



HAL
open science

Adding attractive semio-chemical trait refines the taxonomy of *Alpinobombus* (Hymenoptera: Apidae)

Baptiste Martinet, Nicolas Brasero, Thomas Lecocq, Paolo Biella, Irena Valterová, Denis Michez, Pierre Rasmont

► **To cite this version:**

Baptiste Martinet, Nicolas Brasero, Thomas Lecocq, Paolo Biella, Irena Valterová, et al.. Adding attractive semio-chemical trait refines the taxonomy of *Alpinobombus* (Hymenoptera: Apidae). *Api-dologie*, 2018, 49 (6), pp.838-851. 10.1007/s13592-018-0611-1 . hal-02367480

HAL Id: hal-02367480

<https://hal.science/hal-02367480>

Submitted on 18 Nov 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Adding attractive semio-chemical trait refines the taxonomy of *Alpinobombus* (Hymenoptera: Apidae)

Baptiste MARTINET¹, Nicolas BRASERO¹, Thomas LECOCQ^{1,2}, Paolo BIELLA^{3,4},
Irena VALTEROVÁ⁵, Denis MICHEZ¹, Pierre RASMONT¹

¹University of Mons, Research Institute of Biosciences, Laboratory of Zoology, Place du Parc 20, 7000, Mons, Belgium

²Research Unit Animal and Functionalities of Animal Products (URAFPA), University of Lorraine – INRA, 2 Avenue de la Forêt de Haye, BP 172, 54505, Vandoeuvre-lès-Nancy, France

³Department of Zoology, Faculty of Science, University of South Bohemia, Branišovská 31, 37005, České Budějovice, Czech Republic

⁴Biology Centre, Institute of Entomology, Czech Academy of Sciences, Branišovská 31, 37005, České Budějovice, Czech Republic

⁵Institute of Organic Chemistry and Biochemistry, Academy of Sciences of the Czech Republic, Flemingovo nám 2, CZ-166 10, Prague, Czech Republic

Received 2 February 2018 – Revised 10 July 2018 – Accepted 4 October 2018

Abstract – Species taxonomy of bumblebees (*Bombus* Latreille, 1802) is well known to be problematic due to a potentially high intra-specific variability of morphological traits while different species can converge locally to the same color pattern (cryptic species). Assessing species delimitation remains challenging because it requires to arbitrarily select variable traits whose accuracy continues to be debated. Integrative taxonomic approach seems to be very useful for this group as different independent traits are assessed to propose a rational taxonomic hypothesis. Among operational criteria to assess specific status, the reproductive traits involved in the pre-mating recognition (i.e., the male cephalic labial gland secretions, CLGS) have been premium information. Since these secretions are supposed to be species-specific, these chemical traits can bring essential information where species delimitation is debated. Here, we describe and compare the CLGS of 161 male specimens of nine *Alpinobombus* taxa: *alpinus*, *balteatus*, *helleri*, *hyperboreus*, *kirbiellus*, *navigi*, *neoboreus*, *polaris*, and *pyrrhopygus*. We aim also to test the congruence between this new information (reproductive traits) and published genetic dataset. Our results emphasized six distinct groups with diagnostic major compounds: (a) *alpinus* + *helleri* with hexadec-9-en-1-ol; (b) *polaris* + *pyrrhopygus* with two major compounds hexadec-9-en-1-ol and hexadec-9-enal; (c) *balteatus* with tetradecyl acetate; (d) *kirbiellus* with geranyl geranyl acetate; (e) *hyperboreus* + *navigi* with octadec-11-en-1-ol; (f) *neoboreus* with octadec-9-en-1-ol. Based on this new information, we can confirm the species status of *B. alpinus*, *B. balteatus*, *B. hyperboreus*, *B. kirbiellus*, *B. neoboreus*, and *B. polaris*. We also confirm the synonymy of *helleri* (Alps) with *alpinus* (Sweden). However, the specific status of *navigi* (Alaska) and *pyrrhopygus* (Sweden) is questionable and these taxa do not have specific CLGS composition.

bumblebees / species / cephalic labial gland secretions / arcto-alpine distribution / chemical trait

1. INTRODUCTION

The bumblebees (*Bombus* Latreille, 1802) are eusocial insects widely distributed and included ~ 250 species over the world (reviewed in Williams et al. 2008). Their diversity hotspots are localized in cold and temperate regions (Pekkarinen et al., 1981; Heinrich 2005). They have been studied for

Electronic supplementary material The online version of this article (<https://doi.org/10.1007/s13592-018-0611-1>) contains supplementary material, which is available to authorized users.

Corresponding author: B. Martinet,
baptiste.martinet@umons.ac.be

Manuscript editor: Marina Meixner

a long time by many entomologists because of their large size, attractive coat color pattern, and their economic value as crop pollinators (Michener, 2007; Klein et al. 2007; Patiny et al. 2009; Goulson 2010).

The subgenus *Alpinobombus* (Skorikov, 1914) is a small monophyletic subgenus particularly adapted (i.e., heterothermic thermo-regulation system) and restricted to the coldest areas of the bumblebee distribution such as Alaska, Fennoscandia, N. Siberia, or Greenland (Skorikov 1914; Milliron and Olivier, 1966; Løken 1973; Richards 1973; Heinrich 2005; Berezin 1995). Their presence is crucial for biodiversity and conservation of mountain and tundra habitats (Richards 1973) especially for pollination of arctic wild plants (e.g., *Astragalus* spp., *Bartsia* spp., *Hedysarum* spp., *Pedicularis* spp., *Salix* spp., *Vaccinium* spp.) (Shamurin, 1966; Kevan 1973; Løken 1973; Richards 1973; Stenström & Bergman 1998; Taylor & Rumsey 2003; Eriksen et al. 2006; Biella et al. 2017). Recent studies showed that some of these arcto-alpine populations are in decline and most likely because climate change (Martinet et al. 2015; Rasmont et al. 2015).

Conservation of a species group and its ecosystem service requires theoretical knowledge about its biology, ecology, and systematics. The *Alpinobombus* subgenus has been the focus of systematic studies using morphological characters (e.g., color pattern) that have since been highly criticized (Bickford et al. 2007; Batalha-Filho et al. 2010; Carolan et al. 2012; Lecocq et al. 2015). Indeed, due to their morphological similarities and their circum-polar distribution (i.e., isolated populations with relict populations in southern mountain ranges), many different taxa could show doubtful taxonomic status (Løken 1973; Williams et al. 2015). Several authors have examined the species of *Alpinobombus*, but no consensus on the number of valid species could be found (Richards 1931; Skorikov 1914; Løken 1973; Milliron 1973). Acknowledging the need of taxonomic revision, a recent study has used a more accurate trait than only morphological diagnose: DNA markers (Williams et al. 2015). Williams et al. (2015) proposed the hypothesis of nine putative species and one unnamed species, later

described as *B. kluanensis* Williams et al., 2016 based on genetic barcode and morphology: *B. alpinus* Linnaeus, 1758; *B. balteatus* Dahlbom, 1832; *B. hyperboreus* Schönherr, 1809; *B. kirbiellus* Curtis, 1835; *B. kluanensis* Williams et al., 2016; *B. natvigi* Richards, 1931; *B. neoboreus* Sladen, 1919; *B. polaris* Curtis, 1835; and *B. pyrrhopygus* Friese, 1902 (Figure 1). In addition to the description of the new species *B. kluanensis* (Canada), the nomenclatural operations consisted of splitting the taxon *hyperboreus* into *B. hyperboreus* (Scandinavia and far East Siberia) and *B. natvigi* (Canada, Alaska), the taxon *polaris* into *B. polaris* (Alaska, Canada, Greenland) and *B. pyrrhopygus* (Scandinavia, Russia), and the taxon *balteatus* (Scandinavia, Russia, Mongolia) into *B. balteatus* and *B. kirbiellus* (USA including Alaska, Canada). However, defining the species threshold in DNA barcoding analyses remains challenging despite the development of new statistical techniques such as Bayesian implementation of the general mixed Yule-coalescent model (bGMYC) to recognize species threshold (Pons et al. 2006; Andriollo et al. 2015; Lecocq et al. 2015; Mutanen et al. 2016). This issue can be overcome by integrating the DNA investigations with other pieces of evidence of speciation such as attractive semio-chemical traits or morphology in an integrative framework.

The bumblebee systematic remains challenging (Bertsch & Schweer 2012; Lecocq et al. 2015; Williams et al. 2012; Potapov et al., 2018). Development of an integrative taxonomy approach based on consensus between several independent traits (i.e., DNA, eco-chemical traits) provides an efficient solution to refine bumblebee systematic (Engel 2011; Lecocq et al. 2015). The congruence between independent traits can support the reliability of taxonomic conclusions (de Queiroz 2007). Indeed, although a differentiation in one character can be enough to highlight speciation process (Schlick-Steiner et al. 2010), the species status should be supported only if it is suggested by several lines of evidence. Therefore, the safest way is a strict approach where a taxon deserving a species status is a taxon with differentiation in all studied traits (Lecocq et al. 2015).



Figure 1 Pictures of eight studied *Alpinobombus* taxa. **a** *Bombus alpinus* male PRAS0806 from Sweden. **b** *B. kirbiellus* male PRAS1409 from Yukon (Canada). **c** *B. balteatus* male PRAS1653 from Khanymei (Siberia). **d** *B. hyperboreus* male BMAR0054 from Sweden. **e** *B. navigi* male PRAS1067 from Alaska. **f** *B. polaris* male PRAS1040 from Alaska. **g** *B. neoboreus* male PRAS1435 from Yukon (Canada). **h** *B. pyrrhopygus* male PRAS0929 from Norway (All Photos by P. Rasmont).

Several authors used operational criteria in bumblebee taxonomy such as genetic or semiochemical traits (i.e., cephalic labial gland secretions; CLGS) (e.g., Junior et al. 2015; Lecocq

et al. 2015; Martinet et al. 2018). The bumblebee pre-mating recognition system involves CLGS as attractive compounds for virgin females during nuptial behavior (i.e., “route-flight” or patrolling

behavior) (Ayasse & Jarau 2014). CLGSs are synthesized de novo (Žacek et al., 2013) and consist of a complex mixture of mainly aliphatic compounds (Coppée et al. 2008). Semio-chemical traits are shaped by selective pressure to allow the meeting of conspecific sexual partners and minimize heterospecific hybridization (Paterson 1993; Symonds et al. 2009; Wyatt 2003). Several studies showed that these semio-chemical compounds are highly species-specific (Bergström and Svensson 1973; Svensson 1980; Baer 2003; Bertsch & Schweer 2012; Ayasse & Jarau 2014; Lecocq et al. 2015). Despite the usefulness as chemotaxonomic characters of CLGS in bumblebee taxonomy, the CLGS composition of *Alpinobombus* taxa is only known roughly from Scandinavia and it has been studied with less accurate techniques (Svensson and Bergström, 1979). CLGS are significantly differentiated when we have (a) specific major compounds (abundant relative concentration > 5%), (b) specific indicator compounds, and (c) significant statistical differentiation test (high bootstrap values clustering and differentiation in multivariate statistic test) (Martinet et al. 2018).

Here, we propose to compare the DNA-based taxonomic conclusions on *Alpinobombus* to CLGS differentiation pattern in order to confirm or to reassess the taxonomic conclusions proposed by Williams et al. (2015).

2. MATERIALS AND METHODS

2.1. Sampling

A total of 161 alive bumblebee males collected between 2013 and 2017 were analyzed (Tab. S1). We were able to sample *B. alpinus* ($n = 15$) from Fennoscandia, *B. alpinus helleri* ($n = 5$) from Alps (Italy), *B. polaris* ($n = 21$) from Alaska, *B. pyrrhopygus* ($n = 3$) from Fennoscandia, *B. balteatus* from Sweden ($n = 19$) and from Siberia ($n = 20$), *B. natvigi* from Alaska ($n = 8$), *B. hyperboreus* from Fennoscandia ($n = 7$), *B. neoboreus* from Alaska ($n = 19$) and Yukon ($n =$), and *B. kirbiellus* from Alaska ($n = 20$) and from Yukon ($n = 16$) (Figure 1). Despite efforts to find *B. kluanensis* specimens in its species original location (*locus typicus*, Kluane lake, Yukon), we have not been able to collect it. The individual

bumblebee specimens were killed by freezing at $-20\text{ }^{\circ}\text{C}$. The field work was performed at Abisko (Sweden), Alps (Italy), Khanymey (W-Siberia), Kluane lake (Yukon, Canada), and Toolik field station (Alaska) (Figure 2).

2.2. Reproductive trait description

We focused on the most studied reproductive trait involved in the bumblebee pre-mating recognition (Svensson 1980; Ayasse & Jarau 2014; Baer 2003; Lecocq et al. 2015): the cephalic labial gland secretions (CLGS). CLGS were extracted with 400 μl of *n*-hexane (De Meulemeester et al. 2011). Samples were stored at $-40\text{ }^{\circ}\text{C}$ prior to the analyses. All samples have been extracted just after collection day to avoid alteration bias. Cephalic labial gland secretions have been analyzed on the same year to ensure a good conservation of samples and to avoid analytical bias.

The qualitative composition of the CLGS was determined by gas chromatography mass spectrometry using a Finigan GCQ quadrupole system (GC/MS) with a non-polar DB-5 ms capillary column [5% phenyl (methyl) polysiloxane stationary phase; column length 30 m; inner diameter 0.25 mm; film thickness 0.25 μm]. Double bond positions (C=C) were determined by dimethyl disulfide (DMDS) derivatization (Vincenti et al. 1987). All samples were quantified with a gas chromatograph Shimadzu GC-2010 system (GC-FID) equipped with a non-polar SLB-5 ms capillary column [5% phenyl (methyl) polysiloxane stationary phase; column length 30 m; inner diameter 0.25 mm; film thickness 0.25 μm] and a flame ionization detector. All compounds for which relative abundance (RA) were recorded as less than 0.1% for all specimens were excluded from the analysis (De Meulemeester et al. 2011). The data matrix for each taxa was based (Appendix S2) on the alignment of each relative proportion of compound between all samples performed with GCaligner 1.0 (Dellicour & Lecocq 2013a, b). We defined “main compounds” as compounds that have the highest relative proportion (RA) among all compounds of CLGSs profile of at least one individual of the taxon. To facilitate the alignment of compounds and their identification, before each sample injection, a mixture of



Figure 2 Circum arctic sampling map (Northern Hemisphere polar Lambert Azimuthal projection) after original of Sean Baker on which the red dots indicate the areas where we collected specimens of *Alpinobombus*.

alkanes C10-C40 was injected as a standard for calculation Kovats indices (KI). KI were calculated with GC Kovats 1.0 according to the method described by Dellicour & Lecocq (2013).

2.3. Statistical analysis

We performed statistical comparative analyses of the CLGSs using R (R Development Core Team 2017) to detect CLGS differentiations between *Alpinobombus* taxa. To detect divergence in CLGS

composition, we performed a principal component analysis (PCA) based on correlation distance matrix, and a clustering method, computed with the unweighted pair-group method with average linkage (UPGMA) based on Pearson Phi correlation distance matrix (R package ape; Paradis et al. 2004). We assessed the uncertainty in hierarchical cluster analysis using p values calculated via multiscale bootstrap resampling with 100,000 bootstrap replications (R package pvclust, Suzuki & Shimodaira 2011). When we observed a

differentiation in our clustering and PCA analyses, we checked CLGS differentiations between taxa by performing a permutational multivariate analysis of variance (perMANOVA) followed by pairwise perMANOVA using the Bray-Curtis dissimilarity matrix (R package *vegan*, Oksanen et al. 2017). Prior to these tests, the multivariate homogeneity of within-group covariance matrices was verified using the “betadisper” function. When a significant difference was detected, we performed multiple pairwise comparisons with an adjustment of *p* values (Bonferroni correction). To determine specific compounds of each taxon (indicator compounds, IC, *p* value > 0.70), the indicator-value (IndVal) method was used (Claudet et al. 2006; Dufrene and Legendre, 1997). The statistical significance of indicator compounds (threshold of 0.01) was evaluated with a randomization procedure.

3. RESULTS

3.1. Composition of cephalic labial gland secretions

Overall, within the CLGS, our analyses allowed the detection of 155 compounds: 51 from *B. alpinus* and 51 from *B. alpinus helleri* [IC *alpinus/helleri* = 5], 66 from *B. polaris*, 64 from *B. pyrrhopygus* [IC *polaris/pyrrhopygus* = 11], 66 from *B. balteatus* [IC = 18], 63 from *B. kirbiellus* [IC = 12], 63 from *B. hyperboreus*, 66 from *B. natvigi* [IC *hyperboreus/natvigi* = 22], and 64 from *B. neoboreus* [IC = 13] (Appendix S2). Several minor compounds were shared by all *Alpinobombus* taxa such as octadecenoic acid, tricos-9-ene, pentacosane, heptacos-9-ene, heptacosane, nonacos-9-ene, nonacosane, and hentriacontene. Several other compounds were detected in large amounts (median > 5%) with *Alpinobombus* taxa: (i) hexadec-9-en-1-ol (64.32–89.43%) in *alpinus*; (ii) hexadec-9-enal (0.62–10.86%) in *helleri*; (iii) dodecyl butyrate (3.51–30.47%), tetradecyl butyrate (8.16–28.89%), geranyl citronellol (2.68–27.01%) in *balteatus*; (iv) octadecenoic acid (1.84–20.54%) in *kirbiellus*; (v) dihydrofarnesol (12.29–23.41%), geranyl citronellol (2.87–7.00%) in *hyperboreus*; (vi) dihydrofarnesol (9.05–18.13%), octadecenoic acid (4.59–10.09%),

geranyl citronellol (4.18–12.42%) in *natvigi*; (vii) tetradec-9-en-1-ol (6.58–29.97%), octadecenoic acid (1.19–14.23%), sitosterol (0.40–11.06%) in *neoboreus*; (viii) hexadecenoic acid (3.80–13.62%), octadecenoic acid (2.87–16.20%), hexadecenyl hexadecanoate (3.63–20.74%) in *polaris*; (ix) hexadecenyl hexadecanoate (4.56–6.47%) in *pyrrhopygus*. The full list of identified compounds is presented in Supplementary file 2.

Our hierarchical analyses and principal component analysis (ACP, data matrix of 155 compounds × 161 specimens) emphasized six distinct groups with specific main compounds (Table 1) supported by high multiscale bootstrap resampling values (Figure 3): (i) *B. alpinus* + *B. alpinus helleri* with hexadec-9-en-1-ol (64.32–89.43%) which largely dominate the volatile marking secretion; (ii) *B. polaris* + *B. pyrrhopygus* with hexadec-9-en-1-ol (7.78–43.17%) and hexadec-9-enal (9.62–46.52%); (iii) *B. balteatus* with tetradecyl acetate (13.17–47.17%); (iv) *B. kirbiellus* with geranyl geranyl acetate (28.50–80.59%); (v) *B. hyperboreus* + *B. natvigi* with octadec-11-en-1-ol (23.90–56.80%); (vi) *B. neoboreus* with octadec-9-en-1-ol (25.59–58.11%). The specific recognition signals differed significantly among groups, providing a good indication on the specific status of these taxa. While the typical chromatograms (chemical fingerprint) of these groups were unique, we observed similarities in the spectrum profile between *B. alpinus* and *B. polaris* and between *B. hyperboreus* and *B. neoboreus*. The variability of compound profiles was low for each taxa except for *B. alpinus helleri* showing different relative abundance of hexadec-9-enal (min 0.62%–max 10.86%).

Several significant and specific indicator compounds were revealed by the IndVal method (IndVal > 0.70) (Appendix S2): (a) *alpinus* + *helleri* (IC = 5), (b) *polaris* + *pyrrhopygus* (IC = 11), (c) *balteatus* (IC = 18), (d) *kirbiellus* (IC = 12), (e) *hyperboreus* + *natvigi* (IC = 22), (f) *neoboreus* (IC = 13). Hexadecenyl hexadecanoate (0.22–1.10%) and geranyl citronellyl tetradecanoate (0.07–0.43%) were detected in *B. natvigi* but not in *B. hyperboreus*. Moreover, several compounds such as octadecenoic acid were detected only in

Table 1. List of main compounds identified for *Bombus alpinus*, *B. alpinus helleri*, *B. balteatus*, *B. hyperboreus*, *B. kirbiellus*, *B. natvigi*, *B. neoboreus*, *B. polaris*, and *B. pyrrhopygus* within cephalic labial gland secretions. MW, molecular weight; M, median of relative concentration of compound (%); *n*, number of specimens analyzed. Main compound in italics. Comprehensive information is available in Appendix S2

Compounds (%)	MW	<i>alpinus</i> (<i>n</i> = 15) M	<i>helleri</i> (<i>n</i> = 5) M	<i>balteatus</i> (<i>n</i> = 39) M	<i>hyperboreus</i> (<i>n</i> = 7) M	<i>kirbiellus</i> (<i>n</i> = 36) M	<i>natvigi</i> (<i>n</i> = 8) M	<i>neoboreus</i> (<i>n</i> = 27) M	<i>polaris</i> (<i>n</i> = 21) M	<i>pyrrhopygus</i> (<i>n</i> = 3) M
Hexadec-9-enal	238	0.08	4.36	0.00	0.00	0.00	0.00	0.00	23.86	42.74
Hexadec-9-en-1-ol	240	<i>83.20</i>	<i>62.51</i>	0.00	0.00	0.00	0.00	2.71	27.89	37.79
Tetradecyl acetate	256	0.00	0.00	28.57	0.00	0.00	0.00	0.00	0.00	0.00
Octadec-9-en-1-ol	268	0.02	0.00	0.00	0.00	0.00	0.00	42.29	0.02	0.02
Octadec-11-en-1-ol	268	0.03	0.10	0.00	<i>50.11</i>	0.00	32.16	0.04	0.00	0.00
Geranyl geranyl acetate	332	0.00	0.00	0.00	0.00	57.52	0.00	0.00	0.00	0.00

small amounts in *natvigi* (0.05–3.94%) and in larger amounts in *B. hyperboreus* (4.59–10.09%). Concerning *B. alpinus/helleri*, hexadecanoic acid (0.15–2.02%), ethyl octadec-11-enoate (0.04–1.09%), and X9 (0.10–0.40%) were detected in *helleri* but not in *B. alpinus* and sitosterol (0.00–0.70%) was detected in *B. alpinus* but not in *B. alpinus helleri*.

Statistical test (perMANOVA) confirmed that “taxa factor” has a significant effect on CLGS composition ($F_{8,152} = 209.76$, $p < 0.001$). Significant differences have been found between *balteatus* and *kirbiellus* (pairwise perMANOVA $F_{1,73} = 46.277$, $p < 0.001$), *balteatus* and *hyperboreus/natvigi* ($F_{1,52} = 188.39$, $p < 0.001$), *balteatus* and *alpinus/helleri* ($F_{1,58} = 328.52$, $p < 0.001$), *balteatus* and *polaris/pyrrhopygus* ($F_{1,61} = 273.26$, $p < 0.001$), *balteatus* and *neoboreus* ($F_{1,64} = 59.77$, $p < 0.001$), *neoboreus* and *hyperboreus/natvigi* ($F_{1,40} = 247.91$, $p < 0.001$), *neoboreus* and *alpinus/helleri* ($F_{1,46} = 383.82$, $p < 0.001$), *neoboreus* and *polaris/pyrrhopygus* ($F_{1,49} = 263.66$, $p < 0.001$), *neoboreus* and *kirbiellus* ($F_{1,61} = 418.67$, $p < 0.001$), *kirbiellus* and *alpinus/helleri* ($F_{1,54} = 382.16$, $p < 0.001$), *kirbiellus* and *polaris/pyrrhopygus* ($F_{1,58} = 268.37$, $p < 0.001$), *kirbiellus* and *hyperboreus/natvigi* ($F_{1,49} = 234.44$, $p < 0.001$), *alpinus/helleri* and *polaris/pyrrhopygus* ($F_{1,42} = 93.44$, $p < 0.001$), *alpinus/helleri* and *hyperboreus/natvigi* ($F_{1,33} = 256.79$, $p < 0.001$), and between *hyperboreus/natvigi* and *polaris/pyrrhopygus* ($F_{1,37} = 159.06$, $p < 0.001$).

4. DISCUSSION

For a long time, so-called cryptic species have caused problems for bumblebee taxonomy (Carolan et al. 2012; Williams et al. 2012). Indeed, the lack of separating morphological characteristics was a significant impediment. Although coat coloration is useful for quick and efficient identification of bumblebees in the field (in addition to other structural criteria), critical errors could occur (Bertsch & Schweer, 2011). Within *Alpinobombus* taxa, the problem is that reliable species-specific morphological differences are slight (Williams et al. 2015). In this case, integrating genetic markers, CLGS, and other operational

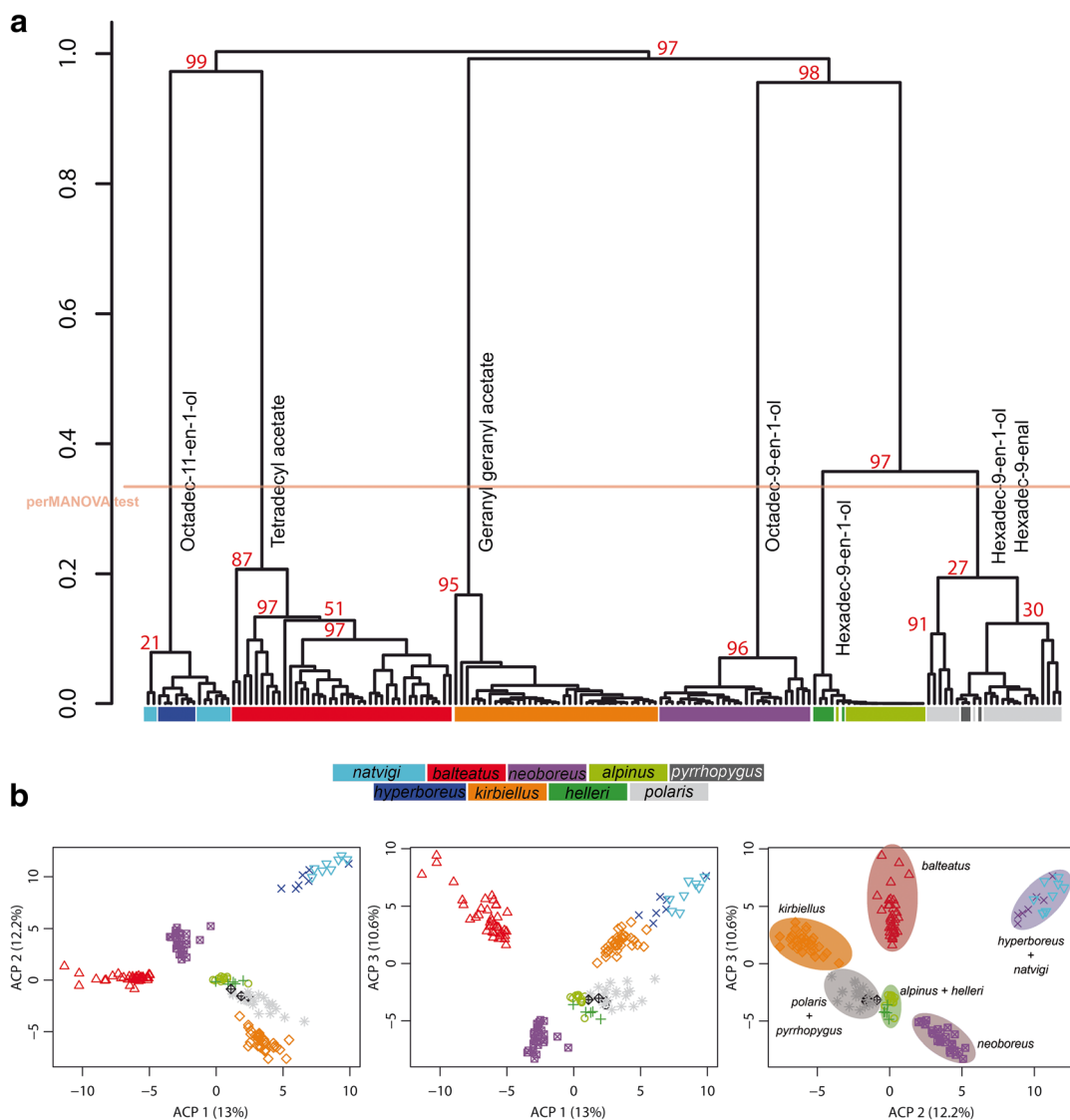


Figure 3 **a** Dendrogram based on cephalic labial gland secretions of *Bombus alpinus*, *B. alpinus helleri*, *B. balteatus*, *B. hyperboreus*, *B. kirbiellus*, *B. natvigi*, *B. neoboreus*, *B. polaris*, and *B. pyrrophygus*. This cluster was obtained by hierarchical clustering using an unweighted pair-group method with arithmetic mean (UPGMA) based on a correlation matrix. The values near nodes represent multiscale bootstrap resampling values. The main compound of each taxa has been annotated on the main branches of the cluster. **b** Principal component analysis (PCA) of cephalic labial gland secretion differentiation on 155 compounds \times 161 specimens within *Alpinobombus* taxa: *Bombus alpinus*, *B. alpinus helleri*, *B. balteatus*, *B. hyperboreus*, *B. kirbiellus*, *B. natvigi*, *B. neoboreus*, *B. polaris*, *B. pyrrophygus*. PC1, PC2, and PC3 are the first, the second, and the third principal component axes.

taxonomic criteria in an integrative framework could bring useful information for biochemical systematics for separation of critical taxa in the context of species delimitation according to an integrative approach sensu de Queiroz (2007).

Nuptial behavior and scent marking of bumblebees have been the subject of several studies (see review in Ayasse & Jarau 2014). For the *Alpinobombus*, Svensson & Bergström (1979) described the patrolling behavior of *B. alpinus*,

B. polaris, *B. balteatus*, and *B. hyperboreus* during their courtship which involved a deposit of long-distance attractive substances on different substrates. Svensson & Bergström (1979) suggested that these CLGS are sex pheromones and are involved in attracting virgin females during patrolling behavior. The gland secretions analyzed in the present study contained the common mixtures of aliphatic compounds and straight-chain fatty acid derivatives (alcohols, esters, aldehydes, and hydrocarbons) already detected in the CLGS of the subgenus *Alpinobombus* (Svensson & Bergström 1979).

4.1. Description of the CLGS of *Bombus kirbiellus*

We found a unique CLGS mixture dominated by geranylgeranyl acetate for *Bombus kirbiellus* (North America), very different from its sister species *Bombus balteatus* (Eurasia) dominated by tetradecyl acetate. Our investigation of CLGS of *kirbiellus* (North America) confirms the results of genetic analysis performed by Williams et al. (2015) (Figure 3). As discussed in Brasero et al. (2015), the geranylgeranyl acetate is also the main compounds identified in *Cullumanobombus* species (Hovorka et al. 2006), in *B. (Alpigenobombus) wurflenii* (Svensson et al. 1984) and *B. (Kallobombus) soroeensis* (Kullenberg et al. 1970). In other insects, Symonds & Elgar (2004) suggested that most chemical components were distributed randomly across the phylogeny in bark beetles (*Dendroctonus*). Our results confirm that the same main compound could be shared by different species phylogenetically and in this case geographically distant.

4.2. Differentiation of *helleri/alpinus*, *hyperboreus/natvigi*, and *polaris* and *pyrrhopygus*

In their recent taxonomic revision, Williams et al. (2015) suggested that (i) populations of *alpinus* from Alps (also called subspecies *helleri*) and N. Scandinavia (*alpinus*) are conspecific, (ii) the heterospecificity between *natvigi* (North America) and *hyperboreus* (Scandinavia, Russia), and the heterospecificity between *polaris*

(Alaska, Canada, Greenland) and *pyrrhopygus* (Sweden, Russia) based on COI and PEPCK gene fragments.

Concerning *alpinus/helleri*, based on CLGS matrix, hierarchical and principal component analyses showed only one group mixing the two populations (Figure 3). However, minor quantitative differences (e.g., variability of the relative abundance of the hexadec-9-enal in *helleri*) can be observed in the spectrum. For *hyperboreus/natvigi*, hierarchical and principal component analyses showed also only one group gathering the two taxa (Figure 2). For these taxa, we observed quantitative differences involving very minor compounds (median < 0.20%) close to our threshold of 0.1% which have probably not a major role in pre-mating recognition (Appendix S2). Brasero et al. (2017) and Terzo et al. (2005) showed intra-specific variability in the CLGS of different bumblebee species. Several compounds could be abundant or absent according to the specimen analyzed. Several authors have shown that CLGS composition may change during the lifetime of the male (age-dependent; Žáček et al. 2009) and even during the day (more active in the morning) (Bergman 1997). However, as discussed by Bertsch & Schweer (2011), these parameters do not modify significantly the species-specific chemical fingerprint. Following our CLGS divergence threshold, we suggest to consider these differences as random intra-specific variability explained by geographical isolation (Coppée et al. 2008; Martinet et al. 2018). We confirmed therefore the hypothesis of Williams et al. (2015) for the taxa *helleri/alpinus* but not for the taxa *hyperboreus/natvigi*.

As for *hyperboreus/natvigi*, the results of CLGS of *polaris* (Alaska) and *pyrrhopygus* (Scandinavia) do not confirm the recent taxonomic hypothesis of Williams et al. (2015) based on genetic analysis (Figure 3). Indeed, hierarchical and principal component analyses emphasize a unique group. Despite the coat color differentiation, all studied specimens seem to belong to the same species, *B. polaris*. As suggested by Martinet et al. (2018), mate recognition system is species-specific and differ even between closely related bumblebee species which have geographical distributions that do not overlap. Moreover, inside these three taxa pairs (*alpinus/helleri*; *hyperboreus/*

natvigi; *polaris*/*pyrrhopygus*), none difference involving main compounds have been observed contrary to what has been observed in the pairs *B. balteatus*/*B. kirbiellus* and *B. hyperboreus*/*B. neoboreus* (closely related species, Williams et al. 2015). However, Symonds et al. (2009) and Symonds & Elgar (2007) suggested in other insect groups that at speciation event, pheromones differ via large shifts in composition (involving abundant compounds) by saltational changes even where other mechanisms exist to ensure reproductive isolation (e.g., allopatry). Subsequently, gradual changes also occur to differentiate taxa (Symonds et al. 2009). Main compounds have been suggested to be strong indicator of potential ethological consequences for pre-mating recognition (Lecocq et al. 2015). However, in bumblebees, several studies have shown that uncontested different species (e.g., *Cullamanobombus* Brasero et al. 2015; Hovorka et al. 2006; *Bombus muscorum* and *Bombus filchnerae* Brasero et al. 2017) have the same main compounds. This could indicate that minor compounds could play also a key role in bumblebee species discrimination as diagnostic character. However, further ethological tests or neural coding studies (e.g., electrophysiological recordings) are needed to assess the role of these minor compounds as neural stimuli in the context of nuptial behavior.

In their taxonomic revision, Williams et al. (2015) have showed that nuclear gene (phosphoenolpyruvate carboxykinase) did not allow to differentiate the ambiguous taxa *polaris*/*pyrrhopygus* and *natvigi*/*hyperboreus* (no unique haplotype). Nevertheless, these couples of taxa were significantly divergent in mitochondrial gene (cytochrome oxidase 1) with bGMYC threshold (Williams et al. 2015). However, recent studies highlighted the high variability of this gene (COI) at intra- and interspecific levels and questioned the COI-based taxonomic conclusions in the absence of corroborations by other taxonomic criteria (e.g., Trunz et al. 2016; Soltani et al. 2017). From a morphological point of view, COI-based taxonomic statuses are not corroborated: there are very few differences between *polaris* and *pyrrhopygus* or between *natvigi* and *hyperboreus* except a slight color pattern differentiation (Williams et al. 2015). However, color

pattern was shown to be a poor taxonomic criterion in bumblebees, making the differentiation doubtful (Bickford et al. 2007; Batalha-Filho et al. 2010; Carolan et al. 2012; Lecocq et al. 2015). Similarly, our CLGS-based conclusions (reproductive traits involved in the pre-mating recognition system) are divergent from COI-based previous study. A hypothesis to explain this divergence could be that these taxa are in a speciation process and that these two data types have different evolution rates (e.g., Symonds et al. 2009). Indeed, according to the species delimitation criterion system used, we assign or not the species status to these allopatric populations, hence the importance to integrate these criteria into an integrative framework to refine our taxonomic hypothesis (higher specificity). Therefore, we advocate to use a more complete integrative taxonomic approach (see such a strict method in Lecocq et al. 2015; Martinet et al. 2018) considering here all available independent operational criteria (genetic, morphological, CLGS datasets) to propose a more supported taxonomic hypothesis. Following this integrative approach, *pyrrhopygus* should be considered as conspecific with *polaris* and *natvigi* conspecific with *hyperboreus*.

When comparing different datasets, involving independent traits commonly used in bumblebee systematic, such as morphology (Williams et al. 2015), genetic (Williams et al. 2015), and chemical (CLGS, present data), it can lead in some cases to an incongruence between different traits in taxonomic conclusions. The integrative taxonomic approach is thus useful for a better supported taxonomic hypothesis. This taxonomic hypothesis depends on the number of criteria used and how to integrate them. Considering that genetic and chemical datasets could have different evolutionary rates (Symonds et al. 2009; Lecocq et al. 2015), it is possible that, according to the criteria used to perform species delimitation, we have or not significant divergence for a taxon involved in a speciation process. Moreover, for the genetic dataset, often only few markers arbitrarily chosen are used (Cruaud et al. 2014). Adding independent operational criteria such as CLGS, which are a very species-specific ecological trait, allow to refine taxonomic hypotheses in an integrative taxonomic approach.

4.3. Impact on zoological nomenclature

When taxa are genetically and chemically differentiated, our analysis supports the next nomenclatural situation in *Alpinobombus* subgenus:

Bombus alpinus (L., 1758) [status confirmed]
 ssp. *alpinus* (L., 1758)
 ssp. *helleri* von Dalla Torre, 1882

Bombus balteatus Dahlbom, 1832 [status confirmed]

Bombus hyperboreus Schönherr, 1809
 ssp. *hyperboreus* Schönherr, 1809
 ssp. *navigi* Richards, 1931 (= *Bombus navigi* Richards 1931, sensu Williams et al. 2015)

Bombus kirbiellus Curtis, 1835 [status confirmed]

Bombus kluanensis Williams et al., 2016 [not assessed]

Bombus neoboreus Sladen, 1919 [status confirmed]

Bombus polaris Curtis, 1835
 ssp. *polaris* Curtis, 1835
 ssp. *pyrrhopygus* Friese, 1902 (= *Bombus pyrrhopygus* Friese 1902, sensu Williams et al. 2015)

4.4. Evolution of CLGS in *Alpinobombus*

The variation in the main compound differentiation within *Alpinobombus* taxa could be due to a switching off and on of specific metabolic pathways (Glaser et al. 2014) and/or by activation of an enzyme transcript present in a common ancestor (Roelofs et al., 2002). For instance, between CLGS of *B. hyperboreus* and *B. neoboreus* (two closely related species), the difference in the main compounds lies in the position of the C=C double bond (octadec-11-en-1-ol vs octadec-9-en-1-ol). This differentiation may arise by the activity of the enzyme D11-desaturase in *B. hyperboreus* and D9-desaturase in *B. neoboreus* (previously described *Bombus lucorum* by Matoušková et al. (2008), or in other taxa Löfstedt 1993; Brasero et al. 2015). Diagnostic compounds in the male labial gland secretions of many bumblebee species are primary alcohols (Bertsch & Schweer 2012). Therefore, it is likely that they play a key role in pre-mating recognition (Ayasse & Jarau. 2014).

Between *B. polaris* and *B. alpinus* which share the main compound (hexadec-9-en-1-ol), the CLGS differentiation of the second major compound in *B. polaris* present in equal amount (hexadec-9-enal) and in much lower concentration in *B. alpinus* could result of a large increase of a minor compound shared by closely related species as suggested by Terzo et al. (2005).

ACKNOWLEDGEMENTS

The authors thank the Abisko (Sweden), Tarfala (Sweden), Toolik field (USA), Kluane lake (Canada), and Khanymey (W-Siberia) scientific stations for their hospitality and their help in material collection. We acknowledge Hannele Savela (Oulu University, INTERACT administration) for her help in administration process. We thank also K. Urbanová (Institute of Organic Chemistry and Biochemistry ASCR) for her help in chemical analyses. The research leading to these results received funding from the European Union's Horizon 2020 project INTERACT, under grant agreement no. 730938. BM contributes as PhD student granted by the FRS-FNRS (Fonds de la Recherche Scientifique). PB contributes as a PhD student funded by the Czech Science Foundation (GACR GP14-10035P) and by the University of South Bohemia (GA JU 152/2016/P). The authors thank the Parco Nazionale dello Stelvio for granting permission to collect in their respective territories to the author PB.

AUTHOR CONTRIBUTION

Conceived and designed the experiments: BM NB TL PR. Material Collection: BM NB PB PR. Experimentation: BM. Analyzed the data: BM IV. Wrote the paper: BM NB TL PB IV DM PR

L'ajout d'un trait sémio-chimique attractif améliore la taxonomie de l'*alpinobombus* (hyménoptères: apidés)

Bourdons / espèces / sécrétions des glandes labiales céphaliques / distribution arcto-alpine / caractère chimique.

Die Zusatzinformation bezüglich attraktiver Semikommunikationsmerkmale verbessert die Taxonomie von *Alpinobombus* (Hymenoptera: Apidae)

Hummeln / Arten / Kopfblabdrüsensekrete / arktisch-alpine Verbreitung / chemisches Merkmal

REFERENCES

- Andriollo T., Naciri Y., Ruedi M. (2015) Two mitochondrial barcodes for one biological species: the case of European Kuhl's Pipistrelles (Chiroptera). *Plos One*, **10**, e0134881.
- Ayasse M., Jarau S. (2014) Chemical ecology of bumble bees. *Annu. Rev. Entomol.* **59**, 299–319.
- Baer B. (2003) Bumblebees as model organisms to study male sexual selection in social insects. *Behav. Ecol. Sociobiol.* **54**, 521–533.
- Batalha-Filho H., Waldschmidt A., Campos L.A.O., Tavares M.G., Fernandes-Salomao T. (2010) Phylogeography and historical demography of the Neotropical stingless bee *Melipona quadrifasciata* (Hymenoptera, Apidae): incongruence between morphology and mitochondrial DNA. *Apidologie*, **4**, 534–547.
- Berezin M.V. (1995) Geographical Diversity, Species Correlation, Population Structure and Cenotic Interactions of Arctic Bumble Bees (Apidae, *Bombus*), in: Grönlund E., Melander O. (ed.) Swedish-Russian Tundra Ecology. Expedition-94. Swedish Polar Research Secretariat, Stockholm, pp. 205–215.
- Bergman P. (1997) Chemical Communication in Bumblebee Premating Behaviour. PhD thesis, Göteborg University, Sweden.
- Bertsch A., Schweer H. (2011) Labial gland marking secretions of male *Bombus lucorum* bumblebees from Europe and China reveal two separate species: *B. lucorum* (Linnaeus 1761) and *Bombus minshanicola* (Bischoff 1936). *Biochem.Syst Ecol.* **39** (4-6), 587-593.
- Bertsch A., Schweer H. (2012) Cephalic labial gland secretions of males as species recognition signals in bumblebees: are there really geographical variations in the secretions of the *Bombus terrestris* subspecies? *Beiträge zur Entomologie*, **62**, 103–124.
- Bergström G., Svensson B.G. (1973) Studies on natural odoriferous compounds VIII. Characteristic marking secretions of two forms *lapponicus* and *scandinavicus* of *B. lapponicus* Fabr. (Hymenoptera, Apidae). *Chemica Scripta*, **4**, 231–239.
- Bickford D., Lohman D.J., Sodhi N.S., Ng P.K.L., Meier R., Winker K., Ingram K.K., Das I. (2007) Cryptic species as a window on diversity and conservation. *Trends Ecol. Evol.* **22**, 148–155.
- Biella P., Bogliani G., Cornalba M., Manino A., Neumayer J., Porporato M., Rasmont P., Milanese P. (2017) Distribution patterns of the cold adapted bumblebee *Bombus alpinus* in the Alps and hints of an uphill shift (Insecta: Hymenoptera: Apidae). *J. Ins. Conserv.* **21**(2), 357-366.
- Brasero N., Martinet B., Urbanová K., Valterová I., Torres A., Hoffmann W., Rasmont P., Lecocq T. (2015) First chemical analysis and characterization of the male species-specific cephalic labial gland secretions of South American bumblebee. *Chem. Biodivers.* **12** (10), 1535-1546.
- Brasero N., Lecocq T., Martinet B., Valterová I., Urbanová K., de Jonghe R. (2017) Variability in Sexual Pheromones Questions their Role in Bumblebee Pre-Mating Recognition System. *J. Chem. Ecol.* <https://doi.org/10.1007/s10886-017-0910-4>.
- Carolan J.C., Murray T.E., Fitzpatrick U., Crossley J., Schmidt H., Cederberg B., McNally L., Paxton R.J., Williams P.H., Brown M.J.F. (2012) Colour patterns do not diagnose species: quantitative evaluation of a DNA barcoded cryptic bumblebee complex. *PLoS One*, **7**, e29251.
- Claudet J., Pelletier D., Jouvenel J.Y., Bachel F., Galzin R. (2006) Assessing the effects of Marine Protected Area (MPA) on a reef fish assemblage in a northwestern Mediterranean marine reserve: identifying community-based indicators. *Biol. Conserv.* **130**, 346-369.
- Coppée A., Terzo M., Valterova I., Rasmont P. (2008) Intraspecific variation of the cephalic labial gland secretions in *Bombus terrestris* (L.) (Hymenoptera: Apidae). *Chem. Biodivers.* **5**, 2654–2661.
- Cruaud A., Gautier M., Galan M., Foucaud J., Sauné L., Genson G., Dubois E., Nidelet S., Deuve T. and Rasplus J.-Y. (2014) Empirical Assessment of RAD Sequencing for Interspecific Phylogeny. *Mol. Biol. Evol.* **31**, 1272–1274.
- Dellicour S., Lecocq T. (2013a) GCALIGNER 1.0 and GCKOVATS 1.0 – Manual of a Software Suite to Compute a Multiple Sample Comparison Data Matrix from Eco-chemical Datasets Obtained by Gas Chromatography. University of Mons, Mons.
- Dellicour S., Lecocq T. (2013b) GCALIGNER 1.0: an alignment program to compute a multiple sample comparison data matrix from large eco-chemical datasets obtained by GC. *J. Sep. Sci.* **36**, 3206–3209.
- De Meulemeester T., Gerbaux P., Boulvin M., Coppee A., Rasmont P. (2011) A simplified protocol for bumble bee species identification by cephalic secretion analysis. *Insectes Sociaux* **58**, 227–236.
- De Queiroz K. (2007) Species concepts and species delimitation. *Syst. Biol.* **56**, 879–886.
- Dufrene M., Legendre P. (1997) Species assemblages and indicator species: The need for a flexible asymmetrical approach. *Ecol. Monogr.* **67**, 345–366.
- Engel M.S. (2011) Systematic melittology: where to from here? *Syst. Entomol.* **36**, 2–15.
- Eriksen B., Molau U., Svensson M. (2006) Reproductive strategies in two arctic *Pedicularis* species (Scrophulariaceae). *Ecography*, **16** (2), 154-166
- Glaser N., Frérot B., Leppik E., Monsempes C., Capdevielle-Dulac C., Le Ru B., Lecocq T., Harry M., Jacquin-Joly E., Calatayud P.-A. J. (2014) Similar Differentiation Patterns Between PBP Expression

- Levels and Pheromone Component Ratios in Two Populations of *Sesamia nonagrioides*. *Chem. Ecol.* **40**, 923–927.
- Goulson D. (2010) *Bumblebees: Behaviour Ecology, and Conservation*. Oxford University Press, Oxford, 336 p.
- Heinrich B. (2005) *Bumblebee economics*. Harvard University Press, Cambridge, London, England, 245p.
- Hovorka O., Valterová I., Rasmont P., Terzo M. (2006) Male cephalic labial gland secretions of two bumblebee species of the subgenus *Cullumanobombus* (Hymenoptera: Apidae: *Bombus* Latreille) and their distribution in Central Europe. *Chem. Biodivers.* **3**, 1015–1022.
- Kevan P.G. (1973) Flowers, insects, and pollination ecology in the Canadian high Arctic. *Polar Record*, **16**, 667–674.
- Klein A.M., Vaissière B.E., Cane J.H., Steffan-Dewenter I., Cunningham S.A., Kremen C., Tscharntke T. (2007) Importance of pollinators in changing landscapes for world crops. *Proc. R. Soc. B Biol. Sci.* **274**, 303–313.
- Kullenberg B., Bergström G., Stallberg-Stenhagen S. (1970) Volatile Components of the Cephalic Marking Secretion of Male Bumble Bees. *Acta Chem. Scand.* **24**, 1481.
- Lecocq T., Dellicour S., Michez D., Dehon M., Dewulf A., De Meulemeester T., Brasero N., Valterová I., Rasplus J.Y., Rasmont P. (2015) Methods for species delimitation in bumblebees (Hymenoptera, Apidae, *Bombus*): towards an integrative approach. *Zool. Scripta* **44**, 281–297.
- Löfstedt C. (1993) Moth pheromone genetics and evolution. *Philos. Trans. R. Soc. London Ser. B: Biol. Sci.* **340**, 167–177.
- Løken A. (1973) Studies on Scandinavian bumble bees Hymenoptera, Apidae. *Norsk Entomologisk Tidsskrift* **20**, 1–218.
- Martinet B., Lecocq T., Smet J. and Rasmont P. (2015) A Protocol to Assess Insect Resistance to Heat Waves, Applied to Bumblebees (*Bombus Latreille*, 1802). *PLoS ONE*, **10**(3), e0118591.
- Martinet B., Lecocq T., Brasero N., Biella P., Urbanová K., Valterová I., Cornalba M., Gjershaug J.O., Michez D., Rasmont P. (2018) Following the Cold: Geographic Differentiation between Interglacial Refugia and Speciation in Arcto-Alpine Species Complex *Bombus monticola* (Hymenoptera: Apidae). *Syst. Entomol.* **43**, 200–217.
- Matoušková P., Luxová A., Matoušková J., Jiroš P., Svatoš A., Valterová I., Pichová I. (2008) A $\Delta 9$ desaturase from *Bombus lucorum* males: Investigation of biosynthetic pathway of marking pheromones. *ChemBioChem*, **9**, 2534–2541.
- Michener, C.D. (2007) *The Bees of the World. Second Edition*. Johns Hopkins University, Baltimore, 1016 pp.
- Milliron H.E., Olivier D.R. (1966) Bumblebees from Northern Ellesmere Island, with observations on usurpations by *Megabombus hyperboreus* (Schönherr) (Hymenoptera: Apidae). *Can. Entomol.* **98**, 207–213.
- Milliron H.E. (1973) A monograph of the western hemisphere bumblebees (Hymenoptera: Apidae: Bombinae). II. The genus *Megabombus* subgenus *Megabombus*. *Mem. Ent. Soc. Can.* **89**, 81–237.
- Mutanen M., Kivelä S.M., Vos R.A., Doorenweerd C., Ratnasingham S., Hausmann A., Huemer P., Dincă V., van Niekerken E.J., Lopez-Vaamonde C., Vila R., Aarvik L., Decaëns T., Efetov K.A., Hebert P.D., Johnsen A., Karsholt O., Pentinsaari M., Rougerie R., Segerer A., Tarmann G., Zahiri R., Godfray H.C. (2016) Species-Level Para- and Polyphyly in DNA Barcode Gene Trees: Strong Operational Bias in European Lepidoptera. *Syst. Biol.* **65**, 1025–1040.
- Oksanen J., Blanchet F.G., Friendly M., Kindt R., Legendre P., McGinn D., Minchin P.R., O'Hara R.B., Simpson G.L., Solymos P., Stevens M.H.H., Szoecs E., Wagner H. (2017) Tertiary Vegan: Community Ecology Package. <https://cran.r-project.org/web/packages/vegan/index.html>. Accessed 5 Oct 2018.
- Paradis E., Claude J., Strimmer K. (2004) APE: analyses of phylogenetics and evolution in R language. *Bioinformatics*, **20**, 289–290.
- Patiny S., Michez D., Rasmont P. (2009) A survey and review of the status of wild bees in the West-Palaeartic region. *Apidologie*, **40**, 313–331.
- Paterson H.E.H. (1993) *Evolution and the Recognition Concept of Species*. The Johns Hopkins University Press, Baltimore, Maryland.
- Pekkarinen A., Teräs I., Viramo J., Paatela J. (1981) Distribution of bumblebees (Hymenoptera, Apidae: *Bombus* and *Psithyrus*) in eastern Fennoscandia. *Notulae Entomologicae* **61**, 71–89.
- Potapov G.S., Kondalov A.V., Spitsyn V.M., Filippov B. Yu., Kolosova Yu S., Zubrii N.A., Bolotov I.N. (2018) An integrative taxonomic approach confirms the valid status of *Bombus glacialis*, an endemic bumblebee species of the High Arctic. *Polar Biology*, **41**: 629–642.
- Rasmont P., Franzen M., Lecocq T., Harpke A., Roberts S.P.M., Biesmeijer K., Castro L., Cederberg B., Dvorak L., Fitzpatrick U., Gonthier Y., Haubruge E., Mahé G., Manino A., Michez D., Neumayer J., Odegaard F., Paukkunen J., Pawlikowski T., Potts S.G., Reemer M., Settele J., Straka J. and Schweiger O. (2015) Climatic risk and distribution atlas of European bumblebees. *BioRisk*, **10**, 1–236.
- R Development Core Team. (2017) R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna. [WWW document]. URL <http://www.R-project.org/> [accessed on 17 October 2017].
- Richards O.W. (1931) Some notes on the humble-bees allied to *Bombus alpinus*, L. *Tromsø Museums Årshefter*, **50**, 1–32.
- Richards K.W. (1973) Biology of *Bombus polaris* Curtis and *B. hyperboreus* Schönherr at lake Hazen, north-west territories (Hymenoptera: Bombini). *Questiones entomologicae* **9**, 115–57.
- Roelofs W.L., Liu W., Hao G., Jiao H., Rooney A.P., Linn Jr. C.E. (2002) Evolution of moth sex pheromones via ancestral genes. *Proc. Natl. Acad. Sci. U.S.A.* **99**, 13621–13626.

- Schlick-Steiner B.C., Steiner F.M., Seifert B., Stauffer C., Christian E., Crozier R.H. (2010) Integrative taxonomy: a multisource approach to exploring biodiversity. *Annu. Rev. Entomol.* **55**, 421–438.
- Shamurin V.F. (1966) Rol' nasekomikh-opilitelei v tundrovikh soobshchestvakh. [The role of insects in tundra communities]. *Organizmi iprirodnayasreda. Voprosi geographii*
- Skorikov A.S. (1914) Les formes nouvelles des bourdons (Hymenoptera, Bombidae). VI. Russkoe éntomologicheskoe Obozrénie *14*, 119–129.
- Soltani G.G., Bénon D., Alvarez N., Praz J.C. (2017) When different contact zones tell different stories: putative ring species in the *Megachile concinna* species complex (Hymenoptera: Megachilidae). *Biol. J. Linn. Soc.* **121(4)**, 815–832.
- Suzuki R., Shimodaira H. (2011) Pvcust: Hierarchical Clustering with P-values via Multiscale Bootstrap Resampling. Contributed package. Version 1-1.10. R Foundation for Statistical Computing, Vienna. [WWW document]. URL <http://www.R-project.org> [accessed on 17 October 2017].
- Stenström M., Bergman P. (1998) Bumblebees at an alpine site in northern Sweden: temporal development, population size, and plant utilization. *Ecography* **21(3)**, 306–316.
- Svensson B.G. (1980) Species-isolating mechanisms in male bumblebees (Hymenoptera, Apidae). *Acta Universitatis Upsaliensis* **549**, 1–42.
- Svensson B.G., Bergström G. (1979) Marking pheromones of *Alpinobombus* males. *J. Chem. Ecol.* **5**, 603–615.
- Svensson B.G., Appelgren M., Bergstrom G. (1984) Geranylgeranyl acetate and 2 heptadecanone as the dominant marking secretion components of the labial glands in the bumblebee *Alpigenobombus wurfleini* Nov. *Acta Regiae Soc. Sci.* **3**, 145–148.
- Symonds M. R. E. and Elgar M. A. (2004) The mode of pheromone evolution: evidence from bark beetles. *Proc. R. Soc. Lond. B.* **271**, 839–846.
- Symonds M. R. E. and Elgar M. A. (2007) The evolution of pheromone diversity. *Trends Ecol. Evol.* **23**, 220–228.
- Symonds M.R.E., Moussalli A., Elagar M.A. (2009) The evolution of sex pheromones in an ecologically diverse genus of flies. *Biol J Linn Soc* **9**, 594–603.
- Taylor K., Rumsey F.J. (2003). *Bartsia alpina* L. *J. Ecol.* **91(5)**, 908–921.
- Terzo M., Urbanova K., Valterova I., Rasmont P. (2005). Intra and interspecific variability of the cephalic labial glands' secretions in male bumblebees: the case of *Bombus (Thoracobombus) ruderarius* and *B. (Thoracobombus) sylvorum* Hymenoptera, Apidae. *Apidologie*, **36**, 85–96.
- Trunz V., Packer L., Vieu J., Arrigo N., Praz C.J. (2016) Comprehensive phylogeny, biogeography and new classification of the diverse bee tribe Megachilini: Can we use DNA barcodes in phylogenies of large genera? *Mol. Phylogenetics Evol.* **103**, 245–259.
- Vincenti M., Guglielmetti G., Cassani G., Tonini C. (1987) Determination of Double Bond Position in Diunsaturated Compounds by Mass Spectrometry of Dimethyl Disulfide Derivatives. *Anal. Chem.* **59**, 694–699.
- Williams P.H., Cameron S.A., Hines H.M., Cederberg B., Rasmont P. (2008) A simplified subgeneric classification of the bumblebees (genus *Bombus*). *Apidologie*, **39**, 46–74.
- Williams P.H., Brown M.J.F., Carolan J.C., An J., Goulson D., Aytekin A.M., Best L.R., Byvaltsev A.M., Cederberg B., Dawson R., Huang J., Ito M., Monfared A., Raina R.H., Schmid-Hempel P., Sheffield C.S., Sima P., Xie Z. (2012) Unveiling cryptic species of the bumblebee subgenus *Bombus* s. str. worldwide with COI barcodes (Hymenoptera: Apidae). *Syst. Biodivers.* *10(1)*, 21–56.
- Williams P.H., Byvaltsev A.M., Cederberg B., Berezin M.V., Ødegaard F., Rasmussen C., Richardson L.L., Huang J., Sheffield C.S., Williams S.T. (2015) Genes suggest ancestral colour polymorphisms are shared across morphologically cryptic species in arctic bumblebees. *PLoS One*, **10(2)**, e0144544.
- Williams P.H., Cannings S.G., Sheffield C.S. (2016) Cryptic subarctic diversity: a new bumblebee species from the Yukon and Alaska (Hymenoptera: Apidae). *J. Nat. Hist.* **50 (45–46)**, 2881–2893.
- Wyatt T.D. (2003) Pheromones and animal behaviour: communication by smell and taste. Cambridge University Press, Cambridge.
- Žacek P., Prchalova-Homakova D., Tykva R., Kindl J., Vogel H., Svatoš A., Pichová I., Valterová I. (2013) De novo biosynthesis of sexual pheromone in the labial gland of bumblebee males. *ChemBioChem*, **14**, 361–371.
- Žáček P., Kalinová B., Šobotník J., Hovorka O., Ptáček V., Coppée A., Verheggen F., Valterová I. (2009) Comparison of age-dependent quantitative changes in the male labial gland secretion of *Bombus terrestris* and *Bombus lucorum*. *J. Chem. Ecol.* **35**, 698–705.