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Introduction to Femtochemistry: Excited State Proton Transfer from Pyranine to Water Studied by Femtosecond Transient Absorption

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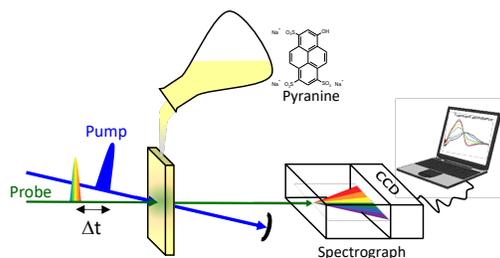
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ABSTRACT

In order to introduce students to the fascinating field of femtochemistry, we propose here a practical laboratory training course conceived for second-year master students in chemistry. We describe the use of a broadband femtosecond transient absorption (pump-probe) experiment for monitoring a fast light-triggered chemical reaction in solution. The experiments are performed on the pyranine photoacid, which upon photoexcitation at 390 nm undergoes a proton transfer to the solvent in about 90 ps. While this practical course involves advanced equipment and techniques, the measured transient absorption data allow easy analysis and interpretation. The transient absorption spectra at a few selected delay times can be analyzed qualitatively in terms of bleach, induced absorption and stimulated emission. Likewise, the transient absorption signals at a few chosen wavelengths can be quantitatively analyzed and explained with simple kinetic models to determine the time constant of the proton transfer reaction. This training aims at giving the students the opportunity to face some of the current challenges in contemporary chemistry by learning the basics of ultrafast spectroscopy.

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GRAPHICAL ABSTRACT



KEYWORDS

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Graduate Education/Research, Physical Chemistry, Photochemistry, UV-vis spectroscopy, Kinetics, lasers, pH, reaction, Proton transfer

INTRODUCTION

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The formation or the breaking of chemical bonds, electron or proton transfers within or between molecules are elementary chemical reactions that can be extremely fast.

Their precise characterization is crucial to address fundamental questions in chemistry and biology. Using light to probe such mechanisms has proven an outstanding means to this end and photophysics, photochemistry and photobiology are today well-

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established fields by their own.^{1, 2} In this regard, pump-probe methods are powerful techniques for measuring the formation of short-lived reaction intermediates produced from an optical excitation. The basic principle of these methods is to use a strong optical "pump" pulse to trigger a photoinduced reaction in the studied medium and a weak "probe" light to monitor the pump-induced change.³⁻⁵ These techniques emerged

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in the middle of the twentieth century with the development of flash photolysis experiments by Eigen, Norrish and Porter (Nobel Prize 1967) for the study of electronic triplet states and free radicals.⁶ Flash photolysis originally used powerful flash lamps for the sample excitation and continuous light source as the probe, providing a time resolution of about one microsecond. The time resolution of such experiments depends

50 mainly on the pump pulse duration and the electronics of the probe detection.
Therefore, by using pulsed laser excitation, nanosecond time resolution can easily be
obtained.⁴ In order to reach (sub-)picosecond resolution and to overcome the limits
imposed by the detector response, purely optical pump-probe techniques using two
short laser pulses generated from the same laser source were developed towards the
55 end of the twentieth century. The successful application of these techniques to the
studies of molecular dynamics opened the field of Femtochemistry (A. H. Zewail, Nobel
Prize 1998, see ref. 5 and references therein). With these techniques, the time
characteristics of the electronics used for the detection of the probe pulse intensity has
no consequence. The time-resolution is only limited by the temporal overlap of the
60 probe and pump pulses and the time-scale is defined by introducing an optical delay
between the pump and the probe pulses. This is described more in detail below.

To introduce students to ultrafast spectroscopy, the third semester of the SERP+
master's program at the Paris-Saclay University ([http://www.master-
serp.eu/programme/](http://www.master-serp.eu/programme/)) offers an overview of the various optical techniques used for the
65 study of photoinduced reactivity of molecules and molecular assemblies at short time
scales (from femtoseconds to nanoseconds). In order to initiate students to the state-of-
the-art instrumentation now widely used in physical chemistry laboratories, we have
developed a half-day laboratory training devoted to the use of femtosecond transient
absorption for the study of an excited-state proton transfer (ESPT) reaction in aqueous
70 solution. To this end, we use pyranine (also known as trisodium 8-hydroxy-pyrene
1,3,6-trisulfonate, HPTS), a well-known photoacid commonly used as pH indicator in
biology.^{7, 8} Pyranine is also a coloring agent widely used for drugs, cosmetics and inks
due to its strong yellow-green fluorescence.⁹ It exhibits a strong absorption around
400 nm, in the spectral region of the second harmonic of femtosecond titanium
75 sapphire lasers, making this compound particularly well suited for femtosecond

spectroscopy. Several time-resolved studies of pyranine in aqueous solution using different experimental approaches have provided evidence that ESPT from the photoacid to the solvent occurs with a time constant of ca. 90 ps.¹⁰⁻¹⁵ The present training consists of the experimental observation of the formation of the deprotonated form of pyranine by broadband femtosecond transient absorption spectroscopy. This training
80 which has been carried out in a research laboratory on a routine basis for several years, aims to (i) consolidate the knowledge acquired during the physical chemistry coursework, (ii) perform a quantitative and critical analysis of experimental data and (iii) learn how to write a scientific report in the form of a scientific article. This training
85 introduces the students to some of the challenges of modern chemistry by both teaching them the concepts and the applications of ultrafast spectroscopy. These important issues are rarely addressed together in the teaching of masters of chemistry.¹⁶ In the following, we present the principle and the practical details of the experiment on pyranine, the interpretation of the observations and a simple kinetic
90 analysis of the results. These elements constitute a protocol that can be adapted in any laboratory equipped for UV-visible time-resolved laser spectroscopy.

BACKGROUND

Basics of UV-visible femtosecond transient absorption spectroscopy

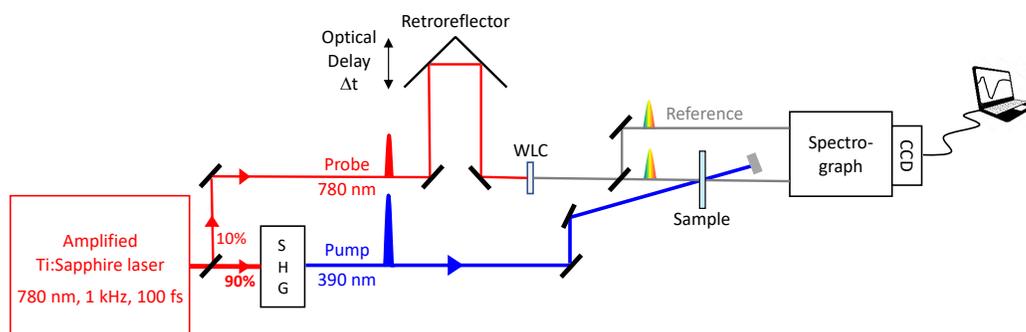
95 Transient absorption spectroscopy measures the change in the absorbance of a sample following photo-excitation by a short light flash or laser pulse at time $t = 0$. The differential absorbance before and after photo-excitation, $\Delta A(\lambda, t)$, at wavelength λ and time t is usually defined as:

$$\Delta A(\lambda, t) = A(\lambda, t) - A_0(\lambda) \quad (1)$$

100 with $A(\lambda, t)$ being the sample absorbance at time t after the photo-excitation and $A_0(\lambda)$ the sample absorbance without the photo-excitation. Note that $A_0(\lambda)$ corresponds to the

steady-state absorbance of the sample that can be measured with a classical spectrophotometer.

According to the timescale of the induced processes to study, two main techniques
105 are available: flash photolysis spectrometers for nanoseconds to milliseconds time range
and ultrafast pump-probe laser set-ups for femtoseconds to nanoseconds time range.
Flash photolysis systems are nowadays simple to handle and affordable. Indeed, several
experiments have already been proposed for training purposes in a physical chemistry
laboratory.¹⁷⁻²⁰ For the femto- and picosecond time scales, “pump-probe” laser
110 techniques are required. For this purpose, amplified self-mode locked pulsed titanium
sapphire lasers are currently the most widely used technology, typically providing 50-
100 fs pulses at kHz repetition rates, with a wavelength around 800 nm and a pulse
energy of a few millijoules.²¹



115 Figure 1. Principle of the “pump-probe” laser technique using a white light continuum as probe. SHG: second harmonic generation. WLC: white light continuum.

As illustrated in Figure 1, the fundamental laser IR pulses are first split into two
coherent replicas of different intensities and propagating along different optical paths.
120 The stronger one, called the “pump”, is used to excite the sample after optional
wavelength conversion. This can be second harmonic generation (SHG) or third
harmonic generation (THG) in non-linear crystals or optical parametric amplification
(OPA) with subsequent second harmonic generation for a tuning range between 250 nm

and 650 nm. The strong pump beam is focused into the sample cell. The weaker one,
125 called the “probe”, is used to measure the pump-induced changes in the sample
absorbance. The probe pulse is optically delayed relative to the pump pulse by deviating
its optical path via a retro-reflector mounted on a motorized translation stage. Since the
speed of light is constant, the delay between the pump and the probe pulses, Δt , can be
determined as $\Delta t = 2\Delta x/c$, where Δx is the spatial displacement of the translation stage
130 and c is the speed of light. For example, a displacement of 150 μm corresponds to a
temporal delay of 1 ps. The “time zero”, $t = 0$, corresponds to the position of the
translation stage for which the pump and probe pulses arrive simultaneously at the
sample. The pump and probe beams are spatially overlapped in the sample cell, in order
to probe the excited volume.⁴

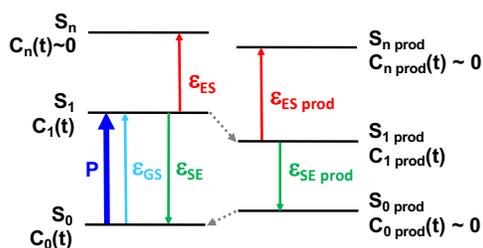
135 For broadband transient absorption measurements, a white light continuum (WLC)
probe is used which serves to measure the temporal change in the absorbance spectra
associated to the electronic states created by the pump. The WLC is generated by self-
phase modulation by tightly focusing the weak laser fundamental probe beam into a
transparent Kerr medium, often a thin plate of CaF_2 , silica, YAG, or sapphire.^{22, 23} In
140 order to correct for any intensity fluctuations of the probe beam, the weak WLC beam is
split in two equal beams, the probe and the reference beams. The reference beam,
which only serves to measure the WLC stability, is by-passing the sample while the
probe beam is collimated into the sample cell, overlapping with the pump beam in order
to probe only the excited volume in the sample. The probe and the reference beams are
145 then focused on the entrance slit of a small spectrograph, dispersed spectrally and
detected simultaneously by a CCD camera. The probe and reference spectra are
recorded by the CCD, with and without the pump pulse on, for each delay line position,
i.e. each time t , so ΔA is calculated as:

$$\Delta A(\lambda, t) = \log\left(\frac{I_{\text{ref/pump on}}(\lambda, t)}{I_{\text{probe/pump on}}(\lambda, t)}\right) - \log\left(\frac{I_{\text{ref/pump off}}(\lambda, t)}{I_{\text{probe/pump off}}(\lambda, t)}\right) \quad (2)$$

150 where $I_{\text{probe/pump on}}$ and $I_{\text{ref/pump on}}$ are the probe and the reference intensities measured with the pump pulse on. $I_{\text{probe/pump off}}$ and $I_{\text{ref/pump off}}$ are the probe and the reference intensities measured with the pump pulse off. The measurement of ($I_{\text{probe/pump on}}$; $I_{\text{ref/pump on}}$) and ($I_{\text{probe/pump off}}$; $I_{\text{ref/pump off}}$) for each pump-probe delay, both pairs stemming from the same coherent replica, allows to correct for the fluctuations of the WLC during the
155 acquisition procedure. This increases the accuracy of the ΔA signals, typically providing a sensitivity of 10^{-3} - 10^{-4} .

Interpretation of UV-visible transient absorption spectra

Measurements of the ΔA spectra of the sample for different pump-probe delays allow
160 to follow the formation of directly photo-excited states as well as intermediate states or photoreaction products.



165 Figure 2. 3-population model showing the formation of an emissive product ($S_{1 \text{ prod}}$) from the excited state S_1 produced by the pump pulse P. Dotted grey arrows represent the non-radiative conversion: $S_1 \rightarrow S_{1 \text{ Prod}}$ and $S_{0 \text{ Prod}} \rightarrow S_0$. Note that the higher excited states S_n and $S_{n \text{ prod}}$ are not populated by the pump pulse and $S_{0 \text{ Prod}} \rightarrow S_0$ conversion is much faster than $S_{1 \text{ Prod}} \rightarrow S_{0 \text{ prod}}$ deactivation. Under these conditions, the concentrations $c_n(t)$, $c_{n \text{ prod}}(t)$ and $c_{0 \text{ prod}}(t)$ are zero.

Figure 2 illustrates a 3-population model involving the formation of an emissive product
170 state from the singlet excited state. In such a case, the differential absorbance of the sample reads as:

$$\Delta A(\lambda, t) = [\epsilon_{\text{ES}}(\lambda) - \epsilon_{\text{SE}}(\lambda) - \epsilon_{\text{GS}}(\lambda)] \cdot l \cdot c_1(t) + [\epsilon_{\text{ES prod}}(\lambda) - \epsilon_{\text{SE prod}}(\lambda) - \epsilon_{\text{GS}}(\lambda)] \cdot l \cdot c_{1 \text{ prod}}(t) \quad (3)$$

with $\epsilon_{ES}(\lambda)$, $\epsilon_{GS}(\lambda)$ and $\epsilon_{ES_{prod}}(\lambda)$, the molar absorption coefficients at wavelength λ of the initial excited state (S_1), the ground state (S_0) and the product excited state ($S_{1_{prod}}$), respectively. $\epsilon_{SE}(\lambda)$ and $\epsilon_{SE_{prod}}(\lambda)$ are the stimulated emission coefficients of S_1 and $S_{1_{prod}}$. $c_1(t)$ and $c_{1_{prod}}(t)$ are the concentrations of the S_1 and $S_{1_{prod}}$ states at time t and l is the optical path. The ΔA spectra are composed of several overlapping bands corresponding to the contributions of the different states created by the pump pulse. Negative contributions are expected from the ground-state bleach (GSB), due to the pump-induced depopulation of the ground state population, and the stimulated emission (SE) from the excited-state leading to an increase in the probe intensity. In contrast, positive contributions are expected from the diminution of the probe intensity due to the excited-state absorption (ESA) and the product state absorption. Figure 3 illustrates one example of these different contributions to the ΔA signal.

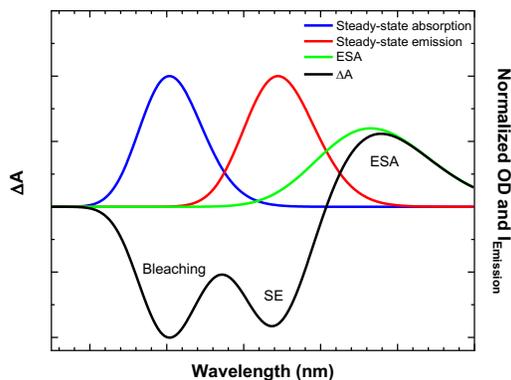


Figure 3. Illustration of three different contributions to the differential absorbance (ΔA) after the sample excitation: ESA, bleaching and SE. The ΔA spectrum is represented by the black line. The steady-state absorption and emission spectra of the sample are illustrated by the blue and red lines, respectively. The ESA is shown by the green line.

Excitation and Acido-Basic Properties of Pyranine

Pyranine (3sPyOH, Trisodium 8-hydroxy-pyrene 1,3,6-trisulfonate, HPTS, Figure 4 (a)) is an aromatic photoacid. It becomes 10 million times more acidic in its first excited state with respect to its ground state ($pK_a = 7.5$ in the ground state and $pK_a^* = 0.4 \pm 1.4$

in the excited state).²⁴ Thus, upon photoexcitation, the neutral pyranine, 3sPyOH,
195 undergoes an intermolecular ESPT from the hydroxyl group to a neighboring solvent
molecule. The reaction can be described by the thermodynamic Förster cycle shown in
Figure 4 (b).

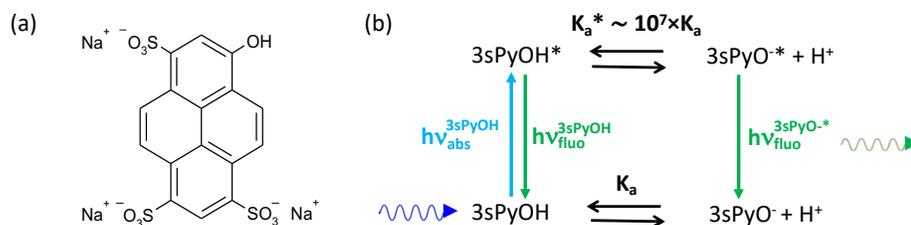


Figure 4. (a) Molecular structure and (b) Förster cycle of pyranine, 3sPyOH.

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The ESPT dynamics of 3sPyOH have been widely studied in different environments.^{25, 26}

In moderately acidic aqueous solution at pH = 4, the protonated form, 3sPyOH,
dominates in the ground state while, upon photoexcitation, ESPT occurs very efficiently
and generates the deprotonated pyranine anion, 3sPyO⁻*. Figure 5 illustrates the

205 steady-state absorption and fluorescence spectra of pyranine in aqueous solutions at

different pH. The absorption spectrum of 3sPyOH exhibits two peaks centered around
370 nm and 400 nm recently associated with the ¹L_b and ¹L_a transitions of pyranine,
respectively.²⁷ As previously reported,¹⁰ the fluorescence spectrum of 3sPyOH* displays

a maximum around 430 nm and a shoulder at ca. 460 nm while the spectrum of the

210 deprotonated form, 3sPyO⁻* is considerably red shifted and shows a maximum at ca.

520 nm.

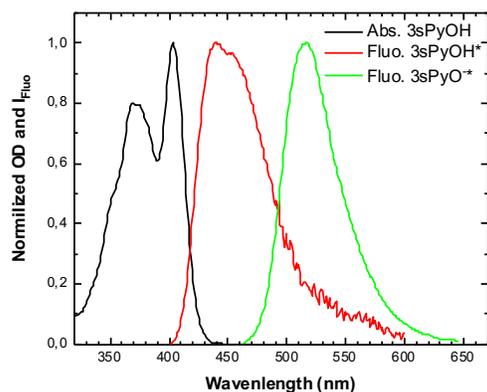


Figure 5. Steady-state absorption spectrum 3sPyOH and fluorescence spectra of 3sPyOH* and 3sPyO** in aqueous solutions.

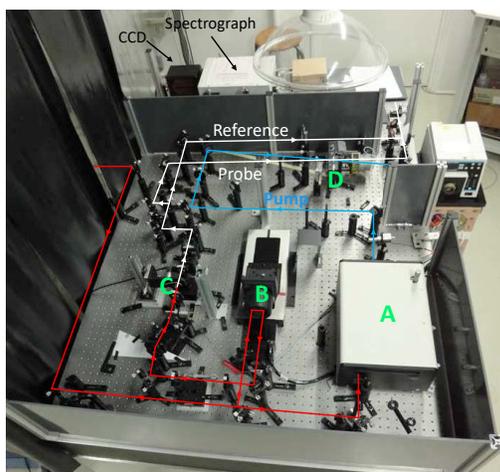
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The ESPT mechanism of pyranine in water is known to be quite complex. It involves two ultrafast steps of <math>< 1\text{ ps}</math> and ca. 3 ps, respectively, prior to the relatively slow proton-transfer step in ca. 90 ps.^{10, 12, 28} The explanation of the two fast steps is still a matter of debate in the literature. Solvation dynamics and the formation of an intramolecular charge-transfer state have been invoked^{11, 28-30} as well as the formation/recombination of contact and separated intermolecular ion pairs before diffusion.^{12, 13, 15} In the present training, the physical meanings of these fast steps are not discussed in detail since the emphasis is on the slower proton transfer process. Accordingly, experimental data are interpreted using a simplified model involving only the ground and excited states of the protonated and deprotonated forms of pyranine shown in Figure 4 (b).

MATERIALS AND EXPERIMENT

Measurements of the transient absorption spectra of pyranine in an aqueous solution at pH 4 are performed with a femtosecond pump-probe set-up (see Figures 1 and 6) based on a commercial amplified titanium sapphire laser (Amplitude Laser) that delivers <math>< 100\text{ fs}</math> duration pulses at 780 nm, with an energy of 2 mJ at a 1 kHz

repetition rate. 640 μJ of the output laser energy is used for the pump-probe set-up. The pump beam at 390 nm is generated from 90% of the incoming 780 nm beam by SHG in a 2 mm-thick BBO crystal. The pump energy is adjusted to 20 μJ and focused to a 0.5 mm diameter beam in the sample cell. The remaining 10% of the incoming 780 nm beam travels through the optical delay line (Microcontrole) before being focused in a 2 mm rotating fused silica plate, for the WLC generation. A diaphragm and a combination of a half-wave plate and reflecting polarizers are used to adjust the energy of the 780 nm beam prior to the WLC generation. A broadband beam-splitter is then used to divide the WLC into a probe and a reference beam. In order to avoid any contribution of rotational diffusion from the sample to the ΔA signals, the polarizations of the pump and probe beams are set at a magic "angle" (54.7°) with a half-wave plate placed in the pump beam just before the sample cuvette. The probe and the reference beams are transported and focused onto the entrance slit of an imaging planar polychromator (Princeton Instruments equipped with a grating having a blaze angle optimized for 500 nm) equipped with a CCD camera (1340 \times 400 pixels, 16 bits, Princeton Instruments) and interfaced to a PC computer. The intensities of the probe and the reference beams are monitored in real time using Winspec/32 software. A shutter, located on the pump path, allows monitoring the probe and the reference beams for each pump-probe delay alternatively with and without the pump excitation. The transient absorption experiments are controlled by a homemade acquisition program written in Visual Basic (v. 6).



255 **Figure 6.** Picture of the experimental pump-probe set-up used for the laboratory training at Institut de Chimie Physique at the University of Paris Saclay. A: SHG; B: optical delay line; C: WLC generation; D: sample cell. The red lines indicate the beam paths of the laser beam at 790 nm, the blue line indicates the beam path of the pump after SHG and the white lines indicate the beam paths of the probe and reference after WLC generation.

260 Preparation of the pyranine sample is described in details in Instructor's notes.

HAZARDS

The femtosecond laser produces high intensity radiations. Even if the laser beams are below eye level and contained on the experimental table by non-reflective screens, people in the room have to wear safety goggles certified to block the potentially dangerous IR and UV light.

The aqueous solution of pyranine at pH4 is prepared prior to experiments, so students do not manipulate chemicals (see Instructor's notes). However, as the solution is acid, it may cause irritations if not washed off the skin reasonably quickly with a large amount of water until no evidence of the chemical remains.

RESULTS AND DISCUSSION

Laboratory sessions

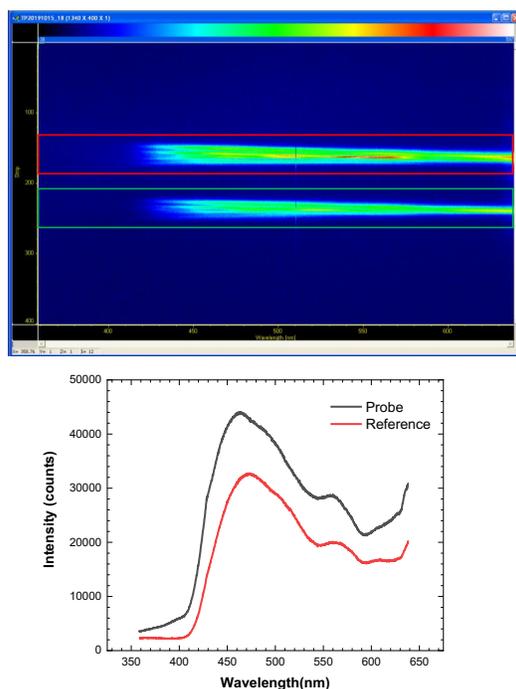
This introduction to femtosecond laser spectroscopy is organized in two laboratory sessions on different days. The first laboratory session of 2h consists of a visit of the

laser laboratory for the entire class of ca. 15 students, with the laser turned off for safety reasons. At this occasion, the set-up with all the instruments is explained (see Instructor's notes). The second laboratory session of 4h is dedicated to the transient absorption measurement of the ESPT reaction of pyranine, by groups of 3 to 4 students.

280 During this session, the laser is switched on. For this reason, the session starts with a short overview of the security rules in a laser lab. Only then, with all students and teachers wearing protective eyewear, are the optical paths of the pump and the probe beams retraced on the optical table, outlining the various optics and apparatuses.

Subsequently, the probe and the reference spectra are visualized in real time with the

285 CCD in full image mode using the camera software. Figure 7 (Top) represents a spectrally dispersed image of the probe and reference spots on the (vertical) input slit of the spectrograph.

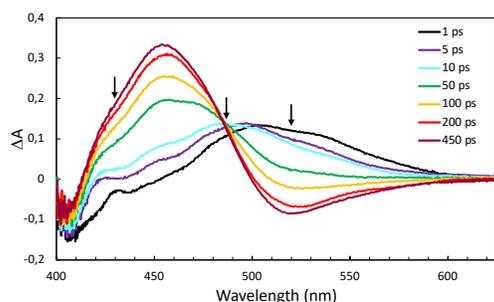


290 Figure 7. (Top) Visualization of the dispersed probe (lower trace) and reference (upper trace) beams with the pump beam on measured with the CCD camera in image mode. The vertical axis represents the position on the input slit. The light intensity is indicated by false colors, ranging from blue to red with increasing intensity. (Bottom) Corresponding spectra of the probe and the reference beams after integration over the two regions of interest defined by red and the green boxes, respectively, around the probe and the reference images in (Top), respectively.

295 While in “live mode”, the limits of the two horizontal “regions of interest” to be recorded during the experiments are defined in order to obtain the intensity variations of the probe and the reference beams separately, as shown in Figure 7. The acquisition parameters are chosen to maximize the signal-to-noise ratio and to avoid the saturation of the detector (see Instructor’s notes). Before starting the measurements, the spatial
300 overlap between the pump and the probe beams in the sample is optimized by observing the pump-induced changes on the probe intensity in the spectral region of 3sPyO* SE around 520 nm. This requires that the probe pulse arrives after the pump pulse in the sample which can be verified by a search for the “time zero” by manually moving the optical delay line. Also note that for a good spatial overlap, the diameter of the pump
305 beam is adjusted to be twice as large as that of the probe.

Data acquisition and analysis

Measurements of the ΔA spectra of pyranine are performed for different pump-probe delays, typically between -5 and 450 ps (see Instructor’s notes). For each delay time t ,
310 the values of the wavelength λ , $I_{\text{probe/pump on}}(\lambda, t)$, $I_{\text{probe/pump off}}(\lambda, t)$, $I_{\text{ref/pump on}}(\lambda, t)$ and $I_{\text{ref/pump off}}(\lambda, t)$ are recorded in a separate file. The ΔA spectra at all the measured pump-probe delays are calculated after the acquisition procedure according to eq. (2) using a home-made program. From these data, students are asked to choose and plot several ΔA spectra of pyranine at representative delay times during the ESPT reaction. An
315 example is given in Figure 8. Using this figure, the first important task for the students is to discuss the temporal variation of their chosen spectra and to attribute the different bands to the different species involved in the ESPT reaction by using the Förster scheme of Figure 4 (b).



320 Figure 8. ΔA spectra of pyranine in an aqueous solution at pH 4 measured for different pump-probe delays, after excitation by a femtosecond laser pulse at 390 nm. Black arrows indicate the three characteristic wavelengths at which kinetic analysis is performed in Figure 9.

Students should notice that directly after the excitation, the ΔA spectra are

325 dominated by the contribution of the protonated form of pyranine. The broad positive band in the spectral region around 520 nm and the negative one in the spectral region between 430 and 450 nm are attributed to ESA and SE of $3s\text{PyOH}^*$, respectively. In the spectral region below 430 nm, the negative contribution to the ΔA spectra can be attributed to GSB due to the depletion of the ground-state population induced by the

330 pump pulse. Note that the ESA band of $3s\text{PyOH}^*$ exhibits a blue shift within 5 ps stemming from solvation dynamics and the earliest steps of the ESPT reaction mentioned above. Both the ESA and the SE bands associated with $3s\text{PyOH}^*$ disappear in a few tens of picoseconds. From 5 ps and onwards, an isosbestic point (*i.e.* a wavelength where ΔA remains constant), can be observed at about 485 nm. This can be

335 taken as an indication of a precursor-successor relationship of two populations, namely the excited state of the protonated and the deprotonated forms of pyranine respectively ($3s\text{PyOH}^* \rightarrow 3s\text{PyO}^*$). After 50 ps, the ΔA spectra become dominated by the contribution of $3s\text{PyO}^*$. The SE and the ESA bands associated with $3s\text{PyOH}^*$ are replaced by the positive band peaking at 455 nm and the negative one centered around

340 520 nm arising from the ESA and SE of $3s\text{PyO}^*$, respectively.

In a second step of the experimental data analysis, the students are asked to select three characteristic wavelengths for the features mentioned above and to plot the kinetic traces, as shown in Figure 9. The observed kinetic traces have then to be discussed. For instance, just after excitation, the kinetics at 430 nm is dominated by the negative contribution of the SE of 3sPyOH*. After a few picoseconds, it becomes positive indicating that the ESA of 3sPyO-* becomes dominant. Inversely, at 520 nm the ΔA is positive just after excitation due to the main contribution of the ESA of 3sPyOH*, then it becomes gradually negative due to the increasing contribution of the SE of 3sPyO-*. The last “flat” kinetic trace illustrates the temporal variation of ΔA in the spectral region of the isosbestic point of the time-resolved spectra.

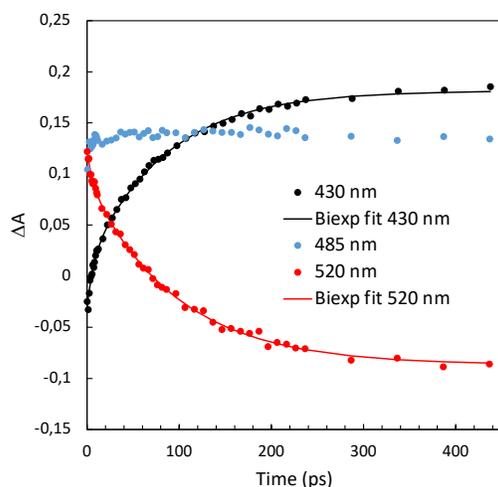


Figure 9. ΔA kinetic traces at three representative wavelengths extracted from the data of Figure 8. The solid lines correspond to data fitted by bi-exponential model functions with Excel (for parameters see Table 1 below).

For the last step of the experimental data analysis, students are asked to perform a non-linear fit of the two selected kinetics in the spectral regions around 430 nm and 520 nm in order to determine the ESPT rate constant. After having imported the experimental data in an Excel table, the students have to define the appropriate fitting function. The measured kinetic traces in the spectral regions of 430 nm and 520 nm can be reasonably well fitted by using a sum of exponential functions, defined as follow:

$$f(t) = \sum_{i=1}^N A_i \times \exp(-t/\tau_i) + C \quad (4)$$

with τ_i being the time constants, A_i the preexponential factor and C a constant representing the signal of the long-lived excited state of 3sPyO^{*}. Note that both A_i and C can be positive or negative. In the present training, mono- and bi-exponential functions can be used. After having chosen the initial parameters and calculated the model values for all the measured optical delay times, students have to apply the least-squares regression algorithm. To this purpose, they have to calculate the error function S , defined as the sum of the squared residuals:

$$S = \sum_{i=1}^N [y(t_i) - f(t_i, \theta)]^2 \quad (5)$$

with $y(t_i)$, the experimental data points and $f(t_i, \theta)$ the model function values for a parameter set $\theta = \{A_i, \tau_i, C\}$.

Table 1. Parameters (τ_i time constants, A_i pre-exponential factors and C constant) of the mono-exponential and biexponential functions (eq. 4) used to fit the kinetic traces at the two selected wavelengths shown in Figure 9. S is the sum of the squared residues (eq. 5).

λ / nm	τ_1 / ps	A_1	τ_2 / ps	A_2	C	S
Mono-exponential fits:						
430	66	-0.187	-	-	0.175	0.0019
520	83	0.192	-	-	-0.083	0.0010
Bi-exponential fits:						
430	6	-0,048	84	-0,169	0,183	0.0003
520	6	0,029	95	0,184	-0,087	0,0004

The “best” fitting parameter set can then be obtained by using the “solver” option in Excel. An explicit example is given in Instructor’s notes together with the procedure for calculating the errors on the fitting parameters (optional). These simple numerical calculations using an Excel-sheet provide an excellent means to illustrate a non-linear parameter regression. This knowledge can evidently serve the students in their future

career. Table 1 gathers the optimized parameters for mono- and bi-exponential fits of the kinetic traces at 430 nm and 520 nm shown in Figure 9.

The time constants of the mono-exponential fits reflect mainly the ESPT reaction. These fits reveal, however, slightly distinct time constants at 430 nm and 520 nm arising from the short time events preceding the ESPT reaction. Notably, the blue shift of the ESA band associated with 3sPYOH* within the first five picoseconds leads to a significant change in the ΔA signals at 430 nm and to a lesser extent at 520 nm as shown in Figure 8. Such spectral changes lead to complex kinetic behavior that requires a sum of exponentials to be properly fitted. The students can optionally discuss this point in their report and compare mono- and bi-exponential fits of their chosen kinetics. The accuracy of bi-exponential fits is in general found to be better. Bi-exponential fits of the two kinetic traces of Figure 9 lead to two time constants (see Table 1), a short one of 6 ps (τ_1) and a longer one of about 90 picoseconds (τ_2) that are comparable to those reported in the literature.^{10, 12, 28} As previously mentioned, there has been much debate regarding the attribution of the shortest time components. Therefore, the detailed discussion about the origin of the 6 ps time constant found here is beyond the scope of this laboratory training. In this context, τ_2 which contributes mainly to the kinetic traces at 430 nm and 520 nm can be attributed to the ESPT reaction. Note that a very good reproducibility of this reaction time during has been observed during the different training sessions over the years (see Instructor's notes).

For the last task of the training session, the students are asked to compile their experimental data and their interpretations in the form of a research paper. The principles and the functioning of the transient absorption set-up and experiments have to be explained in the experimental part. A small bibliographical work on pyranine is requested for the writing of the introduction. In the present case, the students are also

asked to compare their results with time-resolved fluorescence measurements of an aqueous solution of 3sPyOH at pH = 4, performed in another training session. These measurements, made by time-correlated single photon counting (TCSPC), inform mainly
410 on the excited state lifetime of the 3sPyO^{-*} anion, which is several nanoseconds, but a “fast” component can also be observed and directly compared with the ESPT time constant obtained from the transient absorption measurements. The manuscript is then used to evaluate students on both their practical session and their knowledge of ultrafast spectroscopy acquired during their classes.

415 CONCLUSION

This laboratory training is an introduction to femtosecond spectroscopy for master students in chemistry. The basic concepts of ultrafast spectroscopy are addressed through the study of the fast proton-transfer reaction from the photoexcited pyranine
420 molecule to the solvent in aqueous solution using broadband femtosecond transient absorption spectroscopy. This laboratory training is organized in two sessions, a first one of 2h which consists of a visit of the laser laboratory and a second one of 4h dedicated to the transient absorption measurements. This training aims to give students hands-on experience in ultrafast spectroscopy and data analysis, to fill the
425 gap between theoretical and practical knowledge of femtosecond spectroscopy. The students are expected to provide a written report describing the experimental setup, explanations of the instruments used and a brief recall of the chemical reaction investigated, *i.e.* the pyranine molecule and its photophysical properties. This report should be written in the style of a scientific article, respecting the quality and the rigor
430 required. Moreover, students need to perform a quantitative analysis of the data that requires some simple programming.

This training introduces students to the experimental research activity in a physical chemistry laboratory in general and to modern time-resolved spectroscopic techniques in particular. Moreover, it provides an example of a fundamental photochemical
435 reaction that is ubiquitous in chemistry and biology. Even if students do not perform the experiment fully by themselves (all the experiments are executed in a group of 3 to 4 students supervised by a professional researcher), the experiment helps them to understand the concepts and to realize the complexity and the difficulty to acquire experimental data. In this sense, this laboratory training is not limited to the particular
440 problem of a proton transfer reaction but it also confronts the students with the more general problem of making a critical analysis of experimental results. In a wider perspective, this practical laboratory training may serve as a first introduction to the “real” activity in a research laboratory. Going beyond a simple laboratory visit, the students get the opportunity to participate in the running of an advanced experiment,
445 interact with the operators and treat their “own” data set.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available on the ACS Publication website at DOI:
450 XXX.

Instructor’s notes detailing preparation of the pyranine solution, real-time visualization of the white light continuum with a CCD camera, data acquisition, non-linear fitting procedure with Excel and experimental reproducibility (PDF)

Practical training instructions for students describing the basic principles of the
455 experiments and the tasks to be achieved (PDF)

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Notes

460 The authors declare no competing financial interest

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