. (2020)
Chemical activation	I) Impregnation (ZnCl_2 , 80 °C, 2 h) II) Pyrolysis (700 °C, 1 h)	Lovegrass (Poaceae)	Acetylsalicylic acid	pH = 4.0 $C_0 = 200 \text{ mg L}^{-1}$ $V = 20 \text{ mL}$, $m = 60 \text{ mg}$ $T = 298 \text{ K}$, $t = 45 \text{ min}$	Liu	Pseudo-second-order	221.7	Cimirro et al. (2020)
Chemical activation	I) Impregnation (H_3PO_4 , 24 h) II) Microwave (N_2 , 20 min, 1 kW)	Reynoutria japonica	Diclofenac	pH = 7.0 $C_0 = 300 \text{ mg L}^{-1}$ $V = 10 \text{ mL}$, $m = 20 \text{ mg}$ $T = 296 \text{ K}$, $t = 90 \text{ min}$	Sips	Pseudo-second-order	87.09	Koutník et al. (2020)
Chemical activation	I) Impregnation (H_3PO_4 , 24 h) II) Microwave (N_2 , 20 min, 1 kW)	Reynoutria japonica	Paracetamol	pH = 7.0 $C_0 = 300 \text{ mg L}^{-1}$ $V = 10 \text{ mL}$, $m = 20 \text{ mg}$ $T = 296 \text{ K}$, $t = 90 \text{ min}$	Sips	Pseudo-second-order	136.61	Koutník et al. (2020)

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Table 2 (continued)

Activation Method	Modification conditions	Carbon source	Adsorbate	Adsorption conditions	Adsorption isotherm	Adsorption kinetic	$q_{\max}$ (mg g ⁻¹)	Reference
Chemical activation	I) Impregnation (ZnCl ₂ , R.T., 24 h) II) Carbonization (500 °C, 2 h)	Sugar cane bagasse	Diclofenac sodium	pH = 2.0 C ₀ = 50 mg L ⁻¹ V = 25 mL, m = 0.4 g L ⁻¹ T = 298 K, t = 15 min	Langmuir	Pseudo-second-order	315.0	Abo El Naga et al. (2019)
Chemical activation	I) Pyrolysis (300 °C, 1 h, N ₂) II) Impregnation (K ₂ CO ₃ , 105 °C, 8 h) III) Carbonization (900 °C, 1h, N ₂)	Walnut shell	Metronidazole	pH = 6.0 C ₀ = 40 mg L ⁻¹ V = - mL, m = 10 mg T = 293 K, t = 48 h	Langmuir	—	107.4	Teixeira et al. (2019)
Chemical activation	I) Pyrolysis (300 °C, 1 h, N ₂) II) Impregnation (K ₂ CO ₃ , 105 °C, 8 h) III) Carbonization (900 °C, 1h, N ₂)	Walnut shell	Sulfamethoxazole	pH = 6.0 C ₀ = 40 mg L ⁻¹ V = - mL, m = 10 mg T = 293 K, t = 48 h	Langmuir	—	93.5	Teixeira et al. (2019)
Chemical activation	I) Carbonization (500 °C, 2h, N ₂) II) Impregnation (KOH, 120 °C) III) Pyrolysis (550 °C, 1h, N ₂)	Date palm leaflets	Chlorpheniramine	pH = 7.0 C ₀ = 50 mg L ⁻¹ V = 25 mL, m = 0.06 g T = 298 K, t = 48 h	Langmuir	Pseudo-second-order	100.2	(S. N. F. Ali et al., 2019)
Chemical activation	I) Carbonization (500 °C, 2h, N ₂) II) Impregnation (KOH, 120 °C) III) Pyrolysis (550 °C, 1h, N ₂)	Date palm leaflets	Ibuprofen	pH = 7.0 C ₀ = 50 mg L ⁻¹ V = 25 mL, m = 0.06 g T = 298 K, t = 48 h	Langmuir	Pseudo-second-order	52.4	(S. N. F. Ali et al., 2019)
—	—	Babassu coconut	Diclofenac	pH = 7.0 C ₀ = 50 mg L ⁻¹ V = 50 mL, m = 0.05 g T = 298 K, t = 30 h	Freundlich	Intraparticle diffusion	71.150	Viotti et al. (2019)
Chemical activation	I) Impregnation with ZnCl ₂ II) Pyrolysis (600 °C, 30 min, N ₂) III) Leaching out with HCl (Reflux, 1h)	Brazil nutshells	Acetaminophen	pH = 7.0 C ₀ = 800 mg L ⁻¹ V = - mL, m = 1.5 g L ⁻¹ T = 298 K, t = 30 min	Liu	Avrami fractional-order	411.0	Lima et al. (2019)
Chemical activation	I) Impregnation (H ₃ PO ₄ , 85 °C, 2h) II) Pyrolysis (500 °C, 2h, N ₂)	Kenaf	Paracetamol	pH = 7.0 C ₀ = 120 mg L ⁻¹ V = 15 mL, m = 10 mg T = 298 K, t = 300 min	—	Pseudo-second-order	—	Macías-García et al. (2019)
Chemical activation	I) Mechanical grinding II) Heat (105 °C, 4h) III) Carbonization (350–500 °C, 1h) IV) Impregnation (NaOH, 130 °C, 4h)	Waste lemon peel	Ranitidine	pH = 6.0 C ₀ = 10 mg L ⁻¹ V = 100 mL, m = 200 mg T = 313 K, t = 60 min	—	Pseudo-second-order	—	Bhattacharyya et al. (2019)
Chemical activation	I) Carbonization (500 °C, 2h, N ₂) II) Impregnation (NaOH, 800 °C, 1h) III) Dehydration (105 °C, 24h)	Rice husk	Acetaminophen	pH = 5.8 C ₀ = 40 mg L ⁻¹ V = 100 mL, m = 0.15 g T = 298 K, t = 60 min	Redlich-Peterson	Pseudo-second-order	50.25	Paredes-Laverde et al. (2019)
Chemical activation	I) Impregnation (ZnCl ₂ , 28 °C, 24h) II) Carbonization (600 °C, 2h, N ₂)	Tea waste	Sodium diclofenac	pH = 6.5 C ₀ = 30 mg L ⁻¹ V = 100 mL, m = 300 mg L ⁻¹ T = 303 K, t = 8 h	Langmuir	Pseudo-second-order	91.2	Malhotra et al. (2018)
Chemical activation	I) Impregnation with ZnCl ₂ (r = 1.5) II) Carbonization (600 °C)	Caesalpinia ferrea seed pod	Captopril	pH = 7.0 C ₀ = 600 mg L ⁻¹ V = 20 mL, m = 30 mg L ⁻¹ T = 298 K, t = 60 min	Liu	General-order kinetic	535.5	Kasperiski et al. (2018)
Chemical treatment	I) Dehydration (105 °C, 24h) II) Carbonization (600 °C, 1h) III) Chemical-thermal activation	Quercus Brantii (oak) acorn	Acetaminophen	pH = 3.0 C ₀ = 100 mg L ⁻¹ V = 100 mL, m = 100 mg T = 298 K, t = 150 min	Freundlich	Pseudo-second-order	45.45	Nourmoradi et al. (2018)

Chemical activation	I) Dehydration (105 °C, 24h) II) Carbonization (600 °C, 1h) III) Chemical-thermal activation	Quercus Brantii (oak) acorn	Ibuprofen	pH = 3.0 $C_0 = 100 \text{ mg L}^{-1}$ $V = 100 \text{ mL}$, $m = 100 \text{ mg}$ $T = 298 \text{ K}$, $t = 120 \text{ min}$	Freundlich	Pseudo-second-order	96.15	Nourmoradi et al. (2018)
Chemical activation	I) Impregnation with FeCl_3 II) Pyrolysis (800 °C, 2 h, N_2)	Tara gum	Antipyrine	pH = 7.0 $C_0 = 50 \text{ mg L}^{-1}$ $V = - \text{mL}$, $m = 0.2 \text{ g L}^{-1}$ $T = 290 \text{ K}$, $t = 60 \text{ min}$	Langmuir	Hyperbolic	307.7	Bedia et al. (2018)
Chemical activation	I) Carbonization (700 °C, 1h, N_2) II) Impregnation (NH_4Cl , 105 °C, 24h) III) Pyrolysis (800 °C, 2.5 h, N_2)	Pomegranate wood	Chlortetracycline	pH = 6.0 $C_0 = 100 \text{ mg L}^{-1}$ $V = 100 \text{ mL}$, $m = 0.2 \text{ g L}^{-1}$ $T = 293 \text{ K}$, $t = 60 \text{ min}$	Redlich-Peterson	Ritchie second order Pseudo-second order	482.47	Alahabadi et al. (2017)
Chemical activation	I) Impregnation (H_3PO_4 , R.T., 12h) II) Carbonization (450 °C, 2h) III) Dehydration (70 °C, 24h)	Luffa sponge	Ofloxacin	pH = 6.0 $C_0 = 30 \text{ mg L}^{-1}$ $V = 50 \text{ mL}$, $m = 0.5 \text{ g L}^{-1}$ $T = 308 \text{ K}$, $t = 60 \text{ min}$	Freundlich	Pseudo-second order	132	Kong et al. (2017)
Chemical activation	—	Apricot shells	Atrazine	pH = 6.0 $C_0 = 90 \text{ mg L}^{-1}$ $V = 50 \text{ mL}$, $m = 20 \text{ mg}$ $T = 293 \text{ K}$, $t = 5 \text{ h}$	Freundlich	Pseudo-second-order	46.30	Wei et al. (2018)
Chemical activation	—	Walnut shells	Atrazine	pH = 6.0 $C_0 = 90 \text{ mg L}^{-1}$ $V = 50 \text{ mL}$, $m = 20 \text{ mg}$ $T = 293 \text{ K}$, $t = 5 \text{ h}$	Freundlich	Pseudo-second-order	294.12	Wei et al. (2018)
Chemical activation	—	Wood	Atrazine	pH = 6.0 $C_0 = 90 \text{ mg L}^{-1}$ $V = 50 \text{ mL}$, $m = 20 \text{ mg}$ $T = 293 \text{ K}$, $t = 5 \text{ h}$	Freundlich	Pseudo-second-order	303.00	Wei et al. (2018)
Chemical activation	I) Carbonization (950 °C, 3h) II) HCl treatment	Casuarina wood	Chlorothalonil	pH = 7.0 $C_0 = 480 \text{ mg L}^{-1}$ $V = 200 \text{ mL}$, $m = 0.5 \text{ g L}^{-1}$ $T = 298 \text{ K}$, $t = 40 \text{ min}$	Freundlich	Pseudo-first-order	218.4	Gar Alalm and Nasr (2018)
Chemical activation	I) Impregnation (KOH, 105 °C, 1h) II) Carbonization (700 °C, 2h, N_2)	Orange peel	2-(2,4-Dichlorophenoxy) propionic acid	pH = 3.0 $C_0 = 50 \text{ mg L}^{-1}$ $V = 50 \text{ mL}$, $m = 50 \text{ mg}$ $T = 313 \text{ K}$, $t = 210 \text{ min}$	Langmuir	Pseudo-second-order	515.5	Pandiarajan et al. (2018)
Chemical activation	I) Impregnation (KOH, 105 °C, 1h) II) Carbonization (700 °C, 2h, N_2)	Orange peel	2-Methyl-4-chlorophenoxy acetic acid	pH = 3.0 $C_0 = 50 \text{ mg L}^{-1}$ $V = 50 \text{ mL}$, $m = 50 \text{ mg}$ $T = 313 \text{ K}$, $t = 210 \text{ min}$	Langmuir	Pseudo-second-order	215.5	Pandiarajan et al. (2018)
Chemical activation	I) Impregnation (KOH, 105 °C, 1h) II) Carbonization (700 °C, 2h, N_2)	Orange peel	2,4,5-Trichlorophenoxy acetic acid	pH = 3.0 $C_0 = 50 \text{ mg L}^{-1}$ $V = 50 \text{ mL}$, $m = 50 \text{ mg}$ $T = 313 \text{ K}$, $t = 210 \text{ min}$	Langmuir	Pseudo-second-order	416.6	Pandiarajan et al. (2018)
Chemical activation	I) Impregnation (KOH, 105 °C, 1h) II) Carbonization (700 °C, 2h, N_2)	Orange peel	2-Methyl-4-chlorophenoxy acetic acid	pH = 3.0 $C_0 = 50 \text{ mg L}^{-1}$ $V = 50 \text{ mL}$, $m = 50 \text{ mg}$ $T = 313 \text{ K}$, $t = 210 \text{ min}$	Langmuir	Pseudo-second-order	414.9	Pandiarajan et al. (2018)
Chemical activation	I) Impregnation (KOH, 105 °C, 1h) II) Carbonization (700 °C, 2h, N_2)	Orange peel	Methyl chlorophenoxy propionic acid	pH = 3.0 $C_0 = 50 \text{ mg L}^{-1}$ $V = 50 \text{ mL}$, $m = 50 \text{ mg}$ $T = 313 \text{ K}$, $t = 210 \text{ min}$	Langmuir	Pseudo-second-order	574.7	Pandiarajan et al. (2018)
Chemical activation	I) Impregnation (H_3PO_4 , 24h) II) Carbonization (600 °C, 1h)	Spent coffee grounds	Carbendazim	pH = 4.8 $C_0 = 5 \text{ mg L}^{-1}$ $V = 50 \text{ mL}$, $m = 1 \text{ g L}^{-1}$ $T = 298 \text{ K}$, $t = 20 \text{ min}$	Langmuir	Pseudo-second-order	11.918	Hgeig et al. (2019)
Chemical activation	I) Impregnation (H_3PO_4 , 24h) II) Carbonization (600 °C, 1h)	Spent coffee grounds	Linuron	pH = 4.8 $C_0 = 5 \text{ mg L}^{-1}$	Langmuir	Pseudo-second-order	5.834	Hgeig et al. (2019)

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Table 2 (continued)

Activation Method	Modification conditions	Carbon source	Adsorbate	Adsorption conditions	Adsorption isotherm	Adsorption kinetic	$q_{\max}$ (mg g ⁻¹)	Reference
Chemical activation	I) Impregnation with Na ₂ CO ₃ II) Pre-carbonization (300 °C, 2h) III) Impregnation (KOH, 400 °C, 2h)	Starch	Pyraclostrobin	V = 50 mL, m = 1 g L ⁻¹ T = 298 K, t = 20 min pH = 7.0 C ₀ = 0.5 mg L ⁻¹ V = 5 mL, m = 50 mg T = 298 K, t = 5 min	Langmuir	Pseudo-second-order	66.2	Suo et al. (2019)
Chemical activation	I) Impregnation (H ₃ PO ₄ , 100 °C, 24h) II) Carbonization (500 °C, 1h)	Tangerine peels	Acetamiprid	pH = 5.0 C ₀ = 25 mg L ⁻¹ V = 100 mL, m = 0.1 g T = 308 K, t = 240 min pH = 7.0 C ₀ = 10 mg L ⁻¹ V = 10 mL, m = 10 mg T = 303 K, t = 12 min	Langmuir	Pseudo-second-order	35.7	Mohammad et al. (2020)
Chemical activation	I) Impregnation (H ₃ PO ₄ , 24h) II) Carbonization (700 °C, 4h)	Tangerine seed	Bendiocarb	pH = 7.0 C ₀ = 10 mg L ⁻¹ V = 10 mL, m = 10 mg T = 303 K, t = 12 min	Langmuir	Pseudo-second-order	7.97	(Y. Wang et al., 2020)
Chemical activation	I) Impregnation (H ₃ PO ₄ , 24h) II) Carbonization (700 °C, 4h)	Tangerine seed	Metolcarb	pH = 7.0 C ₀ = 10 mg L ⁻¹ V = 10 mL, m = 10 mg T = 303 K, t = 12 min	Langmuir	Pseudo-second-order	9.11	(Y. Wang et al., 2020)
Chemical activation	I) Impregnation (H ₃ PO ₄ , 24h) II) Carbonization (700 °C, 4h)	Tangerine seed	Isoprocab	pH = 7.0 C ₀ = 10 mg L ⁻¹ V = 10 mL, m = 10 mg T = 303 K, t = 12 min	Langmuir	Pseudo-second-order	13.95	(Y. Wang et al., 2020)
Chemical activation	I) Impregnation (H ₃ PO ₄ , 24h) II) Carbonization (700 °C, 4h)	Tangerine seed	Pirimicarb	pH = 7.0 C ₀ = 10 mg L ⁻¹ V = 10 mL, m = 10 mg T = 303 K, t = 12 min	Langmuir	Pseudo-second-order	39.37	(Y. Wang et al., 2020)
Chemical activation	I) Impregnation (H ₃ PO ₄ , 24h) II) Carbonization (700 °C, 4h)	Tangerine seed	Carbaryl	pH = 7.0 C ₀ = 10 mg L ⁻¹ V = 10 mL, m = 10 mg T = 303 K, t = 12 min	Langmuir	Pseudo-second-order	44.64	(Y. Wang et al., 2020)
Chemical activation	I) Impregnation (H ₃ PO ₄ , 24h) II) Carbonization (700 °C, 4h)	Tangerine seed	Methiocarb	pH = 7.0 C ₀ = 10 mg L ⁻¹ V = 10 mL, m = 10 mg T = 303 K, t = 12 min	Langmuir	Pseudo-second-order	93.46	(Y. Wang et al., 2020)
Chemical activation	I) Impregnation with ZnCl ₂ II) Carbonization (500 °C, 2h, N ₂)	Orange pulp	2,4-Dichlorophenoxy acetic acid	pH = 6.22 C ₀ = 100 mg L ⁻¹ V = 100 mL, m = 0.3 g T = 298 K, t = 300 min	Langmuir	Pseudo-second-order	71.94	Angin and Güneş (2020)
Physical activation	I) Carbonization (800 °C, 1h) II) Activation (Steam, 200 °C)	Sugarcane bagasse	2,4-Dichlorophenoxy acetic acid	pH = 6.86 C ₀ = 5 mg L ⁻¹ V = 25 mL, m = 10 mg T = 300 K, t = 48 h	Redlich-Peterson Langmuir	Pseudo-second-order	153.9	Brito et al. (2020)
Physical activation	I) Carbonization (800 °C, 1h) II) Activation (Steam, 200 °C)	Coconut shell	2,4-Dichlorophenoxy acetic acid	pH = 6.86 C ₀ = 5 mg L ⁻¹ V = 25 mL, m = 10 mg T = 300 K, t = 48 h	Redlich-Peterson Langmuir	Pseudo-second-order	233.0	Brito et al. (2020)
Physical activation	I) Carbonization (800 °C, 1h) II) Activation (Steam, 200 °C)	Endocarp of babassu	2,4-Dichlorophenoxy acetic acid	pH = 6.86 C ₀ = 5 mg L ⁻¹ V = 25 mL, m = 10 mg T = 300 K, t = 48 h	Redlich-Peterson Langmuir	Pseudo-second-order	235.5	Brito et al. (2020)

ibuprofen ($\log K_{ow} = 3.97$), and triclosan ($\log K_{ow} = 4.76$), have been evaluated for adsorption onto commercial powdered activated carbon from aqueous solutions (Kaur et al., 2018). The removal efficacy and adsorption capacity were established in the order caffeine < ibuprofen < triclosan, which correlates to the octanol-water partition coefficient ($\log K_{ow}$) and the decreased solubility of the micro-pollutants in water. The obtained results established that the lipophilicity of micro-pollutants displays an important role in adsorption onto activated carbons.

The activated carbon fiber has been deployed for the adsorption of non-ionic carbamazepine, anionic diclofenac, and cationic propranolol from aqueous solutions (Zhao et al., 2020). The activated carbon fiber sorption capability was evaluated by the pH, isotherm, kinetic, and ionic strength experiments. The carbamazepine adsorption onto activated carbon fiber was independent of ionic strength and pH, while the anionic diclofenac adsorption was reduced at high concentrations of NaCl and alkaline pHs. The kinetic studies revealed the fast sorption of cationic propranolol and uncharged carbamazepine, and relatively slow sorption of anionic diclofenac onto activated carbon fiber. Based on the isotherm studies, the calculated q_{max} (mmol g^{-1}) of activated carbon fiber for diclofenac (0.95 ± 0.08), propranolol (1.07 ± 0.08), and carbamazepine (1.27 ± 0.06) exhibited the potent ability of activated carbon fiber for removing the pharmaceutical pollutants from real samples.

The activated carbon prepared from *Reynoutria japonica* invasive herb was reported for the xenobiotics (diclofenac and paracetamol) sorption from water (Fig. 7) (Koutník et al., 2020). The activated carbons were synthesized by microwave treatment using sodium methanolate (MeNa), NaOH, and H_3PO_4 chemicals. The type of activating agent was an important factor with a significant effect on the result of sorbents' texture and the sorbent's surface chemical nature. The activated carbon- H_3PO_4 displayed the highest sorption capacity for diclofenac (87.09 mg g^{-1}) and paracetamol (136.61 mg g^{-1}). The activating agent selection, the π - π interactions, and the interactions between the functional groups of activated carbon and sorbate played an important role in the adsorption rate.

The hierarchical porous activated carbon (SCB-AC) with a high specific surface area ($1145 \text{ m}^2 \text{ g}^{-1}$) was produced from sugar cane bagasse via a single-step ZnCl_2 activation and then applied for the adsorption of diclofenac sodium (DFC) from contaminated aqueous solutions in a batch system (Abo El Naga et al., 2019). The adsorption experiments exhibited that the fast adsorption of DFC onto SCB-AC occurred with an adsorption capacity of 315.0 mg g^{-1} . The SCB-AC not only demonstrated a rapid removal rate and high maximum adsorption capacity for DFC but also displayed high stability and prominent recyclability through simple washing of the DFC-saturated sorbent with acetone. Also, the pH effect on the sorbent surface charge and DCF sorption was studied for predicting the mechanism. At optimal adsorption pH 2, pH value lower than the DFC pK_a (4.2) and SCB-AC pH_{PZC} (7.3), the high DFC adsorption was related to non-electrostatic adsorption mechanisms, including hydrogen bonding, hydrophobic, and π - π electron donor-acceptor (EDA) interactions.

The activated carbon has been synthesized by chemical activation of Tara gum using FeCl_3 for antipyrine removal (Fig. 8); synthesis was performed with different ratios of activating agent and precursor ($r = \text{FeCl}_3/\text{precursor}$, 0.5–3.0 wt%) at variable temperatures from 400 to 1000 °C. The highly porous structure was obtained at $r = 2$ and 800 °C. The kinetic and isotherm studies of the adsorption process exhibited that the experimental adsorption data fitted well with the hyperbolic kinetic and Langmuir isotherm equations. The adsorption results affirmed that the activated carbon derived from Tara gum with $q_{max} = 307.7 \text{ (mg g}^{-1}\text{)}$ was useful

for the adsorption of antipyrine (Bedia et al., 2018).

Activated carbon has also been synthesized from pomegranate wood wastes using an innovated NH_4Cl -induced activation process (NH_4Cl -modified activated carbon; NAC) and was evaluated for chlortetracycline adsorption from aqueous solutions (Fig. 9) (Alahabadi et al., 2017). The adsorption ability of NAC was compared with standard activated carbon (SAC) for chlortetracycline adsorption. The NAC comprised a mesoporous granular particle with higher functional groups, greater surface area, and larger pore volume with much greater chlortetracycline removal and adsorption capacity compared to SAC under similar optimum conditions. The NH_4Cl -induced chemical treatment generated more crystallinity with highly ordered pores for NAC, probably due to the NH_4Cl explosive characteristics. The studied isotherm and kinetic models exhibited more favourability of chlortetracycline sorption onto NAC through the physical π - π EDA interaction.

Paredes-Laverde et al. prepared activated carbons (CH- ZnCl_2 activated carbon and RH- NaOH activated carbon) from coffee husk (CH) and rice husk (RH), via impregnation with activating agents ZnCl_2 and NaOH (Fig. 10) (Paredes-Laverde et al., 2019). The synthesized activated carbons were evaluated for acetaminophen removal from both the distilled water and synthetic urine. The RH- NaOH activated carbon with high carbon content displayed the best results in both media due to the creation of micro-porosity in the activation process and higher surface area. The evaluation of temperature influence in the range of 500–800 °C on the preparation of RH- NaOH activated carbons (RH- NaOH -500, RH- NaOH -650, and RH- NaOH -800) revealed that the functional groups and elemental composition of activated carbons were altered with temperature changes. Furthermore, the rise in temperature during the activation process helped to increase the porosity and surface area. Thus, the RH- NaOH -800 activated carbon showed the highest adsorption efficiency and removed the acetaminophen from aqueous solutions via hydrogen bonding and π - π interactions. Table 2 lists some of the reported papers regarding drug adsorption by activated carbons.

4.2.1.2. Pesticides adsorption onto activated carbons. Many researchers have investigated the elimination of pesticides from aqueous solutions using activated carbons, e.g., metaldehyde (Rolph et al., 2018), paclobutrazol (Grant et al., 2018), 11 pesticides (Grant et al., 2019), iodosulfuron (Ahmad, 2019), carbendazim and linuron (Hgeig et al., 2019), carbendazim (T. Wang et al., 2019), and 2,4-dichlorophenoxyacetic acid (2,4-D) (Amiri et al., 2020).

Two commercial activated carbons (GAB; $S_{Mic} = 580 \text{ m}^2 \text{ g}^{-1}$, $V_{Mic} = 0.27 \text{ cm}^3 \text{ g}^{-1}$, $\text{pH}_{PZC} = 7.46$ and CBP; $S_{Mic} = 99 \text{ m}^2 \text{ g}^{-1}$, $V_{Mic} = 0.04 \text{ cm}^3 \text{ g}^{-1}$, $\text{pH}_{PZC} = 4.76$) were utilized for the adsorption of 4-chloro-2-methylphenoxyacetic acid (MCPA; $\text{pK}_a = 3.07$) and 2,4-dichlorophenoxyacetic acid (2,4-D; $\text{pK}_a = 2.73$) from aqueous solutions (Spaltro et al., 2018). The microporous GAB exhibited higher adsorption capacity ($q_{max} = 367.15 \text{ mg g}^{-1}$) in comparison to CBP ($q_{max} = 273.07 \text{ mg g}^{-1}$). The results of adsorption evaluation demonstrated that ionic strength and pH played a decisive role in the adsorption rate, which was enhanced for both the pesticides by increasing the ionic strength. According to the pH_{PZC} of activated carbons and pesticides pK_a , the MCPA and 2,4-D removal were reduced with increasing of pH ($\text{pH} > 4.76$). The obtained adsorption data declare that electrostatic interaction between the activated carbons surface and pesticides were governing the adsorption mechanism.

The adsorption of diazinon from the aqueous solution was investigated using batch and fixed-bed column techniques onto a walnut shell activated carbon modified by impregnation with H_3PO_4 (WSAC; $S_{BET} = 1434 \text{ m}^2 \text{ g}^{-1}$) (Bayat et al., 2018). The adsorption results in the batch technique confirmed that the

Fig. 7. Preparation of activated carbon by microwave heating using H₃PO₄, NaOH, and sodium methanolate (MeNa) as the chemical agents, the activated carbon structure and its applicability for xenobiotics removal at optimal conditions (pH = 7, activated carbon dosage = 20 mg, time = 90 min, and temperature = 296 K). Reproduced with modification with permission from (Koutník et al., (2020)).

Fig. 8. Synthesis of activated carbons (ACs) from Tara Gum by FeCl₃-activation at different temperatures (400–1000 °C) for antipyrine adsorption at optimal conditions (pH = 7, antipyrine concentration = 50 mg L⁻¹, AC dose = 0.2 g L⁻¹, time = 60 min, and temperature = 290 K). Reproduced with modification with permission from (Bedia et al., (2018)).

pseudo-second-order kinetic and the Freundlich isotherm were the best models to analyze the experimental adsorption data. The adsorption study, due to the high adsorption capacity of WSAC in batch performance ($q_{\max} = 169.49 \text{ mg g}^{-1}$), was carried out via a fixed-bed column for compliance with the industrial scale. In the continuous fixed-bed study, the flow rate and bed length were the effective experimental parameters on the adsorption efficiency; bed $q_{\max}$ of 34.98 mg g^{-1} was achieved with flow rates = 12 L h^{-1} and bed length = 30 cm.

Sou et al. reported the starch-derived mesoporous activated carbon that was used for the elimination of 11 pesticides from aqueous solutions (Suo et al., 2019). The examined pesticides comprised atrazine, thiacloprid, pymetrozine, difenoconazole, imazalil, azoxystrobin, pyraclostrobin, trifloxystrobin, chlorantraniliprole, acetamiprid, and diuron. The adsorption results displayed that the starch-derived AC removed the pesticides very fast (5 min) and with good adsorption efficiency (atrazine = 84%,

thiacloprid = 96%, pymetrozine = 99%, difenoconazole = 92%, imazalil = 95%, azoxystrobin = 90%, pyraclostrobin = 94%, trifloxystrobin = 85%, chlorantraniliprole = 82%, acetamiprid = 96% and diuron = 96%). The evaluation of the adsorption mechanism validated that oxygen or nitrogen-containing functional groups and π -bonding networks of benzene in the AC structure promoted pesticide adsorption.

The orange peel activated carbon (OPAC) was prepared from orange peel waste for removal of chlorophenoxyacetic acid pesticides (2,4-dichlorophenoxyacetic acid; 2,4-D, 2-(2,4-dichlorophenoxy)propionic acid; 2,4-DP, 2-methyl-4-chlorophenoxyacetic acid; MCPA, methylchlorophenoxypropionic acid; MCPP and 2,4,5-trichlorophenoxyacetic acid; 2,4,5-T) from aqueous solutions (Fig. 11a) (Pandiarajan et al., 2018). The OPAC with high surface area ($592.471 \text{ m}^2 \text{ g}^{-1}$) exhibited good $q_{\max}$ for the adsorption of chlorophenoxyacetic acid pesticides (2,4-D: $q_{\max} = 515.46$, 2,4-DP: $q_{\max} = 215.52$, MCPA: $q_{\max} = 414.94$, 2,4,5-T:

Fig. 9. Preparation of NH₄Cl-modified activated carbon (NAC) from pomegranate wood wastes, the adsorption mechanism of chlortetracycline onto NAC, the presentation of surface morphology of activated carbons (NAC and SAC: standard activated carbon), and evaluating and comparing of their adsorption rate. Reproduced with modification with permission from (Alahabadi et al., (2017)).

$q_{\max} = 416.67$, and MCPP: $q_{\max} = 574.71$ mg g⁻¹). The investigation of isotherm and kinetic models proved that the experimental adsorption results fitted well with the Langmuir isotherm and pseudo-second-order kinetic. Besides, the obtained mean sorption energy ($E < 8$ kJ mol⁻¹) from Dubinin-Radushkevich isotherm suggested that the physical sorption occurred in the adsorption process.

The application of orange (*Citrus sinensis* L.) pulp activated carbon (OPAC) was also reported for the elimination of 2,4-D from aqueous media (Angin and Güneş, 2020). The adsorption results showed that OPAC with $S_{\text{BET}} = 1779$ (m² g⁻¹) and $q_{\max} = 71.94$ (mg g⁻¹) displayed good ability for the elimination of 2,4-D pesticide.

The tangerine seed activated carbons (TSAC1-9) were prepared via a one-step pyrolysis method and used for the removal of carbamate pesticides (bendiocarb, metolcarb, isoprocab, pirimicarb, carbaryl, and methiocarb) from complex solutions (Fig. 11b) (Y. Wang et al., 2020). The evaluation of carbonization temperature and time effects on adsorption efficiency showed that TSAC8 (carbonization temperature = 600 °C and carbonization time = 4 h) with a high specific surface area ($S_{\text{BET}} = 659.62$ m² g⁻¹) and large pore volume (0.6203 cc g⁻¹) displayed the fast adsorption rate. The isotherm, kinetic and thermodynamic studies exhibited that the Langmuir and pseudo-second-order models were fitted best with the experimental results, and spontaneous and

exothermic carbamate adsorption occurred onto TSAC.

An innovative approach was reported for the diuron adsorption onto different types of activated carbon fiber felt (ACFF; ACFF-CO₂(250), ACFF-CO₂(200), and ACFF-CO₂(200)/H₂O) supported on the microfiltration membrane in a gravitational filtration system (de Camargo Lima Beluci et al., 2021). Tang et al. reported the removal of trace drugs and pesticides (TDPs; pharmaceuticals: trimethoprim, sulfamethoxazole, roxithromycin, mefenamic acid, ibuprofen, diphenhydramine hydrochloride, clarithromycin, carbamazepine, and diclofenac acid, and pesticides: thiamethoxam, prometryn, prochloraz, imidacloprid, dipterex, dimethomorph, difenoconazole, chlorpyrifos, carbendazim, azoxystrobin, atrazine, and acetamiprid) from secondary effluent by four types of typical granular activated carbon (GAC, coal-based GAC, shell GAC, coconut shell GAC, and wooden GAC) (Tang et al., 2020). The shell GAC with multiple functional group contents (Boehm acidic groups = 900 (μmol g⁻¹); carboxyl: 275, lactones: 325, phenolic hydroxyl: 300, and total Boehm basic groups = 720 μmol g⁻¹), high surface area ($S_{\text{BET}} = 399.06$ m² g⁻¹) and good adsorption capacity showed a promising efficiency for removing of TDPs via physical and chemical interactions. The recently reported literature on the pesticide adsorption onto activated carbons are presented in Table 2.

Fig. 10. Preparation of activated carbons from rice and coffee husk using NaOH or ZnCl_2 as activating agents under activation temperatures of 500–800 °C for acetaminophen removal at optimal conditions (pH = 5.8, AC dosage = 0.15 g, acetaminophen concentration = 40 mg L⁻¹, time = 60 min, and temperature = 298 K). Reproduced with permission from (Paredes-Laverde et al., (2019)).

4.2.2. Graphene-based materials

Graphene is a two-dimensional (2D) crystal made of a monolayer of sp^2 hybridized carbon atoms placed in a hexagonal package that was theoretically predicted in 1947 (Wallace, 1947) but prepared in 2004 (Novoselov et al., 2004). It is a building block of many other carbonic materials such as 3D graphite, carbon nanotubes, and Buckyball's, or fullerenes (Fig. 12a) and is prepared from graphite via a mechanical cleavage, to generate the different forms of graphene, namely pristine graphene, graphene oxide (GO), and reduced graphene oxide (Shanmugham et al., 2020). The graphene oxide has a 2D structure as graphene that is created during chemical exfoliation of graphite flakes or oxidation of graphene (Fig. 12b). In the graphene oxide structure, a single sheet of carbon atoms on the basal plane, and the sides are covalently modified with oxygen-containing functional groups (carbonyl, carboxyl, epoxide, or hydroxyl) (Obodo et al., 2019). The reduced graphene oxide is a graphene oxide in which its oxygen contents are reduced by chemical, electrochemical, or thermal reduction (Fig. 12b). The reduction method is developed to distort the honeycomb hexagonal lattice of graphene oxide and also to improve its electrical conductivity (Tarcan et al., 2020). The pristine graphene with an ultra-high specific surface area, but no porosity, naturally repels water. So, the creation of narrow pores and the production of porous graphene allows rapid water permeation (Vijaya Bhaskar Reddy et al., 2019). The porous graphene sheets have been investigated for water filtration due to their capability to prevent the passage of pollutants but only allowing water to permeate through the filter (Ersan et al., 2017). The convenient properties of porous graphene help to prepare lightweight, eco-friendly, and energy-efficient water filters (Vijaya Bhaskar Reddy et al., 2019). The graphene oxides with hydrophilic functional groups are well dispersed in water and are good adsorbents in water remediation

(Shanmugham et al., 2020). Thus, graphene and its derivatives with various applications (Yusuf et al., 2019) are attractive adsorbents for the removal of inorganic and organic (Yusuf et al., 2017, 2016, 2015) pollutants from water because of their tunable properties, high surface area, and delocalized π -electron framework.

4.2.2.1. Adsorption of drugs onto graphene-based materials. Graphene is considered an ideal adsorbent for water remediation because of its high surface area and hydrophobic property that adsorbs the organic pollutants with a strong affinity.

The adsorption of 2-arylpropionic acids (carprofen, ibuprofen, indoprofen, ketoprofen, and naproxen) has been investigated via experimental and theoretical studies (Z. Wang et al., 2019).

The pH-dependent experiments exposed that the 2-arylpropionic acids adsorption onto graphene was considerably controlled by the solvent-solute interactions, besides the interactions between sorbate and adsorbent surface. Moreover, the incorporation of molecular docking study with equilibrium adsorption experiments revealed that the interactive strength of 2-arylpropionic acids with graphene was significantly correlated to the aromatic moieties rather than the hydrophobicity of sorbate molecules. Results showed that the dominant interaction for the 2-arylpropionic acids adsorption onto graphene was π - π stacking between the graphene bulk π -system and 2-arylpropionic acids aromatic moieties. The binding energy of adsorbed 2-arylpropionic acids onto graphene followed the order: carprofen < indoprofen < naproxen < ketoprofen < ibuprofen.

The GO revealed higher adsorption capacity and increased adsorptive affinity towards the polar contaminants rather than the graphene due to the presence of polar groups, hydrophilic nature, and good dispersion in water. The GO adsorption rate is related to the degree of oxidation so that highly oxidized GO displayed a

Fig. 11. (a) Adsorption of chlorophenoxyacetic acid onto the orange peel activated carbon (OPAC) obtained from orange peel waste. Reproduced with modification with permission from (Pandiarajan et al., 2018). (b) The adsorption of carbamate pesticides onto activated carbon derived from the tangerine seed waste (TSAC). Reproduced with modification with permission from (Y. Wang et al., 2020). (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)

superior adsorption rate than the GO with a lower oxidation degree (Thangavel and Venugopal, 2014). In recent years, many research investigations have been conducted for the removal of drugs using GO, for example, adsorption of trimethoprim and isoniazid antibiotics (Çalışkan Salihi et al., 2020), norfloxacin (Moreira et al., 2020), naproxen (Cigeroğlu et al., 2020), cephalixin (Wernke et al., 2020), sodium diclofenac (Guerra et al., 2019), and fluoroquinolones (ciprofloxacin, norfloxacin, and ofloxacin) (Yadav et al., 2018) from aqueous solutions.

The graphene oxide nanosheet (GOS) has been deployed for the

removal of ciprofloxacin, doxycycline, and tetracycline from aqueous streams (Rostamian and Behnejad, 2018). A comprehensive adsorption evaluation of antibiotics onto GOS revealed that the sequence of q_{max} was in the following order: tetracycline < doxycycline << ciprofloxacin. The mechanism for the adsorption of ciprofloxacin, doxycycline, and tetracycline onto GOS was assumed to occur via cation-π bonding and π-π interaction, which occurred between the ring moiety of antibiotics and the hexagonal cells of GOS. While, the functional groups of antibiotics interacted with GOS electron-rich surfaces via cation-π bonding.

Fig. 12. (a) Structures of some carbon-based nanomaterials including graphene. Reprinted with permission from (Ali et al., 2018). (b) The structural images and possible ways for the preparation of graphene oxide and reduced graphene oxide. Reprinted with permission from (Selvaraj et al., (2020)).

Although GO showed good adsorption ability for organic pollutants, its collection from the solution is relatively difficult due to its aggregation tendency because of the strong van der Waals force between single layers and its hydrophilicity due to the excess oxygen-containing groups. So, the covalent or non-covalent modification of the GO surface has recently been used to address the dispersibility and aggregation issues (Lawal et al., 2020; Shan et al., 2017).

The porous graphene oxide (PGO) has been synthesized by linking the decafluorobiphenyl (DFB) with GO sheets via a nucleophilic substitution reaction (Fig. 13a) (Shan et al., 2017) where the GO hydroxyl groups reacted with fluorine atoms of DFB to fabricate the PGO adsorbent with enlarged pore size and surface area. The GO-DFB not only displayed an improved sorption capability in comparison to GO and 3D-reduced graphene oxide, but also could be easily separated from the solution. The adsorption efficacy of GO-DFB was evaluated for the removal of sulfamethoxazole, sulfadiazine, phenacetin, paracetamol, ibuprofen, and carbamazepine from aqueous solutions. The adsorption results revealed that GO-DFB adsorbent was desirable for typical pharmaceuticals adsorption with $q_{\max}$ carbamazepine = 340.5, $q_{\max}$

sulfamethoxazole = 428.3, $q_{\max}$ sulfadiazine = 214.7, $q_{\max}$ ibuprofen = 224.3, $q_{\max}$ paracetamol = 350.6 and $q_{\max}$ phenacetin = 316.1 $\mu\text{mol g}^{-1}$. The chemical intercalating of rigid molecules proved to be a suitable method to achieve the hydrophilic and layer-stacked PGO for the effective adsorption of pharmaceuticals.

The sulfamethoxazole adsorption onto graphene oxide modified with imidazolium-based ionic liquid (GO-IL) has been reported by Ogunleye et al. (2020). The GO was prepared via the Tour method, then GO-IL was synthesized by co-precipitation of 1-tetradecyl,3-butylimidazolium ionic liquid (IL) and GO. The adsorption efficiency of GO-IL was much improved in comparison with GO due to the presence of IL long-chain carbon tail on the GO-IL surface, which increased the hydrophobicity of GO-IL and enhanced its dispersibility in aqueous solutions. The mechanism of sulfamethoxazole sorption onto GO-IL was attributed to hydrophobic effects, hydrogen bonding, and electrostatic- and π - π interactions.

The GO and non-covalent GO functionalized ionic liquid (IL: 1-hexyl,3-decahexyl imidazolium) were prepared and were applied for the elimination of carbamazepine, ketoprofen, and sulfamethoxazole from aqueous media (Fig. 13b) (Lawal et al., 2020). The

adsorption capacity and adsorption mechanism were determined through theoretical, isotherm, and kinetic studies. The ketoprofen and sulfamethoxazole adsorption onto both graphene oxide and graphene oxide modified with 1-hexyl,3-decahexyl imidazolium was significantly controlled by pH, while the carbamazepine adsorption was not considerably affected by pH variation. The graphene oxide modified with 1-hexyl,3-decahexyl imidazolium displayed superior adsorption efficacy compared to GO for the removal of carbamazepine, ketoprofen, and sulfamethoxazole. The obtained results revealed that the IL could be favorably used in GO functionalizing to improve its adsorption capacity.

4.2.2.2. Adsorption of pesticides onto graphene-based materials. The effect of graphene-based structures on the elimination of chlorpyrifos and dimethoate from the water has been investigated by Lazarević-Pašti et al. (2018). The obtained results showed that pesticide adsorption onto graphene-based-adsorbents was extremely dependent on the sorbent and sorbate structural properties, and the surface area was not the principal factor controlling the removal efficiency. The aliphatic dimethoate was preferably adsorbed onto hydrophilic oxidized graphene surfaces, whereas the graphene basal plane, containing the π electron system and high structural order, removed the chlorpyrifos selectively with aromatic moiety. The intermediate concentration of oxygen functional groups on the surface of graphene-based adsorbent led to the elimination of both the chlorpyrifos and dimethoate.

Recently, the employment of GO as an adsorbent was

investigated for the removal of toxic pesticides such as atrazine (de Souza Antônio et al., 2021), ametryn (Khoshnam et al., 2019), chlorpyrifos, and malathion (Yadav et al., 2019) from water and wastewater media.

The GO nanoplatelets were applied for the adsorption of chloridazon and its degradation metabolites (desphenyl-chloridazon: DC and methyl-desphenylchloridazon: MDC) from aqueous solutions (Yan et al., 2020). The good adsorption capacity of GO for chloridazon (67.18 mg g^{-1}), DC (34.30 mg g^{-1}), and MDC (36.85 mg g^{-1}) was established for efficient remediation via hydrogen bonding and hydrophobic- and π - π interactions.

In addition to adsorption experiments, theoretical insights into the performance of graphene-based materials for the adsorption of pesticides have been explored. The molecular adsorption mechanisms for carbaryl, catechol, and fluridone onto GO were evaluated by Wang et al. (H. Wang et al., 2020). The DFT calculation, molecular dynamics simulation, and binding free energy calculation confirmed that the van der Waals forces and π - π stacking accounted for the major adsorption interactions between the pesticides and GO.

The ability of graphene and its defected forms, hexagonal boron nitride and carbon doped hexagonal boron nitride, have been explored for the adsorption of atrazine through DFT calculation (Goli et al., 2020). The molecular orbital interactions and energy calculations were assayed to reveal the affinity of graphene, hexagonal boron nitride, and carbon doped hexagonal boron nitride towards atrazine and to estimate the magnitude of the interaction.

Fig. 13. (a) Synthesis of porous graphene oxide (GO-DFB) via the chemical intercalating of the decafluorobiphenyl molecule and representation of its adsorption capability for drug removal from aqueous media. Reproduced with modification with permission from (Shan et al., (2017)). (b) The graphene oxide preparation via Tours' method (graphite powder was reacted with the mixture of concentrated $\text{H}_3\text{PO}_4/\text{H}_2\text{SO}_4$ and KMnO_4 at 50°C for 12 h), the preparation of non-covalent GO functionalized ionic liquid (GO-IL: the GO and IL ratio = 5:1, sonication for 6 h, heating at 35°C for 12 h), and the adsorption studies of drugs onto GO-IL. Reproduced with modification with permission from (Lawal et al., (2020)).

The obtained results indicated that the physical interactions occurred between the atrazine and adsorbents, and the principal role for stabilizing the adsorbed atrazine was related to the dispersive interactions. The mobility of adsorbed atrazine onto graphene adsorbent was affected by the relatively low rotational and translational energy barriers. Whereas, in hexagonal boron nitride, the atrazine movement was limited by electrostatic attractions and polar bonds due to the high translational energy barrier. Table 3 summarizes the adsorption of drugs and pesticides onto graphene-based adsorbents from aqueous solutions.

4.2.3. Carbon nanotubes

Carbon nanotubes (CNTs), one-dimensional allotropes of carbon, are the ordered, hollow, and enrolled cylindrical graphene-based materials, comprising sp^2 -hybridized carbon atoms at the corners of the hexagon. The CNTs can be categorized into two types: I) single-walled CNTs (SWCNTs) that contain a single carbon sheet rolled into a tubular structure, and II) multi-walled CNTs (MWCNTs) that consist of concentric nested SWCNTs with a mutual longitudinal axis (Rawtani et al., 2018). The CNTs have nanometer-sized diameters (SWCNTs < 2 nm and 2 < MWCNTs < 100 nm), micrometer-sized lengths and length-to-diameter ratio > 1000 (Sarkar et al., 2018). The SWCNTs were first reported in 1952 by Radushkevich et al. (Radushkevich and Lukyanovich, 1952), and the preparation of multi-walled carbon nanotubes was described by Iijima in 1991 (Iijima, 1991). The employed techniques for the preparation of CNTs preparation include laser ablation, arc discharge, pyrolysis, chemical vapor deposition, and plasma-enhanced chemical vapor deposition (G. Rahman et al., 2019). The potential sites in CNTs responsible for the adsorption of organic micro-pollutants are I) internal sites, II) interstitial channels, III) grooves and IV) the outside surface (Jung et al., 2015). In recent years, CNTs have garnered immense interest to remove organic micro-pollutants from aqueous media. The CNTs, due to their hollow and layered structure, high surface area, small pore size, unique chemical structure, and chemical/thermal stability, serve as excellent sorbents for organic micro-pollutants adsorption (Ahmad et al., 2019). An overview of the adsorption-related information for drugs and pesticides onto CNTs from aqueous solutions is presented in this section.

4.2.3.1. Adsorption of drugs onto carbon nanotubes. The CNTs have been deployed to eliminate a wide range of drugs from water because of their excellent adsorption efficiency; commercial MWCNT was applied for the adsorption of caffeine and diclofenac from aqueous media (Gil et al., 2018). The effect of adsorption parameters such as the drug initial concentration (C_0), MWCNT dosage (m), contact time (t), and temperature (T), has been evaluated for the adsorption behavior of MWCNT. The obtained results showed that the maximum adsorption capacities of MWCNT ($q_{\max}$ caffeine = 10.1 and $q_{\max}$ diclofenac = 8.5 mg g⁻¹) were greatly affected by optimization of adsorption parameters (C_0 = 15 mg dm⁻³, m = 50 mg, t = 30 min, and T = 25 °C).

The optimization effects of adsorption parameters on the removal of carbamazepine and dorzolamide from water using MWCNT were also investigated by Ncibi et al. (Ncibi and Sillanpää, 2017). The results showed that ultrasound application in the adsorption process improved the $q_{\max}$ of MWCNT for carbamazepine (224.6 mg g⁻¹) and dorzolamide (78.8 mg g⁻¹) removal (21 and 30% enhancement) under the optimal conditions.

The MWCNT was prepared with a particle size of 10–40 nm and a specific surface area of 9.1 m² g⁻¹ by ALOthman et al. (ALOthman et al., 2019). The 4-tert-octylphenol endocrine disruptor was adsorbed onto the synthesized MWCNT from the water at a high

adsorption rate (94%); isotherm, kinetic and thermodynamic studies were performed for evaluating the adsorption mechanism. The adsorption results revealed that fast, spontaneous, and exothermic removal of 4-tert-octylphenol occurred onto MWCNT under a liquid film diffusion mechanism.

The adsorption behavior of propranolol (PRO) onto MWCNT was investigated by evaluation of ionic strength, pH, and humic acid (HA) effects on the adsorption process (Nie et al., 2020). Also, the influences of metoprolol tartrate and carbamazepine drugs on PRO removal were evaluated and compared, including the PRO adsorption mechanisms onto MWCNT. The adsorption results exhibited that the PRO adsorption was significantly changed under different pHs, and it was accelerated in the presence of HA without showing any significant changes with varying solution ionic strength. The obtained results of carbamazepine and metoprolol tartrate effects on PRO adsorption showed that the mechanism of PRO adsorption onto MWCNT was mainly attributed to hydrogen bond, and hydrophobic and π - π EDA interactions.

The mechanism of sulfamethoxazole (SMZ) interaction with various MWCNTs (MWCNT10, MWCNT15, MWCNT15-OH, MWCNT15-COOH, and N-doped MWCNTs) were investigated in the presence of HA in water (Wang et al., 2017). The SMZ adsorption onto CNTs was reduced with the increase of HA concentration due to the binding interaction between free HA and SMZ molecules in water, besides the competitive occupation of MWCNTs adsorption sites by the HA.

The MWCNT was used for the adsorption of PPCPs (acetaminophen, carbendazim, caffeine, and triclosan) from wastewater via pre-coagulation technique (Wang et al., 2018). The wastewater pre-coagulation effect was evaluated on adsorptive filtration of PPCP by using the MWCNT membrane. Effluent organic matters (biopolymers, and primarily humic substances) in the wastewater competed with PPCPs for adsorption sites of MWCNT. Thus, the MWCNT adsorption capacity was markedly reduced for PPCPs in wastewater remediation. The pre-coagulation of wastewater effectively diminished the competitive adsorption of organic matters, and improved the adsorption efficiency of acetaminophen, carbendazim, caffeine, and triclosan onto MWCNT. These results exhibited that the pre-coagulation initially removed the biopolymers and especially humic substances, then the PPCPs adsorption occurred onto MWCNT. The adsorption capacity of MWCNT-membrane for PPCPs removal was improved with the removal of humic substances; whereas, it was diminished with biopolymer removal due to the reduced membrane permeability and relieved membrane fouling. This finding demonstrated that pre-coagulation is a viable pretreatment method for the filtration of PPCPs by MWCNT from wastewater.

The influence of molecular structure on the sulfonamides sorption affinity (sulfametoxydiazine, sulfamethazine, sulfamerazine, sulfanilamide, sulfadimethoxine, and sulfadiazine) onto hydroxylated MWCNT (HO-MWCNT) was evaluated via batch technique and DFT calculations (Liu et al., 2020). The sulfonamides adsorption onto HO-MWCNT was controlled by chemisorption, and the $q_{\max}$ of HO-MWCNT for six sulfonamides was in the order: sulfametoxydiazine > sulfamethazine > sulfadiazine > sulfadimethoxine > sulfamerazine > sulfanilamide. The sulfonamides with higher polarity were adsorbed onto polar HO-MWCNT with higher adsorption affinity, based on the similarity and inter-miscibility theory. The results showed that the sulfonamides adsorption onto HO-MWCNT was critically affected by the molecular structure.

The oxidized MWCNT (O-MWCNT) has been synthesized and used for the removal of pefloxacin (PEF) and Cu(II) from contaminated water (Zhou et al., 2019). The mutual effects of PEF and Cu(II) adsorption onto O-MWCNT were investigated with single and binary systems by evaluation of adsorption isotherm, kinetic, and

Table 3

The recent drugs and pesticide adsorption onto carbon nanotubes and graphene-based adsorbents.

Adsorbent	Adsorbate	Adsorption conditions	Adsorption isotherm	Adsorption kinetic	$q_{\max}$ (mg g ⁻¹)	Adsorption mechanism	Reference
GO ^a	Ciprofloxacin	pH = 7.0 $C_0 = 5 \text{ mg L}^{-1}$ $V = 100 \text{ mL}$, $m = 50 \text{ mg}$ $T = 298 \text{ K}$, $t = 50 \text{ min}$	Langmuir	Pseudo-second-order	18.65	Hydrogen bond	Yadav et al. (2018)
GO	Norfloxacin	pH = 7.0 $C_0 = 5 \text{ mg L}^{-1}$ $V = 100 \text{ mL}$, $m = 50 \text{ mg}$ $T = 298 \text{ K}$, $t = 50 \text{ min}$	Langmuir	Pseudo-second-order	24.93	Hydrogen bond	Yadav et al. (2018)
GO	Ofloxacin	pH = 4.0 $C_0 = 5 \text{ mg L}^{-1}$ $V = 100 \text{ mL}$, $m = 50 \text{ mg}$ $T = 298 \text{ K}$, $t = 50 \text{ min}$	Langmuir	Pseudo-second-order	40.65	Hydrogen bond	Yadav et al. (2018)
GO	Ciprofloxacin	pH = 6.0 $C_0 = - \text{mg L}^{-1}$ $V = 10 \text{ mL}$, $m = 10 \text{ mg}$ $T = 299 \text{ K}$, $t = 100 \text{ min}$	Hill Toth	Elovich	173.4	Cation- π bonding π - π interaction	Rostamian and Behnejad (2018)
GO	Doxycycline	pH = 6.0 $C_0 = - \text{mg L}^{-1}$ $V = 10 \text{ mL}$, $m = 10 \text{ mg}$ $T = 299 \text{ K}$, $t = 200 \text{ min}$	Hill Toth	Pseudo-second-order	131.9	Cation- π bonding π - π interaction	Rostamian and Behnejad (2018)
GO	Tetracycline	pH = 6.0 $C_0 = - \text{mg L}^{-1}$ $V = 10 \text{ mL}$, $m = 10 \text{ mg}$ $T = 299 \text{ K}$, $t = 200 \text{ min}$	Hill Toth	Pseudo-second-order	96.98	Cation- π bonding π - π interaction	Rostamian and Behnejad (2018)
GO	Sodium diclofenac	pH = 6.2 $C_0 = 10 \text{ mg L}^{-1}$ $V = - \text{mL}$, $m = 250 \text{ mg L}^{-1}$ $T = 298 \text{ K}$, $t = 300 \text{ min}$	Langmuir	Pseudo-second-order	128.74	Electrostatic attraction Hydrogen bond Hydrophobic interaction π - π bond	Guerra et al. (2019)
GO	Metformin	pH = 6.26 $C_0 = 521 \text{ mg L}^{-1}$ $V = - \text{mL}$, $m = 126.02 \text{ mg L}^{-1}$ $T = 318.5 \text{ K}$, $t = - \text{min}$	Sips Hill	Pseudo-first-order	122.61	Van der Waals forces π - π interactions	Balasubramani et al. (2020)
GO	Cephalexin	pH = 7.0 $C_0 = 100 \text{ mg L}^{-1}$ $V = 25 \text{ mL}$, $m = 10 \text{ mg}$ $T = 298 \text{ K}$, $t = 420 \text{ min}$	Langmuir	Pseudo-second-order	164	—	Wernke et al. (2020)
GO	Trimethoprim	pH = 8.0 $C_0 = 50 \text{ mg L}^{-1}$ $V = - \text{mL}$, $m = 15 \text{ mg}$ $T = 298 \text{ K}$, $t = 60 \text{ min}$	Freundlich	Pseudo-second-order	204.08	Cation- π and π - π dispersion interactions	Çalışkan Salihi et al. (2020)
GO	Isoniazid	pH = 2.0 $C_0 = 25 \text{ mg L}^{-1}$ $V = - \text{mL}$, $m = 15 \text{ mg}$ $T = 298 \text{ K}$, $t = 60 \text{ min}$	Langmuir	Intraparticle diffusion	13.89	Electrostatic attraction	Çalışkan Salihi et al. (2020)
GO	Norfloxacin	pH = 7.0 $C_0 = 30 \text{ mg L}^{-1}$ $V = 10 \text{ mL}$, $m = 200 \text{ mg L}^{-1}$ $T = 303 \text{ K}$, $t = 30 \text{ min}$	Sips	Pseudo-second-order	374.9	Van der Waals forces π - π interaction Hydrogen bond	Moreira et al. (2020)
GO	Naproxen	pH = 4.0 $C_0 = 10 \text{ mg L}^{-1}$ $V = - \text{mL}$, $m = 30 \text{ mg}$ $T = 298 \text{ K}$, $t = 60 \text{ min}$	Dubinin-Radushkevich	Pseudo-second-order	15.16	Chemical adsorption	Cigeroğlu et al. (2020)
PG ^b	Atenolol	pH = 7.5 $C_0 = 10 \text{ mg L}^{-1}$ $V = 20 \text{ mL}$, $m = 5 \text{ mg}$ $T = 295 \text{ K}$, $t = 120 \text{ min}$	Toth	Pseudo-first-order	1455.4	Physical adsorption	Khalil et al. (2020)
PG	Carbamazepine	pH = 7.5 $C_0 = 10 \text{ mg L}^{-1}$ $V = 20 \text{ mL}$, $m = 5 \text{ mg}$ $T = 295 \text{ K}$, $t = 120 \text{ min}$	Freundlich	Pseudo-second-order	191.67	Physical adsorption	Khalil et al. (2020)
PG	Ciprofloxacin	pH = 7.5 $C_0 = 10 \text{ mg L}^{-1}$ $V = 20 \text{ mL}$, $m = 5 \text{ mg}$ $T = 295 \text{ K}$, $t = 120 \text{ min}$	Temkin	Pseudo-second-order	409.15	Physical adsorption	Khalil et al. (2020)
PG	Diclofenac	pH = 7.5 $C_0 = 10 \text{ mg L}^{-1}$ $V = 20 \text{ mL}$, $m = 5 \text{ mg}$ $T = 295 \text{ K}$, $t = 120 \text{ min}$	Sips	Pseudo-second-order	91.59	Physical adsorption	Khalil et al. (2020)
PG	Gemfibrozil	pH = 7.5 $C_0 = 10 \text{ mg L}^{-1}$ $V = 20 \text{ mL}$, $m = 5 \text{ mg}$ $T = 295 \text{ K}$, $t = 120 \text{ min}$	Sips	Pseudo-second-order	160.86	Physical adsorption	Khalil et al. (2020)

(continued on next page)

Table 3 (continued)

Adsorbent	Adsorbate	Adsorption conditions	Adsorption isotherm	Adsorption kinetic	$q_{\max}$ (mg g ⁻¹)	Adsorption mechanism	Reference
PG	Ibuprofen	pH = 7.5 $C_0 = 10$ mg L ⁻¹ V = 20 mL, m = 5 mg T = 295 K, t = 120 min	Langmuir	Pseudo-second-order	110.30	Physical adsorption	Khalil et al. (2020)
GO-IL ^c	Sulfamethoxazole	pH = 6.28 $C_0 = 10$ mg L ⁻¹ V = 10 mL, m = 0.9 g L ⁻¹ T = 299 K, t = 60 min	Freundlich	Pseudo-second-order	27.25	Hydrophobic interaction π - π interaction Hydrogen bond Electrostatic attraction	Ogunleye et al. (2020)
GO	Carbamazepine	pH = 2.0 $C_0 = 50$ mg L ⁻¹ V = - mL, m = 10 mg T = 299 K, t = 30 min	Langmuir	Pseudo-second-order	28.54	π - π interactions Hydrophobic interaction	Lawal et al. (2020)
GO-IL	Carbamazepine	pH = 2.0 $C_0 = 50$ mg L ⁻¹ V = - mL, m = 10 mg T = 299 K, t = 10 min	Redlich-Peterson	Pseudo-second-order	61.88	π - π interactions Hydrophobic interaction	Lawal et al. (2020)
GO	Ketoprofen	pH = 4.0 $C_0 = 50$ mg L ⁻¹ V = - mL, m = 10 mg T = 299 K, t = 30 min	Langmuir	Pseudo-second-order	16.45	π - π interactions Hydrophobic Interaction Electrostatic attraction	Lawal et al. (2020)
GO-IL	Ketoprofen	pH = 8.0 $C_0 = 50$ mg L ⁻¹ V = - mL, m = 10 mg T = 299 K, t = 10 min	Redlich-Peterson	Pseudo-second-order	87.88	π - π interactions Hydrophobic interaction Electrostatic attraction	Lawal et al. (2020)
GO	Sulfamethoxazole	pH = 4.0 $C_0 = 50$ mg L ⁻¹ V = - mL, m = 10 mg T = 299 K, t = 30 min	Langmuir	Pseudo-second-order	21.33	π - π interactions Hydrophobic Interaction Electrostatic attraction	Lawal et al. (2020)
GO-IL	Sulfamethoxazole	pH = 6.0 $C_0 = 50$ mg L ⁻¹ V = - mL, m = 10 mg T = 299 K, t = 10 min	Redlich-Peterson	Weber-Morris	99.16	π - π interactions Hydrophobic Interaction Electrostatic attraction	Lawal et al. (2020)
MWCNT ^d	Propranolol	pH = 6.5 $C_0 = 25$ mg L ⁻¹ V = 40 mL, m = 10 mg T = 299 K, t = 30 min	Sips	Pseudo-second-order	380.22	π - π EDA interaction Hydrophobic interaction Hydrogen bond	Nie et al. (2020)
N-CNT ^e	Dimetridazole	pH = 7.0 $C_0 = -$ mg L ⁻¹ V = 40 mL, m = 10 mg T = 299 K, t = 40 min	—	—	40.93	π - π dispersive interactions Electrostatic interaction hydrophobic interactions	Carrales-Alvarado et al. (2020)
SWCNT ^f	Dimetridazole	pH = 7.0 $C_0 = -$ mg L ⁻¹ V = 40 mL, m = 10 mg T = 299 K, t = 40 min	—	—	81.86	π - π dispersive interactions Electrostatic interaction hydrophobic interactions	Carrales-Alvarado et al. (2020)
MWCNT	Dimetridazole	pH = 7.0 $C_0 = -$ mg L ⁻¹ V = 40 mL, m = 10 mg T = 299 K, t = 40 min	—	—	45.16	π - π dispersive interactions Electrostatic interaction hydrophobic interactions	Carrales-Alvarado et al. (2020)
MWCNT-COOH ^g	Dimetridazole	pH = 7.0 $C_0 = -$ mg L ⁻¹ V = 40 mL, m = 10 mg T = 299 K, t = 40 min	—	—	32.46	π - π dispersive interactions Electrostatic interaction hydrophobic interactions	Carrales-Alvarado et al. (2020)
N-CNT	Metronidazole	pH = 7.0 $C_0 = -$ mg L ⁻¹ V = 40 mL, m = 10 mg T = 299 K, t = 40 min	—	—	56.49	π - π dispersive interactions Electrostatic interaction hydrophobic interactions	Carrales-Alvarado et al. (2020)
SWCNT	Metronidazole	pH = 7.0 $C_0 = -$ mg L ⁻¹ V = 40 mL, m = 10 mg T = 299 K, t = 40 min	—	—	100.98	π - π dispersive interactions Electrostatic interaction hydrophobic interactions	Carrales-Alvarado et al. (2020)
MWCNT	Metronidazole	pH = 7.0 $C_0 = -$ mg L ⁻¹ V = 40 mL, m = 10 mg T = 299 K, t = 40 min	—	—	46.21	π - π dispersive interactions Electrostatic interaction hydrophobic interactions	Carrales-Alvarado et al. (2020)
MWCNT-COOH	Metronidazole	pH = 7.0 $C_0 = -$ mg L ⁻¹ V = 40 mL, m = 10 mg T = 299 K, t = 40 min	—	—	46.21	π - π dispersive interactions Electrostatic interaction hydrophobic interactions	Carrales-Alvarado et al. (2020)
MWCNT-OH ^h	Sulfanilamide	pH = 3.0 $C_0 = 20$ mg L ⁻¹ V = 50 mL, m = 1 g L ⁻¹ T = 298 K, t = 20 min	Langmuir	General order	67.15	Electrostatic attraction Hydrogen bond EDA interaction Lewis acid-base interaction	Liu et al. (2020)
MWCNT-OH	Sulfamerazine	pH = 3.0 $C_0 = 20$ mg L ⁻¹	Liu	Pseudo-second-order	115.74	Electrostatic attraction Hydrogen bond	Liu et al. (2020)

Table 3 (continued)

Adsorbent	Adsorbate	Adsorption conditions	Adsorption isotherm	Adsorption kinetic	$q_{\max}$ (mg g ⁻¹)	Adsorption mechanism	Reference
MWCNT-OH	Sulfadimethoxine	V = 50 mL, m = 1 g L ⁻¹ T = 298 K, t = 20 min pH = 3.0 C ₀ = 20 mg L ⁻¹	Langmuir	General order	128.62	EDA interaction Lewis acid-base interaction Electrostatic attraction Hydrogen bond	Liu et al. (2020)
MWCNT-OH	Sulfadiazine	V = 50 mL, m = 1 g L ⁻¹ T = 298 K, t = 20 min pH = 3.0 C ₀ = 20 mg L ⁻¹	Langmuir	General order	132.33	EDA interaction Lewis acid-base interaction Electrostatic attraction Hydrogen bond	Liu et al. (2020)
MWCNT-OH	Sulfamethazine	V = 50 mL, m = 1 g L ⁻¹ T = 298 K, t = 20 min pH = 3.0 C ₀ = 20 mg L ⁻¹	Liu	Pseudo-second-order	141.55	EDA interaction Lewis acid-base interaction Electrostatic attraction Hydrogen bond	Liu et al. (2020)
MWCNT-OH	Sulfametoxydiazine	V = 50 mL, m = 1 g L ⁻¹ T = 298 K, t = 20 min pH = 3.0 C ₀ = 20 mg L ⁻¹	Liu	Pseudo-second-order	177.54	EDA interaction Lewis acid-base interaction Electrostatic attraction Hydrogen bond	Liu et al. (2020)
MWCNT	4-Tert-octylphenol	V = 50 mL, m = 1 g L ⁻¹ T = 298 K, t = 20 min pH = 10.0 C ₀ = 0.3 mg L ⁻¹	Temkin	Elovich	0.178	Liquid film diffusion	ALothman et al. (2019)
MWCNT	Pefloxacin	V = 5 mL, m = 2.5 mg T = 298 K, t = 70 min pH = 7.0 C ₀ = 50 mg L ⁻¹	Langmuir	Pseudo-second-order	45.16	π - π interactions Hydrogen bond	Zhou et al. (2019)
O-MWCNT ⁱ	Pefloxacin	V = - mL, m = - mg T = 298 K, t = 240 min pH = 6.0 C ₀ = 50 mg L ⁻¹	Freundlich	Pseudo-second-order	61.11	π - π interactions Hydrogen bond Electrostatic interaction	Zhou et al. (2019)
MWCNT	Caffeine	V = - mL, m = 50 mg T = 298 K, t = 30 min pH = 7.0 C ₀ = 5 mg L ⁻¹	Toth	Pseudo-second-order	10.1	Chemical adsorption	Gil et al. (2018)
MWCNT	Diclofenac	V = 10 mL, m = 100 mg T = 298 K, t = 30 min pH = 7.0 C ₀ = 15 mg L ⁻¹	Toth	Pseudo-first-order	8.5	Chemical adsorption	Gil et al. (2018)
MWCNT	Carbamazepine	V = 10 mL, m = 0.1 g L ⁻¹ T = 298 $\pm$ 2 K, t = 30 min pH = 5.8 C ₀ = 50 mg L ⁻¹	Brouers-Sotolongo	—	224.6	Hydrophobic Interaction π - π interaction	Ncibi and Sillanpää (2017)
MWCNT	Dorzolamide	V = 10 mL, m = 0.1 g L ⁻¹ T = 298 $\pm$ 2 K, t = 30 min pH = 8.1 C ₀ = 50 mg L ⁻¹	Brouers-Sotolongo	—	78.8	Hydrophobic Interaction π - π interaction	Ncibi and Sillanpää (2017)
F-g-CN ^j	Diquat dibromide	V = 50 mL, m = 10 mg T = 298 K, t = 30 min pH = 7.0 C ₀ = 40 mg L ⁻¹	Langmuir	Pseudo-second-order	159.3	Electrostatic attractions Chemical adsorption	Liang et al. (2021)
NSGO ^k	Atrazine	V = 20 mL, m = 10 mg T = 298 K, t = 24 h pH = 9.0 C ₀ = 10 mg L ⁻¹	Langmuir	Pseudo-second-order	23.84	Van der Waals force Hydrogen bond	de Souza Antônio et al. (2021)
GO	Chloridazon	V = 10 mL, m = 5 mg T = 298 K, t = 24 h pH = 6.0 C ₀ = 125 mg L ⁻¹	Langmuir	—	67.18	Hydrogen bond Hydrophobic interaction π - π interaction	Yan et al. (2020)
NSGO	Ametryn	V = 25 mL, m = 15 mg T = 303 K, t = 17 min pH = 5.5 C ₀ = 10 mg L ⁻¹	Langmuir	Pseudo-second-order	47.2	—	Khoshnam et al. (2019)
GO	Chlorpyrifos	V = 100 mL, m = 500 mg L ⁻¹ T = R.T., t = 15 min pH = 7.0 C ₀ = 27.96 mg L ⁻¹	Langmuir	Pseudo-second-order	98.04	Hydrogen bond	Yadav et al. (2019)
GO	Malathion	V = 100 mL, m = 500 mg L ⁻¹ T = R.T., t = 30 min pH = 7.0 C ₀ = 61.5 mg L ⁻¹	Langmuir	Pseudo-second-order	1666.67	Hydrogen bond	Yadav et al. (2019)
MWCNT	Diuron	V = 100 μ g L ⁻¹ pH = 7.0 C ₀ = 100 μ g L ⁻¹	Langmuir	Liquid film diffusion	0.108	Hydrogen bond Hydrophobic interaction π - π interactions	Al-Shaalan et al. (2019)

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Table 3 (continued)

Adsorbent	Adsorbate	Adsorption conditions	Adsorption isotherm	Adsorption kinetic	$q_{\max}$ (mg g ⁻¹)	Adsorption mechanism	Reference
MWCNT	Fenuron	V = 10 mL, m = 20 mg T = 298 K, t = 60 min pH = 7.0 C ₀ = 100 µg L ⁻¹ V = - mL, m = 2 g L ⁻¹ T = 298 K, t = 60 min	Freundlich	Liquid film diffusion	0.107	Physical adsorption	(I. Ali et al., 2019a)
MWCNT	2,4-Dichloro phenoxyacetic acid	pH = 7.0 C ₀ = 100 mg L ⁻¹ V = 100 mL, m = 0.1 g T = 298 ± 2 K, t = - min	Thomas Yan	Pseudo-second-order	208.19	—	Bahrami et al. (2018)
CNT	2,4-Dichloro phenoxyacetic acid	pH = -C ₀ = 102 mg L ⁻¹ V = - mL, m = - g T = 283 K, t = 50 min	Langmuir	Pseudo-second-order	84.03	Physical adsorption	Hue et al. (2018)
CNT	Diuron	pH = -C ₀ = 20 mg L ⁻¹ V = 25 mL, m = 0.4 g L ⁻¹ T = 303 K, t = 240 min	Hill	Pseudo-second-order	40.37	Physical adsorption	Deokar et al. (2017)
PMWCNT	Thiamethoxam	pH = 9.0 C ₀ = 150 mg L ⁻¹ V = - mL, m = - g T = 298 K, t = 5 min	Langmuir	Pseudo-second-order	38.18	Hydrogen bond Hydrophobic interaction π-π interactions Electrostatic attractions	Panic et al. (2017)
FMWCNT ^d	Thiamethoxam	pH = 9.0 C ₀ = 150 mg L ⁻¹ V = - mL, m = - g T = 298 K, t = 210 min	Freundlich	Pseudo-second-order	95.43	Hydrogen bond Hydrophobic interaction π-π interactions Electrostatic attractions	Panic et al. (2017)

a) **GO**: Graphene oxide.

b) **PG**: Porous graphene.

c) **GO-IL**: Graphene oxide modified with imidazolium-based ionic liquid.

d) **MWCNT**: Multi-walled carbon nanotube.

e) **N-CNT**: Nitrogen-doped carbon nanotube.

f) **SWCNT**: Single-walled carbon nanotube.

g) **MWCNT-COOH**: MWCNT functionalized with carboxylic groups.

h) **MWCNT-OH**: Hydroxylated MWCNT.

i) **O-MWCNT**: Oxidized MWCNT.

j) **F-g-CN**: Functionalized graphite carbon nitride.

k) **NSGO**: Nanosheet graphene oxide.

l) **FMWCNT**: Functionalized MWCNT.

thermodynamic. There were site enhancement and competition between PEF and Cu(II) on O-MWCNT in a binary system; removal of PEF/Cu(II) was promoted at a low concentration of PEF/Cu(II) (≤ 25 ppm) and was inhibited at a higher concentration. The results exhibited that the PEF/Cu(II) functioned as a bridge between O-MWCNT and PEF/Cu(II). The PEF was adsorbed onto O-MWCNT with hydrogen bond and π - π interactions.

The influence of surface morphology and chemistry of CNTs (SWCNT, CNT doped with nitrogen: N-CNT, MWCNT, and carboxylic acid functionalized MWCNT: MWCNT-COOH) on their sorption affinity towards metronidazole and dimetridazole from aqueous solutions has been studied (Carralés-Alvarado et al., 2020). The adsorption efficiency relied remarkably on the surface area (S_{BET} SWCNT = 404, S_{BET} N-CNT = 371, S_{BET} MWCNT = 144, and S_{BET} MWCNT-COOH = 115 m² g⁻¹) and declined linearly with the concentration of carboxylic and phenolic sites per unit surface area. The SWCNT, with the highest surface area and basic surface in comparison to other CNTs with the acidic surface, presented the highest $q_{\max}$ towards dimetridazole (84 mg g⁻¹) and metronidazole (101 mg g⁻¹); adsorption of dimetridazole and metronidazole onto CNTs was mainly attributed to the π - π stacking interactions, which were promoted by basic groups.

The O-MWCNT has been deployed as an adsorbent for the miniaturized solid-phase extraction of progestins (19-norethisterone acetate, medroxyprogesterone acetate, and levonorgestrel) from the water before their HPLC-UV determination (Aguinaga Martínez et al., 2020). The O-MWCNT with polar functional groups exhibited an excellent adsorption performance for progestins. Attractive features of solid-phase extraction method for

the adsorption of progestins onto O-MWCNT included low usage of organic solvents and the pre-concentration factor (up to 100).

The pristine and HCl-treated MWCNTs have been used as solid-phase extraction adsorbent for the separation and enrichment of pharmaceuticals (azithromycin, bisoprolol, carbamazepine, cilazapril, clopidogrel, diazepam, diclofenac, enalapril, erythromycin, lorazepam, metoprolol, simvastatin, sulfamethoxazole, 4-acetylaminoantipyrine, and 4-formylaminoantipyrine) from the surface and ground waters, before liquid chromatography-tandem mass spectrometry analysis (Lalović et al., 2017). The adsorption efficiency of selected pharmaceuticals was favorably affected by the chemical treatment of MWCNT due to a decrease in the electron acceptor groups of treated MWCNT surface.

The molecular dynamics simulations in water were applied for the adsorption of ciprofloxacin in the neutral and zwitterionic forms onto SWCNT (Veclani and Melchior, 2020). The strongly negative values of interaction energies (E_{int}) suggested preferential sorption on the SWCNT internal wall. Also, the calculated free energy exhibited that the ciprofloxacin adsorption spontaneously occurred onto SWCNT; adsorbed ciprofloxacin were frequently oriented parallel to the interacting surface through π - π interactions. The ciprofloxacin-ciprofloxacin H-bonds analysis in water showed that ciprofloxacin was dispersed on the SWCNT. However, eight molecules of ciprofloxacin were gathered inside the SWCNT, which resulted in some aggregation.

4.2.3.2. Adsorption of pesticides onto carbon nanotubes. Besides drugs, pesticides are another class of pollutants that have been extensively removed using CNTs in aqueous solutions. The

synthesized CNT from plastic waste was applied as an effective adsorbent for the diuron elimination from aqueous media (Deokar et al., 2017). This research established that the plastic waste derived CNT with $q_{\max} = 103.73 \text{ mg g}^{-1}$ could be a suitable adsorbent for the water treatment.

The low-cost MWCNT has been prepared by using Ni/MgO compound in particle sizes ranging from 10.0 to 40.0 nm and a surface area of $9.10 \text{ m}^2 \text{ g}^{-1}$ (Al-Shaalan et al., 2019). Efficient diuron removal (90.5%) was achieved using the synthesized MWCNT ($q_{\max} = 132.5 \text{ mg g}^{-1}$) from water at optimized conditions ($C_{\text{diuron}} = 100.0 \text{ } \mu\text{g L}^{-1}$, $t = 60 \text{ min}$, $C_{\text{MWCNT}} = 2.0 \text{ g L}^{-1}$, $\text{pH} = 7.0$, and $T = 25 \text{ }^\circ\text{C}$). The statistical adsorption data confirmed the prompt adsorption of diuron onto MWCNT in an endothermic manner. The supra-molecular adsorption mechanism was designated by modeling readings and calculating the energy and binding affinity of diuron with MWCNTs. The interactions between diuron and MWCNT involved the π -donor hydrogen bonds, π -alkyl bonds, π - π stacked, and π - π T-shaped bonds. These results established that the physisorption of diuron and the creation of diuron-MWCNT composite comprised sixteen hydrophobic connections and two hydrogen bonds.

Likewise, the Ni/MgO-derived MWCNT has been deployed for the adsorption of fenuron from water through simulation studies for mechanistic insight (I. Ali et al., 2019a). The results revealed that the physisorption of fenuron occurred onto synthesized MCNT, and there was one π - π T-shaped, seven π - π stacked, three π -alkyl, and one π - σ type of hydrophobic interactions between fenuron and MWCNT.

In another study, the isotherm, kinetic, and thermodynamic of thiamethoxam adsorption onto MWCNT were evaluated by Pani et al. for predicting the adsorption mechanism (Panic et al., 2017). To clarify the effect of nanotube structure on the sorption mechanism, the adsorption process was evaluated for both the pristine and nitric acid-modified MWCNT. The introduction of functional groups possessing an oxygen atom in the structure of MWCNT positively affected the thiamethoxam removal from water. Based on the adsorption results, the thiamethoxam removal by MWCNTs occurred with physicochemical properties, and the electron donor-acceptor interactions (aromatic π - π interactions) being the dominant mechanism for the adsorption of thiamethoxam onto MWCNTs. The thermodynamic data showed the spontaneous and exothermic nature of thiamethoxam removal, regardless of the MWCNTs type.

The optimization of malathion removal from water using MWCNT has been reported by Dehghani et al. (2017). Based on the obtained results from both the statistical and three-factor response surface modeling (RSM) analyses for 26 steps of experiments, the MWCNTs could be efficiently applied for the removal of ~100% malathion at the optimal conditions ($C_0 = 6 \text{ mg L}^{-1}$, $C_{\text{MWCNT}} = 0.5 \text{ g L}^{-1}$, $t = 30 \text{ min}$, $\text{pH} = 7$, and $T = 24 \pm 2 \text{ }^\circ\text{C}$).

Also, the effect of adsorption optimization on the diazinon removal was investigated using MWCNT from the water in a batch technique (Dehghani et al., 2019). Optimizing of the adsorption parameters showed that MWCNT adsorbed 100% of diazinon from water under optimal conditions ($C_{\text{diazinon}} = 0.3 \text{ mg L}^{-1}$, $C_{\text{MWCNT}} = 0.1 \text{ g L}^{-1}$, $t = 15 \text{ min}$ at $\text{pH} = 4$ and 7). The MWCNT efficiency for diazinon adsorption in acidic pH could be improved because of the hydroxyl group's protonation.

The adsorption of 2,4-D from aqueous solution onto CNT ($S_{\text{BET}} = 267 \text{ m}^2 \text{ g}^{-1}$) was investigated via batch technique (Hue et al., 2018). The spontaneous and exothermic adsorption of 2,4-D has occurred onto CNT with $q_{\max} = 84.03 \text{ mg g}^{-1}$ under optimal conditions ($C_0 = 102 \text{ mg L}^{-1}$, $t = 50 \text{ min}$, and $T = 283 \text{ K}$). Likewise, the performance of MWCNT ($S_{\text{BET}} = 200 \text{ m}^2 \text{ g}^{-1}$ and $\text{pH}_{\text{PZC}} = 4.1$) was explored for the adsorption of 2,4-D in a fixed-bed column system

(Bahrami et al., 2018). The higher removal of 2,4-D ($\text{pK}_a = 2.64$) occurred at the smaller flow rate (0.5 mL min^{-1}), lower concentration of 2,4-D (50 mg L^{-1}), larger bed depth (9 cm), and lower pH (2). The results established that the MWCNTs could be an excellent choice for the adsorption of pesticide but the adsorption equilibrium time is a significant factor.

Documented literature described the adsorption of chlorophenols (2,4,6-trichlorophenol: 2,4,6-TCIP, 2,6-dichlorophenol: 2,6-DCIP, 3,4-dichlorophenol: 3,4-DCIP, 2,4-dichlorophenol: 2,4-DCIP, 4-chlorophenol: 4-CIP, and 2-chlorophenol: 2-CIP) onto MWCNTs (MWCNT 8–15 nm in outer diameter (OD): MWCNT > 50 nm OD, helical MWCNT 100–200 nm OD, MWCNT-COOH < 8 nm OD, and MWCNT-COOH > 50 nm OD) via DFT method (Watkins et al., 2017). The MWCNT-COOH < 8 nm OD has revealed the highest adsorption for all the investigated chlorophenols. The DFT results affirmed that the physisorption or non-covalent interaction between the chlorophenols benzene ring and CNTs was the principal sorption mechanism, while charge transfer from the functional groups was considered less effective in the adsorption mechanism. Moreover, five structure-property relationship models reliably defined features of the intramolecular interactions.

The use of carbon nanotube as adsorbents for the elimination of drugs and pesticides from aqueous solutions is summarized in Table 3.

4.2.4. Carbon and graphene quantum dots

Carbon quantum dots are tiny fluorescent carbon nanoparticles with dimensions of less than 10 nm in diameter, and graphene quantum dots are a subset of carbon quantum dots generally derived from graphene/graphene oxide. The graphene quantum dots are small graphene sheets with lateral dimensions less than 10 nm with less than 10 graphene layers (Facure et al., 2020). The carbon quantum dots and graphene quantum dots, known as zero-dimensional carbon based nanomaterials, are synthesized by using bottom-up and top-down methods (Irvani and Varma, 2020). They are commonly composed of H, C, N, and O, of which O and C are the most abundant elements because of the -COOH functional group existence. The carbon quantum dots and graphene quantum dots with their eminent physical-chemical properties including potent chemical inertness, low toxicity, plenty of edge functionalization sites, and high surface areas have been attracting increasing attention in various fields such as sensing (M. Z. Li et al., 2019a,b), detection (Chu et al., 2020), extraction (Mohebbi et al., 2020), and membrane fabrication (Zhao and Chung, 2018), along with wastewater treatment via adsorption (Anand et al., 2020) and photocatalytic (Lee et al., 2020) techniques.

The graphene quantum dots were synthesized by using the microwave-assisted hydrothermal method, and applied as an adsorbent ($q_{\max} = 130.10 \text{ mg g}^{-1}$) for the removal of 'oxamyl' carbamate pesticide from aqueous solutions (Agarwal et al., 2016). The isotherm, kinetic, and thermodynamic of adsorption process were investigated for proposing the adsorption mechanism. The results revealed that the adsorption isotherm and kinetic data were well fitted with the Langmuir isotherm and pseudo-second-order kinetic model. The oxamyl adsorption was occurred via feasible, spontaneous and endothermic process, and the adsorption mechanism was comprised the ionic interactions and electrostatic interactions between graphene quantum dots and oxamyl cation.

Despite the outstanding properties of carbon quantum dots and graphene quantum dots for removal of organic contaminants from water, very few researches have been reported for drugs and pesticides adsorption.

Fig. 14. Schematic illustration for the adsorption mechanism of drugs and pesticides onto metal oxides and carbon-based materials.

5. The adsorption mechanism for drugs and pesticides

Comprehending the adsorption mechanism is an important challenge and remarkable phase that can offer improvements in adsorption/desorption conditions or the design of adsorbent. The efficacy and mechanism of adsorption rely on the adsorbents' characteristics (e.g., surface area, porosity, and surface functional groups) (Tong et al., 2019), molecular properties of organic contaminants (e.g., molecular size, aliphatic vs. aromatic, and hydrophobicity) (Ai et al., 2019), background solution properties (ionic strength, pH, natural organic matter, and temperature) (Xiang et al., 2019), and the interactions among functional groups of adsorbent and adsorbate (Kim et al., 2018). The adsorption mechanisms reported in the literature are exclusively either chemisorption or physisorption, based on the strength of interactions between sorbate and sorbent (Gusain et al., 2020). The physisorption occurs due to the reversible weak intermolecular physical interactions such as diffusion, π - π stacking, hydrophobic interactions, London, dipole-dipole and van der Waals forces, and hydrogen bonding with multilayer formation of adsorbate onto the adsorbent. While, the chemisorption involves the irreversible monolayer chemical interactions between the sorbate and sorbent by sharing or transfer of electrons, including covalent bonding, chelation, complex formation, proton displacement, and the redox reactions.

The solution pH is one of the important factors that control the adsorption process because the surface charges of adsorbent and adsorbate are managed by it. The adsorption regulation could be achieved by measuring the point of zero charges (PZC) of sorbents using the zeta potential (Fallah et al., 2020); pH_{PZC} is a pH value at which the electrical charge density of a surface is zero.

At $pH > pH_{PZC}$, the functional groups are deprotonated and the adsorbent surface is negatively charged. Hence the sorbent successfully removes the cationic sorbates via electrostatic attraction from the aqueous solution. At $pH < pH_{PZC}$, the protonation of functional groups leads to a positively charged adsorbent surface and the removal process is favorable for the anionic sorbates. If $pH > pKa$, the sorbate surface charge is negative; however, at $pH < pKa$, the sorbate exists in a positively charged surface. Thus, the adsorption capacity of the adsorbent can be altered by changing the solution pH concerning the adsorbent pH_{PZC} and adsorbate pKa values.

The evaluation of adsorption efficiency using isotherm, kinetic models, and thermodynamic parameters is very effective in predicting the adsorption mechanism (Fallah et al., 2019). Considering the adsorption properties and equilibrium data of sorbent and sorbate, the adsorption isotherm models can define the interaction mechanism of sorbate onto the sorbent at a specific solution pH, time, and temperature. To predict the multilayers or monolayer and heterogeneous or homogenous adsorption mechanism, several isotherm models including Freundlich (1906), Langmuir (Langmuir, 1917, 1916), Temkin (Temkin and Pyzhev, 1940), Dubinin-Radushkevich (Dubinin and Radushkevich, 1947), Sips (1948), Redlich-Peterson (Redlich and Peterson, 1959), Jovanovich (Kudziniski and Wojciechowski, 1977), Flory-Huggins (Nikitas, 1984), and Khan (Khan et al., 1997), have been extended to fit experimental results. However, the Freundlich, Langmuir, and Dubinin Radushkevich models are commonly used for evaluating the isotherm studies. The Langmuir model is applied for computing the adsorbent maximum adsorption capacity (q_{max} ; mg g⁻¹) and illustrates monolayer adsorption onto the homogenous sorbent surface, whereas the Freundlich model reveals the multilayer sorption onto the heterogeneous adsorbent surface. The Dubinin-Radushkevich isotherm estimates the mean free adsorption energy of adsorbent (E , J mol⁻¹), where the E value smaller than 8 is suggesting the physisorption, $8 < E < 16$ corresponding to ion exchange, and $16 < E < 40$ indicating a chemisorption mechanism.

The kinetic models are investigated to explore the adsorption rate and adsorption rate-determining step at a specific solution pH, concentration, and temperature. To determine the order of the adsorption process, diverse kinetic models have been suggested, including inter/intra-particle diffusion (Weber and Morris, 1963), pseudo-first-order (Ho, 2004), pseudo-second-order (Ho, 2006; Ho and McKay, 1999), and Elovich (Wu et al., 2009) models. The kinetic study of the adsorption process is typically evaluated by using the pseudo-first-order, pseudo-second-order, and intra-particle diffusion models. The physical interactions of adsorption at the primary stage are explained by the pseudo-first-order kinetic model, whereas the chemisorption rate-determining step of an adsorption process is determined by the pseudo-second-order kinetic model (Ahmed and Hameed, 2019). The diffusion process occurs in three consecutive steps: I) bulk diffusion, II) film diffusion: transportation from the bulk solution to the sorbent surface across the boundary

layer by diffusion, and III) pore diffusion (intra-particle diffusion): diffusion from the outer surface into the adsorbent pores. The film diffusion and pore diffusion steps are mainly controlled by the diffusion process with slow speed (L. Wang et al., 2020).

The thermodynamic parameters, e.g., free energy changes (ΔG°), standard enthalpy changes (ΔH°), and standard entropy changes (ΔS°), play a crucial role in determining the spontaneity and favorability of the sorption process, which can be calculated at a constant concentration, time and different temperatures (Fallah et al., 2018). The ΔG° is determined from ΔS° and ΔH° values obtained by Van't Hoff plot. The adsorption process occurs spontaneously when ΔG° is a negative value. If ΔH° is a negative value, the adsorption reaction occurs in exothermic form, and if ΔH° is a positive value, the adsorption reaction will be endothermic. The adsorption affinity of sorbate onto the sorbent is revealed by the positive value of ΔS° , which can suggest the increase of randomness.

The activation energy (E_a ; kJ mol⁻¹), computed by the Arrhenius equation, provides the adsorption rate and the chemical or physical nature of adsorption. The E_a values of physical and chemical adsorptions are usually from 5 to 40 to 40–800 kJ mol⁻¹, respectively (Fallah et al., 2019). The chemisorption with higher activation energy and lower adsorption rate leads to efficient and strong interactions between sorbate and sorbent active sites.

Therefore, isotherms and kinetics models, along with thermodynamic parameters and activation energy, can clarify the physiosorption or chemisorption characteristics of an adsorption process. Different methods of drugs or pesticide adsorption on metal oxide adsorbents or their composites and the adsorption mechanisms are schematically presented in Fig. 14.

6. Conclusion and outlook

In this article, a discussion on the toxicological effects of pharmaceutical drugs and pesticides on human life and ecology has been expounded. The residual drugs in the human body cause various serious issues such as genetic disorders, sickness, and cell death, and ecological drug contamination affects wildlife. The presence of pesticides in the body leads to Parkinson's disease through disruption of mitochondrial function, and they are even carcinogenic. Subsequently, we have discussed the recent advances in drug and pesticide removal via adsorption processes using various metal oxides and carbon-based nanostructures. Metal oxides have been employed as effective adsorbents due to their low toxicity, thermal stability, high surface area, porous structure, easy recovery, and the presence of Lewis acid-base sites in their structures. Besides, the utilization of assorted carbon-based materials such as activated carbon, graphene and its various derivatives, carbon nanotubes, and carbon/graphene quantum dots for water detoxification has also been presented in detail. They exhibit some extraordinary characteristics, including high thermal and mechanical stability, corrosion resistance, large surface area, high reactivity, hardness, and low density. Finally, the mechanism of adsorption of drugs and pesticides on the adsorbent surface has also been discussed, which is dependent upon adsorbents characteristics, molecular properties of drugs and pesticides, solution properties, and the type of interactions between functional groups of adsorbent and adsorbate. Thus, the presented review provides a critical study of the toxicological effect of drugs and pesticides and their probable removal from water bodies using metal oxides and carbon nanostructures via adsorption and hopefully stimulate further sustainable exploitation of earth-abundant materials in the future.

Disclaimer

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Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.chemosphere.2021.130055>.

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Supplementary Material

Toxicity and remediation of pharmaceuticals and pesticides using metal oxides and carbon nanomaterials

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Fig. 1S. Major sources and fate of pharmaceuticals and pesticides. Reprinted with permission (Kar et al., 2020).

Fig. 2S. General pathways of drug metabolism. CYP450: Cytochrome P450. Reproduced with permission from (Jaladanki et al., 2020).

Fig. 3S. (a) Catalytic site for cytochromes and (b) the generalized catalytic cycle of a cytochrome P450 enzyme. Three regions of catalytic site for cytochromes are stated as base, wall, and dome. The location of the drug inside the cavity is determined by the chemical characteristics of these regions. The dome region plays a major role in this regard so that hydrophobic groups with a viewpoint of metabolism are drawn to the Fe center. CYP: cytochromes; NADPH: nicotinamide adenine dinucleotide phosphate (reduced form); RH: drug. Reprinted with permission from (Jaladanki et al., 2020).

Fig. 4S. The equations of pesticide oxidation catalyzed by cytochrome P450 (a,b) Normal oxidation cause and inefficacious cycling for superoxides production; (c,d) useless oxidative metabolism of a pesticide through CYP450s, create an uncoupling reaction and superoxide generation. CYP450: Cytochrome P450. Reprinted with modification with permission from (Gray et al., 2013).

Fig. 5S. Summary of postulated relationships between persistent organic pollutants (POP) and their relationships on the risk of type 2 diabetes (T₂D). Reprinted with permission from (Lee et al., 2014).

Table 1S. Human health risk ratios and acceptable daily intake of some reported drugs.(Chinnaiyan et al., 2018; Cunningham et al., 2009)

Therapeutic class of drugs	Generic name	ADI ^{a)} [µg/kg/day]	PEC ^{b)} / PNEC ^{c)} [North America]	PEC / PNEC [Europe]	PEC / PNEC [India]
Anticancer	Topotecan hydrochloride	0.0021	1.6E-04	3.0E-04	-
Anticancer	Mercaptopurine monohydrate	0.0021	2.4E-02	3.8E-02	-
Anticancer	Vinorelbine	0.0021	6.7E-04	1.3E-03	-
Anticancer	Melphalan hydrochloride	0.0021	2.6E-04	2.0E-04	-
Benign prostatic hypertrophy	Dutasteride	0.0016	2.9E-03	2.3E-03	-
Cardiovascular	Digoxin	0.071	2.0E-03	6.4E-04	-
Antiviral (HIV)	Lamivudine	15.9	1.8E-03	5.0E-04	-
Antiviral	Acyclovir/valacyclovir	190	3.7E-04	1.9E-04	-
Antiviral	Abacavir hemisulfate	57.1	7.6E-05	2.4E-05	-
Antiviral (HIV)	Amprenavir + fosamprenavir	133	4.7E-06	1.3E-06	-
Antiviral	Zanamavir	3.14	8.9E-07	2.9E-06	-
Antiviral	Zidovudine	14.3	1.5E-03	4.3E-04	-
Antigastric acid	Ranitidine hydrochloride	10.7	5.3E-03	1.9E-02	-
Anti-inflammatory; vasoconstrictor	Betamethasone valerate	0.24	1.2E-03	2.7E-03	-
Anti-inflammatory; antipyretics	Acetyl salicylic acid	87	-	-	0.07
Corticosteroids	Budesonide	68	-	-	-
Antidepressant	Bupropion hydrochloride	51.7	2.4E-04	4.6E-06	-
Antidiabetic	Metformin hydrochloride	79.4	1.6E-02	2.0E-02	-
AntiParkinson's	Ropinirole hydrochloride	0.12	1.2E-04	7.5E-05	-
Antimalarial	Atovaquone	238	2.1E-06	2.1E-05	-

Antiasthma	Albuterol (salbutamol)	0.05	6.1E-05	2.6E-04	-
Antidepressant	Fluoxetine	77	-	-	0.06
Antidepressant	Amitriptyline	166	-	-	0.13

^{a)} **ADI:** Acceptable daily intake; ^{b)} **PEC:** Predicted environmental concentration; and ^{c)} **PNEC:** Predicted no-effect concentration.

Table 2S. Some pesticides and their health hazards.

Compounds	Impact	Action mechanism	Reference
3,5,6-trichloro-2-pyridinol	Endocrine disruptors for androgen signaling	Reproductive disorders	(Hazarika et al., 2019)
Terbufos (OP), Fonofos (OP)	The significant interaction between exposure and genetic variants,	Prostate cancer	(Gray et al., 2013)
Carbendazim	Possible Carcinogen, show Mutagen	Human Ovarian Granulose-Like Tumor Cell Line (KGN)	(Morinaga et al., 2004)
Chlorantraniliprole	Possibly Eye irritant	Damage in the midgut of Bombyx mori	(Bhandari et al., 2019; Hu et al., 2019)
Chlorpyrifos	Neurotoxicant and psychiatric disorders	Human liver hepatocellular carcinoma cells (HepG2), neurotoxicity in rats	(Mahmoud et al., 2019; Zhou and Li, 2018)
Dichlorvos	Dyspnea, salivation, convulsion, and other neurological phenomena	Chicken brain tissues, a gastric tissue cancer	(Okoroiwu and Iwara, 2018; Xiao et al., 2020)
Dimethoate	Low dermal toxicity	Liver and brain of rats	(Sharma et al., 2005)
Emamectin	Apoptosis of QSG7701 cells and induced the DNA damage	Human liver	(Zhang et al., 2017)
Imidacloprid	Genotoxicity	Liver	(Costa et al., 2009; Schulz-Jander et al., 2002)
Omethoate	Homozygous/heterozygous mutant genotypes	Alteration in the gene expression in the process of	(Duan et al.,

		cervical cancer	2017)
Profenofos	Neurotoxicant, skin irritant, and possibly eye irritant	Testicular toxicity or male reproductive toxicants	(Bhandari et al., 2019; Moustafa et al., 2007)
Quinalphos	Neurotoxicant, skin and eye irritant	Alzheimer's disease and male sterility in humans.	(Bhandari et al., 2019; Govindarajan et al., 2019)
Triazophos	Neurotoxicant, skin and eye irritant	Blood, liver, and brain tissue	(Bhandari et al., 2019; Jain et al., 2011)

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