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Multifaceted roles of efflux proteins in prokaryotes

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Abstract

Multidrug resistance is a globally increasing problem that has become an alarming threat to antibiotic therapy. A principal resistance mechanism that prokaryotic organisms have evolved is active multidrug efflux, whereby antibiotics and other xenobiotics are exported to the external environment by transport proteins in the cell membrane. Such proteins are especially abundant in Gram-negative bacteria that are responsible for a large proportion of hospital-acquired infections. Based on amino acid sequence similarity, substrate specificity and the energy source used for exporting substrates, prokaryotic organisms contain seven major families of distinct multidrug efflux transporters (ABC, MFS, RND, SMR, MATE, AbgT, PACE). Efflux proteins also have roles in biofilm formation, quorum sensing, resistance to heavy metals and biocides, cell homeostasis and in bacterial pathogenicity and virulence. Prokaryotic efflux proteins are therefore highly important targets for advancements in antibiotic therapy and for ongoing experimental characterisation of their structures, functions, molecular mechanisms and regulation.

Key words : active transport, biofilms, efflux proteins, Gram-negative bacteria, multidrug resistance

INTRODUCTION

All over the world, the rate of multidrug resistance is dramatically increasing and the mechanisms of resistance are varied and complicated. Diverse mechanisms contribute to intrinsic, acquired and phenotypic resistance to antimicrobial compounds. Active multidrug efflux systems are one of the major mechanisms of bacterial resistance to drugs and are an alarming threat to antibiotic therapy.^[1-11] Gram-negative bacteria (e.g. *Acinetobacter baumannii*, *Escherichia coli*, *Klebsiella pneumonia*, *Pseudomonas aeruginosa*) are commonly intrinsically more resistant to many antibiotics and biocides as a result of their cell structure and the activity of multidrug efflux pumps^[8,12-15] and they are responsible for over 30% of hospital-acquired infections.^[16-20] Efflux pumps are proteinaceous transporters that can extrude antibiotics and other xenobiotics from the cytoplasm or surrounding membranes of cells to the outside environment. They are found in all microorganisms, including both Gram-negative and Gram-positive bacteria^[21-24] and also in eukaryotic organisms.^[25-32] The active transport mechanism of efflux pumps is driven by an energy source. In this respect, primary active transporters exploit ATP hydrolysis, whilst secondary active transporters use the electrochemical potential difference created by proton pumping or sodium ions. These transporters may be highly specialised for one compound or may be highly promiscuous, transporting a broad range of structurally dissimilar substrates.

FAMILIES OF MULTIDRUG EFFLUX PROTEIN

In the prokaryotic kingdom there are currently seven major families of distinct bacterial multidrug efflux transporters: adenosine triphosphate (ATP)-binding cassette (ABC) superfamily, major facilitator superfamily (MFS), resistance-nodulation-division (RND) family, small multidrug resistance (SMR) family, multidrug and toxic compound extrusion (MATE)

family, p-aminobenzoyl-glutamate transporter (AbgT) family, proteobacterial antimicrobial compound efflux (PACE) family (Figure 1). These families are classified on the basis of their amino acid sequence similarity, substrate specificity and the energy source used to export their substrates. Protein structural organisation in the large majority of the families, including the number of transmembrane helices (TMH), has been confirmed by high resolution structure determination for at least one member.

ABC superfamily proteins^[33-36] are the only primary transporters, using energy from ATP hydrolysis to expel substrates. They are comprised of a transmembrane domain that forms the extrusion pathway and a cytoplasmic nucleotide-binding domain that binds and hydrolyses ATP. A homodimeric structural organisation (2 x 6TMH) in bacterial ABC proteins is represented by multidrug efflux transporter Sav1866 from *Staphylococcus aureus* (PDB 2HYD).^[37]

All of the other families are secondary transporters that use the proton or sodium gradient as the energy source in an antiport manner. The MFS proteins^[38-43] have 12 or 14 TMH and are structurally described by *E. coli* multidrug resistance transporter MdfA (PDB 4ZOW), which confers resistance to chloramphenicol.^[44] MFS transporters are proton-driven and typically contain a 12TMH core consisting of two pseudo-symmetrical six-helix domains. A central cavity between these two domains forms the substrate-transport path and the protein undergoes a rocker-switch mechanism that involves cycling between outward-facing and inward-facing conformations. RND family proteins^[45-49] are proton-driven and function in a tripartite assembly with an outer membrane protein and a periplasmic adapter protein that connects them together. Structural organisation in RND proteins is represented by AcrB from *E. coli* (PDB 2DRD),^[50-52] which functions in complex with AcrA and TolC. Each AcrB protomer consists of a 12TMH

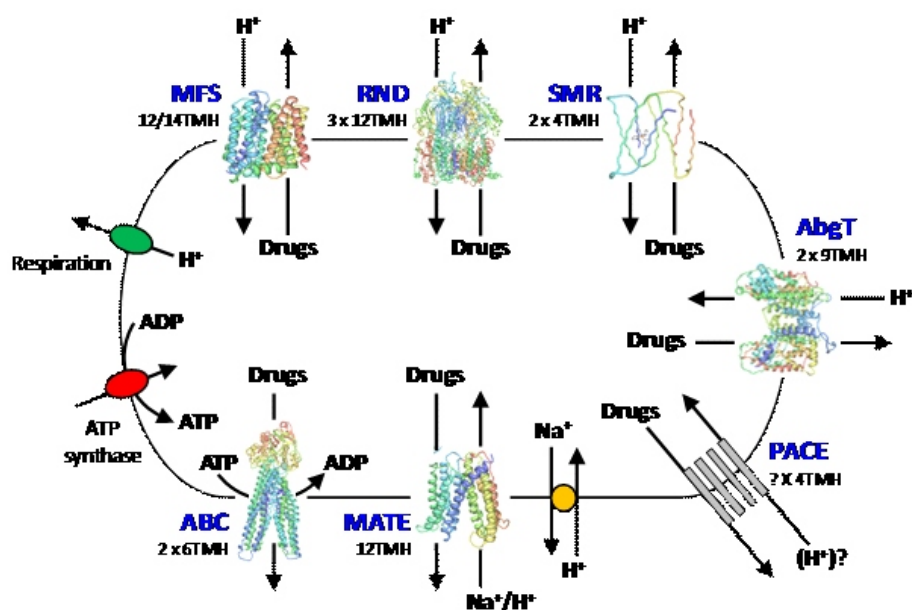


Fig. 1: Schematic illustration for the energisation and structural organisation of seven distinct families of multidrug efflux protein found in bacteria. The large oval represents a bacterial cell. An electrochemical H^+ gradient (proton-motive-force) across the cytoplasmic membrane is generated by respiration (green). Energy from the proton-motive-force drives secondary active transport proteins, such as those in the MFS, RND, SMR and AbgT (super)families. PACE family transporters are also likely to be driven by the H^+ gradient. Sodium/proton antiporters (orange) exploit the proton-motive-force to generate a Na^+ gradient that drives transport by other multidrug efflux proteins, including those in the MATE family. ATP production by ATP-synthase (red) is also driven by the proton-motive-force and ATP is used to drive transport by primary active transporters of the ABC superfamily. The structural organisation in each family is illustrated by a picture of a crystal structure of a representative protein as follows: ABC Sav1866 from *Staphylococcus aureus* (PDB 2HYD); MFS MdfA from *Escherichia coli* (PDB 4ZOW); RND AcrB from *Escherichia coli* (PDB 2DRD); SMR EmrE from *Escherichia coli* (PDB 3B5D); MATE NorM from *Vibrio cholerae* (PDB 3MKT); AbgT MtrF from *Neisseria gonorrhoeae* (PDB 4R11); PACE no structure available;. Structures were drawn using the relevant PDB entry and PDB Workshop 3.9. The number of TMH based on structural characterisation is also given.

transmembrane domain and a headpiece that protrudes into the periplasm. SMR family proteins are proton-driven and highly hydrophobic.^[53-56] They have a homodimeric structural organisation (2 x 4 TMH), described by EmrE from *E. coli* (PDB 3B5D),^[57,58] in which a single membrane-embedded and highly conserved charged residue (Glu14 in TM1) is essential for transport activity. MATE family proteins^[59-61] are driven by either the sodium or proton gradient and their 12 TMH structural organisation is represented by NorM from *Vibrio cholerae* (PDB 3MKT) or by DinF from *Bacillus halodurans* (PDB 4LZ6).^[62-65] The two most recently identified types of bacterial multidrug efflux transporters are from the AbgT family, which confer resistance to sulfonamide antimetabolite drugs^[66-67] and from the PACE family, which confer resistance to the bisbiguanide antiseptic chlorhexidine.^[68-70] AbgT transporters are proton-driven and their homodimeric structural organisation (2 x 9 TMH) has been described by crystal structures of YdaH from *Alcanivorax borkumensis* (PDB 4R0C) and MtrF from *Neisseria gonorrhoeae* (PDB 4R11).^[71-73] PACE family proteins are likely to be proton-driven and they putatively contain 4 TMH that probably function as an oligomer. High-resolution structural information is yet to be obtained for any member of the PACE family.

Simultaneous expression of members from all seven families in the same organism is entirely feasible and a significant challenge for overcoming multidrug resistance.

ABUNDANCE AND MULTIPLE ROLES OF EFFLUX PROTEINS

Up to 30% of all the genes in most sequenced genomes encode membrane proteins.^[74-76] In prokaryotic genomes, up to 50% of these are transport proteins^[77] and a large proportion of these encode efflux proteins, further reflecting their importance. Efflux proteins not only play a key role in drug resistance but also perform many other physiological functions in bacteria.^[9,78-80] Efflux proteins are highly active in bacterial biofilms and they are involved in the formation and maintenance of biofilms.^[81-87] For example, there is a potential role of the AdeFGH efflux pump in the synthesis and transport of autoinducer molecules during biofilm formation of *Acinetobacter baumannii*.^[88] Related to biofilm formation, there is interdependence between efflux proteins and bacterial quorum sensing. Efflux proteins can transport out quorum sensing molecules that can induce expression of genes relating to processes such as biofilm formation and virulence.^[89-93] For example, active efflux influences the potency of quorum sensing inhibitors in *Pseudomonas aeruginosa*.^[94] Efflux proteins also contribute to heavy metal resistance in prokaryotes.^[95,96] This is exemplified by the structurally characterised RND-type *E. coli* CusC(F)BA complex, which exports and confers resistance to copper(I) and silver(I) ions involving a methionine shuttle.^[97-100] There is a developing understanding about co-selection of resistance

mechanisms in bacteria to antibiotics, biocides and metals that includes efflux proteins.^[101-104] Other transport proteins capable of exporting metal ions also contribute to metal ion homeostasis in prokaryotes^[105-114] and this includes members of the ABC, RND, CDF (cation diffusion facilitator),^[115-117] MntX (transporter mediating manganese export)^[118] and CorA (cobalt resistance protein A)^[119-122] transporter families. Efflux proteins also have wider roles in bacterial pathogenicity and virulence to humans, other animals and plants.^[80,123]

CONCLUSION

The high importance of prokaryotic efflux proteins, not least in the global problem of antibiotic resistance, makes them prime targets for drug development and this requires a rigorous ongoing characterisation and understanding of their structures, functions, molecular mechanisms and regulation using genetic approaches and by application of chemical, biochemical, biophysical and computational techniques. Proper high-resolution three-dimensional structure elucidation (using X-ray crystallography, NMR spectroscopy, electron microscopy) in the first instance requires successful gene cloning and amplified expression of the target protein in *E. coli*,^[124-132] followed by production of sufficient quantities of high quality membranes and/or stable and active purified protein. Such studies contribute information that could lead to increased susceptibility to antibiotics (e.g. by inhibition of multidrug efflux pumps), development of new antibiotics or the reversal of bacterial resistance mechanisms.

CONFLICTS OF INTEREST

The authors declare that they have no conflicts of interest.

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REFERENCES

1. Levy SB. Active efflux mechanisms for antibiotic resistance. *Antimicrob. Agents Chemother.* 1992; 36(4):695-703
2. Nikaido H. Prevention of drug access to bacterial targets: permeability barriers and active efflux. *Science* 1994; 264(5157):382-388.
3. Bolhuis H, van Veen HW, Poolman B, Driessen AJ, Konings WN. Mechanisms of multidrug transporters. *FEMS Microbiol. Rev.* 1997; 21(1): 55-84.
4. Nikaido H. Molecular basis of bacterial outer membrane permeability revisited. *Microb. Mol. Biol. Rev.* 2003; 67(4):593-656.
5. Webber MA, Piddock LJ. The importance of efflux pumps in bacterial antibiotic resistance. *J. Antimicrob. Chemother.* 2003; 51(1):9-11.
6. Kaatz GW. Bacterial efflux pump inhibition. *Curr. Opin. Investig. Drugs* 2005; 6(2):191-198.
7. Li XZ, Nikaido H. Efflux-mediated drug resistance in bacteria. *Drugs* 2004; 64(2):159-204.
8. Kumar S, Varela MF. Biochemistry of bacterial multidrug efflux pumps. *Int. J. Mol. Sci.* 2012; 13(4):4484-4495.
9. Sun J, Deng Z, Yan A. Bacterial multidrug efflux pumps: mechanisms, physiology and pharmacological exploitations. *Biochem. Biophys. Res. Commun.* 2014; 453(2):254-267.
10. Spengler G, Kincses A, Gajdacs M, Amaral L. New roads

leading to old destinations: Efflux pumps as targets to reverse multidrug resistance in bacteria. *Molecules* 2017; 22(3) E468.

11. Yılmaz Ç, Özcengiz G. Antibiotics: Pharmacokinetics, toxicity, resistance and multidrug efflux pumps. *Biochem. Pharmacol.* 2017; 133:43-62.
12. Pagès JM, Sandrine AF, Mahamoud A, Bolla JM, Davin-Regli A, Chevalier J, Garnotel E. Efflux pumps of gram-negative bacteria, a new target for new molecules. *Curr. Top. Med. Chem.* 2010; 10(18):1848-1857.
13. Nikaido H, Pagès JM. Broad-specificity efflux pumps and their role in multidrug resistance of Gram-negative bacteria. *FEMS Microbiol. Rev.* 2012; 36(2):340-363.
14. Blair JM, Richmond GE, Piddock LJ. Multidrug efflux pumps in Gram-negative bacteria and their role in antibiotic resistance. *Future Microbiol.* 2014; 9(10):1165-1177.
15. Li XZ, Plésiat P, Nikaido H. The challenge of efflux-mediated antibiotic resistance in Gram-negative bacteria. *Clin. Microbiol. Rev.* 2015; 28(2): 337-418.
16. Kunz AN, Brook I. Emerging resistant Gram-negative aerobic bacilli in hospital-acquired infections. *Chemotherapy* 2010; 56(6): 492-500.
17. Peleg AY, Hooper DC. Hospital-acquired infections due to gram-negative bacteria. *New Eng. J. Med.* 2010; 362(19):1804-1813.
18. Chelazzi C, Pettini E, Villa G, De Gaudio AR. Epidemiology, associated factors and outcomes of ICU-acquired infections caused by Gram-negative bacteria in critically ill patients: an observational, retrospective study. *BMC Anesthesiol.* 2015; 15: 125.
19. Mehrad B, Clark NM, Zhanel GG, Lynch JP 3rd. Antimicrobial resistance in hospital-acquired gram-negative bacterial infections. *Chest* 2015; 147(5):1413-1421.
20. Ruppé É, Woerther PL, Barbier F. Mechanisms of antimicrobial resistance in Gram-negative bacilli. *Ann. Intensive Care* 2015; 5(1):61.
21. Saier MH Jr, Paulsen IT, Sliwinski MK, Pao SS, Skurray RA, Nikaido H. Evolutionary origins of multidrug and drug-specific efflux pumps in bacteria. *FASEB J.* 1998; 12(3): 265-274.
22. Van Bambeke F, Balzi E, Tulkens PM. Antibiotic efflux pumps. *Biochem. Pharmacol. Biochem Pharmacol.* 2000; 60(4):457-470.
23. Poole K, Srikumar R. Assessing the activity of bacterial multidrug efflux pumps. *Methods Mol. Med.* 2001; 48: 211-214.
24. Van Bambeke F, Glupczynski Y, Plésiat P, Pechère JC, Tulkens PM. Antibiotic efflux pumps in prokaryotic cells: occurrence, impact on resistance and strategies for the future of antimicrobial therapy. *J. Antimicrob. Chemother.* 2003; 51(5):1055-1065.
25. Van Bambeke F, Michot JM, Tulkens PM. Antibiotic efflux pumps in eukaryotic cells: occurrence and impact on antibiotic cellular pharmacokinetics, pharmacodynamics and toxicodynamics. *J. Antimicrob. Chemother.* 2003; 51(5): 1067-1077.
26. Prasad R, Gaur NA, Gaur M, Komath SS. Efflux pumps in drug resistance of *Candida*. *Infect. Disord. Drug Targets* 2006;

6(2): 69-83.

27. Cannon RD, Lamping E, Holmes AR, Niimi K, Baret PV, Keniya MV, Tanabe K, Niimi M, Goffeau A, Monk BC. Efflux-mediated antifungal drug resistance. *Clin. Microbiol. Rev.* 2009; 22(2):291-321
28. Fletcher JI, Haber M, Henderson MJ, Norris MD. ABC transporters in cancer: more than just drug efflux pumps. *Nat. Rev. Cancer.* 2010; 10(2):147-156.
29. Misaka S, Müller F, Fromm MF. Clinical relevance of drug efflux pumps in the gut. *Curr. Opin. Pharmacol.* 2013; 13(6):847-852.
30. Prasad R, Rawal MK. Efflux pump proteins in antifungal resistance. *Front. Pharmacol.* 2014; 5: 202.
31. Holmes AR, Cardno TS, Strouse JJ, Ivnitski-Steele I, Keniya MV, Lackovic K, Monk BC, Sklar LA, Cannon RD. Targeting efflux pumps to overcome antifungal drug resistance. *Future Med. Chem.* 2016; 8(12):1485-1501.
32. Saidijam M, Karimi Dermeni F, Sohrabi S, Patching SG. Efflux proteins at the blood-brain barrier: review and bioinformatics analysis. *Xenobiotica* 2017 [Epub ahead of print] DOI: 10.1080/00498254.2017.1328148.
33. Bates SE. Solving the problem of multidrug resistance: ABC transporters in clinical oncology. In *ABC proteins: From bacteria to man*, Holland IB, Cole SPC, Kuchler K, Higgins CF (Eds). Academic Press: New York, 2003, p. 359-391.
34. Chang G. Multidrug resistance ABC transporters. *FEBS Lett.* 2003; 555(1):102-105.
35. Lubelski J, Konings WN, Driessen AJ. Distribution and physiology of ABC-type transporters contributing to multidrug resistance in bacteria. *Microbiol. Mol. Biol. Rev.* 2007; 71(3):463-476.
36. Kerr ID, Jones PM, George AM. Multidrug efflux pumps: the structures of prokaryotic ATP-binding cassette transporter efflux pumps and implications for our understanding of eukaryotic P-glycoproteins and homologues. *FEBS J.* 2010; 277(3): 550-563.
37. Dawson RJ, Locher KP. Structure of a bacterial multidrug ABC transporter. *Nature* 2006; 443 (7108):180-185.
38. Pao SS, Paulsen IT, Saier MH Jr. Major facilitator superfamily. *Microbiol. Mol. Biol. Rev.* 1998; 62(1):1-34.
39. Saidijam M, Benedetti G, Ren Q, Xu Z, Hoyle CJ, Palmer SL, Ward A, Bettaney KE, Szakonyi G, Meuller J, Morrison S, Pos MK, Butaye P, Walravens K, Langton K, Herbert RB, Skurray RA, Paulsen IT, O'reilly J, Rutherford NG, Brown MH, Bill RM, Henderson PJ.. Microbial drug efflux proteins of the Major Facilitator Superfamily. *Curr. Drug Targets* 2006; 7(7):793-811.
40. Reddy VS, Shlykov MA, Castillo R, Sun EI, Saier MH Jr..The major facilitator superfamily (MFS) revisited. *FEBS J.* 2012; 279(11):2022-2035.
41. Kumar S, Mukherjee MM, Varela MF. Modulation of bacterial multidrug resistance efflux pumps of the Major Facilitator Superfamily. *Int. J. Bacteriol.* 2013; 204141.
42. Ranaweera I, Shrestha U, Ranjana KC, Kakarla P, Willmon TM, Hernandez AJ, Mukherjee MM, Barr SR, Varela MF. Structural comparison of bacterial multidrug efflux pumps of the Major Facilitator Superfamily. *Trends Cell. Mol. Biol.* 2015; 10:131-140.
43. Kumar S, He G, Kakarla P, Shrestha U, Ranjana KC, Ranaweera I, Willmon TM, Barr SR, Hernandez AJ, Varela MF. Bacterial multidrug efflux pumps of the Major Facilitator Superfamily as targets for modulation. *Infect. Disord. Drug Targets* 2016; 16(1):28-43.
44. Heng J, Zhao Y, Liu M, Liu Y, Fan J, Wang X, Zhao Y, Zhang XC. Substrate-bound structure of the *E. coli* multidrug resistance transporter MdfA. *Cell Res.* 2015; 25(9):1060-1073.
45. Tseng TT, Gratwick KS, Kollman J, Park D, Nies DH, Goffeau A, Saier MH Jr. The RND permease superfamily: an ancient, ubiquitous and diverse family that includes human disease and development proteins. *J. Mol. Microbiol. Biotechnol.* 1999; 1(1):107-125.
46. Routh MD, Zalucki Y, Su CC, Zhang Q, Shafer WM, Yu EW. Efflux pumps of the resistance-nodulation-division family: a perspective of their structure, function, and regulation in gram-negative bacteria. *Adv. Enzymol. Relat. Areas Mol. Biol.* 2011; 77:109-146.
47. Anes J, McCusker MP, Fanning S, Martins M. The ins and outs of RND efflux pumps in *Escherichia coli*. *Front. Microbiol.* 2015; 6:587.
48. Venter H, Mowla R, Ohene-Agyei T, Ma S. RND-type drug efflux pumps from Gram-negative bacteria: molecular mechanism and inhibition. *Front. Microbiol.* 2015; 6: 377.
49. Puzari M, Chetia P. RND efflux pump mediated antibiotic resistance in Gram-negative bacteria *Escherichia coli* and *Pseudomonas aeruginosa*: a major issue worldwide. *World J. Microbiol. Biotechnol.* 2017; 33(2): 24.
50. Seeger MA, Diederichs K, Eicher T, Brandstätter L, Schiefner A, Verrey F, Pos KM. The AcrB efflux pump: conformational cycling and peristalsis lead to multidrug resistance. *Curr. Drug Targets* 2008; 9(9):729-749.
51. Yamaguchi A, Nakashima R, Sakurai K. Structural basis of RND-type multidrug exporters. *Front. Microbiol.* 2015; 6: 327.
52. Daury L, Orange F, Taveau JC, Verchère A, Monlezun L, Gounou C, Marreddy RK, Picard M, Broutin I, Pos KM, Lambert O. Tripartite assembly of RND multidrug efflux pumps. *Nat. Commun.* 2016; 7:10731.
53. Paulsen IT, Skurray RA, Tam R, Saier MH Jr, Turner RJ, Weiner JH, Goldberg EB, Grinius LL.. The SMR family: a novel family of multidrug efflux proteins involved with the efflux of lipophilic drugs. *Mol. Microbiol.* 1996; 19(6): 1167-1175.
54. Chung YJ, Saier MH Jr. SMR-type multidrug resistance pumps. *Curr. Opin. Drug Discov. Dev.* 2001; 4(2):237-245.
55. Bay DC, Rommens KL, Turner RJ. Small multidrug resistance proteins: a multidrug transporter family that continues to grow. *Biochim. Biophys. Acta* 2008; 1778(9):1814-1838.
56. Bay DC, Turner RJ. Diversity and evolution of the small multidrug resistance protein family. *BMC Evol. Biol.* 2009; 9: 140.
57. Chen YJ, Pornillos O, Lieu S, Ma C, Chen AP, Chang G. X-ray structure of EmrE supports dual topology model. *Proc. Natl. Acad. Sci. U. S. A.* 2007; 104(48):18999-19004.

58. Korkhov VM, Tate CG.. An emerging consensus for the structure of EmrE. *Acta Crystallogr. D*, 2009; 65(Pt 2):186-192.
59. Omote H, Hiasa M, Matsumoto T, Otsuka M, Moriyama Y. The MATE proteins as fundamental transporters of metabolic and xenobiotic organic cations. *Trends Pharmacol. Sci.* 2006; 27(11):587-593.
60. Kuroda T, Tsuchiya T. Multidrug efflux transporters in the MATE family. *Biochim. Biophys. Acta* 2009; 1794(5):763-768.
61. Nie L, Grell E, Malviya VN, Xie H, Wang J, Michel H. Identification of the high-affinity substrate-binding site of the Multidrug and Toxic Compound Extrusion (MATE) family transporter from *Pseudomonas stutzeri*. *J. Biol. Chem.* 2016; 291(30):15503-15514.
62. He X, Szewczyk P, Karyakin A, Evin M, Hong WX, Zhang Q, Chang G. Structure of a cation-bound multidrug and toxic compound extrusion transporter. *Nature* 2010; 467(7318):991-994.
63. Lu M, Radchenko M, Symersky J, Nie R, Guo Y. Structural insights into H⁺-coupled multidrug extrusion by a MATE transporter. *Nat. Struct. Mol. Biol.* 2013; 20(11):1310-1317.
64. Lu M, Symersky J, Radchenko M, Koide A, Guo Y, Nie R, Koide S. Structures of a Na⁺-coupled, substrate-bound MATE multidrug transporter. *Proc. Natl. Acad. Sci. U. S. A.* 2013; 110(6):2099-2104.
65. Lu M. Structures of multidrug and toxic compound extrusion transporters and their mechanistic implications. *Channels (Austin)* 2016; 10(2):88-100.
66. Delmar JA, Yu EW. The AbgT family: A novel class of antimetabolite transporters. *Prot. Sci.* 2016; 25(2):322-337.
67. Chitsaz M, Brown MH. The role played by drug efflux pumps in bacterial multidrug resistance. *Essays Biochem.* 2017; 61(1):127-139.
68. Hassan KA, Jackson SM, Penesyan A, Patching SG, Tetu SG, Eijkelkamp BA, Brown MH, Henderson PJ, Paulsen IT. Transcriptomic and biochemical analyses identify a family of chlorhexidine efflux proteins. *Proc. Natl. Acad. Sci. U. S. A.* 2013; 110(50):20254-20259.
69. Hassan KA, Elbourne LD, Li L, Gamage HK, Liu Q, Jackson SM, Sharples D, Kolstø AB, Henderson PJ, Paulsen IT.. An ace up their sleeve: a transcriptomic approach exposes the AceI efflux protein of *Acinetobacter baumannii* and reveals the drug efflux potential hidden in many microbial pathogens. *Front. Microbiol.* 2015; 6:333.
70. Hassan KA, Liu Q, Henderson PJ, Paulsen IT. Homologs of the *Acinetobacter baumannii* AceI transporter represent a new family of bacterial multidrug efflux systems. *MBio* 2015; 6(1):e01982-14.
71. Bolla JR, Su CC, Delmar JA, Radhakrishnan A, Kumar N, Chou TH, Long F, Rajashankar KR, Yu EW. Crystal structure of the *Alcanivorax borkumensis* YdaH transporter reveals an unusual topology. *Nat. Commun.* 2015; 6:6874.
72. Su CC, Bolla JR, Kumar N, Radhakrishnan A, Long F, Delmar JA, Chou TH, Rajashankar KR, Shafer WM, Yu EW. Structure and function of *Neisseria gonorrhoeae* MtrF illuminates a class of antimetabolite efflux pumps. *Cell Rep.* 2015; 11(1):61-70.
73. Vergara-Jaque A, Fenollar-Ferrer C, Mulligan C, Mindell JA, Forrest LR. Family resemblances: A common fold for some dimeric ion-coupled secondary transporters. *J. Gen. Physiol.*, 2015; 146(5):423-434.
74. Wallin E, von Heijne G. Genome-wide analysis of integral membrane proteins from eubacterial, archaean, and eukaryotic organisms. *Prot. Sci.* 1998; 7(4):1029-1038.
75. Liu J, Rost B. Comparing function and structure between entire proteomes. *Prot. Sci.* 2001; 10(10):1970-1979.
76. Fagerberg L, Jonasson K, von Heijne G, Uhlén M, Berglund L. Prediction of the human membrane proteome. *Proteomics* 2010; 10(6):1141-1149.
77. Daley DO, Rapp M, Granseth E, Melén K, Drew D, von Heijne G. Global topology analysis of the *Escherichia coli* inner membrane proteome. *Science* 2005; 308(5726):1321-1323.
78. Piddock LJ. Multidrug-resistance efflux pumps - not just for resistance. *Nat. Rev. Microbiol.* 2006; 4(8):629-636.
79. Li XZ, Nikaido H. Efflux-mediated drug resistance in bacteria: an update. *Drugs* 2009; 69(12):1555-1623.
80. Blanco P, Hernando-Amado S, Reales-Calderon JA, Corona F, Lira F, Alcalde-Rico M, Bernardini A, Sanchez MB, Martinez JL. Bacterial multidrug efflux pumps: Much more than antibiotic resistance determinants. *Microorganisms* 2016; 4(1):E14.
81. Kvist M, Hancock V, Klemm P. Inactivation of efflux pumps abolishes bacterial biofilm formation. *App. Environ. Microbiol.* 2008; 74(23):7376-7382.
82. Matsumura K, Furukawa S, Ogihara H, Morinaga Y. Roles of multidrug efflux pumps on the biofilm formation of *Escherichia coli* K-12. *Biocontrol Sci.* 2011; 16(2):69-72.
83. Soto SM. Role of efflux pumps in the antibiotic resistance of bacteria embedded in a biofilm. *Virulence* 2013; 4(3):223-229.
84. Baugh S, Phillips CR, Ekanayaka AS, Piddock LJ, Webber MA. Inhibition of multidrug efflux as a strategy to prevent biofilm formation. *J. Antimicrob. Chemother.* 2014; 69(3):673-681.
85. Yamasaki S, Wang LY, Hirata T, Hayashi-Nishino M, Nishino K. Multidrug efflux pumps contribute to *Escherichia coli* biofilm maintenance. *Int. J. Antimicrob. Agents* 2015; 45(4):439-441.
86. Yoon EJ, Chabane YN, Goussard S, Snesrud E, Courvalin P, Dé E, Grillot-Courvalin C. Contribution of resistance-nodulation-cell division efflux systems to antibiotic resistance and biofilm formation in *Acinetobacter baumannii*. *MBio* 2015; 6(2):e00309-15.
87. Bay DC, Stremick CA, Slipski CJ, Turner RJ. Secondary multidrug efflux pump mutants alter *Escherichia coli* biofilm growth in the presence of cationic antimicrobial compounds. *Res. Microbiol.* 2017; 168(3):208-221.
88. He X, Lu F, Yuan F, Jiang D, Zhao P, Zhu J, Cheng H, Cao J, Lu G. Biofilm formation caused by clinical *Acinetobacter baumannii* isolates is associated with overexpression of the AdeFGH efflux pump. *Antimicrob. Agents. Chemother.* 2015; 59(8):4817-4825.
89. Chan YY, Bian HS, Tan TM, Mattmann ME, Geske GD,

- Igarashi J, Hatano T, Suga H, Blackwell HE, Chua KL. Control of quorum sensing by a *Burkholderia pseudomallei* multidrug efflux pump. J. Bacteriol. 2007; 189(11):4320-4324.
90. Varga, Z.G., A. Armada, P. Cerca, L. Amaral and M.A. Mior Ahmad Subkiet al., 2012. Inhibition of quorum sensing and efflux pump system by trifluoromethyl ketone proton pump inhibitors. In Vivo 26: 277-285.
91. Kalia VC, Wood TK, Kumar P. Evolution of resistance to quorum-sensing inhibitors. Microb. Ecol. 2014; 68(1):13-23.
92. Chen YF, Liu SY, Liang ZB, Lv MF, Zhou JN, Zhang LH. Quorum sensing and microbial drug resistance. Yi Chuan 2016; 38(10): 881-893.
93. Liang ZB, Chen YM, Chen Y, Cheng YY, Zhang LH. RND efflux pump and its interrelationship with quorum sensing system. Yi Chuan 2016; 38(10): 894-901.
94. Moore JD, Gerdt JP, Eibergen NR, Blackwell HE. Active efflux influences the potency of quorum sensing inhibitors in *Pseudomonas aeruginosa*. Chembiochem 2014; 15(3):435-442. Erratum in Chembiochem 2014; 15(5):634.
95. Nies DH. Efflux-mediated heavy metal resistance in prokaryotes. FEMS Microbiol. Rev. 2003; 27(2-3): 313-339.
96. Voica DM, Bartha L, Banciu HL, Oren A. Heavy metal resistance in halophilic Bacteria and Archaea. FEMS Microbiol. Lett. 2016; 363(14): fnw146.
97. LongLong F, Su CC, Zimmermann MT, Boyken SE, Rajashankar KR, Jernigan RL, Yu EW. Crystal structures of the CusA efflux pump suggest methionine-mediated metal transport. Nature 2010; 467(7314):484-488.
98. Su CC, Long F, Zimmermann MT, Rajashankar KR, Jernigan RL, Yu EW. Crystal structure of the CusBA heavy-metal efflux complex of *Escherichia coli*. Nature 2011; 470(7335):558-562.
99. Delmar JA, Su CC, Yu EW. Structural mechanisms of heavy-metal extrusion by the Cus efflux system. Biometals 2013; 26(4):593-607.
100. Delmar JA, Su CC, Yu EW. Heavy metal transport by the CusCFBA efflux system. Prot. Sci. 2015; 24(11):1720-1736.
101. Pal C, Bengtsson-Palme J, Kristiansson E, Larsson DG. Co-occurrence of resistance genes to antibiotics, biocides and metals reveals novel insights into their co-selection potential. BMC Genomics 2015; 16: 964.
102. Wales AD, Davies RH. Co-Selection of resistance to antibiotics, biocides and heavy metals, and its relevance to foodborne pathogens. Antibiotics (Basel) 2015; 4(4):567-604.
103. Pal C, Asiani K, Arya S, Rensing C, Stekel DJ, Larsson DGJ, Hobman JL. Metal resistance and its association with antibiotic resistance. Adv. Microb. Physiol. 2017; 70:261-313.
104. Romero JL, Grande Burgos MJ, Pérez-Pulido R, Gálvez A, Lucas R. Resistance to antibiotics, biocides, preservatives and metals in bacteria isolated from seafoods: Co-selection of strains resistant or tolerant to different classes of compounds. Front. Microbiol. 2017; 8: 1650.
105. Rosch JW, Gao G, Ridout G, Wang YD, Tuomanen EI. Role of the manganese efflux system mntE for signalling and pathogenesis in *Streptococcus pneumoniae*. Mol. Microbiol. 2009; 72(1):12-25.
106. Kirsten A, Herzberg M, Voigt A, Seravalli J, Grass G, Scherer J, Nies DH. Contributions of five secondary metal uptake systems to metal homeostasis of *Cupriavidus metallidurans* CH34. J. Bacteriol. 2011; 193(18):4652-4663.
107. López G, Latorre M, Reyes-Jara A, Cambiazo V, González M. Transcriptomic response of *Enterococcus faecalis* to iron excess. Biometals 2012; 25(4):737-747.
108. Guilhen C, Taha MK, Veyrier FJ. Role of transition metal exporters in virulence: the example of *Neisseria meningitidis*. Front. Cell. Infect. Microbiol. 2013; 3:102.
109. Frawley ER, Fang FC. The ins and outs of bacterial iron metabolism. Mol. Microbiol. 2014; 93(4):609-616.
110. Raimunda D, Elso-Berberián G. Functional characterization of the CDF transporter SMc02724 (SmYiiP) in *Sinorhizobium meliloti*: Roles in manganese homeostasis and nodulation. Biochim. Biophys. Acta 2014; 1838(12):3203-3211.
111. Kim HM, Ahn BE, Lee JH, Roe JH. Regulation of a nickel-cobalt efflux system and nickel homeostasis in a soil actinobacterium *Streptomyces coelicolor*. Metallomics 2015; 7(4):702-709.
112. Latorre M, Low M, Gárate E, Reyes-Jara A, Murray BE, Cambiazo V, González M. Interplay between copper and zinc homeostasis through the transcriptional regulator Zur in *Enterococcus faecalis*. Metallomics 2015; 7(7):1137-1145.
113. Chandrangu P, Rensing C, Helmann JD. Metal homeostasis and resistance in bacteria. Nat. Rev. Microbiol. 2017; 15(6):338-350.
114. Quintana J, Novoa-Aponte L, Argüello JM. Copper homeostasis networks in the bacterium *Pseudomonas aeruginosa*. J. Biol. Chem. 2017; 292(38): 15691-15704.
115. Paulsen IT, Saier MH Jr. A novel family of ubiquitous heavy metal ion transport proteins. J. Membr. Biol. 1997; 156(2):99-103.
116. Montanini B, Blaudez D, Jeandroz S, Sanders D, Chalot M. Phylogenetic and functional analysis of the Cation Diffusion Facilitator (CDF) family: improved signature and prediction of substrate specificity. BMC Genomics 2007; 8: 107.
117. Kolaj-Robin O, Russell D, Hayes KA, Pembroke JT, Soulimane T. Cation Diffusion Facilitator family: Structure and function. FEBS Lett. 2015; 589(12):1283-1295.
118. Veyrier FJ, Boneca IG, Cellier MF, Taha MK. A novel metal transporter mediating manganese export (MntX) regulates the Mn to Fe intracellular ratio and *Neisseria meningitidis* virulence. PLoS Pathog. 2011; 7(9):e1002261.
119. Eng BH, Guerinot ML, Eide D, Saier MH Jr. Sequence analyses and phylogenetic characterization of the ZIP family of metal ion transport proteins. J. Membr. Biol. 1998; 166(1): 1-7.
120. Kehres DG, Lawyer CH, Maguire ME. The CorA magnesium transporter gene family. Microb. Comp. Genomics 1998; 3(3): 151-169.
121. Niegowski D, Eshaghi S. The CorA family: structure and function revisited. Cell. Mol. Life Sci. 2007; 64(19-20):2564-2574.
122. Guskov A, Eshaghi S. The mechanisms of Mg²⁺ and Co²⁺

transport by the CorA family of divalent cation transporters. *Curr. Topics Membranes* 2012; 69: 393-414.

123. Alcalde-Rico M, Hernando-Amado S, Blanco P, Martínez JL. Multidrug efflux pumps at the crossroad between antibiotic resistance and bacterial virulence. *Front. Microbiol* 2016; 7: 1483.

124. Henderson P, Hoyle C, Ward A. Expression, purification and properties of multidrug efflux proteins. Portland Press Limited 2000.

125. Ward A, Sanderson NM, O'Reilly J, Rutherford NG, Poolman B, Henderson PJF. The amplified expression, identification, purification, assay and properties of hexahistidine-tagged bacterial membrane transport proteins. In *Membrane transport - a practical approach*, Baldwin SA (Ed), Oxford: Blackwell, p. 141-166.

126. Morrison S, Ward A, Hoyle CJ, Henderson PJ. Cloning, expression, purification and properties of a putative multidrug resistance efflux protein from *Helicobacter pylori*. *Int. J. Antimicrob. Agents* 2003; 22(3):242-249.

127. Szakonyi G, Leng D, Ma P, Bettaney KE, Saidijam M, Ward A, Zibaei S, Gardiner AT, Cogdell RJ, Butaye P, Kolsto AB, O'Reilly J, Hope RJ, Rutherford NG, Hoyle CJ, Henderson PJ. A genomic strategy for cloning, expressing and purifying efflux proteins of the Major Facilitator Superfamily, *J. Antimicrob. Chemother.* 2007; 59(6):1265-1270.

128. Bettaney KE, Sukumar P, Hussain R, Siligardi G, Henderson PJ, Patching SG. A systematic approach to the amplified expression, functional characterization and purification of inositol transporters from *Bacillus subtilis*. *Mol. Membr. Biol.* 2013; 30(1): 3-14.

129. Rosano GL, Ceccarelli EA. Recombinant protein expression in *Escherichia coli*: advances and challenges. *Front. Microbiol.* 2014; 5:172.

130. Kroeger JK, Hassan K, Vörös A, Simm R, Saidijam M, Bettaney KE, Bechthold A, Paulsen IT, Henderson PJ, Kolstø AB. *Bacillus cereus* efflux protein BC3310 - a multidrug transporter of the unknown major facilitator family, UMF-2. *Front. Microbiol.* 2015; 6: 1063.

131. Ma P, Patching SG, Ivanova E, Baldwin JM, Sharples D, Baldwin SA, Henderson PJ. Allantoin transport protein, PucI, from *Bacillus subtilis*: evolutionary relationships, amplified expression, activity and specificity, *Microbiology* 2016; 162(5):823-836.

132. Ahmad I, Nawaz N, Darwesh NM, ur Rahman S, Mustafa MZ, Khan SB, Patching SG. Overcoming challenges for amplified expression of recombinant proteins using *Escherichia coli*. *Protein Expr. Purif.* 2017 [Epub ahead of print] DOI: 10.1016/j.pep.2017.11.005.