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Submitted on 6 Nov 2020

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Efficient one-pot, three components procedure to prepare new α-
 amino phosphonate and phosphonic acid acyclic nucleosides

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Efficient one-pot, three components procedure to prepare new α-amino phosphonate and phosphonic acid acyclic nucleosides

An efficient one-pot three-component Kabachnik-Fields reaction of aldehydes (acyclic nucleosides), amines (or amino acid), and triethylphosphite proceeded for the synthesis of amino phosphonates using natural phosphate coated with iodine (I$_2$@NP) as a catalyst. The novel α-aminophosphonate and phosphonic acid acyclic nucleosides were tested for their anti-HCV and anti-HIV activities. The molecular docking showed that the non-activity of these compounds may be due to the absence of hydrophobic pharmacophores.

Keywords: acyclic nucleosides; α-Aminophosphonates; Kabachnik-Fields reaction; catalysis; Natural phosphate

Introduction

The viral disease is a major health problem worldwide. Indeed, there is still a need to discover new potent, safe, and selective antiviral drugs. Among these, α-aminophosphonates have attracted the attention of medicinal chemists due to their potential antiviral activity [1]. The α-aminophosphonates are structurally similar to α-amino acids, known to have good cell permeability[2], and physiological stability as the phosphorus-carbon bond, which is not susceptible to enzymatic degradation by phosphatases [3-5]. Moreover, acyclic nucleoside phosphonates, such as adefovir and tenofovir, are now in clinical use for the treatment of viral infections (HIV, HBV, CMV). These drugs possess a phosphonomethyl ether moiety allowing them to bypass the first phosphorylation step [6, 7]. The after-mentioned properties prompted many research groups including ours to synthesize acyclic nucleoside phosphonates varying notably the length and substitution pattern of the phosphonate link [8-12].

The synthesis of α-aminophosphonates has gained more attention due to their broad spectrum of biological activity like anticancer, antiviral, herbicides, fungicides, bactericides, peptide mimetic, and enzyme inhibitor [13-18]. Three-component one-pot Kabachnik-Fields condensation of carbonyl compounds with amines and phosphites is perhaps the most exploited reaction for the preparation of structurally diverse α–aminophosphonates [19, 20]. While early procedures were limited to simple starting
materials and newer wide-scope syntheses catalysed by Lewis acids, have been reported [21-27]. Recently, several solids catalysts have been used in the preparation of α-aminophosphonates such as SbCl₃/Al₂O₃ (28), Mg(ClO₄)₂[29], Na₂CaP₂O₇[30], KF doped natural phosphate (NP) [31], and PEG-SO₃H [32]. In general, these three-component reactions may take place via an imine or an α-hydroxy-phosphonate intermediate [33]. During the formation of imine, water is formed which could either deactivate or decompose the Lewis acid catalyst thereby limiting the scope of carrying out this reaction in one-pot [34]. Among supports reported in the literature, natural phosphate was found [31] to be an efficient Lewis acid to catalyse the synthesis of α-aminophosphonates in one-pot due to its ability to tolerate the water generated during the course of the reaction. Moreover, it was used widely as support because of its ionic substitution ability, structural stability, and high adsorption capacity, making it an attractive cost-effective catalyst for several chemical transformations [35-37]. Meanwhile, iodine has emerged as a very efficient Lewis acid catalyst for various organic transformations and is relatively inexpensive compared to other Lewis acids, including rare earth metal triflates, and is more tolerant in comparison to typical Lewis acids/bases [38-40]. Furthermore, Zahouily et al. showed that iodine supported on natural phosphate could be effectively employed for the protection of carbonyl compounds as their thioacetals in good yield at ambient temperature and mild conditions [41].

In the light of these successes, here we report a one-pot three-component reaction of α-aminophosphonates using a catalytic amount of I₂@NP as a catalyst. A simple protocol was followed in the preparation of novel α-amino phosphonate and phosphonic acid acyclic nucleosides with both pyrimidine and purine nucleobases. Finally, all newly synthesized compounds were evaluated for their anti-HCV and anti-HIV activities.

Results and discussion

In the aim to synthesis a novel conjugated acyclic nucleoside-α-amino acid phosphonate 5-9 and their phosphonic analogs10-14. Our study started with the synthesis of various acyclic nucleoside N-acetaldehyde, which were obtained by treatment of the unprotected nucleobases with bromoacetaldehyde diethyl acetal in the presence of potassium carbonate. Then, aldehydes 4a-d were obtained by hydrolysis of the acetal group3a-d using NP/HCl as an acidic medium (Table 1) [42].
To evaluate the influence of the catalyst, we prepared natural phosphates coated with different Lewis acids using aldehyde 4a, aniline, and triethyl phosphite as a model experiment (Table 2). The best results were obtained with I$_2$@NP in refluxing acetonitrile which yielded α-aminophosphonate 5a in 77% yields (Table 2, entry 4).

**Table 2: Optimal conditions for the preparation of amino phosphonates 5a**

<table>
<thead>
<tr>
<th>Entry</th>
<th>Catalyst</th>
<th>Yield (%)$^a$</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>---</td>
<td>20</td>
</tr>
<tr>
<td>2</td>
<td>NP</td>
<td>0</td>
</tr>
<tr>
<td>3</td>
<td>I$_2$</td>
<td>30</td>
</tr>
<tr>
<td>4</td>
<td>I$_2$@NP</td>
<td>77</td>
</tr>
<tr>
<td>5</td>
<td>ZnCl$_2$@NP</td>
<td>65</td>
</tr>
<tr>
<td>6</td>
<td>ZnBr$_2$@NP</td>
<td>40</td>
</tr>
<tr>
<td>7</td>
<td>CF$_3$SO$_2$H@NP</td>
<td>25</td>
</tr>
<tr>
<td>8</td>
<td>Zn(OTf)$_2$@NP</td>
<td>51</td>
</tr>
<tr>
<td>9</td>
<td>SnCl$_4$@NP</td>
<td>38</td>
</tr>
</tbody>
</table>

$^a$ Isolated yield after gel chromatography purification.
As presented in table 3, the optimized conditions were generalized using acyclic nucleoside acetaldehydes 4a-d, triethyl phosphite, and various amines, including amino acids. The acyclic nucleoside diethyl α-aminophosphonates obtained 5-9(a-d) were further treated with TMSBr in DMF to yield the corresponding α-aminophosphonic acids 10-14(a-d). The reaction tolerates aromatic and aliphatic amines as well as amino acids. The results provide the first examples of conjugated nucleoside-α-aminophosphonates and pave the way for the design of new potential bioactive compounds through a simple and eco-friendly transformation.

Table 3: Results of the preparation of novel conjugated nucleoside-α-aminophosphonates 5-9(a-d) and their phosphonic analogs 10-14 (a-d)

<table>
<thead>
<tr>
<th>Entry</th>
<th>B</th>
<th>R</th>
<th>Phosphonate, Yield (%)&lt;sup&gt;a&lt;/sup&gt;</th>
<th>Phosphonic, Yield (%)&lt;sup&gt;a&lt;/sup&gt;</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>U</td>
<td>Ph</td>
<td>5a, 77</td>
<td>10a, 60</td>
</tr>
<tr>
<td>2</td>
<td>T</td>
<td>Ph</td>
<td>5b, 45</td>
<td>10b, 60</td>
</tr>
<tr>
<td>3</td>
<td>6-AzaU</td>
<td>Ph</td>
<td>5c, 40</td>
<td>10c, 50</td>
</tr>
<tr>
<td>4</td>
<td>A</td>
<td>Ph</td>
<td>5d, 40</td>
<td>10d, 70</td>
</tr>
<tr>
<td>5</td>
<td>U</td>
<td>Ph-CH&lt;sub&gt;2&lt;/sub&gt;</td>
<td>6a, 50</td>
<td>11a, 60</td>
</tr>
<tr>
<td>6</td>
<td>T</td>
<td>Ph-CH&lt;sub&gt;2&lt;/sub&gt;</td>
<td>6b, 60</td>
<td>11b, 35</td>
</tr>
<tr>
<td>7</td>
<td>6-AzaU</td>
<td>Ph-CH&lt;sub&gt;2&lt;/sub&gt;</td>
<td>6c, 60</td>
<td>11c, 35</td>
</tr>
<tr>
<td>8</td>
<td>A</td>
<td>Ph-CH&lt;sub&gt;2&lt;/sub&gt;</td>
<td>6d, 62</td>
<td>11d, 25</td>
</tr>
<tr>
<td>9</td>
<td>U</td>
<td>CH₃-CH-CH₂-CH₃</td>
<td>7a, 66</td>
<td>12a, 25</td>
</tr>
<tr>
<td>10</td>
<td>T</td>
<td>CH₃-CH-CH₂-CH₃</td>
<td>7b, 60</td>
<td>---&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>11</td>
<td>6-AzaU</td>
<td>CH₃-CH-CH₂-CH₃</td>
<td>7c, 50</td>
<td>12c, 20</td>
</tr>
<tr>
<td>12</td>
<td>A</td>
<td>CH₃-CH-CH₂-CH₃</td>
<td>7d, 50</td>
<td>---&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>13</td>
<td>U</td>
<td>CH₂-CO₂Et</td>
<td>8a, 55</td>
<td>13a, 40</td>
</tr>
<tr>
<td>14</td>
<td>T</td>
<td>CH₂-CO₂Et</td>
<td>8b, 60</td>
<td>13b, 20</td>
</tr>
<tr>
<td>15</td>
<td>6-AzaU</td>
<td>CH₂-CO₂Et</td>
<td>8c, 50</td>
<td>13c, 20</td>
</tr>
<tr>
<td>16</td>
<td>A</td>
<td>CH₂-CO₂Et</td>
<td>8d, 55</td>
<td>13d, 20</td>
</tr>
<tr>
<td>17</td>
<td>U</td>
<td>CH₃-CH-CO₂Me</td>
<td>9a, 40</td>
<td>14a, 35</td>
</tr>
<tr>
<td>18</td>
<td>T</td>
<td>CH₃-CH-CO₂Me</td>
<td>9b, 40</td>
<td>14b, 25</td>
</tr>
</tbody>
</table>
Next, we evaluated the antiviral activity of these new phosphonic acids in an HCV sub-genomic RNA replicon using Huh 5.2 cells. Unfortunately, none of the compounds exhibited any significant antiviral activity. On the other hand, these new phosphonic acids were subjected to a standard in vitro antiviral screening using HIV-1 and HIV-2. None of the compounds exhibited any significant antiviral activity.

In the aim to understand the lack of activity of synthesized compounds and suggests some modifications to improve the antiviral activity, molecular docking was carried out. The HCV NS3 protease plays a pivotal role in the replication of the HCV virus. Its inhibition has proven effective in reducing viral loads in humans [43]. Furthermore, several phosphonate derivatives were reported as potential inhibitors of HCV NS3 protease [44-47]. Meanwhile, HIV reverse transcriptase (RT) is an interesting target for the treatment of HIV disease. Over the past years, various nucleoside phosphonates were described to inhibit RT [48-50]. Considering these facts, the compound 14a was docked into the active sites of HCV NS3 protease (PDB: 1W3C) [51] and HIV reverse transcriptase (PDB: 2RF2) [52]. Before, to determine the validity of the docking protocol, first, the self-docking experiments were carried out. The root-mean-square deviations (RMSD) between the predicted and the native poses found to be 2.37Å (1W3C; DN1) and 0.27Å (2RF2; MRX). These results indicated that the adopted docking protocol is good for the reproduction of the native poses.

The docking results are summarized in Table 5 and figures 1-4. Regarding HCV NS3 protease, the compound 14a showed a low affinity with an estimated binding energy of -3.55Kcal/mol. It has reported that the hydrophobic interactions with amino acid residues Val132, Cys159, and Val158 enhanced the inhibitory effect of HCV NS3 protease inhibitors [53]. As shown in figure 2, the hydrophile phosphonic acid group of ligand 14a is located in the hydrophobic cavity (brown region) of protease protein, leading to a decrease in the inhibitory effect (absence of hydrophobic interactions). Thus, the introduction of aromatic hydrophobic pharmacophore (such as biphenyl) [53] will increase the hydrophobic interactions, especially with CYS159, which improve the
antiviral activity against HCV. On the other hand, HIV RT showed also low affinity for compound $14a$ with the estimated free energy of binding $-6.63$ kcal/mol. This can be explained by the lack of hydrophobic and aromatic interactions. Also, the hydroxyl group (phosphonate) is located in the hydrophobic region, leading to repulsion between the protein and the ligand.

Table 5: Detail of binding interactions

<table>
<thead>
<tr>
<th>Protein</th>
<th>Ligand</th>
<th>Energy of binding (Kcal/mol)</th>
<th>RMSD (Å)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1W3C</td>
<td>DN1</td>
<td>-8.79</td>
<td>2.37</td>
</tr>
<tr>
<td></td>
<td>$14a$</td>
<td>-3.55</td>
<td>----</td>
</tr>
<tr>
<td>2RF2</td>
<td>MRX</td>
<td>-10.55</td>
<td>0.27</td>
</tr>
<tr>
<td></td>
<td>$14a$</td>
<td>-6.63</td>
<td>----</td>
</tr>
</tbody>
</table>

Figure 1: 2D interactions of compound $14a$ in the binding sites of HCV NS3 protease
Figure 2: The position of ligand 14a in the hydrophobic cavity of HCV NS3 protease
Conclusion

In conclusion, I$_2$@NP was found to be an efficient catalyst for the one-pot three-component reaction of acyclic nucleoside acetaldehydes, amines, and triethyl phosphites. New $\alpha$-aminophosphonates and $\alpha$-aminophosphonic acids were obtained in good yields with an eco-friendly, inexpensive, and efficient catalyst. This work opens a way to explore the chemical diversity offered by this methodology.

Experimental

Chemistry

Melting points were measured using a Büchi B-545 digital capillary melting point apparatus and used without correction. Reactions were checked with TLC using aluminium sheets with silica gel 60 F254 from Merck. The spectra of 1H NMR and 13C NMR were recorded in solution in DMSO-d$_6$ or CDCl$_3$ on a Bruker Advance 300 spectrometer at 300 and 75 MHz, respectively. The chemical shifts are expressed in parts per million (ppm) by using DMSO-d$_6$ as internal reference. The multiplicities of the signals are indicated by the following abbreviations: s, singlet; d, doublet; t, triplet; q, quadruplet; and m, multiplet, and coupling constants are expressed in Hertz. FAB mass spectra were recorded on a Varian MAT 311A spectrometer. Mass spectra were
collected using an API 3200 LC/MS/MS system, equipped with an ESI source. The chemical reagents used in synthesis were purchased from Fluka, Sigma and Aldrich. Column chromatography was performed on silica gel (30–60 mm). All solvents were distilled and dried before using.

**General Procedure for the preparation of aldehydes 1-4**

To a solution of the base (0.80 mmol) and K₂CO₃ (0.5 eq) in DMF (15 mL) was added 1.5 eq. of bromo acetaldehyde diethylacetal and the resulting mixture was refluxed for a time ranging from 3 to 4 hours. After completion of the reaction as indicated by TLC, the reaction mixture was quenched with acetic acid in water (10 % v/v) and filtered. Evaporation of the solvent followed by purification (chromatography column on silica gel, CH₂Cl₂/MeOH) afforded the corresponding aldehydes.

**1,1-diethoxy-2-(thymin-1-yl) acetaldehyde 3a**
Yield: 56 %; ¹H NMR (300 MHz, CDCl₃) δ (ppm) 1.10 (t, J=7.03 Hz, 6H), 1.85 (s, 3H, CH₃), 3.50 (q, J=7.03Hz, 2H), 3.65 (q, J=7.04Hz, 2H), 3.65 (d, J=5.23Hz, 2H), 4.55 (t, J=5.25 Hz, 1H), 7.05 (s, 1H), 9.55 (sb, 1H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 12.17, 15.27, 50.83, 64.27, 100.35, 109.89, 142.07, 151.30, 164.62. MS (FAB+), m/z = 243 [M+H]⁺. HMRS calcd for C₁₁H₁₈N₂O₄ [M+H]⁺: 243.2716 found 243.2722

**1,1-diethoxy-2-(uracil-1-yl) acetaldehyde 3b**
Yield: 60%; ¹H NMR (300 MHz, CDCl₃) δ (ppm) 1.10 (t, 6H, J=7.02Hz, 2CH₃), 3.50 (q, 2H, J=7.03Hz, OCH₂), 3.65 (q, 2H, J=7.03Hz, OCH₂), 3.70 (d, 2H, J=5.23Hz, N-CH₂), 4.55 (t, 1H, J=5.25Hz, CH-CH₂), 5.50 (d, 1H, J=7.93Hz, H-5), 7.20 (d, 1H, J=7.9Hz, H-6), 9.55 (s, 1H, N3-H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 12.17, 15.27, 50.83, 64.27, 100.35, 109.89, 142.07, 151.30, 164.62. MS (FAB+), m/z = 229 [M+H]⁺. HMRS calcd for C₁₀H₁₆N₂O₄ [M+H]⁺: 229.2450 found 229.2458

**1,1-diethoxy-2-(azauracil-1-yl) acetaldehyde 3c**
Yield: 60%. ¹H NMR (300 MHz, CDCl₃) δ (ppm) 1.10 (t, 6H, J=7.02Hz, 2CH₃), 3.50 (q, 2H, J=7.03Hz, OCH₂), 3.65 (q, 2H, J=7.02Hz, OCH₂), 4.00 (d, 2H, J=5.78Hz, N-CH₂), 4.85 (t, 1H, J=5.80Hz, CH-CH₂), 7.35 (s, 1H, H-5), 9.70 (s, 1H, N-3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 15.17 (2CH₃), 41.36 (CH₂-CH₂), 62.00 (2CH₂-CH₃), 97.80
CH₂), 135.36(C5), 149.26(C4), 155.93(C2). MS (FAB+), \( m/z = 230 \ [M+H]^+ \). HMRS calcd for C9H15N3O4 [M+H]^+: 230.2331 found 230.2325

1, 1-diethoxy-2-(adenin-9-yl) acetaldehyde 3d

Yield: 60%. \(^1\)H NMR (300 MHz, CDCl₃) \( \delta \) (ppm) 1.10 (t, 6H, 7.02Hz, 2CH₃), 3.48 (q, 2H, \( J=7.03Hz \), OCH₂-CH₃), 3.70 (q, 2H, \( J=7.03Hz \), OCH₂-CH₃), 4.20 (d, 2H, \( J=5.22Hz \), N-CH₂), 4.65 (t, 1H, \( J=5.23Hz \), CH₂-CH-O), 5.50 (s, 2H, NH₂), 7.80 (s, 1H, H-2), 8.30 (s, 1H, H-8). \(^{13}\)C NMR (100 MHz, CDCl₃) \( \delta \) (ppm) 15.22 (2CH₃), 46.28 (CH₂-CH), 63.96 (2CH₂-CH₃), 100.37 (CH-CH₂), 119.26 (C5), 141.77 (C6), 150.28 (C4), 153.02 (C2), 155.29 (C8). MS (FAB+), \( m/z = 252 \ [M+H]^+ \). HMRS calcd for C11H17N5O2 [M+H]^+: 252.2849 found 252.2858

Catalyst preparation:

Natural phosphate coated with iodine (I₂@NP):

To a solution of iodine (759 mg) in CH₂Cl₂ (5 mL) was added natural phosphate (3 g) and stirred for 15 min and evaporated to dryness.

Natural phosphate coated with ZnBr₂

To a solution of ZnBr₂ (25 mg) in water (5 mL) was added natural phosphate (175 mg). The mixture was stirred for 15 min and evaporated to dryness.

Natural phosphate coated with ZnCl₂

To a solution of ZnCl₂ (25 mg) in water (5 mL) was added natural phosphate (175 mg). The mixture was stirred for 15 min and evaporated to dryness.

Natural phosphate coated with CF₃SO₃H

To a solution of CF₃SO₃H (1 mL) in methylene chloride (5 mL) was added natural phosphate (3 g). The mixture was stirred for 15 min and evaporated to dryness.

Natural phosphate coated with SnCl₄

A mixture of 1g of SnCl₄ (0.45 mL) in 5 mL water was stirred for 5 minutes, and then 1g of natural phosphate was added. The mixture was stirred for 15 min and evaporated to dryness.

General Procedure for the Synthesis of \( \alpha \)-amino, aminoacid phosphonate acyclonucleosides
To a suspension of the desired acetal (0.41 mmol) in acetonitrile (8 mL), was added 400mg of NP/HCl and water (2 mL) and the mixture was heated (80°C) for 2h. After filtration, the solid residue was washed by CH₃CN and the filtrate was evaporated to yield the corresponding aldehyde 4(a-d) quantitatively. To the crude aldehyde was added acetonitrile (5 mL), 1 equivalent of the amine/aminoacid, triethylphosphite (0.9 eq, 0.6 mL), and 0.2 equivalent of natural doped phosphate (I₂@NP, 104 mg; ZnCl₂@NP, 92mg; ZnBr₂@NP, 160 mg; CF₃SO₃H @NP, 40 mg; Zn(OTf)₂@NP, 280mg; SnCl₄@NP,44 mg). The mixture was refluxed for 3 h and the resulting suspension was filtered and washed with acetonitrile. The filtrate was then evaporated and the residue was dissolved in CH₂Cl₂, washed with a solution of Na₂S₂O₃ (1M) and the organic phase was dried over Na₂SO₄. After filtration and evaporation, the crude product was purified by silica gel chromatography (eluent: CH₂Cl₂/MeOH).

**N¹ (2-anilino-2-diethoxyphosphinyl-ethan-1-yl) uracil 5a**

Yield: 77%. ¹H NMR(400MHz, CDCl₃) δ (ppm) 1.10 (t, 3H, J=7.02Hz, CH₃), 1.20 (t, 3H, J=7.02Hz, CH₃), 3.80-4.10 (m, 4H, 2 OCH₂-CH₃ ), 4.18 (m, 2H, N1-CH₂-CH), 4.40 (m, 1H, CH-CH₂); 4.60 (s, 1H, NH-Ph), 5.50 (d, 1H, J = 7.87Hz, H5), 6.50-7.00 (m, 5H, Ph), 7.05 (d, 1H, J = 7.87Hz ,H6), 10.10 (s, 1H, N3-H). ¹³CNMR(100MHz, CDCl₃)δ (ppm) 15(2CH₃), 48.91(CH₂-CH), 61.80-63.12(2CH₂-CH₃), 100.69 (CH-CH₂), 112.28 (C5), 128.41-144(Ph), 145.23 (C6), 150.39(C4) 163.24(C2). MS (FAB+), m/z =368 [M+H]+. HMRS calcd for C₁₆H₂₂N₃O₅P [M+H]+: 368.3367 found 368.3375

**N¹ (2-anilino-2-diethoxyphosphinyl-ethan-1-yl) thymine 5b**

Yield: 45% ¹H NMR(400 MHz, CDCl₃) δ (ppm) 1.10 (t, 3H, CH₃), 1.20 (t, 3H, J=7.03 Hz, CH₃), 1.70 (s, 3H, CH₃), 3.80-4.10 (m, 4H, 2 OCH₂-CH₃), 4.20 (m, 2H, N1-CH₂-CH), 4.40 (m, 1H, CH-CH₂); 4.60 (s, 1H,NH- Ph), 6.50-7.00 (m, 5H, Ph), 7.05 (s, 1H, H6), 9.80 (s, 1H, N3-H). ¹³CNMR(100MHz, CDCl₃)δ (ppm) 12.11 (CH₃); 16.35 (2CH₃), 48.91 (CH₂-CH), 62.74(CH₂-CH₃), 63.99(CH₂-CH₃), 100.01(CH-CH₂), 113.35 (C5),129.32-141(Ph), 146.43 (C6), 151.46 (C4), 164.56(C2).MS (FAB+), m/z = 382 [M+H]^+.HMRS calcd for C₁₇H₂₄N₃O₅P [M+H]^+: 382.3633 found 382.3640

**General Procedure for the Synthesis of α–amino, aminoacid phosphonate acyclonucleosides**

To a suspension of the desired acetal (0.41 mmol) in acetonitrile (8 mL), was added 400mg of NP/HCl and water (2 mL) and the mixture was heated (80°C) for 2h. After
filtration, the solid residue was washed by CH$_3$CN and the filtrate was evaporated to yield the corresponding aldehyde quantitatively. To the crude aldehyde was added acetonitrile (5 mL), 1 equivalent of the amine/amino acid, triethylphosphite (0.9 eq, 0.06 mL), and 0.2 equivalent of natural doped phosphate(NP/I$_2$, 104 mg). The mixture was refluxed for 3 h and the resulting suspension was filtered and washed with acetonitrile. The filtrate was then evaporated and the residue was dissolved in CH$_2$Cl$_2$, washed with a solution of Na$_2$S$_2$O$_3$ (1M) and the organic phase was dried over Na$_2$SO$_4$. After filtration and evaporation, the crude product was purified by silica gel chromatography (eluent: CH$_2$Cl$_2$/MeOH).

N$^1$(2-anilino-2-diethoxyphosphinyl-ethan-1-yl) uracil 5a

Yield: 77%. $^1$H NMR(400MHz, CDCl$_3$) $\delta$ (ppm) 1.10 (t, 3H, $J$=7.02Hz, CH$_3$), 1.20 (t, 3H, $J$=7.02Hz, CH$_3$), 3.80-4.10 (m, 4H, 2 OCH$_2$-CH$_3$), 4.18 (m, 2H, N1-CH$_2$-CH), 4.40 (m, 1H, CH-CH$_2$); 4.60 (s, 1H, NH-Ph), 5.50 (d, 1H, $J$ = 7.87Hz, H5), 6.50-7.00 (m, 5H, Ph), 7.05 (d, 1H, $J$ = 7.87Hz ,H6), 10.10 (s, 1H, N3-H).$^{13}$CNMR(100MHz, CDCl$_3$) $\delta$ (ppm) 15(2CH$_3$), 48.91(CH$_2$-CH), 61.80-63.12(2CH$_2$-CH$_3$), 100.69 (CH-CH$_2$), 112.28 (C5), 128.41-144(Ph), 145.23 (C6), 150.39(C4) 163.24(C2). MS (FAB+), $m/z$ =368 [M+H]$^+$.HMRS calcd for C16H22N3O5P [M+H]$^+$: 368.3367 found 368.3375

N$^1$(2-anilino-2-diethoxyphosphinyl-ethan-1-yl) thymine 5b

Yield: 45% $^1$H NMR(400 MHz, CDCl$_3$) $\delta$ (ppm) 1.10 (t, 3H, CH$_3$), 1.20 (t, 3H, $J$=7.03 Hz, CH$_3$), 1.70 (s, 3H, CH$_3$), 3.80-4.10 (m, 4H, 2 OCH$_2$-CH$_3$), 4.20 (m, 2H, N1-CH$_2$-CH), 4.40 (m, 1H, CH-CH$_2$); 4.60 (s, 1H,NH- Ph), 6.50-7.00 (m, 5H, Ph), 7.05 (s, 1H, H6), 9.80 (s, 1H, N3-H).$^{13}$CNMR(100MHz, CDCl$_3$) $\delta$ (ppm) 12.11 (CH$_3$); 16.35 (2CH$_3$), 48.91(CH$_2$-CH), 62.74(CH$_2$-CH$_3$), 63.99(CH$_2$-CH$_3$), 100.01(CH-CH$_2$), 113.35 (C5),129.32-141(Ph), 146.43 (C6), 151.46 (C4), 164.56(C2).MS (FAB+), $m/z$ = 382 [M+H]$^+$.HMRS calcd for C17H24N3O5P [M+H]$^+$: 382.3633 found 382.3640

N$^1$(2-anilino-2-diethoxyphosphinyl-ethan-1-yl) azauracil 5c

Yield: 40%. $^1$H NMR(400MHz, CDCl$_3$) $\delta$ (ppm) 1.10 (t,3H, CH$_3$), 1.22(t, 3H, $J$=7.02Hz, CH$_3$), 3.90-4.10 (m, 4H, 2 OCH$_2$-CH$_3$), 4.15 (m, 2H, N1-CH$_2$-CH), 4.40 (m, 1H, CH-CH$_2$), 4.50 (s, 1H, NH-Ph), 7.20 (s, 1H, H5), 6.50-7.10 (m, 5H, Ph), 10.60 (s, 1H,N3-H).$^{13}$CNMR(100MHz, CDCl$_3$)$\delta$(ppm) 14.38(2CH$_3$), 38.22(CH$_2$-CH), 60.83(CH$_2$-CH$_3$), 62.00 (CH$_2$-CH$_3$);111.45(CH-CH$_2$), 113.37 (C5), 127.27-144.53 (Ph), 147.59 (C4),
154.57(C2). MS (FAB+), m/z = 369 [M+H]^+. HMRS calcd for C15H21N4O5P [M+H]^+: 369.3248 found 369.3239

N^9 (2-anilino-2-diethoxyphosphinyl-ethan-1-yl) adenine 5d

Yield: 40%. \(^1\)H NMR(400 MHz, CDCl\(_3\)) \(\delta\) (ppm) 1.10 (t,3H, J=7.06Hz, CH\(_3\)), 1.20 (t, 3H, CH\(_3\)), 3.90-4.10 (m, 4H, 2 OCH\(_2\)-CH\(_3\)), 4.30 (m, 2H, N9-CH\(_2\)-CH), 4.40 (m, 1H, CH-CH\(_2\)), 4.60 (s, 1H, NH-Ph), 5.60 (s, 2H, NH\(_2\)), 6.40-7.05 (m, 5H, Ph), 7.75 (s,1H,H2), 8.35 (s,1H,H8). \(^{13}\)CNMR(100 MHz, CDCl\(_3\))\(\delta\) (ppm) 16.34 (2CH\(_3\)), 44.66(CH\(_2\)-CH), 62.83-63.96 (2CH\(_2\)-CH\(_3\)), 113.52 (CH-CH\(_2\)), 119.52(C5),129.22-140 (Ph),141.34 (C6), 150.28 (C4), 153.02 (C2), 155.35(C8). MS (FAB+), m/z = 391 [M+H]^+. HMRS calcd for C17H23N6O3P [M+H]^+: 391.3766 found 391.3773

N^1 (2-benzylamino-2-diethoxyphosphinyl-ethan-1-yl) thymine 6a

Yield: 50%. \(^1\)H NMR(400 MHz, CDCl\(_3\)) \(\delta\) (ppm) 1.10 (t,3H, J=7.02Hz, CH\(_3\)), 1.20 (t, 3H, CH\(_3\)), 3.20 (m,1H,Ha),3.50 (m, 1H, Hb), 3.80-3.90 (m, 2H, N1-CH\(_2\)-CH), 4.00-4.10 (m, 4 H, 2 OCH\(_2\)-CH\(_3\)), 4.10 (m, 1H, CH-CH\(_2\)), 4.25 (s, 1H, NH-Ph), 7.05 (s, 1H, H6), 7.20 (d, 1H, J = 7.87 Hz, H5), 6.50-7.00 (m, 5H, Ph), 9.80 (s, 1H, N3-H). \(^{13}\)CNMR(100 MHz, CDCl\(_3\))\(\delta\) (ppm): 12.19 (CH\(_3\)), 16.44(2CH\(_3\)), 48.02(CH\(_2\)-CH), 52.31(CH\(_2\)-Ph), 62.57(CH\(_2\)-CH\(_3\)), 62.77 (CH\(_2\)-CH\(_3\)), 109 (CH-CH\(_2\)), 127 (C5), 141.9 (C6), 128-139 (Ph), 151.1 (C4), 164.64(C2). MS(ES+), m/z = 382 [M+H]^+. HMRS calcd for C17H24N3O5P [M+H]^+: 382.3633 found 382.3640

N^1 (2-benzylamino-2-diethoxyphosphinyl-ethan-1-yl) azauracil 6b

Yield: 60%. \(^1\)H NMR(400 MHz, CDCl\(_3\)) \(\delta\) (ppm) 1.10 (t,3H, J=7.03 Hz, CH\(_3\)), 1.30 (t, 3H, J=7.03 Hz, CH\(_3\)), 1.80 (s, 3H, CH\(_3\)), 3.10 (m, 1H, Ha ), 3.50 (m, 1H, Hb), 3.80-3.90 (m, 2H, N1-CH\(_2\)-CH), 4.00-4.10 (m, 4 H, 2 OCH\(_2\)-CH\(_3\)), 4.10 (m, 1H, CH-CH\(_2\)), 4.25 (s, 1H, NH-Ph), 7.05 (s, 1H, H6), 7.20 (m, 5H, Ph), 9.80 (s, 1H,N3-H). \(^{13}\)CNMR(100 MHz, CDCl\(_3\))\(\delta\) (ppm): 12.19 (CH\(_3\)), 16.44(2CH\(_3\)), 48.02(CH\(_2\)-CH), 52.31(CH\(_2\)-Ph), 62.57(CH\(_2\)-CH\(_3\)), 62.77 (CH\(_2\)-CH\(_3\)), 109 (CH-CH\(_2\)), 127 (C5), 141.9 (C6), 128-139 (Ph), 151.1 (C4), 164.64(C2). MS(ES+), m/z = 396 [M+H]^+. HMRS calcd for C18H26N3O5P [M+H]^+: 396.3899 found 396.3906

N^1 (2-benzylamino-2-diethoxyphosphinyl-ethan-1-yl) azauracil 6c

Yield: 60%. \(^1\)H NMR(400 MHz, CDCl\(_3\)) \(\delta\) (ppm) 1.30 (t,3H, J=7.02Hz, CH\(_3\)), 1.40 (t, 3H, J=7.02Hz, CH\(_3\)), 3.40 (m, 1H, Ha), 3.80 (m, 1H, Hb), 3.90-4.10 (m, 2H, N1-CH2-
CH), 4.30 (m, 4H, 2 OCH₂-CH₃), 4.50 (m, 1H, CH-CH₂), 4.55 (s, 1H,NH-CH₂- Ph), 7.20 (s, 1H, H5), 7.40 (m, 5H, Ph), 10.60 (s, 1H,N3-H).¹³CNMR(100 MHz, CDCl₃) δ (ppm) 16.60 (2CH₃), 39.80(CH₂-CH), 51.77(CH₂-Ph),62.51(CH₂-CH₃), 62.72(CH₂-CH₃), 127.23(CH-CH₂), 135.14 (C5), 128-135 (Ph), 149.48(C4), 156.26 (C2). MS(ES+), m/z = 383 [M+H]⁺.HMRS calcd for C₁₆H₂₃N₄O₅P [M+H]⁺: 383.3513 found 383.3506

N⁹ (2-benzylamino-2-dioxyphosphinyl-ethan-1-yl) adenine 6d

Yield: 62%.¹H NMR(400 MHz, CDCl₃) δ (ppm) 1.20 (t,3H, J=7.06Hz, CH₃), 1.30 (t, 3H, J=7.06Hz, CH₃), 3.30 (m, 1H, Ha), 3.90 (m,1H, Hb), 3.70- 3.90 (m, 2H, N₉-CH₂-CH₃), 4.20 (m, 4H, 2 OCH₂-CH₃), 4.25 (s,1H,NH-CH₂-Ph), 5.90 (s, 2H, NH₂), 7.30 (m, 5H, Ph), 7.90 (s,1H,H₈).¹³CNMR(100 MHz, CDCl₃) δ (ppm) 16.44 (2CH₃), 44.11(CH₂-CH), 62.74-62.83(2CH₂-CH₃), 119.26 (CH-CH₂), 52.18(CH₂-Ph), 127.19 (C5), 128-138 (Ph), 141.86 (C6), 150.16(C4), 152.75 (C2), 155.28 (C8). MS(ES+), m/z = 405 [M+H]⁺.HMRS calcd for C₁₈H₂₅N₆O₃P [M+H]⁺: 405.4032 found 405.4040

N¹ (2-butyramino-2-dioxyphosphinyl-ethan-1-yl) uracil 7a

Yield: 66%.¹H NMR(400 MHz, CDCl₃) δ (ppm) 0.80 (t,3H,CH₃-CH₂), 1.20 (m, 6H, 2CH₃-CH₂), 1.30 (q,2H,CH₂-CH₃), 1.40 (m,2H,CH₂-CH₂-CH₃), 2.60-2.80 (m, 2H, NH-CH₂), 3.00 -3.60 (m, 2H, Ha and Hb), 4.10 (m, 1H, CH-CH₂), 4.25 (m, 4H, 2 OCH₂-CH₃ ), 4.30 (m, 1H, N-H), 5.60 (d, 1H, J= 7.87Hz, H₅), 7.30 (d, 1H, J= 7.87Hz , H₆), 9.00 (s, 1H,N₃-H).¹³CNMR(100 MHz, CDCl₃) δ (ppm) 13.87 (CH₃), 16.45(CH₃), 16.50(CH₃), 20.08 (CH₂-CH₃), 28.92(CH₂-CH₂-CH₃), 48.82 (CH₂-CH₂-CH₂-CH₃), 49.84(CH₂-CH), 62.47-62.88(2CH₂-CH₃), 100.92 (CH-CH₂), 128.81 ( C5), 146.48 (C6), 150.92 ( C4), 163.91 (C2). MS(ES+), m/z = 348 [M+H]⁺.HMRS calcd for C₁₄H₂₆N₃O₅P [M+H]⁺: 348.3471 found 348.3480

N¹ (2-butyramino-2-dioxyphosphinyl-ethan-1-yl) thymine 7b

Yield: 60%.¹H NMR(400 MHz, CDCl₃) δ (ppm) 0.80 (t,3H, CH₃-CH₂), 1.30 (t, 6H,2 CH₃-CH₂), 1.80 (s, 3H, CH₃), 1.35 (m, 2H,CH₂-CH₃), 1.40 (m,2H,CH₂-CH₂-CH₃), 2.90 (m,2H,NH-CH₂), 3.10 (m, 2H, Ha and Hb), 3.90-4.10 (m, 6H, 2 OCH₂-CH₃,CH-CH₂ and N-H), 7.20 (s, 1H, H₆), 8.70 (s, 1H,N₃-H).¹³CNMR(100 MHz, CDCl₃) δ (ppm) 12.19 (CH₃), 13.87 (CH₃-CH₂),16.45(CH₃), 16.50(CH₃), 20.08 (CH₂-CH₃),
28.92(CH₂-CH₂-CH₃), 48.82 (CH₂CH₂-CH₂-CH₃), 49.84(CH₂-CH), 62.47-62.88(2CH₂-CH₃), 100.92( CH-CH₂), 128.81 (C5), 146.48 (C6), 150.92 ( C4), 163.91 (C2). 

MS(ES+), \textit{m/z} = 362 [M+H]^+.HMRS calcd for C15H28N3O5P [M+H]^+: 362.3736 found 362.3742

\( \text{N}^1 (2\text{-butylamino-2-diethoxyphosphinyl-ethan-1-yl) azauracil 7c} \)

Yield: 47%. \( ^1 \text{H NMR}(400 MHz, CDCl₃) \delta (ppm): 0.80 (t,3H,CH₃), 1.20 (m,6H, 2CH₃-CH₂, 1.30 (q, 2H,CH₂-CH₃), 1.40 (m,2H,CH₂-CH₂-CH₃), 2.50-2.80 ( m,2H,NH-CH₂), 3.40 (m, 2H, Ha and Hb), 4.00-4.30 (m, 6H, 2 OCH₂, CH-CH₂, N-H), 7.30 (s, 1H, H5), 10.60 (s, 1H,N3-H). \( ^{13} \text{CNMR}(100 MHz, CDCl₃) \delta (ppm) 13.87(CH₃-CH₂), 16.49-16.54 (2CH₃), 48(CH₂-CH₂), 20.01 (CH₂-CH₃), 32.35 (CH₂-CH₂-CH₂-CH₃), 62.22(CH₂-CH₂), 62.67(CH₂-CH₃), 100(CH-CH₂), 135.06 (C5), 149.51 (C4), 156.42 (C2). MS(ES+), m/z = 349 [M+H]^+.HMRS calcd for C13H25N4O5P [M+H]^+: 349.3351 found 349.3360

\( \text{N}^9 (2\text{-butylamino-2-diethoxyphosphinyl-ethan-1-yl) adenine 7d} \)

Yield: 50%. \( ^1 \text{H NMR}(400 MHz, CDCl₃) \delta (ppm) 0.80 (t,3H,CH₃), 1.20 (m,6H, 2CH₃-CH₂, 1.20-1.40 (q, 2H,CH₂-CH₃), 1.40-1.50 (m,2H,CH₂-CH₂-CH₃), 2.50-2.70 ( m,2H,NH-CH₂), 3.10 (m, 2H, Ha and Hb), 4.10 (m,4H, 2OCH₂-CH₂, 4.25 (m, 1H, N-H), 4.30 (m,1H,CH-CH₂), 6.10 (s,2H, NH₂), 8.10 (s, 1H, H2), 8.30 (s,1H, H8). \( ^{13} \text{CNMR}(100 MHz, CDCl₃) \delta (ppm) 13.53 (CH₃-CH₂), 16.44 (2CH₃),20.04 (CH₂-CH₃), 29.35 (CH₂-CH₂-CH₃), 40.02 (CH₂-CH₂-CH₂-CH₃)44.17(CH₂-CH), 63.02(CH₂-CH₃), 118.99 (CH-CH₂), 120 (C5), 142.31 (C6), 150.08 (C4), 152.7 (C2), 155.2 (C8). MS(ES+), m/z = 371 [M+H]^+.HMRS calcd for C13H25N4O5P [M+H]^+: 349.3351 found 349.3360

\( \text{N}^1(2\text{-glycinoethylester-2 diethoxyphosphinyl-ethan-1-yl) uracil 8a} \)

Yield: 60%. \( ^1 \text{H NMR}(400 MHz,CDCl₃) \delta (ppm) 1.13 (t, 3H,J=7.13Hz,CH₃), 1.30(t,6H,J=7.06Hz,2CH₃), 3.25(t,1H,J=4.6Hz,CH), 3.50-3.60(m,2H,CH₂), 3.70(d, 2H,J=4.6Hz,CH₂), 4.10(m,7H,3CH₂, NH), 5.60(d,1H,J=7.9Hz,H-5), 7.20 (d, 1H, J= 7.9 Hz, H-6), 9.55 (sd, 1H, NH). \( ^{13} \text{C NMR}(100 MHz,CDCl₃) \delta (ppm) 14.15(CH₃), 16.45 (CH₃), 16.49 (CH₃), 49.40 (CH), 52.98 (CH₂), 54.45 (CH₂), 60.90 (CH₂), 62.77 (CH₂), 62.73 (CH₂), 101.39(C5), 146.06( C6),151.35 (C4), 164.11 (C2), 173.63 (CO, ester). MS(ES+), m/z =378 [M+H]^+.HMRS calcd for C14H24N3O7P [M+H]^+: 378.3300 found 378.3310
N^1(2-glycinoethylester-2-diethoxyphosphinyl-ethan-1-yl)thymine 8b

Yield: 50%. \(^1H\) NMR (400 MHz, CDCl\(_3\)) δ(ppm) 1.15 (t, 3H, J=7.15Hz, CH\(_3\)), 1.30(t, 6H, J=7.05Hz, 2CH\(_3\)), 1.80 (s, 3H, CH\(_3\)), 3.25(t, 1H, J=4.5Hz, CH), 3.50-3.60(m,2H,CH\(_2\)), 3.70(d, 2H, J=4.5Hz, CH\(_2\)), 4.10(m,7H,3CH\(_2\), NH), 7.10(s,1H,H-6), 9.20 (s, 1H, NH). \(^13C\) NMR (100 MHz, CDCl\(_3\)) δ(ppm) 14.15(CH\(_3\)), 16.45(CH\(_3\)), 16.67(CH\(_3\)), 44.70(CH\(_2\)), 44.80(CH\(_2\)), 54.46(CH\(_3\)), 56.48(CH\(_2\)), 61.81(CH\(_2\)), 64.27(CH), 119.82(C5), 143.71(C6), 151.28(C4), 153.63(C2), 157.11(C8), 173.19(CO, ester). MS(ES+), m/z =392 [M+H]^+. HMRS calcd for C\(_{15}\)H\(_{26}\)N\(_3\)O\(_7\)P[M+H]^+: 392.3566 found 392.3574

N^1(2-glycinoethylester-2-diethoxyphosphinyl-ethan-1-yl)6-Aza uracil 8c

Yield: 50%. \(^1H\) NMR (400 MHz, CDCl\(_3\)) δ(ppm) 1.15 (t, 3H, J=7.13Hz, CH\(_3\)), 1.30(t, 6H, J=7.06Hz, 2CH\(_3\)), 3.25(t, 1H, J=3.6Hz, CH), 3.50-3.60(dd,2H, J=3.6Hz, CH\(_2\)), 4.00-4.20(m,9H,4CH\(_2\), NH), 7.20(s,1H,H-5), 10.50 (s, 1H, NH). \(^13C\) NMR (100 MHz, CDCl\(_3\)) δ(ppm) 14.15(CH\(_3\)), 16.53(CH\(_3\)), 16.48(CH\(_3\)), 39.50(CH), 49.01(CH\(_2\)), 52.02(CH\(_2\)), 54.07(CH\(_2\)), 60.72(CH\(_2\)), 62.70(CH\(_2\)), 135.00(C5), 150.09(C4), 156.97(C2), 172.65(CO, ester). MS(ES+), m/z =379 [M+H]^+. HMRS calcd for C\(_{13}\)H\(_{23}\)N\(_4\)O\(_7\)P[M+H]^+: 379.3181 found 379.3175

N^9(2-glycinoethylester-2-diethoxyphosphinyl-ethan-1-yl)adenine 8d

Yield: 55%. \(^1H\) NMR (400 MHz, CD\(_3\)OD) δ(ppm) 1.20 (t, 3H, J=7.15Hz, CH\(_3\)), 1.30(t, 6H, J=7.05Hz, 2CH\(_3\)), 3.20(t,1H, J=3.6Hz, CH), 3.50-3.60(dd,2H, J=3.6Hz, CH\(_2\)), 4.00-4.20(m,9H,4CH\(_2\), NH), 7.20(s,1H,H-5), 10.50 (s, 1H, NH). \(^13C\) NMR (100 MHz, CD\(_3\)OD) δ(ppm): 9.31(CH\(_3\)), 14.46(CH\(_3\)), 16.67(CH\(_3\)), 44.70(CH\(_2\)), 44.80(CH\(_2\)), 54.46(CH\(_2\)), 56.48(CH\(_2\)), 61.81(CH\(_2\)), 64.27(CH), 119.82(C5), 143.71(C6), 151.28(C4), 153.63(C2), 157.11(C8), 173.19(CO, ester). MS(ES+), m/z =401 [M+H]^+. HMRS calcd for C\(_{15}\)H\(_{25}\)N\(_6\)O\(_5\)P[M+H]^+: 401.3700 found 401.3710

N^1(2-alaninomethylene-2-diethoxyphosphinyl-ethan-1-yl) uracil 9a

Yield: 50%. \(^1H\) NMR (400 MHz, CDCl\(_3\)) δ(ppm) 1.11 (d, 3H, J=7.13Hz, CH\(_3\)), 1.16(m, 9H,3CH\(_3\)), 3.25 (t, 1H, J=4.6Hz, CH), 3.50-3.60(q,1H, J=7.13Hz, CH), 3.70(d, 2H, J=4.6Hz, CH\(_2\)), 4.01(m,5H,2CH\(_2\), NH), 5.60(d,1H, J=7.9Hz, H-5), 7.20(d,1H, J=7.9Hz, H-6), 9.55(sd, 1H, NH). \(^13C\) NMR (100 MHz, CDCl\(_3\)) δ(ppm) 16.42(CH\(_3\)), 18.71(CH\(_3\)), 19.06(CH\(_3\)), 46.41(CH\(_2\)), 49.30(CH), 50.70(CH\(_3\)), 51.90(CH), 62.60(CH\(_2\)), 62.70(CH\(_2\)), 101.30(C5), 146.10(C6), 151.10(C4), 164.40(C2), 174.70
**N<sup>1</sup> (2-alaninomethyl ester-2-diethoxy phosphinyl-ethan-1-yl) Thymine 9b**

Yield: 55%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ (ppm) 1.00 (d, 3H, J = 7.10 Hz, CH₃), 1.30 (m, 12H, 4CH₃), 3.25 (t, 1H, J = 4.60 Hz, CH), 3.50-3.60 (q, 1H, J = 7.10 Hz, CH), 3.70 (d, 2H, J = 4.60 Hz, CH₂), 4.01 (m, 5H, 2CH₂, NH), 7.10 (s, 1H, H-6), 9.60 (s, 1H, NH). ¹³C NMR (100 MHz, CDCl<sub>3</sub>) δ (ppm) 11.00 (CH₃), 16.60 (CH₃), 18.80 (CH₃), 19.20 (CH₃), 49.10 (CH₂), 49.60 (CH), 50.90 (CH₃), 52.09 (CH), 55.30 (CH₂), 62.70 (CH₂), 109.20 (C5), 141.90 (C6), 151.30 (C4), 164.90 (C2), 174.80 (CO, ester). MS (ES+), m/z = 406[M+H]<sup>+</sup>. HMRS calcd for C₁₆H₂₈N₃O₇P [M+H]<sup>+</sup>: 406.3832 found 406.3842

**N<sup>1</sup> (2-alaninomethyl ester-2-diethoxy phosphinyl-ethan-1-yl) 6aza uracil 9c**

Yield: 52%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ (ppm) 1.00 (d, 3H, J = 7.10 Hz, CH₃), 1.30 (m, 9H, 3CH₃), 3.25 (t, 1H, J = 4.60 Hz, CH), 3.50-3.60 (q, 1H, J = 7.10 Hz, CH), 3.70 (d, 2H, J = 4.60 Hz, CH₂), 4.01 (m, 5H, 2CH₂, NH), 7.20 (s, H-5), 9.60 (s, 1H, NH). ¹³C NMR (100 MHz, CDCl<sub>3</sub>) δ (ppm) 11.00 (CH₃), 16.60 (CH₃), 18.80 (CH₃), 19.20 (CH₃), 49.10 (CH₂), 49.60 (CH), 50.90 (CH₃), 52.09 (CH), 55.30 (CH₂), 62.70 (CH₂), 109.20 (C5), 141.90 (C6), 151.30 (C4), 164.90 (C2), 174.80 (CO, ester). MS (ES+), m/z = 393[M+H]<sup>+</sup>. HMRS calcd for C₁₄H₂₅N₄O₇P [M+H]<sup>+</sup>: 393.3447 found 393.3440

**N<sup>9</sup> (2-alaninomethyl ester-2-diethoxy phosphinyl-ethan-1-yl) adenine 9d**

Yield: 55%. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD) δ (ppm) 1.00 (d, 3H, J = 7.13 Hz, CH₃), 1.40-1.60 (m, 9H, 3CH₃), 3.10 (t, 1H, J = 4.50 Hz, CH), 3.50-3.70 (q, 1H, J = 7.13 Hz, CH), 4.01 (d, 2H, J = 4.50 Hz, CH₂), 4.35 (m, 5H, 2CH₂, NH), 8.20 (s, 2H, H-2, H-8). ¹³C NMR (100 MHz, CD<sub>3</sub>OD) δ (ppm) 10.00 (CH₃), 16.74 (CH₃), 19.18 (CH₃), 45.01 (CH₂), 48.00 (CH₂), 52.23 (CH₃), 53.07 (CH), 56.17 (CH) 119.82 (C5), 143.75 (C6), 151.08 (C4), 153.70 (C2), 157.17 (C8), 176.24 (CO, ester). MS (ES+), m/z = 415[M+H]<sup>+</sup>. HMRS calcd for C₁₆H₂₇N₆O₅P [M+H]<sup>+</sup>: 415.3965 found 415.3975

**Deprotection of α-amino, and α-aminoacids phosphonates acyclonucleosides**

**General Procedure**

To a suspension of α-aminophosphonate acyclonucleosides<sup>5-9</sup> (0.15 mmol) in acetonitrile (5 mL) was added TMSBr (10 eq), and the resulting mixture was stirred overnight at room temperature. After evaporation of the solvent, water (5 mL) was
added and the resulting suspension was neutralized with an 28% ammonia solution. The water was then removed in vacuo and the residue was purified with on preparative plate (eluent: isopropanol/NH$_4$OH/H$_2$O) and reverse phase HPLC (C18). The final product was obtained after a cation exchange chromatographic on a Na$^+$ DOWEX50WX2. The residue obtained is dried, the purity of products was controlled by HPLC (eluent: CH$_3$CN/H$_2$O, 50/50 v / v).

$N^1$(2-anilino-2-dihydroxyphosphinyl-ethan-1-yl) uracil 10a

Yield: 64%. $^1$H NMR (400 MHz, D$_2$O) δ (ppm) 3.60 (m, 1H, Ha), 3.85 (m, 1H, Hb), 4.35 (m, 1H, CH- CH$_2$), 5.50 (d, 1H, $J$= 7.87 Hz, H5), 6.50-7.00 (m, 5H, Ph), 7.45 (d, 1H, $J$=7.87 Hz,H6). $^{13}$C NMR (100 MHz, D$_2$O) δ (ppm) 51.6(CH$_2$-CH), 100.69(CH-CH$_2$), 112.48 (C5), 129-144 (Ph), 148.23(C6), 152.73 (C4), 167.24(C2). MS(ES+), m/z = 312 [M+H]$^+$.HMRS calcd for C12H14N3O5P [M+H]$^+$: 312.2304 found 312.2312

$N^1$(2-anilino-2-dihydroxyphosphinyl-ethan-1-yl) thymine 10b

Yield: 60%. $^1$H NMR (400MHz, D$_2$O) δ (ppm) 1.50 (s, 3H, CH$_3$), 3.70 (m, 1H, Ha), 4.20 (m, 1H, Hb), 4.35 (m, 1H, CH-CH$_2$), 6.50-7.00 (m, 5H, Ph), 7.30 (s, 1H, H6). $^{13}$C NMR (100MHz, D$_2$O) δ (ppm) 12.11 (CH$_3$), 51.53 (CH$_2$-CH), 110.01 (CH-CH$_2$), 113.42 (C5), 129.32-141.00 (Ph), 143.95 (C6), 152 (C4), 164.56 (C2). MS (ES+), m/z = 326 [M+H]$^+$. HMRS calcd for C13H16N3O5P [M+H]$^+$: 326.2570 found 326.2580

$N^1$(2-anilino-2-dihydroxyphosphinyl-ethan-1-yl) azauracil 10c

Yield: 50%. $^1$H NMR (400 MHz, D$_2$O) δ (ppm) 4.05 (m, 1H, CH-CH$_2$), 4.20 (m, 1H, Ha), 4.40 (m, 1H, Hb), 7.15 (s, 1H, H5), 6.50-7.00 (m, 5H, Ph).$^{13}$C NMR (100 MHz, D$_2$O)δ (ppm) 42.24(CH$_2$-CH), 113.17(CH-CH$_2$), 129.39-141 (Ph), 135.05 (C5), 150.42 (C4), 158.24 (C2). MS(ES+), m/z = 313 [M+H]$^+$. HMRS calcd for C11H13N4O5P [M+H]$^+$: 313.2184 found 313.2178

$N^9$(2-anilino-2-dihydroxyphosphinyl-ethan-1-yl) adenine 10d

Yield: 70%. $^1$H NMR (400MHz, D$_2$O) δ (ppm) 4.35 (m, 1H, Ha), 4.50 (m, 1H, Hb), 4.90 (m, 1H, CH-CH$_2$), 6.30-7.10 (m, 5H, Ph), 7.75 (s, 1H, H2), 8.35 (s, 1H, H8). $^{13}$C NMR (100 MHz, D$_2$O) δ (ppm) 46.48 (CH$_2$-CH), 112.49 (CH-CH$_2$), 117.30 (C5), 128.40-144 (Ph), 142.88 ( C6), 149.06 ( C4), 151.79 ( C2), 154.92 (C8). MS (ES+), m/z = 363 [M+H]$^+$. HMRS calcd for C13H15N8O3P [M+H]$^+$: 363.2837 found 363.2845

$N^1$(2-benzylamino-2- dihydroxyphosphinyl -ethan-1-yl) uracil 11a
Yield: 60%. $^1$H NMR (400 MHz, D$_2$O), $\delta$ (ppm) 3.20 (m, 1H, Ha), 3.90 (m, 1H, Hb), 4.00 (m, 2H, N1-CH$_2$-CH), 4.30 (m, 1H, CH-CH$_2$), 5.60 (d, 1H, $J$= 7.87, H5), 7.30 (m, 5H, Ph), 7.45 (d, 1H, $J$=7.87, H6). $^{13}$C NMR (100MHz, D$_2$O)$\delta$ (ppm) 48.81(CH$_2$-CH), 51.60(CH$_2$-Ph), 102.12(CH-CH$_2$), 129.29 ( C5), 129 -131.11 (Ph), 146.84 ( C6), 153 (C4), 166.45 ( C2). MS(ES+), $m/z$ =326 [M+H]$^+$. HMRS calcd for C13H16N3O5P [M+H]$^+$: 326.2570 found 326.2564

$^{11}$b (2-benzylamino-2-dihydroxyphosphinyl-ethan-1-yl) thymine

Yield: 35%. $^1$H NMR (400 MHz, D$_2$O) $\delta$ (ppm) 1.70 (s, 3H, CH$_3$), 3.30 (m, 1H, Ha), 3.90 (m, 1H, Hb), 4.10- 4.30 (m, 2H, N1-CH$_2$-CH), 4.50 (m, 1H, CH-CH$_2$), 7.40 (m, 5H, Ph), 7.20 (s, 1H, H6). $^{13}$C NMR (100MHz, D$_2$O) $\delta$ (ppm) 11.19 (CH$_3$), 48.50(CH$_2$-CH), 111.39 ( CH-CH$_2$), 127 ( C5), 129-130 (Ph), 142.22 (C6), 153.14 ( C4), 166.64 ( C2). MS (ES+), $m/z$ = 340 [M+H]$^+$. HMRS calcd for C14H18N3O5P [M+H]$^+$: 340.2835 found 340.2841

$^{11}$c (2-benzylamino-2- dihydroxyphosphinyl-ethan-1-yl) azauracil

Yield: 35%. $^1$H NMR (400MHz, D$_2$O) $\delta$ (ppm) 3.40 (m,1H,Ha), 4.10 (m, 1H, Hb), 4.20 (m, 2H, N1-CH$_2$-CH), 4.50 (m, 1H, CH-CH$_2$), 7.30 (m, 5H, H5), 7.40 (m, 5H, Ph). $^{13}$C NMR (100MHz, D$_2$O) $\delta$ (ppm) 39.63 (CH$_2$-CH), 50.13 (CH$_2$-Ph), 128.21 (CH-CH$_2$), 129-131 (Ph), 135.51 (C5), 150.59 (C4), 157.93 ( C2). MS(ES+), $m/z$ = 327 [M+H]$^+$. HMRS calcd for C12H15N4O5P [M+H]$^+$: 327.2457 found 327.2450

$^{11}$d (2-benzylamino-2- dihydroxyphosphinyl-éthan-1-yl) adenine

Yield: 24%. $^1$H NMR (400 MHz, D$_2$O) $\delta$ (ppm): 3.30 (m, 1H, Ha), 3.90 (m, 1H, Hb), 4.00-4.30 (m, 2H, N9-CH$_2$-CH), 4.60 (m, 1H, CH-CH$_2$), 7.00 (m, 5H, Ph), 7.90 (s, 1H, H2), 7.95 (s, 1H, H8). $^{13}$C NMR (100MHz, D$_2$O) $\delta$ (ppm) 44.11 (CH$_2$-CH), 52.18 (CH$_2$-Ph), 119.26 (CH-CH$_2$), 127.15 (C5), 128-130 (Ph), 141.86 (C6), 150.16 (C4), 152.75 (C2), 155.28 (C8). MS(ES+), $m/z$ = 349 [M+H]$^+$. HMRS calcd for C14H17N6O3P [M+H]$^+$: 349.2969 found 349.2977

$^{12}$a (2-butylamino-2-dihydroxyphosphinyl-éthan-1-yl) uracil

Yield: 25%. $^1$H NMR (400MHz, D$_2$O), $\delta$ (ppm) 0.80 (t, 3H, CH$_3$-CH$_2$), 1.20 (q, 2H, CH$_2$-CH$_3$), 1.50 (m, 2H, CH$_2$-CH$_2$-CH$_3$), 3.10 (m, 2H, CH$_2$-(CH$_2$)$_2$-CH$_3$), 3.40-4.10 (m, 2H, N1-CH$_2$), 4.30 (m, 1H, CH-CH$_2$), 5.60 (d, 1H, $J$ = 7.87Hz, H5), 7.30 (d, 1H, $J$ = 7.87Hz, H6). $^{13}$C NMR (100MHz, D$_2$O) $\delta$ (ppm) 12.74 (CH$_3$-CH$_2$), 19.04 (CH$_2$-CH$_3$),
28.03 (CH$_2$-CH$_2$-CH$_3$), 46.42 (CH$_2$-CH$_2$-CH$_2$-CH$_3$), 47.90 (CH$_2$-CH), 102.05 (CH-CH$_2$), 128.81 (C5), 147.21 (C6), 153.21 (C4), 166.72 (C2). MS (ES+), $m/z = 292$ [M+H]$^+$. HMRS calcd for C10H18N3O5P [M+H]$^+$: 292.2407 found 292.2415

N$^1$ (2-butylamino-2-dihydroxyphosphinyl-éthan-1-yl) azauracil 12c

Yield: 20%. $^1$H NMR (400MHz, D$_2$O) $\delta$ (ppm) 0.80 (t, 3H, CH$_3$), 1.20 (q, 2H, CH$_2$-CH$_3$), 1.50 (m, 2H, CH$_2$-CH$_2$-CH$_3$), 3.10 (m, 2H, CH$_2$-(CH$_2$)$_2$-CH$_3$), 3.70 (m, 1H, CH-CH$_2$), 4.30 (m, 2H, N1-CH$_2$), 7.50 (s, 1H, H5).

MS (ES+), $m/z = 293$ [M+H]$^+$. HMRS calcd for C9H17N4O5P [M+H]$^+$: 293.2288 found 293.2295

N$^1$(2-glycinoethylester-2-dihydroxyphosphinyl-ethan-1-yl) uracil 13a

Yield: 40%. $^1$H NMR(400MHz, D$_2$O) $\delta$ (ppm) 1.15 (t, 3H, J=7.10Hz, CH$_3$), 1.25 (t, 1H, J=3.50Hz, CH), 3.70 (m, 2H, CH$_2$), 3.80 (d, 2H, J=3.5Hz, CH$_2$), 4.10 (q, 4H, J=7.10Hz, 2CH$_2$), 5.70 (d, 2H, $J=7.9$Hz, 1H, H-5), 7.60 (d, 2H, $J=7.90$Hz, 1H, H-6). 13CNMR (100MHz, D$_2$O) $\delta$(ppm) 16.45 (CH$_3$), 49.40 (CH), 54.79 (CH$_2$), 56.50 (CH$_2$), 57.39 (CH$_2$), 101.37 (C5), 147.18 (C6), 152.20 (C4), 166.67 (C2), 171.42 (CO, ester). MS (ES+), $m/z = 322$ [M+H]$^+$. HMRS calcd for C10H16N3O7P [M+H]$^+$: 322.2237 found 322.2246

N$^1$(2-glycinoethylester-2-dihydroxyphosphinyl-ethan-1-yl) Thymine 13b

Yield: 20%. $^1$H NMR(400MHz, D$_2$O) $\delta$(ppm) 1.15 (t, 3H, J=7.10Hz, CH$_3$), 1.70 (s, 3H, CH$_3$), 3.25(t, 1H, J=3.50Hz, CH), 3.70 (m, 2H, CH$_2$), 3.80(d, 1H, J=3.45Hz, CH$_2$), 4.10(q, 4H, J=7.10Hz, 2CH$_2$), 7.60s, 1H, H-6). 13CNMR (100 MHz,D$_2$O) $\delta$(ppm) 13.20 (CH$_3$), 16.45 (CH$_3$), 49.40 (CH), 54.79 (CH$_2$), 56.50 (CH$_2$), 57.39 (CH$_2$), 101.37 (C5), 147.18 (C6), 152.20 (C4), 166.67 (C2), 171.42 (CO, ester). MS(ES+), $m/z =336$ [M+H]$^+$. HMRS calcd for C11H18N3O7P [M+H]$^+$: 336.2502 found 336.2512

N$^1$(2-glycinoethylester-2-dihydroxyphosphinyl-ethan-1-yl) azauracil 13c

Yield: 20%. $^1$H NMR(400MHz, D$_2$O) $\delta$(ppm) 1.15 (t, 3H, J=7.10Hz, CH$_3$), 3.25 (t, 1H, J=3.5Hz, CH), 3.70 (m, 2H, CH$_2$), 3.80 (d, 2H, J=3.5Hz, CH$_2$), 4.10 (q, 4H, J=7.10Hz, 2CH$_2$), 7.50 (s, 1H, H-5). 13CNMR (100MHz, D$_2$O) $\delta$ (ppm) 13.16 (CH$_3$), 39.50 (CH), 49.01 (CH$_2$), 52.02 (CH$_2$), 62.57 (CH$_2$), 135.55 (C5), 150.77 (C4), 158.23 (C2), 172.65

**N9(2-glycinoethylester-2-dihydroxyphosphinyl-ethan-1-yl) adenine 13d**

Yield: 20%. $^1$H NMR (400 MHz, D$_2$O) δ(ppm) 1.00 (t, 3H, J=7.13Hz,CH$_3$), 3.20(t, 1H, J=3.6Hz,CH), 3.40(m,2H,CH$_2$), 3.70(d, 2H, J=3.6Hz,CH$_2$), 4.20(q,2H,J=7.13Hz,CH$_2$), 8.10 (s, 2H, H-2, H-8). $^{13}$C NMR (100MHz D$_2$O) δ(ppm) 12.99 (CH$_3$), 44.80 (CH$_2$), 54.46 (CH$_2$), 56.48 (CH$_2$), 62.71 (CH), 119.82 (C5), 143.71 (C6),152.28(C4), 153.63 (C2), 157.11 (C8), 173.19 (CO, ester). MS(ES+), m/z = 345 [M+H]+. HMRS calcd for C11H17N6O5P [M+H]+: 345.2636 found 345.2628

**N1(2-alaninomestyler-2-dihydroxyphosphinyl-ethan-1-yl) uracil 14a**

Yield: 35%. $^1$H NMR(400 MHz, D$_2$O) δ(ppm) 1.15 (d, 3H,J=7.10Hz,CH$_3$), 2.75(t, 1H,J=4.6Hz,CH), 3.40(q,1H,J=7.10Hz,CH),3.60(s,3H,CH$_3$), 4.00(d,2H,J=4.6Hz,CH$_2$), 5.65(d,J=7.90 Hz,1H,H-5), 7.50(d,J=7.90 Hz,1H,H-6).$^{13}$CNMR (100MHz, D$_2$O) δ(ppm) 17.84 (CH$_3$), 48.84 (CH), 52.7 (CH), 55.03 (CH$_3$), 58.49 (CH$_2$), 101(C5), 147.07(C6), 158.50(C4), 174.64(C2), 181.70 (CO, ester). MS(ES+), m/z = 326 [M+H]+. $^{31}$PNMR (75 MHz, D$_2$O) δ(ppm) 14.93, 16.22. HMRS calcd for C11H18N3O7P [M+H]+: 326.2509 found 326.2502

**N1(2-alaninomestyler-2-dihydroxyphosphinyl-ethan-1-yl) thymine 14b**

Yield: 25%. $^1$HNMR(400 MHz, D$_2$O) δ(ppm) 1.15 (d, 3H,J=7.10Hz,CH$_3$), 1.80(s,3H,CH$_3$) 2.75(t, 1H,J=4.60 Hz,CH), 3.40(q,1H,J=7.10Hz,CH),3.60(s,3H,CH$_3$), 4.00(d,2H,J=4.6Hz,CH$_2$), 7.44(s,1H,H-6).$^{13}$C NMR (100MHz, D$_2$O) δ(ppm) 17.83 (CH$_3$), 48.84 (CH), 52.62 (CH), 53.54 (CH$_3$), 58.54 (CH$_2$), 101.55 (C5), 147.06(C6),158.50(C4), 174.60(C2), 181.65 (CO, ester). MS(ES+), m/z = 350 (M+H+). $^{31}$PNMR (75 MHz, D$_2$O) δ(ppm): 14.09, 16.06. HMRS calcd for C12H20N3O7P [M+H]+: 350.2768 found 350.2774

**N1(2-alaninomestyler-2-dihydroxyphosphinyl-ethan-1-yl) azauracil 14c**

Yield: 30%. $^1$H NMR (400 MHz, D$_2$O) δ(ppm) 1.19(d, 3H,J=7.13Hz,CH$_3$), 3.30(t,1H,J=4.4Hz,CH), 3.40(q,1H,J=7.13Hz,CH), 3.60(s,3H,CH$_3$), 4.00(d,2H,J=4.40 Hz,CH$_2$), 7.50(s,1H,H-5).$^{13}$CNMR (100MHz, D$_2$O) δ(ppm) 13.20 (CH$_3$), 39.85 (CH), 47.30 (CH), 53.79 (CH$_3$), 62.63 (CH$_2$), 135.59(C5), 150.78(C4), 158.31(C2), 181.60
N\textsuperscript{9}(2-alaninomethylester-2-dihydroxyphosphinyl-ethan-1-yl) adenine 14d

Yield: 15%. \textsuperscript{1}H NMR (400 MHz, D\textsubscript{2}O) \(\delta\) (ppm) 1.19 (d, 3H, J=7.10Hz,CH\textsubscript{3}), 3.30 (t, 1H, J=4.5Hz,CH), 3.40 (q, 1H, J=7.10Hz,CH), 3.60 (s, 3H, CH\textsubscript{3}), 4.00 (d, 2H, J=4.50 Hz,CH\textsubscript{2}), 8.00 (s, 2H, H-8, H-2). \textsuperscript{13}CNMR (100MHz, D\textsubscript{2}O) \(\delta\) (ppm) 13.20 (CH\textsubscript{3}), 39.85 (CH), 47.30 (CH), 53.79 (CH\textsubscript{3}), 62.63 (CH\textsubscript{2}), 119.82 (C5), 143.71 (C6), 152.28 (C4), 153.63 (C2), 157.11 (C8), 173.19 (CO, ester). MS(ES+), m/z = 359 (M+H\textsuperscript{+}). \textsuperscript{31}PNMR (75 Mhz, D\textsubscript{2}O) \(\delta\) (ppm) 15.06, 15.90. HMRS calcd for C12H19N6O5P \([\text{M+H}]^+\): 359.2902 found 359.2912

**Molecular docking**

In silico computational docking studies were performed using AutoDock4.2 [54]. The X-ray crystallographic structures of HCV NS3 protease and HIV reverse transcriptase were downloaded from the RCSB Protein Data Bank (PDB) 1W3C [51] and 2RF2 [52], respectively. The proteins were prepared separately by removing water and co-crystallized ligands bound with the proteins to make receptor free of any ligand before docking. Then, Polar hydrogens and Gastieger charges were added using the MGL Tools and proteins saved in PDBQT format [55]. Ligand 14a was created separately using ChemDraw Ultra 12.0, energy minimized in Chem3D, torsional bonds of ligand were set flexible and saved in PDBQT format. Next, the receptor was kept rigid, the grid covering all the amino acid residues present inside the active site of proteins was built for 1W3C(grid box size of 40Å X 46Å X 52Å with a spacing of 0.375Å between the grid points and centered at 66.685 (x), 18.626 (y), and 0.736 (z)) and for 2RF2(grid box size of 50Å X 40Å X 56Å with a spacing of 0.375Å between the grid points and centered at 7.845 (x), 13.204 (y), and 15.671 (z)). The best conformers were searched by the Lamarckian genetic algorithm (LGA), the population size was set to 150 and maximum number of energy evaluation was set to 250,000.00. Finally, the results were analyzed and visualized by the discovery studio.

**Acknowledgements**

This project was supported by the comité mixte inter-universitaire Franco-Marocain programme (A.I. N° = Ma/06/143) and by Centre National de Recherche Scientifique et
Technique project RS/2011/01 (Rabat, Morocco). The authors would like to thank Professor J. Balzarini (Rega Institute for Medical Research, Katholieke University Leuven, Leuven, Belgium) for his scientific contribution and help on the realization of this work. Also, we would like to acknowledge the technical staff of the CAC (Centre of Analysis and Characterization) University Cadi Ayyad Marrakech for running the spectroscopic analysis.

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