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Corresponding Authors:
Francisco J. Quintero, IBVF-CSIC, fjquintero@ibvf.csic.es
José M. Pardo, IBVF-CSIC, jose.pardo@csic.es

A Critical Role of Sodium Flux via the Plasma Membrane Na\(^+\)/H\(^+\) Exchanger SOS1 in the Salt Tolerance of Rice

Houda El Mahi\(^{1,11}\), Javier Pérez-Hormaeche\(^{2,11}\), Anna De Luca\(^{2,11}\), Irene Villalta\(^{3,9}\), Joaquín Espartero\(^{1}\), Francisco Gámez-Arjona\(^{4}\), José Luis Fernández\(^{5}\), Mireia Bundo\(^{6}\), Imelda Mendoza\(^{2}\), Delphine Mieulet\(^{7}\), Eric Lalanne\(^{8}\), Sang-Yeol Lee\(^{9}\), Dae-Jin Yun\(^{10}\), Emmanuel Guiderdoni\(^{7}\), Manuel Aguilar\(^{5}\), Eduardo O. Leidi\(^{1}\), José M. Pardo\(^{2,12}\), Francisco J. Quintero\(^{2,12}\)

1. Instituto de Recursos Naturales y Agrobiología (IRNAS), Consejo Superior de Investigaciones Científicas (CSIC), 41012 Seville, Spain.
2. Instituto de Bioquímica Vegetal y Fotosíntesis (IBVF), Consejo Superior de Investigaciones Científicas (CSIC) and University of Seville, 41092 Seville, Spain.
3. Institut de Recherche sur la Biologie de l’Insecte, Université de Tours, Parc de Grandmont, 37200 Tours, France
4. Swiss Federal Institute of Technology in Zurich, Zurich CH-8092, Switzerland
5. Instituto de Investigación y Formación Agraria y Pesquera (IFAPA), Junta de Andalucía, 41200 Seville, Spain.
6. Centre for Research in Agricultural Genomics (CRAG), CSIC-IRTA-UAB-UB, Campus Universitat Autonoma de Barcelona (UAB), 08193 Barcelona, Spain.
7. Centre for International Cooperation on Agricultural Research for Development (CIRAD), Joint Research Unit of Genetic Improvement and Adaptation of Mediterranean and Tropical Plants (UMR-AGAP), 34398 Montpellier, and Université de Montpellier, 34000 Montpellier, France.
Author's Contributions

HEM, AL, JPH, IV, FGA, JE, JLF, IM, DM, EOL, and FJQ performed the experimental work; EL, JPH, IV, and JMP did the transcriptomic analyses; IV, SYL, DJY, EG, MA, EOL, JMP, and FJQ designed the research; all authors contributed to the discussion and interpretation of results; JPH, JMP, and FJQ wrote the article; IV, MB, EG, and EOL revised and edited the manuscript.

One-sentence Summary

The plasma membrane Na⁺/H⁺ exchanger SOS1 plays a major role in the salt tolerance of rice by controlling Na⁺ uptake and root-shoot partitioning.
ABSTRACT

Rice (Oryza sativa) stands among the world's most important crop species. Rice is salt-sensitive and the undue accumulation of sodium ions (Na\(^+\)) in shoots has the strongest negative correlation with rice productivity under long-term salinity. The plasma membrane Na\(^+\)/H\(^+\) exchanger protein SOS1 is the sole Na\(^+\) efflux transporter that has been genetically characterized to date. Here, the importance of SOS1-facilitated Na\(^+\) flux in the salt tolerance of rice was analyzed in a reverse-genetics approach. A sos1 loss-of-function mutant displayed exceptional salt sensitivity that was correlated with excessive Na\(^+\) intake and impaired Na\(^+\) loading into the xylem, thus indicating that SOS1 controls net root Na\(^+\) uptake and long-distance Na\(^+\) transport to shoots. The acute Na\(^+\) sensitivity of sos1 plants at low NaCl concentrations allowed analysis of the transcriptional response to sodicity stress without effects of the osmotic stress intrinsic to high-salinity treatments. In contrast to that in the wild type, sos1 mutant roots displayed preferential down-regulation of stress-related genes in response to salt treatment, despite the greater intensity of stress experienced by the mutant. These results suggest there is impaired stress detection or an inability to mount a comprehensive response to salinity in sos1. In summary, the plasma membrane Na\(^+\)/H\(^+\) exchanger SOS1 plays a major role in the salt tolerance of rice by controlling Na\(^+\) homeostasis and possibly contributing to the sensing of sodicity stress.

KEYWORDS

Rice, sodium transport, SOS1, salinity signaling
INTRODUCTION

Rice (Oryza sativa) is the most important salt-sensitive cereal crop (Hoang et al., 2016). Rice is grown in a diverse range of environments with varying soil-water conditions. Salinity is an intrinsic problem in coastal areas and river deltas where rice farming predominates, but is also a worsening problem in inland areas due to the buildup of secondary salinization of the soil surface as a consequence of excessive use of irrigation with poor quality water and insufficient drainage (Thomson et al., 2010). Excessive amounts of salts in the soil solution cause substantial declines in the productivity of many agricultural plants, and in rice as little as 3.5 dS/m (2.1 g/L of salts) is sufficient to curtail 10% of yield, and 7.2 dS/m (4.3 g/L) producing 50% yield loss (Munns and Tester, 2008; Hoang et al., 2016).

Salinity poses two main challenges to plants. One is osmotic stress due to excess solutes outside the roots that reduce the ability of plants to extract soil water, which affects cell turgor and expansion immediately. Secondly, the ionic stress that is most often caused by excessive influx of Na⁺ ions into the plant leads to interruption of metabolic processes (Hasegawa et al., 2000; Munns and Tester, 2008). Physiological and genetic studies indicate that a range of traits, such as low shoot Na⁺ content, preservation of the K⁺ status of the whole plant, vacuolar compartmentation of salts in older leaves, intrinsic cellular tolerance, and plant vigor, could incrementally improve the ability of rice plants to cope with salinity (Yeo et al., 1990; Thomson et al., 2010). Integration of all these physiological mechanisms for salinity tolerance by pyramiding superior alleles controlling different processes is the long-term goal to increase the level of salt tolerance in rice (Gregorio et al., 2002).

Sustained plant growth under salt stress requires tight control of Na⁺ uptake, Na⁺ redistribution among organs by long-distance transport, and Na⁺ accumulation into cell vacuoles for detoxification and osmotic balance (Munns and Tester, 2008). The ability to manage Na⁺ content has partly a cellular basis since rice protoplasts derived from cultivars with contrasting salt tolerance showed that halotolerance depended on reduced Na⁺ uptake and a fast compartmentation of Na⁺ into vacuoles (Kader and...
Lindberg, 2005), whereas enhancement of Na⁺ efflux yielded salt-tolerant calli (Zhu and Wu, 2008). At the whole-plant level, comparative studies have shown that salinity-tolerant rice cultivars accumulate less Na⁺ in leaves and shoots compared to that in salinity-sensitive rice cultivars (Golldack et al., 2003; Lee et al., 2003; Lin et al., 2004; Ren et al., 2005), and that shoot Na⁺ accumulation had the strongest negative correlation with the performance of contrasting rice genotypes under long-term salinity (Coskun et al., 2013). Hence, the quantitative measure of the Na⁺ and K⁺ concentrations in the shoot in salinized conditions has been often employed as the trait screened in quantitative trait loci (QTL) mapping and breeding programs (Hoang et al., 2016). Known proteins governing the partitioning of Na⁺ between roots and shoots are members of the so-called High-affinity Potassium Transporters (HKT), albeit they have repeatedly been shown to behave as Na⁺ transporters mediating either channel-like Na⁺ uniport (class I or HKT1 group) or Na⁺-K⁺ symport (class II or HKT2 group). Generally, dicot species have only a few HKT genes in their genomes preferably encoding class-I HKT proteins, whereas monocots have multiple HKT genes of both classes. The rice protein OsHKT1;5/SCK1 and the wheat proteins TmHKT1;4/ and TmHKT1;5, all of them class-I proteins, mediate the retrieval of Na⁺ from the xylem sap, thereby restricting the transfer of Na⁺ to shoots. Rice OsHKT1;4 also restricts leaf sheath-to-blade Na⁺ transfer under salinity stress (Cotsaftis et al., 2012; Suzuki et al., 2016).

Although several candidate proteins have been suggested, no major plasma membrane transporters responsible for the bulk influx of Na⁺ into root cells have as yet been conclusively identified at the genetic level and it seems likely that multiple transport proteins and systems incrementally contribute to Na⁺ influx (Kronzucker and Britto, 2011; Nieves-Cordones et al., 2016). In rice, the HKT2;1 protein that is expressed in the root cortex and endodermis may provide an entry pathway for 'nutritional' Na⁺ uptake under K⁺ limitation to support cell expansion and plant growth (Garcia-deblas et al., 2003; Horie et al., 2007). Sodium may also leak into the root stele of rice plants through anatomical discontinuities in the endodermis produced by the emergence of secondary roots (Yeo et al., 1987), or through the lateral roots themselves (Faiyue et al., 2010). For Na⁺ transport in the opposite (efflux) direction,
only the SOS1 transporter (Salt Overly-Sensitive 1) has been genetically characterized (Shi et al., 2002; Oh et al., 2009; Olias et al., 2009). In dicots, SOS1 controls net Na\(^+\) uptake by roots and long-distance transport to shoots (Oh et al., 2009; Olias et al., 2009). Thus, the opposing yet concerted activity of SOS1 in xylem loading and of HKT1-like proteins in xylem unloading might determine the amount of Na\(^+\) that is eventually exported from roots to shoots. Although HKT proteins of cereal crops is well established to play a major role in their salt tolerance, the contribution of Na\(^+\) loading into the xylem by specific transporters remains to be determined. Likewise, the regulatory cross-talk of these complementary transport systems has not been explored (Ji et al., 2013).

Little is known about the regulation of HKT proteins. HKT gene expression is often modulated by K\(^+\) availability and soil salinity, but the post-transcriptional regulation of HKT transporters remains entirely unknown. In rice, the Na\(^+\) uptake gene \textit{HKT2;1} is rapidly repressed in roots by as little as 30 mM NaCl to curtail Na\(^+\) entry (Horie et al., 2007). Surprisingly, \textit{HKT1;4} expression is also down-regulated proportionally to the severity of salinity treatment during the vegetative growth stage, and RNAi plants showed no differences in salt sensitivity or in Na\(^+\) content (Suzuki et al., 2016). A contribution of \textit{HKT1;4} towards the prevention of Na\(^+\) accumulation in flag leaf blades was found in the reproductive stage only (Suzuki et al., 2016). \textit{OsHKT1;1}, involved in Na\(^+\) re-circulation, is expressed mainly in the phloem of leaf blades and up-regulated by salt stress (Wang et al., 2015). The transcription factor OsMYBc binds to the \textit{OsHKT1;1} promoter and loss of OsMYBc resulted in a reduction in NaCl-induced expression of \textit{OsHKT1;1} and salt sensitivity.

More progress has been made towards the understanding of SOS1 regulation. The SOS1 gene is up-regulated by salinity (Shi et al., 2003; Martinez-Atienza et al., 2007) and, in \textit{Arabidopsis thaliana}, the mRNA is stabilized by an ill-defined process mediated by reactive oxygen species (ROS) (Chung et al., 2008). The transport activity of SOS1 is enhanced by protein phosphorylation that counteracts autoinhibition (Quintero et al., 2002; Quintero et al., 2011). The minimal SOS functional module in both dicots and monocots comprises proteins SOS1, SOS2/CIPK24 and SOS3/CBL4 (Quintero et al.,...
SOS2/CIPK24 is a protein kinase that belongs to the SnRK3 family (sucrose non-fermenting-1-related protein kinase-3). SOS3 is a myristoylated Ca\(^{2+}\)-binding protein that likely perceives the increase in cytosolic Ca\(^{2+}\) elicited by excess Na\(^+\) and recruits SOS2 to the plasma membrane to achieve phosphorylation and activation of SOS1 (Quintero et al., 2002; Quintero et al., 2011). In Arabidopsis, SOS3 functions primarily in roots whereas the related protein SCaBP8/CBL10 operates by a similar mechanism in shoots (Quan et al., 2007). CBL10 may also regulate Na\(^+\) uptake into the vacuole (Kim et al., 2007).

The rice complement of SOS proteins has been characterized by heterologous expression in yeast and Arabidopsis (Martinez-Atienza et al., 2007), but a precise description of their role in the physiology and salinity stress management of rice plants has not been achieved. Here we show that a sos1 loss-of-function mutant displays exceptional salt sensitivity that correlates with excessive Na\(^+\) intake and impaired Na\(^+\) loading into the xylem. We took advantage of the acute sensitivity of sos1 plants to Na\(^+\) to inspect the sodicity-stress transcriptome with little interference by the osmotic challenge intrinsic to high salinity treatments. Surprisingly, sos1 mutant roots showed a marked down-regulation of genes despite the greater intensity of the stress they suffered, which suggested impaired stress detection or an inability to mount a comprehensive response to salinity stress in the absence of SOS1.
RESULTS

Selection of mutant lines

Public repositories of rice mutant lines were surveyed for entries with putative T-DNA or Tos17 insertions in genes SOS1 (Os12g44360), CIPK24/SOS2 (Os06g40370) and CBL4/SOS3 (Os05g45810). Mutants in the Nipponbare genetic background were preferred because this temperate japonica cultivar enabled the lines of interest to complete their life cycle in our experimental paddies. Mutant line codes, annotated insertion points of mutagenic DNA, and insertions experimentally confirmed by diagnostic PCR and sequencing of amplicons are listed in Table 1. Detailed description of the molecular characterization of all insertional lines tested herein is given in the Supplemental Information file. As contrasting materials for mutant phenotyping, congeneric plants in which the mutagen was segregated out (null segregants) were kept as controls for subsequent experimentation.

Out of seven mutant lines tested for putative mutagenic insertions at SOS1 (Piffanelli et al., 2007), only AVIB03 carrying an insertion of Tos17 at intron 19 was confirmed not to produce a wild-type SOS1 mRNA (Supplemental Figures S1 to S4). Molecular analyses demonstrated that homozygous plants of this lineage produced a truncated mRNA whose elongation was interrupted by the transposon and that, in contrast to that in other lines (Supplemental Figures S1 to S4), salinity treatment did not promote the correct processing of the mRNA and eviction of the Tos17 insertion (Figure 1B). Of note is the significant induction of SOS1 mRNA by salinity in Nipponbare plants. Sequencing of the RT-PCR amplicon with the intervening Tos17 (Supplemental Figure S4D) demonstrated that transposon insertion in AVIB03 had occurred after nucleotide 9802, relative to the ATG initiation codon in the genomic sequence of SOS1. The in silico read-through translation of the putative mRNA predicted the synthesis of a SOS1 protein with a frame-shift after residue K767 and the addition of a short C-terminal extension of sequence VCFCFKTTLSH. This truncated protein was predicted inactive based on the removal of essential amino acids and functional domains of the wild-type protein that are downstream the truncation point (Figure 1A) (Quintero et al., 2011;
Nuñez-Ramirez et al., 2012). To demonstrate this point, a synonymous mutation was created in the rice SOS1 cDNA by introducing a stop codon after residue K767. Expression of this truncated SOS1 protein (SOS1-Δ767) in the yeast strain AXT3K that is devoid of all endogenous Na⁺ transporters and is salt-sensitive (Quintero et al., 2002) failed to restore salt tolerance upon the co-expression of SOS2 and SOS3 proteins of Arabidopsis (Figure 1C). We have shown elsewhere the competence of the Arabidopsis
SOS2/SOS3 complex to activate the rice SOS1 protein (Martinez-Atienza et al., 2007), and Figure 1C illustrates this fact. By contrast, the rice SOS1 protein with a C-terminal truncation after residue E970 yielded a hyperactive and SOS2/SOS3-independent exchanger (Figure 1C). Truncation at residue E970 in the rice SOS1 protein mimics the Arabidopsis mutant protein SOS1-Δ998, which lacks the autoinhibitory domain of SOS1 that is targeted by the SOS2/SOS3 kinase complex (Quintero et al., 2011).

In accordance with the yeast data demonstrating a defective SOS1 protein in line AVIB03, plants of that line showed a remarkable salt-sensitive phenotype in hydroponic culture that correlated with much greater Na⁺ contents in shoots and roots (Figure 2). Representative pictures of sos1 plants at different time points after stress imposition (25, 50 and 75 mM NaCl) are shown in Figure 2A. The sos1 mutant plants showed growth retardation as early as 5 d after treatment with 50 and 75 mM NaCl. After 10 d, sos1 plants displayed severe symptoms of stress also at 25 mM NaCl, the lowest salt concentration tested. The experiment was terminated after 24 d when all sos1 plants had died irrespective of the salt concentration. Quantitative growth data at 7 d after transfer to 50 mM NaCl confirmed the salt-sensitivity of homozygous sos1 plants compared to azygous and wild-type Nipponbare controls (Figure 2B). At that point, sos1 roots were severely dehydrated and after 14 d sos1 shoots had bleached. Salt sensitivity correlated with greater Na⁺ contents in roots and shoots of the sos1 mutant.

The SOS1 protein activity accounts for ca. 60% of the Na⁺/H⁺ exchange measured in plasma membrane vesicles of Arabidopsis (Qiu et al., 2003). Measurement of Na⁺/H⁺ exchange in purified plasma membrane fractions (Supplemental Figure S5) demonstrated 39% reduction in exchange activity in vesicles derived from the AVIB03 line relative to that in the wild-type plants. Quenching of fluorescence units per mg of protein per minute was 34.73 ± 2.56 and 56.74 ± 11.34, respectively (p=0.107 by One-Way ANOVA test, n=4 technical replicas in two different assays). Together, these results signify that line AVIB03 is deficient in SOS1 activity and hence this lineage was selected for further experimentation.
We next searched for mutants in the rice genes *CIPK24* and *CBL4*, which we have previously described as functional homologues of Arabidopsis *SOS2*(*CIPK24*) and *SOS3*(*CBL4*) (Martinez-Atienza et al., 2007). Mutant lines in the Nipponbare background AMHC11, AMHC12 and AMHE12 were annotated to carry T-DNA insertions 84 nucleotides upstream of the translation initiation codon of locus Os06g40370 (*CIPK24*) (Sallaud et al., 2004). Molecular characterization confirmed identical T-DNA insertion points at the 5'-UTR of *CIPK24* in these lines (Supplemental
Figure S6). Homozygous plants produced a mature CIPK24 mRNA carrying the T-DNA insertion and no wild-type mRNA was detected. Mutant seedlings were sensitive to NaCl in hydroponic culture. Supplemental Figure S6 illustrates the end-point growth of homozygous cipk24 plants (L24H), azygous CIPK24 controls (L24A), and wild-type Nipponbare plants after 7 d in hydroponic medium supplemented with 40 and 80 mM NaCl. These data indicate that the rice kinase CIPK24, which has been shown to activate SOS1 (Martinez-Atienza et al., 2007), also plays a role in the salt tolerance of rice.

At the time this research was conducted, there were no available mutants with mutagenic insertions in the coding region of gene CBL4 (Os05g45810) in the Nipponbare background. Line AJBC12 was annotated to bear a T-DNA insertion right downstream the 3'-UTR of CBL4 (Sallaud et al., 2004). RT-PCR analyses with T2 plants of this lineage showed that the CBL4 transcript was not affected by the adjacent T-DNA insertion and no further experimentation was done with this line.

Salt tolerance assays in rice paddies

To examine in detail the relevance of the SOS system in the salt tolerance of cultivated rice, we next sought to define the performance of sos1 and cipk24 mutant plants in near-agronomical conditions in experimental rice paddies. Homozygous mutant lines of genotype sos1 and cipk24 (denoted henceforth as L1H and L24H, respectively) were compared to congenic segregant lines selected during the genotyping as lacking the mutagenic insertion (denoted here as azygous segregant lines L1A and L24A), and to control plants of the Nipponbare background. In lines L1H and L1A the T-DNA of the original transformation was also segregated out. Four-week-old plants were transferred to paddies irrigated with water from an underground well whose salinity content was on average 0.42 g/L, of which Na\(^+\) was at 5.09 mM, K\(^+\) at 0.14 mM, and Ca\(^{2+}\) at 4.0 mM. For the saline treatment, underground water was supplemented with seawater salts to a final concentration of 1.8 g/L salts during irrigation to mimic frequent salinity episodes in rice farms in Spain. The final concentrations of Na\(^+\), K\(^+\) and Ca\(^{2+}\) in the salinized irrigation water were 27.6, 0.24 and 4.1 mM, respectively.
Chloride was 30.5 mM. As depicted in Figure 3 and Supplemental Figure S7, sos1 plants showed statistically significant growth retardation even at the low salt content of irrigation water, conditions in which the Na⁺ content in sos1 shoots was low but still triple that of the congenic SOS1 control and the wild type Nipponbare (0.035% vs 0.01% of dry matter, respectively). Growth arrest of salinized sos1 plants at 1.8 g/L salt correlated with massive Na⁺ accumulation in shoots and notable K⁺ loss (Figure 3). The cipk24 mutant plants showed a modest growth reduction in these conditions that was statistically significant only at $p = 0.095$ ($n=10$) relative to that in the congenic azygous
control (Supplemental Figure S7), in agreement with the salt sensitivity that mutant cipk24 showed at early developmental stages in hydroponic culture at 80 mM NaCl but not at 40 mM (Supplemental Figure S6).

**Complementation and over-expression lines**

Because only one putative sos1 mutant line was available, we sought to confirm that the exceptional salt-sensitivity of AVIB03 plants was due to the lack of function of SOS1 and not to another uncharacterized mutation carried over through the mutant selection process. Genetically complemented lines were obtained by transformation of AVIB03 plants with the SOS1 cDNA under the control of the constitutive and moderately expressed UBI1 gene promoter of maize. Restoration of salt-tolerance was confirmed by scoring the growth of geneticin-resistant T1 plants (n=32), belonging to four independent transformation lines, in hydroponic culture with 50 and 75 mM NaCl for 3 weeks (Supplemental Figure S8A). This test showed that 13 out 16 plants regained tolerance to 50 mM NaCl, and that 12 out of 16 withstood the 75 mM NaCl treatment. None of the untransformed sos1 plants survived the assay (n=8). To further confirm the basis for the suppression of the salt-sensitive phenotype, the complemented line C11 that had a low expression level of the UBQ1:SOS1 transgene (Supplemental Figure S8B) was chosen for additional analyses. Two-week-old seedlings of wild type, sos1 mutant and transformed line C11 were transferred to 25 and 50 mM NaCl in Miyamoto's medium for 3 weeks, after which plant weight, and Na⁺ and K⁺ contents were determined. Results showed that shoot growth in saline media was restored to wild-type levels, whereas root growth lagged behind that of control plants (Figure 4). Moreover, Na⁺ content in C11 plants largely matched that in wild-type plants.

To test the potential of the SOS1 exchanger to increase the salt tolerance of rice, we also generated Nipponbare transgenic plants with expression of the SOS1 cDNA driven by the strong CaMV 35S gene promoter to create SOS1 overexpression. Transgenic plants showing high levels of transgene expression as determined by RT-PCR were selected and propagated to produce homozygous lines. Salt tolerance was evaluated in
hydroponic cultures with Miyamoto’s medium, which contains low levels of K⁺ (0.20...
mM) and Na⁺ (0.14 mM) and thus could maximize the impact of the ionic component of moderate salt stress (25–50 mM NaCl). Seven-d-old seedlings of two independent 35S:SOS1 lines were cultivated in the presence of 25 and 50 mM NaCl for 4 weeks (n=5 per genotype and treatment). Transgenic plants overexpressing SOS1 showed greater survival under salt treatments than that of control plants transformed with a mock.
empty vector (Figure 5). Plants that survived the treatment at 25 mM were assessed for dry weight and Na⁺ content (control plants surviving 50 mM NaCl had damaged roots and were discarded for further analyses; see Figure SA). Plants overexpressing \textit{SOS1} sustained a robust growth at 25 mM NaCl that correlated with lower Na⁺ contents in both shoots and roots compared to that in control plants (Figure 5C), as should be expected from the near ubiquitous expression of \textit{SOS1} driven by the 35S promoter and the consequently enhanced Na⁺ efflux in many cell types.

\textit{Expression pattern of SOS1}

In dicots, the SOS1 protein has been implicated in Na⁺ fluxes relevant to net ion uptake by roots and to xylem loading for root/shoot ion partition (Shi et al., 2002; Oh et al., 2009; Olias et al., 2009). To understand the critical role of SOS1 in the salt tolerance of rice plants, the expression pattern of the \textit{SOS1} gene was investigated in transgenic rice transformed with the \textit{SOS1} gene promoter (defined as the 3.9 kb genomic fragment upstream the initiation codon) fused to the ß-glucuronidase (\textit{GUS}) reporter gene. Four independent transgenic lines were assayed for GUS expression and they all produced consistent staining patterns, thereby ruling out artifacts by positional effects of the transgene. GUS expression was detected at all developmental stages tested and no significant differences in the expression pattern were found between control and saline conditions (50 mM NaCl, 24 h). Maximal GUS staining occurred in the vasculature, both in shoots and roots (Figure 6). In roots, longitudinal and transverse sections indicated a preferential expression in xylem parenchyma (Figure 6A-C). No expression was detected at the root apex or root hairs (Figure 6D,E). In secondary roots the staining was apparent only after the vascular bundle started to differentiate (Figure 6F-H). Likewise, the expression of \textit{SOS1:GUS} was located in vascular bundles in aerial tissues. In the coleoptile, staining was restricted to undifferentiated cells around the two vascular bundles (Figure 6I). In the flag leaf, all vascular bundles were stained, although the signal was stronger around older xylem vessels (Figure 6J,K). There was no detectable expression in stomata (Figure 6K). Thus, the preferential expression pattern of rice \textit{SOS1} recapitulated the pattern described in Arabidopsis, with the exception that the rice reporter gene \textit{SOS1:GUS} was not strongly expressed in root tips.
as reported for Arabidopsis plants using an equivalent approach (Shi et al., 2002). In conclusion, the expression pattern in rice suggested a role for SOS1 in Na⁺ xylem loading in addition to the cellular-based protection imparted by Na⁺ efflux.

*Function of SOS1 in Na⁺ efflux and xylem loading*

The preferential expression pattern of the SOS1 gene promoter in parenchyma cells associated to the vasculature suggested a function for SOS1 in xylem loading since, at physiological Na⁺ and H⁺ concentrations, a plasma membrane Na⁺/H⁺ exchanger likely mediates the energetically uphill Na⁺ efflux out of parenchyma cells. Scanning Electron
Microscopy coupled to Energy Dispersive X-ray microanalysis (SEM-EDX) was used to
determine the mineral composition of various root cell types in cross-sections of salt-
treated rice plants (Figure 7). In parallel, net tissue contents of Na\(^+\) and K\(^+\) were also
determined. Roots of the sos1 plants treated with 50 mM for 2 d in hydroponic culture
had a Na\(^+\) content that doubled the amount in the wild type (112.95 mM vs. 51.4 mM),
and showed a 37% reduction in the K\(^+\) content (10.0 mM vs. 15.8 mM). In agreement
with these values, the amount of Na\(^+\) measured by SEM-EDX in the sos1 mutant was
greater than that in the corresponding wild-type samples of all cell types tested (Figure
7). In wild-type roots, the maximal Na\(^+\) content was found in the cortex, with a
progressive decline in endodermis and xylem parenchyma cells. This profile is in
accordance with the selectivity filter imposed by the root endodermis (and buildup of
Na\(^+\) content in the cortex relative to that in the endodermis), and with the active
loading of Na\(^+\) ions into the xylem vessels after ions entered the stele (minimal Na\(^+\)
content in xylem parenchyma). By contrast, the Na\(^+\) distribution in the sos1 root
followed an inward-decreasing gradient signifying the unrestricted transit of Na\(^+\)
through all cell layers and the passive leak of Na\(^+\) into the xylem. Together, these
results highlight the critical role that the Na\(^+\)/H\(^+\) exchanger SOS1 plays in Na\(^+\) efflux and
xylem loading in rice plants.

On the other hand, the minimal K\(^+\) content in cortical cells in wild-type roots and the
increasing K\(^+\) values towards the xylem reflected the active uptake of K\(^+\) by
endodermal cells and its translocation towards the vasculature. A key parameter of the
physiological status of individual cells and whole plants under salt stress is the Na\(^+\)/K\(^+\)
ratio. Figure 7B shows that the ion-selective barrier imposed by the endodermis to the
centripetal movement of Na\(^+\) in the root caused a disproportionate Na\(^+\)/K\(^+\) ratio of
11.43 in cortex cells of wild-type roots, in stark contrast with the low 0.49 ratio
measured in xylem parenchyma cells presumably effected by the active Na\(^+\) loading in
xylem vessels. In accordance with the critical role of SOS1 in Na\(^+\) export to the xylem,
the Na\(^+\)/K\(^+\) ratio in the xylem parenchyma of the sos1 mutant escalated to 9.40, a 19-
fold increase relative to wild-type values. This change in the Na\(^+\)/K\(^+\) ratio between wild
type and the sos1 mutant signified the greatest difference among cell types. These
results are also evidence of the profound effects that a deficient Na\(^+\)-specific transport
system may produce on the K\(^+\) status of individual cells and tissues.
Transcriptome of the sos1 mutant

High salinity imposes both a water stress because the osmotic effect of the saline solution outside and an ionic imbalance resulting primarily from the accumulation of Na\(^+\) and the loss of K\(^+\) (Munns and Tester, 2008). Contrary to most analyses of the transcriptional response of plants to high-salinity, in which it is not possible to separate the genetic response to the osmotic and the ionic components of high salinity (Rabbani et al., 2003; Kanwar et al., 2014; Wang et al., 2016), the extreme Na\(^+\)-sensitivity of the sos1 mutant plant offered the unique advantage of inspecting the specific transcriptional response to sodicity stress at low external salt concentrations. Roots are the main defensive barrier against salinity and the organ that first senses this soil-derived stress (Munns and Tester, 2008). Hence, to identify genes responding primarily to the Na\(^+\) toxicity affecting sos1 plants, roots from mutant and control wild-type seedlings were harvested at the same time point (4 d) after a 75 mM NaCl treatment, and compared to roots harvested alongside from untreated seedlings. Total RNA was extracted and used for microarray experiments (see “Materials and Methods” for details).

Based on the standard criteria of greater than 2-fold changes with statistical significance at \(p<0.05\) in \(t\)-tests, we identified differentially regulated genes (DEGs) in sos1 mutant roots under salt and control conditions. Supplemental Table S1 online summarizes the genes up- and down-regulated in the mutant roots when compared to expression levels in the control line under the two treatments. Only 84 DEGs, representing 0.3% of total genes annotated, were detected between the two contrasting lines cultivated in control conditions. This result points to a minor role of SOS1 during the root development in the absence of sodicity stress. Of note is that HKT2;1 (Os06g0701700), encoding a transporter protein that mediates Na\(^+\) influx into K\(^+\)-starved roots and thereby improves growth (Horie et al., 2007), was the second most up-regulated DEG in the sos1 mutant without salinity treatment (Supplemental Table S1). By contrast, many more genes were differentially expressed in the mutant relative to that in the wild type when salt stress was imposed, in accordance with the
substantial impact of salinity on sos1 roots. Of the 5678 total DEGs identified in the
roots of the sos1 mutant under salinity, only 11 genes were commonly up-regulated and 18 were down-regulated in the sos1 mutant relative to expression levels in the wild type irrespective of the salt treatment (Figure 8), indicating that the transcriptional impact of the sos1 mutation was largely specific to the salt-induced response. The large number of DEGs (5633 genes, 19.48% of total) that presented an altered response in sos1 roots relative to that in wild-type roots under salt stress demonstrates the drastic effect of the sos1 mutation in the transcriptional response to sodicity. Notably, the majority (64%, 3608 genes) of these salt-responsive DEGs were down-regulated in the mutant root, which may represent a biased transcriptional response towards either repressed or less-induced transcription in the sos1 mutant in response to salt.

To evaluate the potential biological function of the differentially expressed genes in the sos1 mutant, Gene Ontology (GO) analysis was performed using the AgriGO online service (http://bioinfo.cau.edu.cn/agriGO) for the up-regulated, down-regulated and total DEGs identified in the salt-stressed or control-cultivated roots (Supplemental Table S2). The 5633 salt-responding DEGs fell into 129 significantly enriched GO categories corresponding to 66 biological processes, 28 functional categories, and 35 cellular components. The top over-represented GO categories included "cellular nitrogen compound metabolic process" (137 genes; False Discovery Rate (FDR) of 7.2x10^{-16}), and those related to the biological process "transport" (FDR=2.2x10^{-4}) (Supplemental Table S2). Further analysis showed again that these GO categories were constituted by far more down-regulated than up-regulated genes. This bias explained that most (n=115) of the 129 significant GO-terms were also enriched specifically in the group of the down-regulated DEGs (Figure 8), including the 25 top enriched GO categories (Supplemental Table S3). Moreover, 73 of the significant GO terms identified were only present among the enriched categories of the down-regulated DEGs. For example, among the 97 total DEGs classified under the term “ion transmembrane transporter activity” (GO:0015075), 79 of them were down-regulated DEGs. The GO cellular component "membrane" and "membrane part" ranked top among GO terms enriched in down-regulated DEGs.
The MapMan software (Thimm et al., 2004) was also used to assign genes to functional categories and to further analyze the metabolic pathways and cellular responses altered in sos1 roots. Over-representation analysis of the up- and down-regulated categories (Supplemental Table S4 and Supplemental Figure S9) indicated a general repression of the glycolysis, the tricarboxylic acid (TCA) cycle, nitrogen metabolism, amino acid synthesis, redox metabolism, protein targeting (secretory pathway) and protein degradation (ubiquitin proteasome). Among the genes related to “abiotic stress” responses, only the “drought/salt” and “unspecified” sub-categories were significantly different between the two genotypes, with an over-representation of unspecified stress genes among the up-regulated salt-responding DEGs. Together, these results support the notion that the transcriptional response of the sos1 mutant to salt involves a global alteration of the cell physiology due to preferential down-regulation of genes that otherwise would be part of the normal response of wild-type roots to salinity.

The majority of the DEGs showing the largest variations in transcript abundance corresponded to proteins of unknown function or with sequence similarity to other proteins but whose specific function remains to be determined. Moreover, the differential hybridization method used in microarrays only captures genes whose transcript abundance at sampling are substantially different between samples; genes whose transcriptional dynamics are of similar magnitude between samples are not detected even if they responded to the stimulus. This was best exemplified by CIPK24, which presumably was not identified as a DEG by the microarray data because this gene was similarly induced in wild-type and sos1 roots, as demonstrated by RT-qPCR (Figure 9). Thus, we analyzed in greater detail groups of genes encoding proteins with a known role in processes that are likely to be relevant to the ionic component of salinity stress, mainly Na\(^+\) and K\(^+\) transporters and their regulators CIPKs and CBLs. Among the 100 genes selected for this analysis (Supplemental Table S5), the rice SalT locus was the most transcriptionally induced gene in sos1 roots relative to that in control salt-treated roots (log\(_2\) FC = 4.79). Gene SalT, encoding a 15-kDa jacalin-related mannose-binding lectin, is located on a QTL responsible for the bulk of genetic variation in ion uptake under saline conditions at the seedling stage (Thomson et al.,
Likewise, *SRO1c* (*Os03g0230300*) was highly induced in *sos1* roots in the transcriptomic data set (log2 FC = 4.56) and verified by RT-qPCR (168-fold induction at 7 d, compared to 2.6-fold in the wild type). *SRO1c* encodes a putative poly-[ADP-ribose] polymerase (PARP) and is the most stress-responsive gene within the rice *SRO* family (You et al., 2013). Notably, *SRO1c* is closely related to the Arabidopsis protein RCD1, an inactive PARP that shuttles between nucleus and cytoplasm to interact with SOS1 (Katiyar-Agarwal et al., 2006).
Among the CIPK-encoding genes, CIPK21 showed the largest transcriptional up-regulation by salt in sos1 vs. wild-type roots according to the microarray data (log2 FC = 2.22), which was confirmed by RT-qPCR. A modest 1.93-fold transcriptional induction was measured in wild-type roots after 7 d in salt, whereas in sos1 the 6.05-fold induction after 4 d treatment was followed by a striking 21.47-fold induction after 7 d (Figure 9). CIPK15, which has been implicated in the salt stress response and enhances the salinity tolerance of transgenic rice plants (Xiang et al., 2007), was also up-regulated in the sos1 mutant (log2 FC = 1.75; Supplemental Table S5). CIPK1, 31, 10 and 17 also showed differential up-regulation in sos1 roots relative to that in the wild type, whereas CIPK23, 29, 25 and 4 were down-regulated by at least 2-fold. CBL genes showed little differential responsiveness in microarray data, with CBL9 being the most highly induced (log2 FC = 2.04) and CBL7 the most down-regulated (log2 FC = -2.46). RT-qPCR data confirmed the up-regulation of CBL9 and showed that the down-regulation of CBL7 in sos1 was only transient at 4 d, with a return to pre-stress level after 7 d (Figure 9). Although not identified as statistically significant DEGs, the rice homologs of CBL4/SOS3 and CBL10, the two CBLs regulating CIPK24/SOS2 and SOS1 in Arabidopsis, showed complementary kinetics in sos1 roots so that the transient induction of OsCBL10 was followed by up-regulation of OsCBL4 (Figure 9).

Successful adaptation to salinity requires the accumulation of inorganic and organic osmolytes inside vacuoles for osmoregulation and ion detoxification (Munns and Tester, 2008). Among these osmolytes, Na\(^+\), K\(^+\), Cl\(^-\) and organic acids such as malate are most abundant. In Arabidopsis, NHX1 and NHX2 are vacuolar Na,K/H antiporters that function in planta to drive the uphill accumulation of K\(^+\) into the vacuolar lumen for osmotic regulation, generation of cellular turgor, and stomatal movements (Leidi et al., 2010; Barragan et al., 2012; Andrés et al., 2014). The rice genes encoding exchangers OsNHX1 and OsNHX2 showed a 3- to 4-fold transcriptional up-regulation in sos1 roots. Gene Os09g0484900, encoding a putative protein 75% similar to the tonoplast dicarboxylate transporter of Arabidopsis involved in the uptake of malate into the vacuole (Emmerlich et al., 2003), was also highly induced in the salinized sos1 roots according to both the microarray and RT-qPCR data (8-fold induction at 7 d in the WT vs. 85-fold in sos1). Malate storage within the vacuole allows the plant to accumulate...
this metabolite to very high concentrations (up to >300 mM) and this large gene induction likely reflects the need to accumulate anions in the vacuolar lumen for osmotic adjustment and charge compensation of compartmentalized Na⁺. These results suggest that in response to salinity, the sos1 mutant differentially modulates multiple K⁺ and malate transporters to achieve cytosolic ion homeostasis and osmoregulation in intracellular compartments.

Other genes encoding K⁺-transport proteins showed minor transcriptional responses. No HAK genes showed differential up-regulation in sos1 roots compared to that in the wild type according to microarray data. On the other hand, HAK1, 5 and 16 were down-regulated 3- to 4-fold in sos1 relative to that in the wild type. OsHAK5 mediates high-affinity K⁺ uptake by roots under nutrient limitation and in saline conditions (Yang et al., 2014), whereas OsHAK1 plays a role in K⁺ uptake and translocation to shoots over low- and high-K⁺ concentration ranges (Banuelos et al., 2002; Chen et al., 2015). Closer inspection by RT-qPCR showed that both OsHAK1 and OsHAK5 transcript abundance displayed complex kinetics (Figure 9). OsHAK1 transcripts showed opposite kinetics after stress imposition in control and sos1 plants. A transient induction of OsHAK5 was also observed in control plants, but the up-regulation of OsHAK5 was sustained throughout the treatment in the sos1 mutant. The RT-qPCR data is in agreement with the expected role of OsHAK5 in sustaining K⁺ uptake in a saline environment (Horie et al., 2011; Yang et al., 2014) and probably reflects the need of sos1 roots to re-establish K⁺ homeostasis under salinity.

The HKT proteins of cereals have been involved in the uptake of Na⁺ by roots and in the root-to-shoot partition of Na⁺ and K⁺ (Garcia deblas et al., 2003; Yao et al., 2010; Munns et al., 2012; Hamamoto et al., 2015). Out of eight HKT genes in the rice genome, only HKT8/HKT1;5/SKC1 (Os01t0307500) showed a modest yet statistically significant down-regulation in sos1 roots under salt (log₂ FC = -0.81), whereas HKT1/HKT2;1 (Os06t0701700) was up-regulated in sos1 roots in the absence of stress (log₂ FC = 2.10). The rice transporter HKT8/SKC1 regulates Na⁺/K⁺ levels in shoots by recirculating Na⁺ from shoots to roots (Ren et al., 2005). The significance of the HKT8/HKT1;5/SKC1 down-regulation might be related to the excessive accumulation of
Na$^+$ in the sos1 roots and represent a root-protecting mechanism limiting the back-flow of Na$^+$ to roots.
DISCUSSION

We showed previously that the rice SOS system comprises the OsSOS1 transporter, the OsCIPK24 protein kinase that is the closest homologue to the Arabidopsis SOS2/AtCIPK24, and OsCBL4 that is most similar to SOS3/AtCBL4 (Martinez-Atienza et al., 2007). The fact that rice OsSOS1 is encoded by the single copy gene Os12g44360 allowed a reverse-genetics approach to determine the importance of this protein in the salt tolerance of rice. Here, we have shown that the sos1 mutant plant displays an exceptional salt-sensitivity that is akin to the Arabidopsis sos1 mutant phenotype. By contrast to the uniqueness of OsSOS1, the modest salt-sensitivity of the rice cipk24 mutant relative to sos1 plants is likely due to functional redundancy. The genetic complement of CIPK-CBL modules in the rice genome comprises 10 CBLs and 33 CIPKs, which is similar to the 10 CBLs and 25 CIPKs known in Arabidopsis (Kolukisaoglu et al., 2004; Gu et al., 2008; Kanwar et al., 2014). Most of the CIPK genes of rice showed transcriptional responses to various abiotic stresses including salinity (Xiang et al., 2007; Kanwar et al., 2014). Since transcription of CIPK8, the closest homologue to CIPK24, was induced by salt (Xiang et al., 2007; Kanwar et al., 2014), a functional relatedness of CIPK8 and CIPK24 appears likely regarding the salt-stress response of rice. Our transcriptomic analysis also flagged CIPK15, CIPK21 and CIPK31 as potential players in the sodicity response of roots based on the greater up-regulation in sos1 roots compared to that in the wild type (Supplemental Table S5). Indeed, CIPK15 has been reported to increase the salinity tolerance of transgenic rice (Xiang et al., 2007), whereas a loss-of-function cipk31 mutant was salt sensitive (Piao et al., 2010). There is no reverse genetics data for CIPK21, but RT-qPCR confirmed a 21.47-fold up-regulation of CIPK21 transcripts after 7 d in sos1 roots (Figure 8). Likely, this large transcriptional induction of CIPK21 in the sos1 mutant reflects its major role in Na⁺ homeostasis. Thus, transcriptomic and genetic data, together with the enormous difference between the salt-sensitive phenotype of the cipk24 and the sos1 mutants, suggest that several CIPKs may target SOS1 for post-transcriptional activation in rice.

Recently, a genetic screen for reduced cesium (Cs⁺) accumulation in rice identified a putative loss-of-function allele of SOS2/CIPK24 as the underlying genetic lesion
The lower Cs\(^+\) content in the *lcs1/sos2* mutant correlated with reduced expression of K\(^+\) transporters HAK1, HAK5 and AKT1 under low external K\(^+\)/Na\(^+\) ratios, suggesting that Cs\(^+\) uptake occurred through root K\(^+\) transporters. Similar to our findings, the *lcs1/sos2* mutant accumulated Na\(^+\) in roots and was moderately salt-sensitive.

Entry and redistribution of Na\(^+\)

The salt tolerance imparted by SOS1 is so intimately related to cellular tolerance that overexpression of *SOS1* and resistance to NaCl has been used as selection system for rice transformation (Zhu and Wu, 2008). Here, we show that nearly ubiquitous overexpression of *SOS1* driven by the 35S promoter yielded salt-tolerance that correlated with reduced Na\(^+\) contents in shoots and roots, as should be expected from enhanced efflux of cellular Na\(^+\) (Figure 4). In addition, we have shown that SOS1 also plays a systemic role by re-distributing Na\(^+\) ions between roots and shoots at the xylem-parenchyma boundary (Figure 6). Rice roots develop Casparian bands and suberin lamellae on the walls of the exodermal and endodermal cell layers that act as physical barriers against the apoplastic flow of water and dissolved minerals (Krishnamurthy et al., 2009). However, lateral roots emerge from the pericycle and disrupt the continuity of the endodermal barrier as they grow towards the cortex and exodermis. This rupture of the apoplastic barrier generates sites through which salts could leak into the main root. The contribution of this ‘bypass-flow’ to Na\(^+\) uptake in rice is thought to constitute a major component of the salt sensitivity of this crop, particularly at high transpiration rates (Yeo et al., 1987). Opposing this view, later studies found no evidence indicative of Na\(^+\) entry at the sites of secondary root emergence in rice plants although the tissue still remained open to apoplastic flow tracers (Faiyue et al., 2010). Rather, it was concluded that bypass-flow occurred through the lateral roots themselves (Faiyue et al., 2010). Regardless of whether bypass flow occurs through lateral roots or the endodermal rupture points caused by them, our data support the notion that net Na\(^+\) fluxes into and across the root are far from being an uncontrolled process. The combination of the tissue expression pattern of the *SOS1* gene promoter and the profile of Na\(^+\) content along a cross-section of rice
roots from wild-type and sos1 mutant plants submitted to a moderate salinity stress (2 d in 50 mM NaCl) indicated that SOS1 prevented, in the first place, the excessive accumulation of Na$^+$ in all cell types of the root, and that this protein specifically directed the flow of Na$^+$ past the endodermal filter toward the xylem vessels for the acropetal evacuation of Na$^+$ out of the root (Figures 5 and 6). Nevertheless, it should be emphasized that SOS1 is primarily a cellular-based Na$^+$ efflux system (Shi et al., 2002; Oh et al., 2009) and Na$^+$ will accumulate over time in root tissues to greater levels in the sos1 mutant than that in wild-type plants, which would eventually lead to Na$^+$ leaking into shoots by mass flow. Moreover, discontinuities in the root endodermis may also allow Na$^+$ export out of roots without a biochemical control by dedicated transporters. In other words, Na$^+$ will accumulate in the shoot of the sos1 mutant as a consequence of uncontrolled uptake by roots even if SOS1 contributes to transporter-mediated Na$^+$ loading into the xylem.

The sensitivity of rice plants to salinity is strongly associated with the accumulation of Na$^+$ in the leaves (Coskun et al., 2013). Like SOS1, the gene encoding the Na$^+$-selective transporter SKC1/HKT8/HKT1;5 is preferentially expressed in the parenchyma cells surrounding the xylem vessels, where the HKT protein is thought to mediate the recirculation of Na$^+$ by unloading Na$^+$ from the root xylem (Ren et al., 2005). Our data shows that SOS1 counteracts HKT function by actively exporting Na$^+$ from parenchyma cells into the xylem vessels. Thus, the amount of Na$^+$ that is eventually delivered to shoots will be determined by the interplay between HKT and SOS1 proteins acting antagonistically. The optimal set-points for Na$^+$ root content and the net transfer of Na$^+$ from shoots to roots must be commensurate with the ability of shoot tissues to compartmentalize the incoming Na$^+$ to avert ion toxicity while achieving full osmotic adjustment (Pardo, 2010). The down-regulation of SKC1/HKT8/HKT1;5 in salinized sos1 roots could represent a root-protecting mechanism limiting the back-flow of Na$^+$ to roots that are already exposed to high Na$^+$ contents. The reciprocally balanced activity of HKT and SOS1 proteins has been evidenced also in wheat. Two loci, termed Nax1 and Nax2 encoding proteins HKT1;4 and HKT1;5, respectively, are genetic determinants of enhanced Na$^+$ retrieval from the xylem and reduced rates of Na$^+$ delivery to the shoot (Huang et al., 2006; James et al., 2006; Byrt et al., 2007). Notably,
the Nax loci also reduced the activity and expression level of the SOS1 exchanger in both root cortical and stele tissues, which contributed further towards reducing the xylem Na⁺ load (Zhu et al., 2015). In Arabidopsis, a functional hierarchy of HKT- and SOS1-dependent transport processes was established by comparing the salt sensitivity of hkt1 and sos1-sos3 mutants (Rus et al., 2004). The mutant lacking HKT1 function was sensitive to long-term salinity due to the excessive accumulation of Na⁺ in shoots and the hkt1 mutation suppressed the NaCl hypersensitivity of the sos3 mutant by achieving a balanced distribution of Na⁺ that was closer to that of wild-type plants (Rus et al., 2001; Rus et al., 2004). Together, these data demonstrate that a finely regulated interplay between HKT and SOS systems at the xylem parenchyma is a key component of the salt tolerance of vascular plants.

An unresolved question is the mechanistic basis for the intimate relationship of Na⁺-specific transporters acting at the xylem parenchyma with the K⁺ status of the plant. In rice, both SOS1 and SKC1/HKT8/HKT1;5 selectively transport Na⁺ and yet they affect the accumulation of K⁺ in shoots. Wheat plants carrying the Nax1 and Nax2 loci encoding the Na⁺ transporters HKT1;4 and HKT1;5 also had higher rates of K⁺ transport from roots to shoots (James et al., 2006). Root xylem loading is a key step for the delivery of nutrients to the shoot (Drew et al., 1990; Engels and Marschner, 1992; Gaymard et al., 1998). In Arabidopsis, the voltage-dependent K⁺-channel SKOR mediates xylem loading and root-to-shoot translocation of K⁺ (Gaymard et al., 1998). The SKOR gene is primarily expressed in pericycle and root xylem parenchyma cells. The SKOR channel directs outward K⁺ currents upon membrane depolarization facilitating the release of the K⁺ from the parenchyma cells into the xylem vessels. The Arabidopsis sos1 mutant was initially described as impaired in K⁺ uptake (Wu et al., 1996), but subsequent research has shown that SOS1 is a highly specific Na⁺/H⁺ exchanger with no ability to transport K⁺ (Qiu et al., 2002; Shi et al., 2002; Quintero et al., 2011). Here we show that removal of SOS1 from its preferential expression tissue at the xylem parenchyma cells leads to the accumulation of Na⁺ ions while in transit to the xylem vessels, with the concomitant reduction in K⁺ content in these same cells. The Na⁺/K⁺ ratio in xylem parenchyma cells climbed from 0.49 in the wild type to 9.40 in the sos1 mutant, i.e. a 19-fold change (Figure 6). The coupling of Na⁺ unloading by
the xylem-localized HKT transporters with K⁺ loading into xylem vessels via depolarization-activated K⁺ channels has been hypothesized (Horie et al., 2009). HKT-mediated Na⁺ uptake from xylem vessels could induce membrane depolarization of xylem parenchyma cells, which in turn would activate SKOR, the depolarization-activated outward-rectifying K⁺ channel mediating K⁺ release from xylem parenchyma cells. On the other hand, cytosolic Na⁺ has been shown to inhibit the K⁺ channel AKT1 involved in K⁺ uptake by Arabidopsis roots (Qi and Spalding, 2004). Greater cytosolic Na⁺ concentration in sos1 plants resulted in impaired K⁺ uptake by roots and poor growth under salt stress. Our data suggest a combined scenario in which the high Na⁺/K⁺ ratio in cortex cells may interfere with root K⁺ uptake and the symplastic movement towards the stele (note that AKT1 and AKT2 genes are differentially repressed in sos1 roots; Supplemental Table S5), while the exorbitant Na⁺/K⁺ ratio in xylem parenchyma cells of the sos1 may interfere with K⁺ loading into the xylem for nutrient partitioning between shoots and roots. This model may also explain the initial observation that the growth of the Arabidopsis sos1 mutant was impaired under low K⁺ availability. This hypothesis about the mechanistic links between long-distance transport of Na⁺ and K⁺ nutrition deserves further research.

Transcriptome of the sos1 mutant

Nitrate (NO₃⁻) is the quantitatively most important anion counter-balancing xylem loading of K⁺ (Engels and Marschner, 1993). In Arabidopsis, the low-affinity, bi-directional NO₃⁻ transporter NRT1.5 mediates NO₃⁻ efflux from pericycle cells to the xylem vessels of Arabidopsis, and SKOR and NRT1.5 cooperate to ensure K⁺ delivery to the shoot via xylem (Drechsler et al., 2015). Alternatively, NRT1.5 could load K⁺ into the xylem directly (Li et al., 2017; Ragel et al., 2019). A similarly complex relationship is also apparent between Na⁺ and K⁺ loading into the xylem (Alvarez-Aragon and Rodriguez-Navarro, 2017). Our transcriptomic data of the sos1 mutant suggests a broad molecular connection between Na⁺/K⁺ homeostasis and nitrogen metabolism under salinity stress. The biological process, as categorized by GO terms, "Cellular Nitrogen Compound Metabolic Process" was the most severely down-regulated category in the sos1 mutant under salt relative to that in control roots (FDR= 1.6x10⁻
Key proteins involved in nitrate uptake by roots and nitrate assimilation were repressed in the sos1 mutant. Genes encoding four NRT2-like transporters and two NAR2-like regulators have been identified in the rice genome. The Nitrate Transporter-Activating protein 2.1 (NAR2.1) interacts with the nitrate transporters OsNRT2.1, OsNRT2.2 and OsNRT2.3a to activate the uptake and distribution of nitrate within the plant (Feng et al., 2011). The strong expression of OsNAR2.1, OsNRT2.1 and OsNRT2.2 in root epidermal cells and that OsNAR2.1 knockdown greatly impaired both high- and low-affinity nitrate transport systems demonstrated the key role of these proteins in nitrate uptake (Yan et al., 2011). Notably, these three key genes were substantially repressed in sos1 roots by salinity. The log2 fold change in transcript abundance for NAR2.1 (Os02g0595900), NRT2.1 (Os02g0112100) and NRT2.2 (Os02g0112600) were -4.24, -3.33 and -2.91, respectively. Similarly, the majority of enzymatic genes involved in the nitrate assimilation by its incorporation to glutamine (nitrate and nitrite reductases, and glutamine synthetases) were down-regulated in salt-treated sos1 roots. OsCIPK23 (Os07t0150700), the closest rice homologue to the protein kinase CIKP23 that regulates positively nitrate uptake in Arabidopsis (Ho et al., 2009), was also down-regulated in salinized sos1 roots relative to that in the control (log2 FC = -1.66), in agreement with a general repression of nitrogen uptake and metabolism.

Consequently, the transcriptional repression of these key genes in the sos1 mutant root should have a profound effect on nitrate uptake from the mineral medium and subsequent nitrogen assimilation, as already indicated by the GO term and MapMan analyses. The physiological implications of this response are presently unclear, but might be related to the mutual stimulation of nitrate and Na+ uptakes that has been observed in various species including rice (Kaburagi et al., 2015; Nie et al., 2015; Gao et al., 2016). In Arabidopsis, nitrate-dependent transport systems mediate the uptake and load of Na+ into the xylem, and they may constitute a major pathway for the accumulation of Na+ in Arabidopsis shoots (Alvarez-Aragon and Rodriguez-Navarro, 2017). Thus, repression of nitrate uptake by sos1 roots could have helped to curtail Na+ entry and/or distribution.

An intriguing observation is that the GO term 'Response to Stress' was down-regulated in the sos1 mutant in the two conditions tested, with and without salt, which in
principle meant that the sos1 mutant was less responsive to stress. A number of genes whose function could be reasonably predicted as being beneficial to overcome salinity stress (CBL4, NHX1, NHX2, and the putative vacuolar malate transporter encoded by Os09g0484900) or genes that are known molecular markers of stress intensity (SalT) were more highly expressed in the sos1 mutant than that in wild-type roots. These molecular markers and the severely disturbed Na⁺/K⁺ profiles of sos1 roots demonstrate that the mutant roots were indeed suffering sodicity stress more intensely than the wild type. Thus, the large proportion of genes down-regulated in the sos1 mutant might be the consequence of impaired stress detection or an inability to mount a full-fledged response, despite the greater intensity of the stress imposed upon sos1 roots. Previous studies (Zhu, 2002; Shabala et al., 2005; Ji et al., 2013) have suggested that SOS1 may perform regulatory or sensory functions that may or may not be strictly linked to its function as a Na⁺ transporter. The exceedingly high salt-sensitivity of sos1 mutants of various species (Zhu, 2002; Oh et al., 2009; Olias et al., 2009) and the interaction of the very large C-terminal domain of SOS1 with other stress signaling intermediaries (Katiyar-Agarwal et al., 2006) lend indirect support to this hypothesis. Precedents of this dual function as enzymes and key regulators are the nitrate transporter NTR1.1/CHL1 and the hexokinase HXK1 that serve as sensor proteins of their substrates (Moore et al., 2003; Ho et al., 2009; Bouguyon et al., 2015).

Alternatively, SOS1 may not be a *bonafide* sensor but serve as a molecular scaffold to facilitate the crosstalk of various signaling pathways needed to mount salt tolerance. The SRO (Similar to RCD-One) proteins are a group of plant-specific proteins that have important functions in stress adaptation and development. In Arabidopsis, SOS1 interacts with RCD1, a nucleo-cytoplasmic protein that together with AtSRO1 functions to regulate oxidative stress, hormonal and developmental responses (Katiyar-Agarwal et al., 2006; Jaspers et al., 2010). Moreover, RCD1 Interacts with dehydration-responsive DREB2 proteins and a number of transcription factors belonging to several protein families. The rice genome contains five SRO genes (SRO1α to 1e) and transcription of SRO1c is highly responsive to various stresses including salinity. As in Arabidopsis, rice SRO1c has a negative role in resistance to oxidative stress, which seems to be associated to the repression of ROS-scavenging enzyme genes (You et al.,
Surprisingly, transcriptomic and RT-qPCR data showed a staggering 55- and 168-fold induction of SRO1c in sos1 roots after 4 and 7 d of salt treatment, which runs counterintuitive to the expected output since ROS scavenging is vital to overcome salinity-induced oxidative stress in rice (Hoang et al., 2016) and OsSRO1c-overexpressing rice suffers from greater sensitivity to oxidative stress (You et al., 2013). The observed under-representation of DEGs belonging to the GO Biological Process 'Oxidation Reduction' (FDR = 2.3x10^{-6}, Supplemental Table S2) and the GO Molecular Function 'Oxidoreductase Activity' (FDR = 2.6 10^{-5}) in sos1 roots under salinity are likely related to over-induction of the negative transcription regulator SRO1c. Our MapMan analysis of the sos1 mutant transcriptional response also pointed to a significant enrichment of genes involved in the redox cellular metabolism among the down-regulated DEGs, with 61 down-regulated genes grouped in this category. Detailed analysis of the repressed genes indicates that detoxification enzymes thioredoxins, ascorbate-peroxidases (APXs), catalases and superoxide dismutases were mainly repressed in the mutant relative to that in the wild type. Moreover, further analysis of the mutant transcriptome indicated that the expression of genes grouped in the “drought/salt abiotic stress” category is significantly altered in response to salt (corrected p-value = 0.037). The differential expression was surprisingly not due to a higher level of induction of the genes of this category in response to salt, but again to a general repression in the sos1 mutant relative to the wild-type expression level. Thus, it appears that the lack of SOS1 triggers a run-away process leading to unsuitable repression of ROS-controlling genes by OsSRO1c and exacerbated salt damage. Taken globally, the transcriptomic profile of the sos1 mutant root is evidence of an as yet undefined but superior regulatory role of SOS1 in the salinity stress response.

**MATERIALS AND METHODS**

Plant culture, salinity treatments and chemical analyses

Rice seeds were germinated onto wet filter paper for 5 d and then transferred to decapped and perforated Eppendorf tubes pre-filled with rockwool. Culture was continued in a hydroponics setting with aeration in modified Miyamoto's medium...
(Miyamoto et al., 2001), with weekly transfers to fresh medium. Nutrient medium consisted of 0.09 mM (NH₄)₂SO₄, 0.05 mM KH₂PO₄, 0.05 mM KNO₃, 0.03 mM K₂SO₄, 0.06 mM Ca(NO₃)₂, 0.07 mM MgSO₄, 0.11 mM Fe-EDTA, 4.6 µM H₃BO₃, 1.8 µM MnSO₄, 0.3 µM ZnSO₄, 0.3 µM CuSO₄, pH 5.7, plus 0.3 µM Na₂MoO₄. The actual concentrations of K⁺ and Na⁺ in the medium as measured by Atomic Absorption Spectrophotometry were 0.20 mM K⁺ and 0.14 mM Na⁺. The hydroponic containers were placed in a growth chamber set to a daily light/dark cycle of 16/8 h, photosynthetically active radiation 300 µmol m⁻² s⁻¹, 30°C/22°C day/night temperature, and 40–60% relative humidity. Salt treatments were given by transferring the plants to fresh medium supplemented with the indicated amounts of NaCl.

Contrary to Nipponbare plants, seedlings of mutant (L1H and L24H) and congenic azygous lines (L1A and L24A) showed asynchronous germination and heterogeneous growth rates, which presumably arose from somaclonal variation. Therefore, after one week in hydroponic culture, outlier plants were removed and only those with sizes similar to Nipponbare controls were transferred to fresh media with and without supplemental NaCl.

For halotolerance tests in experimental paddies in the field, seeds were first germinated in pots with peat. Four-week-old plants of mutant genotypes sos1 and cipk24, their respective congenic null segregants, and of wild type Nipponbare were then transferred to paddies with clay-loam soil. Plants were arranged in paddies with 5 rows (one per genotype) of 10 plants each, to have 4 repetitions (individual paddies) of each growth condition (control and salinized) and 40 plants per genotype and treatment. Genotypes were arranged differently in each individual paddy. Control paddies were irrigated with water from an underground well with a salt content of 0.4 g/L. Salinity treatment was imposed by automatic mixing of underground water with a salty brine to give a final concentration of 1.8 g/L of seawater salts in the irrigation water.

Chlorophyll contents were determined with a Minolta SPAD-502 Chlorophyll Meter and values are given as SPAD units. To measure Na⁺ and K⁺ contents in tissues, plants...
exposed to salt stress were separated in shoots and roots and weighed. Roots were washed thoroughly 2–3 times over 2 min with cold distilled water to remove surface-contaminating salts, blotted dry and deep-frozen until use. Dried ground material was autoclaved to extract minerals and the supernatant submitted to Atomic Absorption Spectrophotometry.

*Rice mutant lines*

Nomenclature and genes codes are according to the Rice Genome Annotation Project, version 6.1 (http://rice.plantbiology.msu.edu/). Putative mutant lines annotated to carry T-DNA or Tos17 insertions in genes *SOS1* (Os12g44360), *SOS2/CIPK24* (Os06g40370) and *SOS3/CBL4* (Os05g45810) in the Nipponbare genetic background were screened for in the public repositories OryGenesDB of Génoplante (http://orygenesdb.cirad.fr/), the Rice Functional Genomic Express Database (http://signal.salk.edu/cgi-bin/RiceGE), the Rice Tos17 Insertion Mutant Database (https://tos.nias.affrc.go.jp/), and the Rice Mutants Database (http://rmd.ncpgr.cn/).

Seeds received from repositories were considered as T2 generation, comprising both heterozygous and homozygous individuals. Plants emerging from these primary T2 seed pools were subjected to diagnostic PCR to identify individuals homozygous for the mutagenic insertion based on the presence of the amplicon predicted from the line annotation and by the absence of the wild-type amplicon when oligonucleotides flanking the insertion point were used as primers in the diagnostic PCR. Primers used for diagnostic PCR and RT-PCR are listed in Supplemental Table S6. Congenic null-segregants without the mutagenic insertion (denoted here as azygous plants) were also selected as controls for further experiments to account for possible epigenetic or phenotypic alterations arising during plant transformation and regeneration procedures. Plants selected after diagnostic PCR were used to produce T3 seeds, and T3 plants were subsequently used for RT-PCR and phenotypic analyses.

*Nucleic acids, plasmids and transformation*
For RT-PCR analyses, total RNA was isolated from ca. 100 mg of frozen plant leaves with TRIZOL (Invitrogen). Synthesis of cDNA with integrated genomic DNA removal was done from 1 μg of total RNA using the QuantiTect Reverse Transcription Kit (Qiagen). PCR was done with DNA purified as described in (Murray and Thompson, 1980). For sequencing of amplicons and precise identification of insertion points, RT-PCR was performed with Accuzyme (Bioline Reagents Ltd) and amplicons were ligated to vector pCR-Blunt and transformed into *E. coli*. Recombinant plasmids were purified using the Perfectprep Gel Cleanup kit (Eppendorf), diluted at 100 ng/µL in H2O, and sequenced with M13 Reverse and Forward primers.

Constructs made to test the function of the rice SOS1 protein in yeast were done in vector pDR195 carrying the *PMA1* gene promoter. Co-expression of Arabidopsis SOS2 and SOS3 was achieved with plasmid pFL32T (Quintero et al., 2002). The ORF encoding the truncated protein OsSOS1Δ970 was obtained by PCR using as template the wild-type cDNA (Martinez-Atienza et al., 2007) and with oligonucleotides that included *BamHI* cutting sites and a stop codon after amino acid L969 of SOS1. The resulting amplicon was digested with *BamHI* and inserted in pDR195. Yeast transformation followed the Lithium-Acetate method. The *Saccharomyces cerevisiae* strain AXT3K (*Δena1::HIS3::ena4, Δnha1::LEU2, Δnhx1::KanMX4*) has been described elsewhere (Guo et al., 2004). Sodium tolerance tests were performed in the alkali cation-free AP medium (Rodriguez-Navarro and Ramos, 1984) supplemented with 1 mM KCl and with NaCl as indicated in each experiment.

To determine the expression pattern of *SOS1* a 3.9-kb fragment in the intergenic region between *SOS1* (Os12g44360) and the adjacent gene (Os12g44370) and proximal to the coding region of *SOS1* was amplified by PCR and inserted as a *SalI* and *XbaI* fragment into vector pBIG containing a promoter-less *GUS* reporter gene (Becker, 1990). GUS chemical staining with X-gal in whole tissues was as described in (Jefferson et al., 1987). To test complementation of the sos1 mutant, the full-length cDNA of rice *SOS1* was cloned in a pCAMBIA1300-derived vector containing the maize *UBIQUITIN1* gene promoter and the *nopaline synthase* (*NOS*) terminator (Campo et al., 2014). For *SOS1* overexpression, the cDNA was cloned into the vector pCAMBIA2300 under the
control of the CaMV 35S promoter and terminator and transformed in Nipponbare plants. Rice transformation by *Agrobacterium tumefaciens* EHA105 has been described (Campo et al., 2014).

**Measurement of Na⁺/H⁺ exchange in membrane vesicles**

Plasma membrane vesicles were isolated from 2-month-old rice roots from 24-h salinized (50 mM NaCl) plants using two-phase partitioning as described by Qiu et al., (2003). All steps were carried out at 4°C or on ice. Roots were homogenized in buffer containing 0.33 M sucrose, 10% (w/v) glycerol, 0.2% (w/v) BSA, 5 mM EDTA, 5 mM DTT, 5 mM ascorbate, 0.2% (w/v) casein, 0.6% (w/v) polyvinylpyrrolidone (PVP-10), 1 mM PMSF, 1 µg/mL pepstatin A, and 50 mM HEPES-KOH (pH 7.5). Eight milliliters of homogenization buffer were used per gram of tissue. The homogenate was filtered through two layers of Miraclcloth (Merck) and centrifuged at 10,000 g for 10 min. The supernatant then was centrifuged for 50 min at 100,000 g to obtain a microsomal pellet, that was resuspended in a buffer containing 0.33 M sucrose, 3 mM KCl, 0.1 mM EDTA, 1 mM DTT, 1 mM PMSF, 1 µg/mL pepstatin A, and 5 mM potassium phosphate (pH 7.8). The suspension was added to a phase mixture to obtain a phase system consisting of 6.2% (w/w) Dextran T-500 and 6.2% (w/w) polyethylene glycol 3350 in 5 mM potassium phosphate (pH 7.8), 0.33 M sucrose, 3 mM KCl. Phase separation was facilitated by centrifugation at 1,000 g. The upper phase was then repartitioned twice in this system. The final upper phase was collected, diluted with suspension buffer [0.33 M sucrose, 10% (w/v) glycerol, 0.1 mM EDTA, 2 mM DTT, 1 µg/mL pepstatin A, 20 mM HEPES-KOH (pH 7.5)], and centrifuged for 45 min at 100,000 g. The resulting pellet was collected and resuspended with the above-described suspension buffer containing 1 mM EDTA, frozen in liquid nitrogen and stored at -80°C until use. Before membrane isolation plants were treated for 24 h with 50 mM NaCl in Miyamoto’s medium to induce SOS1 activity. Vesicles purified from non-treated plants produced very low rates of Na⁺/H⁺ antiport activity, in agreement with previous reports (Qiu et al., 2003).

The measurement of Na⁺/H⁺ proton exchange was as described by Qiu et al. (2003), except that fluorescence quenching of 9-amino-6-chloro-2-methoxy-acridine (ACMA)
was used to monitor the formation and dissipation of pH gradients. Fluorescence was
recorded with a Hitachi fluorescence spectrophotometer (FL-2500) in a thermostated
cell (25°C) at excitation and emission wavelengths of 415 and 485 nm, respectively
(slit-width, 10 nm). Purified plasma membranes (50 µg of protein) were added to a
buffer containing 250 mM mannitol, 10 mM bistris-propane-2-(N-morpholine)-
ethanesulphonic acid (BTP-MES) (pH 6.8), 100 mM KCl, 3 mM MgSO4 and 1 µM ACMA
(1 mL final volume). The ∆pH was generated with the activity of the plasma membrane
H+-ATPase and recorded as fluorescence quenching after ATP addition (2.5 mM ATP,
pH 6.8). The Na+-induced dissipation of ∆pH was initiated after adding 100 mM NaCl
into the cuvette, and recorded as the recovery of fluorescence. Exchange rates are

calculated as fluorescence recovery relative to that prior to ATP addition (∆F/Fmax)
per minute and milligram of protein.

Microarray analysis

Control and sos1 seedlings were grown hydroponically in Miyamoto’s medium for 3
weeks in environmentally controlled conditions (12 h of light at 25°C and ca. 450 µmol
quanta m⁻² s⁻¹, and 12 h darkness at 21°C; relative humidity ca. 65%). Salt stress was
imposed by transferring to fresh nutrient solution containing 75 mM NaCl for 4 d.
Control plants received fresh nutrient solution without NaCl. No signs of plant
deterioration were observed at the end of the salinity treatment. Total RNA was
extracted from whole roots pooled from five plants per genotype and treatment. The
quality and concentration of the RNA were determined using the Agilent 2100
Bioanalyzer and samples with an RNA Integrity Number (RIN) less than 6 were
discarded. Three on-chip technical replicates per biological sample were analyzed.
Gene expression profiling was performed by microarray hybridization using a custom
rice transcriptomic array (Oryzon Genomics, Barcelona, Spain) containing 3 x 29450
probes (3 technical replicates per probe) representing 20750 genes. Microarray
construction, probe labeling, and hybridization conditions were as in (Campo et al.,
2014). Results were normalized using self-to-self comparison Nipponbare vs
Nipponbare, and fold change was calculated as log₂ of the ratio of relative expression
of sos1 (sample, Smp) versus wild-type (control, Ctr) plants grown in identical
conditions. Only genes for which replicate spots showed differential expression of $-0.5 \log_2(Smp/Ctr) > 0.5$ and $P$ value $< 0.05$ were considered as differentially expressed. Bootstrap analysis with Significance Analysis of Microarrays was used to identify differentially expressed genes using a cut-off of 2 (Tusher et al., 2001). Significance Analysis of Microarrays calculates the fold change and significance of differences in expression. Data sets for microarray analyses have been deposited in the National Center for Biotechnology Information Gene Expression Omnibus with accession number GSE129844.

Differential expression of a select number of DEGs was confirmed by reverse-transcription quantitative PCR (RT-qPCR). Three-week-old control and sos1 seedlings were treated with 75 mM NaCl in Miyamoto's medium for 4 and 7 d. Total RNA was extracted from roots and 500 ng were used to synthesize the cDNA using the Quantitec Reverse Transcription kit (Qiagen, Hilden, Germany). Quantitative real-time PCR (qRT-PCR) assays were performed using the CFX Connect Real-Time PCR Detection System (Bio-Rad). The PCR mixture (total volume of 20 μl) was composed of 5 μl of diluted cDNA, 10 μl iTaq Universal SYBR Green supermix (2x), 0.3 μM of each primer and H2O. The specific oligonucleotides used are listed in the Supplemental Table S6. Thermal cycling consisted of 95°C for 3 min, followed by 40 cycles of 10 sec at 95°C, 15 sec at 58°C, and 15 sec at 72°C. A melting curve was generated to check the specificity of the amplified fragment. The efficiency of all primers under these conditions was 85–100% for all samples tested. Rice UBIQUITIN5 was used as a house-keeping gene in expression analysis. Relative gene expression was determined using the $2^{-ΔΔCT}$ method (Livak and Schmittgen, 2001). Threshold cycle values were determined using ICYCLER Q software (Bio-Rad).

**Statistical analyses**

Numerical data was analyzed with IBM SPSS Statistics v.21 (Armonk, NY, USA) using a completely randomized design model. Means and variances were compared by One-Way Analysis of Variance (ANOVA) to find significant differences using the Fisher's Least Significant Differences test at $p < 0.01$ level, unless stated otherwise.
**ACCESSION NUMBERS**

Sequence data from this article can be found in the Rice Genome Annotation Project (http://rice.plantbiology.msu.edu/) and UniProt (https://www.uniprot.org) databases under accession numbers *SOS1* (LOC_Os12g44360, Q5ICN3), *CIPK24/SOS2* (LOC_Os06g40370, Q69Q47) and *CBL4/SOS3* (LOC_Os05g45810, Q75KU4).

**SUPPLEMENTAL MATERIAL**

**Supplemental Figure S1.** Molecular characterization of rice lines AKFG08 and AKFB09.

**Supplemental Figure S2.** Molecular characterization of rice line NC2588.

**Supplemental Figure S3.** Molecular characterization of rice line AUJE11.

**Supplemental Figure S4.** Molecular characterization of the rice line AVIB03.

**Supplemental Figure S5.** Na⁺/H⁺ exchange activity in plasma membrane vesicles.

**Supplemental Figure S6.** Molecular characterization of the rice lines AMHC11 and AMHC12.

**Supplemental Figure S7.** Salinity tolerance test in experimental paddies.

**Supplemental Figure S8.** Complementation of the sos1 mutant.

**Supplemental Figure S9.** MapMan analysis of the metabolic and cellular responses of sos1 roots to salt.

**Supplemental Table S1.** Differentially expressed genes (DEGs) in the sos1 mutant roots when compared to the control line (Excel file).
**Supplemental Table S2.** Gene Ontology (GO) analysis of total DEGs identified in the salt-stressed or control-cultivated roots (Excel file).

**Supplemental Table S3.** Top 25 enriched GO categories (Excel file).

**Supplemental Table S4.** MapMap analysis of DEGs identified in the salt-stressed sos1 roots (Excel file)

**Supplemental Table S5.** Differential expression levels of select genes involved in Na⁺ and K⁺ transport processes (Excel file).

**ACKNOWLEDGEMENTS**

This work was supported by grants BIO2015-70946-R to FJQ, and grants BFU2015-64671-R and BIO2016-81957-REDT to JMP from MINECO (co-financed by the European Regional Development Fund), and by the Rural Development Administration, Republic of Korea (SSAC grants PJ01108101 to D-JY and PJ01318205 to JMP). Part of this work was conducted at the Rice Functional Genomics Platform (REFUGE), Montpellier France, supported by an Agropolis Fondation grant to DM and EG. FGA was supported by a 'Juan de la Cierva' aid from MINECO.
Table 1. Insertion points (nt, nucleotide) are expressed relative to the ATG initiation codon in the genomic sequences, corresponding to nucleotides 27508321 in *SOS1*, 24043690 in *CIPK24* and 26535878 in *CBL4*. When confirmed by sequencing of the PCR amplicon containing the mutagenic insertion, the actual insertion point is also given; otherwise it is noted as not determined (n.d.). OTL, Oryza Tag Line database; NIAS, National Institute of Agrobiological Sciences, Japan.

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FIGURE LEGENDS

Figure 1. Functionality of truncated SOS1 protein.
A, Diagram of functional domains of SOS1. The junction domain is the C-terminal portion of SOS1 that has sequence homology with NHX8 and no known functional role; further downstream is the regulatory domain that is specific and characteristic of SOS1-like proteins that has been dissected in three functional modules: the activation domain with sequence similarities to cyclic-nucleotide binding site (amino acids 740–820), the autoinhibitory domain (977–1023) and the CIPK24 phosphorylation site (Ser-1135) that counteracts autoinhibition (Quintero et al., 2011). The arrowhead marks the truncation point after K767 in the mutant protein. B, RT-PCR of SOS1 mRNA of wild-type Nipponbare plants (WT) and homozygous plants of line AVIB03 (sos1), with primers S1-E19-F and S1-E20-R annealing at exons 19 and 20. Detached leaves were treated (salt) or not (unt) with 150 mM NaCl for 24 h. Insertion of Tos17 at the intervening intron 19 prevented the amplification in the sos1 mutant. As control, the cDNA of SOS1 was used as template in the PCR reaction (cDNA); C- indicates a mock reaction without template. C, Truncated SOS1 proteins of rice were expressed in the yeast strain AXT3K, with and without the co-expression of the Arabidopsis SOS2–SOS3 kinase complex, and compared to the wild-type SOS1. Decimal dilutions of saturated liquid cultures in selective media were plated in AP medium with 1 mM KCl (1K) supplemented with the indicated millimolar concentrations of NaCl.

Figure 2. Growth parameters of sos1 mutant in salt.
Five-d-old seedlings of wild type, sos1 mutant, and a congenic segregant without the sos1 mutation, were transferred to hydroponic culture in Miyamoto's medium supplemented with the indicated amounts of NaCl. A, Depicted are three representative plants per genotype of wild type (WT) and sos1 mutant at 5 and 10 d after transfer to 25, 50 and 75 mM NaCl. B, Fresh weight (FW) and dry weight (DW) of shoot and roots, the chlorophyll content (SPAD units), and the shoot and root Na+ content of wild type (black columns), sos1 (white) and the congenic azygous control (grey) before and after one week in 50 mM NaCl. The chlorophyll content at 14 d is also given. Plants after 2 weeks growth in salt were not further processed because
roots of the sos1 mutant were severely damaged. Values are the mean and SD of five plants per genotype. Different letters indicate significantly different values relative to that in the wild type determined by One-Way ANOVA (a= p<0.01).

Figure 3. Plant growth in experimental paddies.
A, Representative plants of sos1 (L1H), SOS1 congenic segregant (L1A) and wild type Nipponbare (WT) genotypes grown for 2 months in experimental paddies irrigated with underground water (left) or supplemented with salt (right). Salt contents in the irrigation water are indicated on top. B, Na+ and K+ content in shoots of control (white columns) and salinized plants (grey columns). Genotypes are labeled as in (A). Plotted are the mean and SD of five plants per genotype and condition after 30 d of saline treatment. Different letters indicate significantly different values at p<0.001 determined by One-Way ANOVA.

Figure 4. Complementation of sos1 mutant.
Ten-d-old seedlings of wild type, sos1 mutant, and the complementation line C11 were transferred to hydroponic culture in Miyamoto’s medium supplemented with the indicated amounts of NaCl. Plants were harvested two weeks later. A, Depicted are representative plants of genotypes Nipponbare (N), the sos1 mutant (S) and line C11 (C). B, Dry weight and Na+ contents in shoots and roots of plants harvested after 7 d of the salinity treatment. Represented are the mean values and SD of wild type (black columns), sos1 (white) and complementation line C11 (grey) of four plants per genotype and growth condition. Letters indicate significantly different values relative to that in the wild type determined by One-Way ANOVA (a= p<0.01, b= p<0.05).

Figure 5. Salt tolerance imparted by SOS1
Seven-d-old seedlings of two independent Nipponbare transgenic lines expressing 35S:SOS1 were cultivated alongside control seedlings transformed with an empty vector (V) in the absence or presence of 25 and 50 mM NaCl for 4 weeks. A, Representative control (V) and transgenic plants (SOS1) are depicted. B, Survival rate of plants after the completion of the experiment. C, Dry weight (DW) and Na+ content (as percentage of DW) of plants surviving the 25-mM NaCl treatment (n=6). Letters
indicate significantly different values relative to that in the wild type determined by One-Way ANOVA (a= p<0.01, b= p<0.05, c= p<0.1).

**Figure 6. Tissue expression pattern of SOS1.**
A to K, Representative images of staining patterns resulting from GUS expression under the control of the SOS1 gene promoter. A and B, Preferential expression around the root vascular bundle. C, Root cross-section. D and E, Lack of staining in undifferentiated tissues near the root cap and in root hairs. F to H, Staining indicates that expression of SOS1 correlates with the differentiation of vascular bundles in secondary roots. I, Scattered expression around the vasculature of the coleoptile. J, Vascular bundles of the flag leaf. K, Strong expression in leaf xylem parenchyma. Arrowheads point to non-stained stomata. The scale bars represent 100 µm.

**Figure 7. Mineral content analysis by SEM-EDX.**
A, Representative SEM cross section of root with labelled arrows indicating the cell types whose elemental composition was analyzed by EDX. Scale bar is 0.1 mm. B, Percentage of total counts of Na⁺ and K⁺, measured in 4–5 cells of indicated cell types from 2 plants of each genotype. Plants were treated with 50 mM NaCl for 2 d. Shown are the mean and SD values. Letters indicate significantly different values relative to that in the wild type for each cellular type determined by One-Way ANOVA (a= p<0.01; b= p<0.05). C, Na/K ratios of net content values given in (B). The experiment was repeated twice with similar results.

**Figure 8. Summary of Differentially Expressed Genes**
A, Four-way Venn diagram indicating the number of up-regulated and down-regulated genes found in the comparison between the sos1 rice mutant and the wild type Nipponbare in the non-stress (left; Control: 0 mM) and salt-stress (right; Salt: 75 mM) treated roots. (B) Three-way Venn diagram indicating the number of significant GO terms from the gene-ontology analysis of the up-regulated, down-regulated and total DEGs found in the sos1 rice mutant and the wild type Nipponbare in salt-stressed roots.
Figure 9. Quantitation of expression levels of genes relevant to salinity tolerance. RT-qPCR analysis was performed using the 2−ΔΔCT method and the gene expression data normalized for the quantity of UBQ5 gene transcript before (0) and 4- and 7-d after salt treatment with 75 mM NaCl. All y axes are mean values of normalized expression levels ± SD. All x axes are time of treatment and genotype.

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