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Unravelling the formation of BAB block copolymer assemblies during PISA in water

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FIGURE FOR TOC

Text for TOC

BAB triblock copolymers prepared using PISA *via* aqueous RAFT dispersion polymerization self-assemble into a transient network of bridged micelles. The slowdown of the exchange of B blocks between micellar cores during the polymerization is highlighted as well as the parameters affecting the polymerization.

Abstract:

The mechanism of formation of associative BAB triblock copolymers through aqueous polymerization-induced self-assembly (PISA) is investigated for the first time, on copolymers constituted of a hydrophilic poly(*N,N*-dimethylacrylamide) block (PDMAc = block A) and a hydrophobic poly(diacetone acrylamide) block (PDAAm = block B). Such BAB copolymers tend to form bridged micelles/networks, which was expected to make the PISA process much more complex than for conventional diblock copolymers. Kinetic monitoring, light scattering analyses and macroscopic observations allowed identifying crucial parameters that influence the polymerization and the final dispersion properties, notably the stirring of the polymerization medium, the macroRAFT agent concentration, its ionization degree (related to the pH) and its Z group alkyl chain length.

Introduction:

BAB copolymers are amphiphilic triblock copolymers, where the inner A block is hydrophilic and covalently connected to two outer B blocks that are hydrophobic. Such copolymers may find applications as thermoplastic elastomers¹, filtration membranes² or hydrogels^{3,4} for biomedical applications. They are usually synthesized in solution in a non-selective organic solvent in which both types of blocks are soluble. However, this strategy requires the use of organic solvents and does not allow the formation of amphiphilic triblock copolymers for which no common solvent can be found.

An attractive pathway for the synthesis of amphiphilic block copolymers is polymerization-induced self-assembly (PISA),^{5–8} which consists in forming the B blocks by chain extension of the A block in a selective solvent for the latter, so that polymerization and self-assembly occur simultaneously. PISA can be achieved in water, a green solvent, and may be applied even if the A and B blocks cannot be dissolved in a common non-selective solvent thereby circumventing the limitations encountered in homogeneous solution. In addition, PISA allows the production of amphiphilic block copolymers in high solids contents (up to 50 wt%) and is scalable to industrial quantities. In contrast to AB and ABA amphiphilic copolymers, only very few BAB triblocks prepared by PISA can be found in the literature. Zhang's group pioneered this field in 2014 and reported the possibility

to synthesize linear BAB triblock copolymer assemblies through a RAFT dispersion PISA process of styrene conducted in alcohol/water mixtures.^{9–12} In more detail, depending on the macroRAFT agent used, spherical particles, complex aggregates or higher order morphologies (and their aggregates) were observed.^{9,11} Recently, our group reported the first successful synthesis of BAB copolymers by PISA in pure water¹³: symmetrical bifunctional poly(*N,N*-dimethylacrylamide) (PDMAc) macroRAFT agents with a central benzoic acid group and two outer alkyl chains as Z-groups were used in the aqueous dispersion polymerization of diacetone acrylamide (DAAm) (see structure in **Scheme S1**). The study notably highlighted the crucial role of the central benzoic acid group in the stabilization of the nano-objects and also revealed an impact of the length of the alkyl chains ends. The scarcity of the examples found in the literature and the problems of colloidal stability and control of the polymerization reported show that the synthesis of BAB triblock copolymers by PISA is much more complex than PISA of AB diblock copolymers and hardly understood.^{14–17}

The self-assembly of BAB triblock copolymers in aqueous medium is well understood when thermodynamic equilibrium can be reached.^{18,19} As for AB diblock copolymers, the B blocks of BAB copolymers self-assemble in aqueous medium in order to shield themselves from water, leading to spherical hydrophobic cores in the simplest and most frequent case. In dilute solution, the two B blocks of one BAB triblock gather into the same hydrophobic core, so that the A block forms a loop, leading to flower-like micelles (see **Scheme 1**). At higher concentration, the two B blocks of a single BAB triblock can enter different hydrophobic cores, bridging flower-like micelles into larger and larger aggregates until a 3D-spanning physical network is eventually formed above a critical percolation concentration C_p .

Scheme 1. Self-assembly of BAB triblocks under thermodynamic equilibrium into **a)** individual flower-like micelles at low concentration, **b)** finite size aggregates of connected flower-like

micelles at intermediate concentration and c) 3D-spanning transient network of connected flower-like micelles above the percolation concentration.

Attraction caused by the bridges may result in macroscopic phase separation between a very dilute aqueous phase and a concentrated transient percolated network where intermicellar bridges are favored over intramicellar loops. Phase separation is disfavored for charged polymers at low ionic strength (because the entropic penalty is high due to the confinement of the counter-ions in the concentrated polymer phase). In contrast phase separation happens more readily for neutral BAB copolymers or for charged BAB copolymers at high ionic strength.^{20–23} It is also disfavored when AB diblock copolymers are added to the BAB triblock copolymers because this reduces attraction.²⁴ The rheological properties of the transient network are controlled by the life-time of the bridges which remains reasonably short (allowing exchange of bridges/B blocks between different hydrophobic cores) if thermodynamic equilibrium is indeed reached.

However, many amphiphilic block copolymers actually form frozen structures in aqueous medium²⁵, meaning that the exchange of B blocks between hydrophobic cores is kinetically hindered within the time scale of observation (because of glassy B blocks and/or because of too hydrophobic B blocks). In that case, self-assembly is impacted by the process and may strongly differ from the scenario described above, where thermodynamic equilibrium is reached. In the case of PISA, the increase of the B block length during polymerization affects self-assembly both in terms of aggregation¹⁴ and exchange dynamics. Longer B blocks indeed favor aggregation and slow down exchange between cores.^{7,25,26} Moreover, the A block may be pre-assembled if the RAFT agent is functionalized by a long alkyl chain.²⁷ Very different organizations at the nano-scale may thus be expected during the synthesis of BAB triblock copolymers by PISA in aqueous medium depending on the initial organization of the bifunctional macroRAFT agent (A block) and on the way the exchange rate of the B blocks and their extent of aggregation are affected by their growth. It is also possible that chain extension of the macroRAFT agent does not happen symmetrically so that the B block on one side reaches a frozen self-assembled state before the other one is large enough to self-assemble, further affecting the behavior of the system. Moreover, if macroscopic phase separation does occur during PISA due to the attraction caused by the bridges, polymerization control may be lost.

We observed in our former PISA study on PDAAm-*b*-PDMAc-*b*-PDAAm particles,¹³ that changing the initial concentration of macroRAFT agent, the ionization degree of the central benzoic acid function on the R group of the macroRAFT agent or the length of its terminal alkyl chains (C₄ or C₁₂) on its Z groups (see **Scheme 2**) could result in the loss of colloidal stability during PISA, and was in some cases accompanied by a loss of control over the polymerization. In contrast, suspensions that remained colloidally stable always resulted in controlled polymerizations. In order to better understand the uninvestigated and obviously complex mechanism of self-assembly of BAB triblock copolymers during PISA, in the present study, we monitored a typical synthesis of BAB triblock copolymers by PISA (**Scheme 2**) and investigated their self-assembly by static and dynamic light scattering as well as qualitative macroscopic observations.

Experimental part:

Materials

1,3,5-Trioxane (99%, Aldrich), 4,4'-azobis(4-cyanopentanoic acid) (ACPA, $\geq 98\%$ Aldrich) and diacetoneacrylamide (DAAm, 99%, Aldrich) were used as received.

Polymer synthesis

The synthesis of the macroRAFT agents (C₁₂-TTC-PDMAc)₂-BA and (C₄-TTC-PDMAc)₂-BA (named respectively in this study C₁₂-macroRAFT agent and C₄-macroRAFT agent) were described in a former study¹³ and detailed in the **SI (Scheme S1, Table S1, Figure S1)**. The macroRAFT agent extensions with DAAm (**Scheme 2**) conducted with stirring were performed according to a formerly developed protocol¹³ at 70°C, in a 2 mL round bottom flask using an olive-shaped stirring bar (5×10 mm) and a stirring speed of 500 rpm as detailed in the **SI (Table S2, Figure S2)**. The macroRAFT agent extensions with DAAm conducted without stirring during the polymerization as well as the kinetic study performed in D₂O in NMR tubes are described in details in the **SI, section 2**.

Sample preparation for static and dynamic light scattering (SLS and DLS)

The macroRAFT agent powders were dissolved and the copolymer dispersions were diluted in pure milli-Q water at room temperature at the desired concentration. After complete

dissolution/dispersion, NaCl concentration was adjusted to 0.01 mol L⁻¹ using concentrated stock solutions to prevent any polyelectrolyte effect²⁸. For the macroRAFT agent samples, the desired ionization degree was then set by addition of concentrated NaOH solution. The ionization degree corresponds to the percentage of charged benzoic acid units: $\alpha = [\text{COO}^-]/([\text{COO}^-] + [\text{COOH}]) = n_{\text{NaOH}}/n_{\text{benzoic acid}}$. Before analysis, the samples were filtered through 0.45 μm pore size filters (except for $C > 20 \text{ g L}^{-1}$ for which the solutions were analyzed without filtration). The NaCl concentration was kept low and the solutions were studied at concentrations where they remained transparent to avoid multiple scattering which would have been detrimental to the data interpretation.

Characterization techniques

¹H NMR. DAAM conversion was determined by ¹H NMR in D₂O at room temperature with a Bruker 300 or 400 MHz spectrometer in 5-mm diameter tubes by the relative integration of the protons of the 1,3,5-trioxane at 5.2 ppm and the vinylic protons of DAAM at 6.3-6.2 and 5.7 ppm.

SEC. Size exclusion chromatography (SEC) analyses of the polymers were carried out with two PSS GRAM 1000 Å columns (8 × 300 mm; separation limits: 1 to 1000 kg mol⁻¹) and one PSS GRAM 30 Å column (8 × 300 mm; separation limits: 0.1 to 10 kg mol⁻¹) coupled with a differential refractive index (RI) detector and a UV detector. DMF (+ LiBr, 1 g L⁻¹) at 60 °C was used as the mobile phase with a flow rate of 0.8 mL min⁻¹. Samples were filtrated on 0.2 μm pore-size membrane before injection. The SEC analysis of the RAFT agent (C₁₂-TTC)₂-BA was carried out with two columns thermostated at 40 °C (PSS SDV, linear M, 8 mm × 300 mm, bead diameter: 5 μm), coupled with a differential refractive index (RI) detector and a UV detector. THF at 40 °C was used as the mobile phase with a flow rate of 1 mL min⁻¹. Samples were filtrated on 0.45 μm pore-size membrane before injection. The dispersity ($D = M_w/M_n$), the number-average molar mass (M_n), and the weight-average molar mass (M_w) were calculated from the RI signals by a calibration curve based on PMMA standards with OmniSEC 5.11 software.

Static and Dynamic Light Scattering (SLS and DLS). Light scattering (LS) measurements were done with a standard ALV-CGS3 system equipped with an ALV-5003 multi tau correlator system (ALV GmbH, Germany) with a vertically polarized helium-neon laser with wavelength $\lambda = 633$

nm as light source. The measurements were done at 20 °C (or 70 °C) over a large range of scattering vectors q varying from ca. $3 \times 10^{-4} \text{ \AA}^{-1}$ to $3 \times 10^{-3} \text{ \AA}^{-1}$. $q = (4\pi n/\lambda) \times \sin(\theta/2)$, with θ the angle of observation and n the refractive index of the solvent.

Static light scattering (SLS). For SLS, the absolute intensity, R_θ in cm^{-1} , scattered by the polymer was determined according to **(Equation 1)**.

$$R_\theta = \frac{I_{\text{solution}}(\theta) - I_{\text{water}}(\theta)}{I_{\text{toluene}}(\theta)} \times \left(\frac{n_{\text{water}}}{n_{\text{toluene}}} \right)^2 R_{\text{toluene}} \quad \textbf{(Equation 1)}$$

With I_{solution} , I_{water} and I_{toluene} the average intensities scattered, respectively, by the solution, the solvent (water), and the reference (toluene) at angle θ ; $n_{\text{water}} = 1.333$ and $n_{\text{toluene}} = 1.496$ correspond to the refractive indexes of the solvent and the reference respectively; and $R_{\text{toluene}} = 1.35 \times 10^{-5} \text{ cm}^{-1}$ the Rayleigh ratio of toluene for a wavelength $\lambda = 633 \text{ nm}$.

The apparent molar mass (M_{app}) of the solute was calculated from the Rayleigh ratio of the solution as:

$$M_{\text{app}} = R_\theta / (K \times C) \quad \textbf{(Equation 2)}$$

$$\text{where } K = (4\pi^2 n_{\text{solvent}}^2) \times (\partial n / \partial C)^2 / (\lambda^4 \cdot N_a)$$

Here $\partial n / \partial C = 0.15 \text{ mL g}^{-1}$ (value known for PDMAc homopolymer in water²⁹) is the specific refractive index increment of the solute and N_a is Avogadro's number. This value will be used for all the copolymers analyzed in this work. When the particles were small compared to q^{-1} and the solutions were sufficiently diluted so that interactions could be neglected, M_{app} corresponded to the weight-average molecular weight of the particles. At high polymer concentrations, M_{app} deviates from the real molecular weight of the particles either due to repulsive interactions, which decrease M_{app} and that are caused by electrostatic interactions and/or excluded volume interactions, or by attractive interactions, which are mainly caused by bridging between flower-like micelles.²⁰ The evolution of M_{app} with C therefore gives precious information about the relative strength of repulsive and attractive interactions between the particles.

Dynamic light scattering (DLS). The normalized electric field autocorrelation functions ($g_1(t)$) obtained by DLS measurements were analyzed in terms of a relaxation time (τ) distribution:

$$g_1(t) = \int A(\tau) e^{\left(\frac{-t}{\tau}\right)} d\tau \quad \textbf{(Equation 3)}$$

The REPES routine³⁰ was used to obtain $A(\tau)$ (the fraction of particles with relaxation time τ defined as a scattering intensity-weighted amplitude) without assuming a specific shape for the distribution. For most polymer solutions, two populations of scattering particles were observed by DLS, a slow and a fast one. In those cases, it was observed that the slow mode of relaxation corresponded to an apparent hydrodynamic radius (R_{app}) value about 10 times as large as for the fast mode of relaxation; implying that the former mode corresponded to particles having a molecular weight at least a hundred times larger than those of the latter mode assuming that the slow mode corresponded to randomly formed fractal aggregates with a fractal dimension close to 2 (note that denser aggregates would exhibit even higher molecular weights, strengthening the conclusion). As a conclusion, even though the slow mode of relaxation contributed significantly to the DLS and SLS signals because of its large molecular weight, its weight percentage within the sample could be neglected. The SLS and DLS data presented in the manuscript therefore correspond to the contribution of the fast mode of relaxation assuming that the contribution of the slow mode in terms of weight concentration is negligible. A similar behavior has already been observed in the literature for many other self-assembling systems and the same treatment was applied to get rid of the contribution of the large aggregates.³¹ The apparent diffusion coefficient D was calculated from the average relaxation rate of the fast relaxation mode as $D = \langle \tau^{-1} \rangle / q^2$ and is related to R_{app} according to **(Equation 4.**

$$R_{app} = \frac{k_B T}{6\pi\eta D} \quad \textbf{(Equation 4)}$$

With k_B Boltzmann constant, T the absolute temperature and η the viscosity of the solvent at the temperature of the experiment. When the particles were small compared to q^{-1} and the solutions were sufficiently diluted so that interactions could be neglected, R_{app} corresponded to the z-average hydrodynamic radius, R_h .

Cryo-TEM. Cryo-transmission electron microscopy (cryo-TEM) was used to determine the morphology and size of the polymer nanoparticles. The samples were diluted in water to 1wt% prior to analysis. According to protocols reported elsewhere,^{32,33} thin liquid films of particle dispersions were prepared at room temperature by depositing 4 μ L of the diluted sample and successive blotting. They were flash frozen in liquid ethane and observed at -180 °C on a JEOL

JEM-2100 LaB6 microscope operating at 200 kV under low-dose conditions (10 electrons $\text{\AA}^{-2} \text{s}^{-1}$). Digital images were recorded on a ultrascan 1000, 2k×2k CCD camera (Gatan, USA).

Results and Discussion

1. Overview of the self-assembly throughout the PISA process

Scheme 2. Reaction scheme for the synthesis of PDAAm-*b*-PDMAc-*b*-PDAAm BAB triblock copolymers by polymerization induced self-assembly (PISA) in water.

Stirring may be expected to affect the organization of BAB triblock copolymers prepared by PISA. Indeed, if a percolated network is formed during the process, it might be broken through stirring and will reform only if the B blocks can exchange rapidly compared to the polymerization time. To evaluate the impact of stirring during the dispersion polymerization, PDAAm-*b*-PDMAc-*b*-PDAAm BAB triblock copolymers were synthesized in typical conditions (70 °C, 10 wt% of DAAm), with or without stirring (see **Scheme 2**, and for the experimental details see **SI** section **2.a**), and **Table S2**). The RAFT dispersion polymerizations of DAAm were conducted with the previously used¹³ C₁₂-macroRAFT agent ($M_{n,\text{NMR}} = 8.6 \text{ kg mol}^{-1}$) targeting a $DP_{n/\text{arm}}$ PDAAm of ~ 80 (see entry B and E in **Table S2**).

Figure 1. Photographs showing the macroscopic aspect of the final polymerization media synthesized with (left) and without (right) stirring (entry B and E, **Table S2**).

As shown in the photographs in **Figure 1**, the absence or presence of stirring during polymerization provided macroscopically very different samples. The polymerization performed under stirring led to a liquid and stable dispersion, whereas the unstirred polymerization phase-separated into a gel-like phase and an excluded liquid phase. The resulting gel fraction could not be dispersed again in water, suggesting the formation of a non-dynamic network (see **Scheme 1**). Interestingly, SEC analysis of the stirred sample and the gel fraction of the unstirred sample (which represents by far the majority of material) revealed a nearly perfect overlay of the SEC chromatograms (see **Figure S2**) and a relatively narrow molar mass distribution ($\mathcal{D} < 1.3$, see **Table S2**), demonstrating that the absence of stirring had no significant impact on the control of the polymerization, at least not in these polymerization conditions. In view of these results, we can conclude that the different visual aspect of the samples in the absence or presence of stirring is not related to intrinsic differences in chemistry of the macromolecular chains. It must therefore be correlated to their supramolecular organization and their dynamics during the polymerization.

In order to investigate in more details what happens at the nano-scale during the synthesis of BAB triblock copolymers by PISA and understand the role of stirring, we used LS to characterize the self-assembly in aqueous medium of the BAB triblock copolymers for different degrees of polymerization of the B blocks (corresponding to different monomer conversions during the PISA process). In our previous study¹³ we had shown that the pH of the solution, *i.e.* the ionization degree (α) of the central benzoic acid group had an impact on the assembly. Here, we set α at a constant value of 28% for a $[\text{DAAm}]_0/[\text{C}_{12}\text{-macroRAFT}]_0 = 100$ (see **Table 1**), which corresponds to

conditions where tiny spherical particles ($d_{\text{cryo-TEM}} \sim 20$ nm) had been obtained. First the C₁₂-macroRAFT agent (see **Table S1**, entry C₁₂-PDMAc-*bis* $M_{n,\text{NMR}} = 8.5$ kg mol⁻¹) was analyzed by SLS and DLS in water at 20 °C (see **Figure 2**). To investigate the self-assembly over a broad range of concentrations (from 0.2 to 70 g L⁻¹), the macroRAFT agent was dissolved at 70 g L⁻¹ and then diluted to the different concentrations.

The evolution of the apparent molar mass (M_{app}) with the polymer concentration corresponded to the typical signature of BAB triblock copolymers self-assembling under thermodynamic equilibrium.²⁰ Indeed, in diluted conditions ($C < 2$ g L⁻¹), M_{app} was constant ($M_{\text{app}} \sim 140$ kg mol⁻¹) and corresponded to the molecular weight of the self-assembled particles. This value gave an aggregation number (N_{agg}) of 14 ($N_{\text{agg}} = M_{\text{app}} (\text{particles}) / M_{\text{w}} (\text{C}_{12}\text{-PDMAc-}i\text{bis})$ with $M_{\text{w}} = M_{n,\text{NMR}} (\text{C}_{12}\text{-PDMAc-}i\text{bis}) \times \bar{D} = 8.5 \times 1.18 = 10.0$ kg mol⁻¹). Moreover, the determined hydrodynamic radius ($R_{\text{h}} \sim 7$ nm) was compatible with the formation of individual flower-like micelles.[†] Above 3 g L⁻¹, a sharp increase of M_{app} with concentration was observed due to attractive interactions, which is typical of the formation aggregates of bridged micelles (see **Scheme 1**). At even higher concentrations ($C > 20$ g L⁻¹) the repulsive excluded volume interactions dominated, leading to a decrease of M_{app} .

This study showed that the C₁₂ alkyl chains were hydrophobic enough to trigger self-assembly of the C₁₂-macroRAFT agent. Moreover, the self-assembly was dynamic (at thermodynamic equilibrium) since the observed variations of M_{app} with concentration could not have been obtained by successive dilutions from 70 g L⁻¹ if the bridges between the flower-like micelles had been frozen. Finally, the evolution of M_{app} with concentration and the increase of the viscosity qualitatively observed with the naked eye above $C = 30$ g L⁻¹ implied that a percolated transient network was formed at 70 g L⁻¹, that is the macroRAFT concentration at which the PISA process was performed. The whole behavior of this polymer was consistent with what was reported for telechelic poly(ethylene oxide) decorated on both sides by C₁₂ alkyl chains.^{21,34} The polymerization temperature being 70 °C, we also performed LS of the macroRAFT agent at this temperature, but no significant differences in aggregation were observed when comparing the LS experiments at

[†] The maximum theoretical length of the stretched PDMAc corona is $R_{\text{h,max}} = 0.25 \times DP_{\text{n/arm PDMAc}} = 11$ nm.

either 20 °C or 70 °C (see **Figure S3**). We could thus reasonably conclude that the polymerization of DAAM started from a dynamic transient percolating network of C₁₂-macroRAFT.

Figure 2. Evolution of the apparent molar mass M_{app} as a function of the polymer concentration obtained by static light scattering at 20 °C in water for the macroRAFT agent C₁₂-PDMAc-*bis* and for block copolymers with $DP_{n/arm} = 12, 21$ and 33 prepared by dispersion polymerization of DAAM in D₂O in the presence of the macroRAFT agent C₁₂-PDMAc-*tris*, at $\alpha = 28\%$, without stirring (see **Table 1**). The solid lines are guides for the eyes. Dotted lines indicate precipitation of the sample caused by the addition of salt used for LS analyses ($[NaCl] = 0.01 \text{ mol L}^{-1}$).

Thereafter, a typical dispersion polymerization of DAAM in the presence of the C₁₂-macroRAFT agent (see **Table S1**, entry C₁₂-PDMAc-*tris*, $M_{n,NMR} = 8.7 \text{ kg mol}^{-1}$) was conducted in D₂O, without stirring (70 °C, 10 wt% of DAAM, $[C_{12}\text{-macroRAFT}]_0 = 70 \text{ g L}^{-1}$, with $\alpha = 28\%$, see experimental details in **Table 1** and protocol in **SI, part 2.b**). The polymerization was stopped at different monomer conversions (corresponding to $DP_{n/arm}$ PDAAm of 12, 21, 33 and 49, see **Table 1**) and the samples were analyzed by SEC and LS. SEC analyses (**Figure S4**) showed the expected shift of the mass distributions with increasing monomer conversion corroborating the formation of block copolymers. ¹H NMR-monitoring of the polymerization in similar conditions (70 °C, 10 wt% of DAAM, $[DAAM]_0/[C_{12}\text{-macroRAFT}]_0 = 100$) indicated the complete consumption of the C₁₂-macroRAFT agent at low conversions (conv. < 5% see **Figure S5A** and corresponding comment in the **SI**) which indicated that a triblock copolymer was indeed formed at very low monomer conversion.

Table 1. Kinetic monitoring of the formation of BAB triblock copolymers: unstirred dispersion polymerizations of DAAM in D₂O in the presence of the symmetric C₁₂-macroRAFT agent at $\alpha = 28\%$ and $[C_{12}\text{-macroRAFT}]_0 = 70 \text{ g L}^{-1}$ ^a

Entry	conv. ^b (%)	C(polym) (g L ⁻¹)	$DP_{n/arm}^c$ PDAAm	$M_{n,th}^c$ (kg mol ⁻¹)	$M_{n,PMMA}^d$ (kg mol ⁻¹)	$\bar{D}^d$	Phase sep.
C ₁₂ -12-28%	25	105	12	12.9	9.3	1.20	no
C ₁₂ -21-28%	42	128	21	15.7	11.1	1.24	no
C ₁₂ -33-28%	66	160	33	19.8	13.1	1.21	no
C ₁₂ -49-28%*	97	193	49	25.3	21.5	1.34	no

^a Polymerization conditions: T = 70 °C, $[DAAM]_0 = 10 \text{ wt\%}$, α (BA) = 28%, $M_{n,NMR}$ (C₁₂-PDMAc-*tris*) = 8.7 kg mol⁻¹, $[C_{12}\text{-macroRAFT}]_0 = 70 \text{ g L}^{-1} = 8 \text{ mmol L}^{-1}$, $[ACPA]_0 = 0.9 \text{ mmol L}^{-1}$. ^b Monomer conversion determined by ¹H NMR. ^c Theoretical number-average degree of polymerization, $DP_{n/arm}$ PDAAm, and number-average molar mass, $M_{n,th}$, calculated using the experimental conversion and using $M_{n,NMR}$ for the PDMAc block (**Table S1**). ^d Number-average molar mass and dispersity, $\bar{D}$, determined by SEC in DMF (+ LiBr 1g L⁻¹) with a PMMA calibration. *For experiment C₁₂-49-28% the polymerization was conducted in water without stirring according to the protocol described in our previous work.¹³ Note that the sample names C₁₂-X-28% stands for a block copolymers of $DP_{n/arm}$ PDAAm = X synthesized at $\alpha = 28\%$ with C₁₂-macroRAFT agent.

Each sample was studied by LS following the same procedure: the concentrated sample withdrawn from the PISA reaction was diluted to different concentrations before measurement. For $DP_{n/arm}$ PDAAm = 12 (25 % monomer conversion), the values of M_{app} and $R_{h,app}$ at low concentration were only slightly higher than those determined for the macroRAFT agent and similar N_{agg} were determined (see **Figure S6**). This suggests that similar bridged flower-like micelles were obtained ($M_{app} = 200 \text{ kg mol}^{-1}$ corresponding to $N_{agg} \sim 13$, $R_{h,app} = 10 \text{ nm}$ which was still smaller than $R_{h,max} = 0.25 \times DP_{n/arm} = 13 \text{ nm}$), see **Figure 2**. It was not possible to investigate the sample at $C \geq 15 \text{ g L}^{-1}$ by LS, because the solutions were turbid in the conditions of the LS analysis ($[NaCl] = 0.01 \text{ mol L}^{-1}$ added before LS measurement), suggesting phase separation due to strong attraction (bridging of flower-like micelles). Note that no turbidity was observed in the polymerization medium for such concentration. This shows that the addition of a small amount of salt was responsible for the turbidity; probably because the presence of salt favors phase separation²⁰ as mentioned in the introduction (see experimental part and **Figure S7** for details). At higher monomer conversions, when the length of the hydrophobic block increased to $DP_{n/arm}$ PDAAm = 21 or 33, dilution of the concentrated solutions of the BAB copolymers to $C < 5 \text{ g L}^{-1}$ produced much larger objects with a M_{app} of $\sim 3.0 \times 10^3 \text{ kg mol}^{-1}$ (corresponding to $N_{agg} \sim 130$) and a $R_{h,app}$ of $\sim$

35 nm (see **Figure 2**). These large values, and especially the fact that $R_{h,app}$ exceeded by far the maximum length expected for fully stretched blocks ($R_{h,max} = 0.25 \times DP_{n/arm} \sim 15$ nm) suggested that dilution of the copolymers did not yield individual flower-like micelles but tiny aggregates of flower-like micelles. The fact that the aggregates of flower-like micelles were not disassembled into individual flower-like micelles even at 0.2 g L^{-1} hinted at the formation of frozen structures unable to reorganize upon dilution. The final sample obtained at almost quantitative monomer conversion (97%) with the longest PDAAm B-blocks ($DP_{n/arm \text{ PDAAm}} = 49$) was a macroscopic gel that could not be dispersed in water afterwards, confirming that a non-dynamic percolated network was obtained for this final DP_n .

From this series of experiments, we concluded that the PISA polymerization of DAAm with $[C_{12}\text{-macroRAFT}] \sim 70 \text{ g L}^{-1}$ and $\alpha = 28\%$ starts in a percolated network of bridged spherical flower-like micelles (see **Scheme 3**). Initially, the exchange of B blocks (which are only C_{12} alkyl chains at 0% conversion) between hydrophobic cores was quite dynamic. However, with the increase of the length of the B blocks during polymerization the exchange rate slowed down. Indeed, the B blocks still obviously exchanged rapidly between hydrophobic cores (at least within the time scale of the dilution of the samples, < 30 min) for $DP_{n/arm \text{ PDAAm}} = 12$, whereas they were certainly frozen for $DP_{n/arm \text{ PDAAm}} = 49$. The transition between a dynamic and a frozen network probably occurred between $DP_{n/arm \text{ PDAAm}} = 12$ and 21 according to the LS results.[‡] This behavior explains the differences between the syntheses performed with or without stirring: the network which was present from the beginning of the reaction remained intact throughout the reaction in the absence of stirring; on the contrary, stirring broke the network into finite size aggregates of flower-like micelles and the latter could no more reconnect into a macroscopic gel once the exchange of B blocks was too slow (see **Scheme 3**). This latter phenomenon explains why cryo-TEM and LS analyses on representative BAB dispersions synthesized with stirring (see **Table S2**, entry C) still revealed the formation of tiny individual particles, which are most probably small aggregates of a few connected flower-like micelles (see **Scheme 3**, **Figure S8** and comments to the figure in the SI).

[‡] We note that the conversion is not total for these DP_n . Residual monomer may swell the hydrophobic core and thereby impact the exchange dynamics of the B blocks. This would shift the DP_n -range where a transition between fast and slow exchange occurs towards higher DP_n values. However, the qualitative conclusion that increasing monomer conversion slows down the exchange dynamics remains valid.

Scheme 3. Schematic representation of the aggregates/network formed with and without stirring during the synthesis of BAB copolymers by PISA.

2. Parameters influencing phase separation and their consequence on polymerization control

In the previous section, we showed that PISA starts in a percolating transient network of C₁₂-macroRAFT agent that gradually becomes frozen during polymerization. In some cases (see for example sample E on **Figure 1** at [C₁₂-macroRAFT]₀ = 33 g L⁻¹ and α = 28%), this was accompanied with a phase separation. We therefore explored in more details the influence of macroRAFT agent concentration, α and the length of the alkyl chain on phase separation in order to understand its origin and determine its role on the control of the polymerization. We stress that understanding these aspects is of outmost importance for the efficient design of BAB triblock copolymers by PISA, both in terms of colloidal stability and control of the polymerization.

2.1. Role of the macroRAFT agent concentration and ionization degree (α)

The role of the macroRAFT agent concentration was investigated first. In an additional series of unstirred experiments, we performed polymerizations at 70 °C, at constant monomer concentration

(10 wt% of DAAM), varying the initial concentration of macroRAFT agent (see **Table S2**, entry D, E and F) but keeping constant the ionization degree of the central benzoic acid moiety ($\alpha = 28\%$) and the final monomer conversion ($\sim 95\%$). These conditions corresponded to targeted $DP_{n/arm}$ PDAAM of 49, 79 and 140 (for $[C_{12}\text{-macroRAFT}]_0 \sim 67, 33$ and 20 g L^{-1} , respectively) at full monomer conversion. Macroscopically, an obvious phase separation into a non-dispersible gel phase and a liquid phase was observed with the two lowest macroRAFT agent concentrations (see $[C_{12}\text{-macroRAFT}]_0 \sim 33$ and 20 g L^{-1} in **Figure S9i**). The volume fraction of the liquid phase increased with decreasing macroRAFT concentration and therefore with increasing length of the PDAAM arms. In contrast, it has been reported in the literature that AB³⁵ and ABA³⁶ PDMAc/PDAAM block copolymers prepared by PISA in water (with block lengths comparable to those in our study) led to stable dispersions. Furthermore, Zhang *et al.* compared AB diblock copolymers with BAB triblock copolymers prepared by PISA in a mixture of alcohol and water: whereas the former resulted in stable dispersions, the BAB system resulted in phase separation.⁹ This corroborates that the observed phase separation for BAB copolymer dispersions is certainly caused by their specific associative behavior.¹⁸ Phase separation indeed increases the proportion of intermicellar bridges over intramicellar loops in the polymer-rich phase; which increases the conformational entropy of the B blocks and is favorable. The higher the propensity of the BAB triblocks to form bridges rather than loops (stronger attraction), the more phase separation will be favored. These results indicated qualitatively that the attraction caused by the bridging of micelles was sufficient at $\alpha = 28\%$ to trigger phase separation for $[C_{12}\text{-macroRAFT}]_0 \leq 33 \text{ g L}^{-1}$, corresponding - in the experimental conditions - to a $DP_{n/arm}$ PDAAM ≥ 79 . At the molecular level, SEC indicated that the polymerization control was maintained in spite of the obvious phase separation for $DP_{n/arm}$ PDAAM = 79, but not for $DP_{n/arm}$ PDAAM = 140 corresponding to the lowest macroRAFT agent concentration (**Figure S9ii**). In contrast, we observed that, the synthesis of copolymers with similar $DP_{n/arm}$ performed under stirring resulted in a controlled polymerization with no phase separation (see **Table S2**, entry A, B and C and **Figure S10**).

Next, the role of α was investigated. In our former study using stirring,¹³ we had observed that colloidal stability and control of the polymerization were lost when the polymerizations were conducted either at a low pH (corresponding to $\alpha < 7\%$) or with a neutral macroRAFT agent (absence of a central benzoic acid moiety in the RAFT agent). As described above, unstirred polymerizations performed with a monomer concentration of 10 wt% and $[C_{12}\text{-macroRAFT}]_0 = 70$

g L^{-1} (corresponding to $DP_{\text{n/arm PDAAm}} = 49$ at full conversion) were well-controlled and colloidally stable at $\alpha = 28\%$ (see **Table 1** and **Figure S4**). As expected, we obtained similar results when the copolymers were synthesized at a higher ionization degree ($\alpha = 45\%$) (see **Table S3** and **Figure S11A**).

To understand the role of α , its impact on the balance between attraction and repulsion was investigated qualitatively by LS on the C_{12} -macroRAFT agent, at $\alpha = 7\%$, 28% and 78% . As shown in **Figure 3**, at low concentrations, varying α hardly affected N_{agg} of the individual flower-like micelles. However, between 5 and 20 g L^{-1} , M_{app} increased much more strongly with concentration for lower α . An increase of M_{app} with increasing polymer concentration C indicates that attraction (due to bridging of flower-like micelles) dominates the behavior of the self-assemblies; the steeper the slope, the stronger the effect. Qualitatively, in the **Figure 3**, the slope in the range 5 to 20 g L^{-1} was greater for lower degree of ionizations α , indicating that attraction was strongly increased at lower α . This was certainly the result of a lower amount of charges at lower α , causing less repulsion.²⁰ It must also be mentioned that the presence of salt - needed for the LS analysis - had a huge impact on the colloidal stability. Indeed, the macroRAFT agent prepared at $\alpha = 7\%$ could not be analyzed by LS at concentrations higher than 16 g L^{-1} due to the formation of a precipitate, whereas solutions of the C_{12} -macroRAFT agent at $\alpha \geq 28\%$ remained transparent up to 70 g L^{-1} at the same NaCl concentration.

Figure 3. Evolution of the apparent molar mass M_{app} of C_{12} -PDMAc-*bis* at $\alpha = 7\%$ ($\blacktriangle$), 28% ($\bullet$) and 78% ($\times$) and of C_4 -PDMAc at $\alpha = 7\%$ ($\triangle$) and 28% ($\circ$) as a function of the polymer concentration obtained by light scattering at 20°C in water. The solid lines are guide for the eyes.

Dotted lines indicate precipitation of the sample caused by the addition of salt for LS analysis ($[\text{NaCl}] = 0.01 \text{ mol L}^{-1}$).

In contrast to the C_{12} -macroRAFT agent, BAB triblock copolymers with $DP_{\text{n/arm PDAAm}} \sim 10$ were turbid in the presence of $[\text{NaCl}] = 0.01 \text{ mol L}^{-1}$ above respectively $C = 10 \text{ g L}^{-1}$ for $\alpha = 28\%$ (sample C_{12} -12-28%) and $C = 20 \text{ g L}^{-1}$ for $\alpha = 45\%$ (sample C_{12} -10-45%) (**Figure S11B**). These observations 1) qualitatively suggest that attraction due to bridging of flower-like micelles increased with the growth of PDAAm outer blocks from the macroRAFT and 2) confirmed that higher α decreased attraction even for short BAB triblock copolymers. Polymers with $DP_{\text{n/arm PDAAm}} = 21$ or 33 were not under thermodynamic equilibrium as explained in the previous part and were therefore not investigated by LS as a function of α .

The results presented in this section confirmed that the loss of colloidal stability during PISA was due to attraction, *i.e* the bridging of flower-like micelles, which resulted in phase separation. This phenomenon was favored by decreasing α of the macroRAFT agent unit, increasing the ionic strength, decreasing the macroRAFT agent concentration and, thereby increasing the length of the PDAAm blocks. The loss of colloidal stability induced by bridging seemed sufficiently strong at $\alpha = 7\%$ to result in a loss of polymerization control. This might be due to a difference in local concentration of the macroRAFT agent combined with a low $[\text{C}_{12}\text{-macroRAFT}]_0/[\text{ACPA}]_0$ ratio leading to the generation of several populations of polymer chains. For $\alpha = 28\%$, control was lost without stirring at 20 g L^{-1} , but was preserved under stirring, suggesting that the heterogeneity of the reaction medium was not critical at this α and could be alleviated by stirring.

2.2. Comparison of the C_4 and C_{12} -macroRAFT agents

In our previous study,¹³ we observed that BAB polymers synthesized in similar conditions using telechelic macroRAFT agents end-capped by either dodecyl or butyl alkyl chains (named C_{12} -macroRAFT agent *vs.* C_4 -macroRAFT agent, see **Scheme 2**) led to significant differences in colloidal stability and polymerization control. These differences were confirmed in the present study, where we performed unstirred PISA experiments with both macroRAFT agents. For $[\text{macroRAFT}]_0 = 70 \text{ g L}^{-1}$ at 70°C and $\alpha = 28\%$ or 45% , the polymerizations in presence of C_4 -

macroRAFT led systematically to phase separation whereas the polymerization medium remained homogeneous when the C_{12} -macroRAFT was used in the same conditions. The results are summarized in **Figure 4** (for details see **Tables 1, S3, S4** and **Figure S13**). At $\alpha = 45\%$ phase separation was observed for $DP_{n/arm} PDAAm \geq 32$ (and persisted when the sample was cooled to room temperature). At lower α (28%), this phase separation starts for lower monomer conversions ($< 20\%$, corresponding to $DP_{n/arm} PDAAm = 10$), and becomes more pronounced with increasing conversion (see **Figure S13B and C**). Remarkably, the phase separation observed during PISA for short DP_n ($DP_n = 10$, sample C_4 -10-28% in **Table S4**) was thermo-reversible (phase-separated at 70°C and monophasic at room temperature).

Figure 4. Summary of the macroscopic aspects of block copolymers dispersions synthesized with 10 wt% of DAAM without stirring either with C_{12} or C_4 -macroRAFT agent with $[\text{macroRAFT}]_0 \sim 70 \text{ g L}^{-1}$ at $\alpha = 28$ and 45% . *Thermo-reversible phase separation observed: homogeneous sample at 25°C that phase-separates reversibly at 70°C .

In order to understand the important divergences observed for the C_4 - and the C_{12} - series, potential differences in the macromolecular chemical structure were first examined. For both series, the

copolymers obtained by PISA were characterized by ^1H NMR and SEC at different monomer conversions, corresponding to $DP_{\text{n/arm PDAAm}}$ of ~ 10 , 20 and 30 (for details see **Table 1**, **S3**, **S4**). Comparable $DP_{\text{n/arm PDAAm}}$ were reached for both series, and SEC analyses revealed that the quality of control was similar with the C_4 and the C_{12} macroRAFT, at least up to $DP_{\text{n/arm PDAAm}} \sim 30$ (see SEC in **Figure S4**, **Figure S11A** and **Figure S12**). Moreover, ^1H NMR analyses indicated that for both macroRAFT agents, the primary transfer reaction occurred on both trithiocarbonates of the macroRAFT agents quantitatively at very low monomer conversion ($< 5\%$), leading to a bilateral polymer growth and the formation of triblock copolymers from the very beginning of the polymerization (see **Figure S5** and explanatory text in the **SI**). In view of these results, the observed macroscopic differences for PISA series C_4 and the C_{12} could not be explained by strong differences in the macromolecular chemical structure of the BAB triblock copolymers.

Potential differences in terms of supramolecular self-assembly were then investigated by LS experiments. At low concentration (**Figure 3**), an M_{app} around 10 kg mol^{-1} was determined for the C_4 -macroRAFT (independent of α), which was very close to the molar mass of the macroRAFT agent determined by ^1H NMR ($M_{\text{n,NMR}} = 8.9 \text{ kg mol}^{-1}$). These experiments revealed that, in contrast to the C_{12} -macroRAFT agent, the C_4 -macroRAFT agent did not self-assemble in water. This conclusion was also confirmed by the difference in viscosity observed for both macroRAFT agents: a distinct increase of viscosity with polymer concentration could be observed for aqueous solutions of C_{12} -macroRAFT agent, whereas no significant increase of viscosity could be observed for C_4 -macroRAFT agent, at least not by the naked eye. This decrease of association with the decrease of the alkyl length is consistent with the literature on telechelic homopolymers decorated on both sides with alkyl chains.²¹ These results show that, unlike the C_{12} -system, the RAFT dispersion polymerization of DAAM with the C_4 -macroRAFT agent does not start in a percolating transient network, but in a non-associative system.

Block copolymers of $DP_{\text{n/arm PDAAm}} \sim 10$ synthesized with both macroRAFT agents in comparable conditions ($\alpha = 45\%$, C_4 -macroRAFT: entry $\text{C}_4\text{-11-45\%}$ in **Table S4**, and C_{12} -macroRAFT: entry $\text{C}_{12}\text{-10-45\%}$ in **Table S3**) were then investigated by LS analyses in order to understand how the assembly evolved upon the growth of the PDAAm side chains (see **Figure 5**). At low concentrations ($C < 10 \text{ g L}^{-1}$), M_{app} was significantly smaller for the C_4 series ($\text{C}_4\text{-11-45\%}$) than for the C_{12} series ($\text{C}_{12}\text{-10-45\%}$); but a progressive increase of aggregation with the concentration

was observed for the C₄-system. M_{app} and $R_{h,app}$ became similar for both polymers at $C = 20 \text{ g L}^{-1}$, implying that they both formed flower-like micelles with a similar extent of aggregation in spite of a seemingly weaker cooperativity of the self-assembly for C₄-11-45%. Unfortunately, it was not possible to follow the evolution of M_{app} at concentrations higher than 20-30 g L^{-1} (which could have given clearer information on attraction) because the addition of 0.01 M NaCl for the LS studies caused significant turbidity. Polymers with $DP_{n/arm \text{ PDAAm}} = 20$ or 30 were not under thermodynamic equilibrium, as explained in the previous parts, and were therefore not investigated either. Overall, the LS experiments indicated that the initial state of the macroRAFT agents was very different for the C₄ and the C₁₂ macroRAFT at $\alpha = 45\%$, but that flower-like micelles with similar extent of aggregation were obtained as soon as the PDAAm arms reached a $DP_{n/arm \text{ PDAAm}} \sim 10$.

Figure 5. Evolution of the apparent molar mass M_{app} as a function of the polymer concentration obtained by light scattering at 20 °C in water for block copolymers with $DP_{n/arm} \sim 10$, prepared by dispersion polymerization of DAAM in D₂O at 70 °C with the C₁₂-macroRAFT agent (see **Table S3**) or C₄-macroRAFT agent (see **Table S4**; the scattered intensity was too low below 1 g L^{-1} to obtain reliable data) at $\alpha = 45\%$ without stirring. The solid lines are guide for the eyes. Dotted lines indicate precipitation of the sample caused by the addition of salt for LS analysis ($[\text{NaCl}] = 0.01 \text{ mol L}^{-1}$).

To summarize, the results suggest that the growth of the B blocks occurred symmetrically on both sides of the macroRAFT agents for both series of polymers. The C₄-macroRAFT was not self-

assembled initially, contrary to the C₁₂ one, but flower-like micelles of similar extent of aggregation and dimensions were observed for both series of BAB triblock copolymers with $DP_{n/arm} PDA_{Am} \sim 10$. This implied that both series of polymers form a percolated network of bridged flower-like micelles starting from $DP_{n/arm} PDA_{Am} = 10$. From this, it was deduced that the phase separation observed at sufficiently high $DP_{n/arm} PDA_{Am}$ for the C₄-series (at both $\alpha = 28\%$ or 45% , see **Figure 4**) was caused by a strong attraction between bridged micelles. In contrast, for the C₁₂-series no phase separation was observed in these conditions ($[macroRAFT]_0 = 70 \text{ g L}^{-1}$), but only for lower macroRAFT concentrations, *i.e.* for higher $DP_{n/arm} PDA_{Am}$, and at low α (28%), as described in part 2.1. Finally, we explain the thermo-sensitive phase-separation observed for C₄-10-28% (no phase separation at 20°C , phase separation at 70°C) by the thermo-sensitive behavior of the PDA_{Am} blocks which has been reported before in the litterature.^{35–37}

The fact that obvious differences between the C₄ and the C₁₂-series were observed in terms of phase separation for $DP_{n/arm} PDA_{Am} \sim 10$ (see **Figure 4**) implies that these differences must have a thermodynamic origin. Indeed, the light scattering experiments presented in part 1 for sample C₁₂-10-28% and the fact that the thermally-induced phase separation observed for sample C₄-10-28% is reversible indicated that both series of polymers with a $DP_{n/arm} PDA_{Am} \sim 10$ are at thermodynamic equilibrium. From the observations we must conclude that the attraction, *i.e.* the propensity to form bridges rather than loops, was stronger for the C₄-series than for the C₁₂-series. One potential origin for the difference of attraction in both series might be the presence of a small but distinct amount of monofunctional polymers (AB diblocks) in the BAB triblocks. It has been reported in the literature that a small fraction of AB copolymers in BAB triblock copolymers reduces attraction.²¹ The formation of a small quantity of AB diblock copolymers is actually inherent to the RAFT mechanism, but as C₄ and C₁₂ series were performed in identical conditions, we may reasonably assume that the observed differences should result from monofunctional RAFT agent present in different quantities in the C₄ and C₁₂ molecular RAFT agent, (C₁₂-TTC-PDMAc)₂-BA and (C₄-TTC-PDMAc)₂-BA). Indeed, the SEC chromatograms of the C₁₂-series (see **Figures S4** and **S11**) do exhibit a more pronounced shoulder in the lower molar mass region than those prepared from the C₄-macroRAFT (**Figure S12**). This suggests the presence of a slightly larger amount of AB diblocks in the C₁₂ than in the C₄ series and could explain a somewhat lower attraction for the former. We also note that for $DP_{n/arm} PDA_{Am} \geq 20$, the exchange of B blocks between micelles becomes extremely slow and the assemblies become frozen. The differences between the C₄ and

the C₁₂-series may therefore not only have a thermodynamic but also a kinetic origin: it is possible that the C₄-series, which starts from an unassembled state for $DP_{n/arm\ PDAAm} = 0$ (C₄-macroRAFT agent), does not have time to rearrange in a homogeneous transient network during polymerization before the exchange rate of PDAAm blocks becomes too slow. This would then result in a different organization compared to the C₁₂-series and potentially in a different ratio of bridges over loops favoring phase separation in the C₄ series.

Conclusion

In this paper, we investigated by light scattering and macroscopic observations the behavior of BAB triblock copolymers prepared by PISA in dispersion in order to understand the main features of these complex polymerizations.

The bifunctional C₁₂-macroRAFT agent was clearly shown to self-assemble into a dynamic percolated network of flower-like micelles at the beginning of the polymerization. Upon polymerization, the growth of the insoluble PDAAm blocks caused a decrease of the exchange rate of the bridges between the micelles until the network eventually became frozen. This occurred for a $DP_{n/arm\ PDAAm}$ ranging between 10 and 20. As a consequence, synthesis of these BAB triblocks under stirring resulted in an irreversible breaking of the network into tiny particles consisting of a few bridged flower-like micelles, whereas synthesis without stirring afforded a non-flowing gel. The bifunctional C₄-macroRAFT agent was initially not self-assembled, but also formed a transient network of bridged flower-like micelles upon growth of the PDAAm blocks.

A second consequence of the synthesis of BAB triblock copolymers instead of AB diblock copolymers was the fact that bridging between flower-like micelles could result into macroscopic phase separation because of the formation of intermicellar bridges instead of intramicellar loops. This loss of colloidal stability could be accompanied with a loss of polymerization control. Attraction caused by bridging of flower-like micelles, resulting eventually in phase separation, was favored by increase of ionic strength, growth of the PDAAm arms, decrease of macroRAFT agent concentration or ionization degree (α). This study explains why it was not possible to obtain colloidally stable dispersions and controlled polymerizations using a neutral bifunctional macroRAFT agent.¹³ It was also observed that replacement of the C₁₂ alkyl chains by C₄ ones

favored phase separation. This might be due to the presence of a smaller amount of monofunctional RAFT agents in the C₄-series limiting the quantity of AB diblock copolymers present in the sample. In contrast, the absence of phase separation for the C₁₂-series could be explained by the presence of a larger amount of AB diblock copolymers, limiting attraction. We believe that this study gives a general picture of the mechanism of assembly of BAB triblock copolymers during PISA and proposes clues to achieve colloidal stability and control during the polymerization.

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Keywords: PISA, aqueous RAFT dispersion polymerization, BAB triblock copolymers, flower-like micelles, transient network, light scattering

References

- 1 W. Wang, W. Lu, A. Goodwin, H. Wang, P. Yin, N.-G. Kang, K. Hong and J. W. Mays, *Progress in Polymer Science*, 2019, **95**, 1–31.
- 2 P. Tyagi, A. Deratani, D. Bouyer, D. Cot, V. Gence, M. Barboiu, T. N. T. Phan, D. Bertin, D. Gigmes and D. Quemener, *Angewandte Chemie International Edition*, 2012, **51**, 7166–7170.
- 3 B. Baroli, *Journal of Pharmaceutical Sciences*, 2007, **96**, 2197–2223.
- 4 C. Tsitsilianis, *Soft Matter*, 2010, **6**, 2372.
- 5 B. Charleux, G. Delaittre, J. Rieger and F. D’Agosto, *Macromolecules*, 2012, **45**, 6753–6765.
- 6 J.-T. Sun, C.-Y. Hong and C.-Y. Pan, *Polymer Chemistry*, 2013, **4**, 873–881.
- 7 J. Rieger, *Macromolecular Rapid Communications*, 2015, **36**, 1458–1471.
- 8 M. J. Derry, L. A. Fielding and S. P. Armes, *Progress in Polymer Science*, 2016, **52**, 1–18.
- 9 C. Gao, S. Li, Q. Li, P. Shi, S. A. Shah and W. Zhang, *Polym. Chem.*, 2014, **5**, 6957–6966.
- 10 C. Gao, J. Wu, H. Zhou, Y. Qu, B. Li and W. Zhang, *Macromolecules*, 2016, **49**, 4490–4500.
- 11 Y. Qu, S. Wang, H. Khan, C. Gao, H. Zhou and W. Zhang, *Polymer Chemistry*, 2016, **7**, 1953–1962.
- 12 W. Wang, C. Gao, Y. Qu, Z. Song and W. Zhang, *Macromolecules*, 2016, **49**, 2772–2781.
- 13 P. Biais, P. Beaunier, F. Stoffelbach and J. Rieger, *Polymer Chemistry*, 2018, **9**, 4483–4491.
- 14 C. Grazon, J. Rieger, N. Sanson and B. Charleux, *Soft Matter*, 2011, **7**, 3482.
- 15 W. Zhang, F. D’Agosto, O. Boyron, J. Rieger and B. Charleux, *Macromolecules*, 2012, **45**, 4075–4084.
- 16 E. R. Jones, O. O. Mykhaylyk, M. Semsarilar, M. Boerakker, P. Wyman and S. P. Armes, *Macromolecules*, 2016, **49**, 172–181.
- 17 F. Brunel, J. Lesage de la Haye, M. Lansalot and F. D’Agosto, *J. Phys. Chem. B*, 2019, **123**, 6609–6617.
- 18 C. Chassenieux, T. Nicolai and L. Benyahia, *Current Opinion in Colloid & Interface Science*, 2011, **16**, 18–26.
- 19 T. Annable, R. Buscall, R. Ettelaie and D. Whittlestone, *Journal of Rheology*, 1993, **37**, 695–726.

- 20 C. Charbonneau, M. M. De Souza Lima, C. Chassenieux, O. Colombani and T. Nicolai, *Physical Chemistry Chemical Physics*, 2013, **15**, 3955.
- 21 F. Laflèche, T. Nicolai, D. Durand, Y. Gnanou and D. Taton, *Macromolecules*, 2003, **36**, 1341–1348.
- 22 A. N. Semenov, J.-F. Joanny and A. R. Khokhlov, *Macromolecules*, 1995, **28**, 1066–1075.
- 23 Q. T. Pham, W. B. Russel, J. C. Thibeault and W. Lau, *Macromolecules*, 1999, **32**, 2996–3005.
- 24 F. Laflèche, D. Durand and T. Nicolai, *Macromolecules*, 2003, **36**, 1331–1340.
- 25 T. Nicolai, O. Colombani and C. Chassenieux, *Soft Matter*, 2010, **6**, 3111.
- 26 A. Halperin and S. Alexander, *Macromolecules*, 1989, **22**, 2403–2412.
- 27 J. Rieger, F. Stoffelbach, C. Bui, D. Alaimo, C. Jérôme and B. Charleux, *Macromolecules*, 2008, **41**, 4065–4068.
- 28 G. G. Cameron, *Polymer International*, 1995, **38**, 106–106.
- 29 S. Michielsen, in *Polymer Handbook*, John Wiley & Sons, Inc., New York, 4. ed., 1999, p. VII/547–627.
- 30 W. Brown, *Dynamic Light Scattering: The Method and Some Applications*, Oxford University Press, New York, 1993.
- 31 J. P. Patterson, M. P. Robin, C. Chassenieux, O. Colombani and R. K. O'Reilly, *Chem. Soc. Rev.*, 2014, **43**, 2412–2425.
- 32 J. Lesage de La Haye, J.-M. Guigner, E. Marceau, L. Ruhlmann, B. Hasenknopf, E. Lacôte and J. Rieger, *Chemistry – A European Journal*, 2015, **21**, 2948–2953.
- 33 J. Dubochet, M. Adrian, J.-J. Chang, J.-C. Homo, J. Lepault, A. W. McDowell and P. Schultz, *Quarterly Reviews of Biophysics*, 1988, **21**, 129.
- 34 C. Chassenieux, T. Nicolai and D. Durand, *Macromolecules*, 1997, **30**, 4952–4958.
- 35 S. J. Byard, M. Williams, B. E. McKenzie, A. Blanz and S. P. Armes, *Macromolecules*, 2017, **50**, 1482–1493.
- 36 G. Mellot, P. Beaunier, J.-M. Guigner, L. Bouteiller, J. Rieger and F. Stoffelbach, *Macromolecular Rapid Communications*, 2018, **40**, 1800315.
- 37 X. Wang, J. Zhou, X. Lv, B. Zhang and Z. An, *Macromolecules*, 2017, **50**, 7222–7232.