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Hepatocellular adenoma in a European flatfish (*Limanda limanda*): genetic alterations in laser-capture micro-dissected tissue and global transcriptomic approach

^{†a}Adélaïde Lerebours, [†]Emma Chapman, [‡]Brett P. Lyons, [‡]John P. Bignell, [‡]Grant D. Stentiford, and [†]Jeanette M. Rotchell*

[†]School of Environmental Sciences, University of Hull, Cottingham Road, Hull, HU6 7RX, United Kingdom

[‡]Centre for Environment, Fisheries and Aquaculture Science, Weymouth Laboratory, Barrack Road, The Nothe, Weymouth, Dorset DT4 8UB, United Kingdom

^aCurrent address: School of Earth and Environmental Sciences, University of Portsmouth, Burnaby Building, Burnadt Road, Portsmouth PO1 3QL, United Kingdom

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Corresponding Author

*Jeanette M. Rotchell, phone number: +44 (0) 1482 465333, fax number: +44 (0) 1482 465458, e-mail: J.Rotchell@Hull.ac.uk

Running title: Molecular changes in dab hepatocellular adenoma samples

Abstract

Liver tumours in flatfish have been diagnosed using histopathology for decades to monitor the impacts of marine pollution. Here we describe the application of specific gene (*retinoblastoma*, *Rb*) profiling in laser capture micro-dissected samples, and a suppression subtractive hybridization (SSH) approach to isolate differentially expressed genes in hepatocellular adenoma (HCA) samples from dab, *Limanda limanda*.

The *Rb* profiles from apparently normal and HCA micro-dissected samples of fish from the North Sea showed no significant difference, and genotypic heterogeneity within defined histological phenotypes was observed. In the SSH, sequences associated with cell signalling, cell cycle, gene expression regulation, protein transport and protein degradation were isolated. These included up-regulation of *arrestin domain containing 3* (*arrdc3*), *Rac-1* and *tribbles*, and down-regulation of *ankyrin repeat/sterile alpha-motif domain-containing protein 1B-like* (*ANKS1B-like*), *c-fos*, *CDKN1B* and *RhoA-like* sequences, previously implicated in mammalian HCA. This study offers new candidates involved in fish liver tumour development.

1. Introduction

Liver pathologies of flatfish dab (*Limanda limanda*) English sole (*Parophrys vetulus*) and European flounder (*Platichthys flesus*) have been used internationally to monitor the effects of exposure to marine pollution since the 1980s (Malins et al., 1985; Vethaak and Reinallt, 1992; Stentiford et al., 2003; Lyons et al., 2015). Such lesions have been associated with exposure to anthropogenic contaminants such as polycyclic aromatic hydrocarbons (PAHs) (Malins et al., 1985; Vethaak and Reinallt, 1992). Dab possess both a similar histopathological tumour profile to humans (Stern and Zon, 2003) and homologs of human cancer genes such as *ras* and retinoblastoma (*Rb*), including mutational alterations of the *Rb* gene in tumour tissues (Du Corbier et al., 2005). In this respect, we have previously proposed that the dab hepatocellular tumour development could act as surrogate for cancer and the tumourigenesis process in human populations (Rotchell et al., 2009). Studies using this species also facilitate a better understanding of chemically-induced carcinogenesis in wild animals (Stentiford et al., 2005; Ward et al., 2006; Southam et al., 2008).

The histopathology of tumours and pre-tumours in dab liver are routinely diagnosed using a quality assured process involving histological tissue sections generated from wax-embedded samples (Feist et al., 2004). This diagnostic approach has recently been coupled with molecular analyses of tumour and surrounding normal tissue (Small et al., 2010). In addition, gross lesions and apparently normal tissues have been resected from the dab liver for molecular investigations such as genetic alterations of neoplastic associated genes (Du Corbier et al., 2005; Rotchell et al., 2009, Lerebours et al., 2014), transcriptomic (Small et al., 2010), proteomic (Stentiford et al., 2005) and metabolomic studies (Stentiford et al., 2005; Southam et al., 2008). Traditional microdissection and molecular analysis of populations of cells presents a challenge in that cellular heterogeneity within tissues samples may result in misleading findings (Cole et al., 1999; Sluka et al., 2008). Laser Capture-Microdissection (LCM) allows the ability to view and dissect target cells microscopically, thereby providing a direct link between a specific histopathological lesion and the molecular profile of that lesion (Gillespie et al., 2001). LCM has previously been applied to pathological studies in

aquatic organisms including fish (Vinas and Piferrer, 2008; Jorgensen et al., 2009; Kitahashi et al., 2009; Lerebours et al., 2013; Nowak et al., 2013; Leguen et al., 2015; Schaeck et al., 2016), crustaceans (Small et al., 2008), and cnidarians (Wiebring et al., 2010).

The study applies the LCM technique to facilitate an investigation of the *Rb* allele status in hepatocellular adenoma (HCA) and normal hepatocytes populations from North Sea sampled dab. *Rb* was selected on the basis that *Rb* mutations have previously been reported in dab liver tumour samples (Du Corbier et al., 2005; Lerebours et al., 2014). In parallel, a global transcriptomic approach using the suppressive subtractive hybridisation (SSH) method was used to analyse the differentially expressed genes between HCA-bearing liver and normal liver samples.

2. Materials and Methods

2.1. Sample collection

L. limanda were captured at UK Clean Seas Environmental Monitoring Programme (CSEMP) sites (Nicolaus et al., 2016) at the Dogger Bank, North Sea during July 2008 and 2009 using 30 min tows of a standard Granton trawl aboard the RV *Cefas Endeavour*. Upon landing, fish were immediately removed from the catch and placed into flow-through tanks containing aerated seawater. The sex, size (total length), weight (g), and Fulton index Condition (FC) ($FC = BW/L^3 \times 100$), and presence of external signs of disease were noted for each fish using methodology specified by the International Council for the Exploration of the Sea (Bucke et al., 1996). Following euthanasia (using a Schedule 1 method, in accordance with National U.K. regulations), the body cavity was opened and the liver was assessed for the presence of macroscopic liver tumours according to the guidelines set out by Feist et al. (2004). For each fish, a standardised cross-section was obtained, placed into a pre-labelled histology cassette and fixed for 24h in 10% neutral buffered formalin before transfer to 70% industrial methylated spirit. An additional tissue cross-section was also obtained from the site immediately adjacent to the formalin-fixed sample. This sample was embedded in Optimum Cutting Temperature (OCT) media (RA Lamb, U.S.A.), frozen immediately

in an iso-pentane cryobath and then stored at -80°C for subsequent laser-capture microscopy and molecular investigations. The sampling protocol allowed for a direct comparison of lesions in formalin fixed and cryopreserved samples of liver (Figure 1). The humane euthanasia of animals and sampling of their tissues, as detailed in this study, was undertaken in accordance with both national regulations and institutional policy.

2.2. Sample processing

Samples were processed according to Lerebours et al. (2013). Formalin-fixed paraffin embedded (FFPE) samples of liver were prepared by vacuum infiltration processing using standard protocols (Feist et al., 2004). Following embedding, sections were cut at 3-5 µm on a rotary microtome (Leica Biosystems, U.K.), and resulting tissue sections were mounted on glass slides before staining with haematoxylin and eosin (HE) (Figure 1). Glass slides were mounted with DePex (Fisher Scientific, U.K.) and analysed by light microscopy using a Nikon Eclipse E800 microscope (Nikon, U.K.). Diagnoses of liver tumours followed guidelines set out by Feist et al. (2004).

Frozen samples were selected according to the presence and identification of lesions in the corresponding FFPE sections. OCT-embedded liver samples were transferred to a cryostat and acclimatised to -20°C. For each liver sample, two frozen sections were cut and collected onto an RNase clean glass slide. These sections were cut at 8-10 µm which has previously shown to give efficient yields of high quality RNA without an excessive increase in tissue opacity, or chance of dissecting unwanted cells (Lerebours et al. 2013). Sections were subsequently stained according to a protocol adapted from Huang et al. (2002). Glass slides were immediately placed into 70% ethanol for 2-4 min and then rinsed in DEPC water. Mayer's hematoxylin stain was applied to the slide surface for 2 min followed by a rinse in DEPC-treated water. The slides were then incubated in Scott's bluing solution (Leica Microsystems, U.K.) for 30s followed by a rinse in DEPC water and a rinse in 70% ethanol. Eosin stain was applied for 45s followed by dehydration in 95% ethanol

for 30s, in 100% ethanol for 1 min and two baths of xylene of 1 min each. Frozen sections were screened to determine whether those lesions previously observed and classified within FFPE liver sections, were also present in the frozen sections (Figure 1). Specifically, the histopathological lesions of interest for the purposes of this study were controls (no abnormalities detected) and HCA.

Upon identification, digital images of histological lesions were obtained using the Lucia G Screen Measurement System (Nikon UK Ltd., U.K.).

2.2. Tissue microdissection

LCMs were performed using the MMI Cellcut system (Molecular Machines and Industries, Switzerland). The system comprised an inverted microscope (Nikon) with a motorised stage attached to a computer with digital camera, panel monitor and a fixed ultraviolet (cold) laser source. Two to five frozen tissue sections (depending on the size of the lesion) were collected onto RNase clean LCM membrane slides from fish that displayed no abnormalities and from lesions of interest in fish displaying tumours. Membrane slides were immediately placed in 70% ethanol on ice prior to staining as described above, followed by allowing to air dry. During this process, interspersed frozen sections were also obtained and collected onto glass slides (in between those collected for LCM) and stained as described above for further comparative histological assessment. These additional sections allowed for the determination of the nature and localisation of the lesions whenever present in the fish analysed (due to the reduced image quality resulting from differential refractive light index of LCM membrane slides and no cover slip). Micro-dissected samples were captured following the laser cutting by mechanically lowering and raising an adhesive isolation cap (attached to a 0.5 mL collection tube) onto the membrane.

2.3. *Rb* cDNA isolation and allele profiling

The samples used for the *Rb* allele profiling were generated from laser microdissected samples cut from 8 µm tissue sections which represents approximately 1/600th of the whole mass of

the liver (and differs from the classically dissected samples used for the SSH analysis described in section 2.4 below) (Figure 1).

Total RNAs from laser micro-dissected samples (using n=7 fish for HCA and n=11 fish for normal tissue *Rb* profile analysis) were extracted using the Absolutely RNA Microprep Kit (Stratagene, La Jolla, U.S.A.) according to the supplier's instructions. Normal samples were those with a visible absence of abnormalities, specifically preneoplastic and neoplastic lesions: vacuolated foci of cellular alteration (FCA), eosinophilic FCA, basophilic FCA, HCA, and hepatocellular carcinoma (HCC). The optional DNase treatment was found to decrease the concentration of RNAs and was not suitable for our analyses. The elution step was repeated 2 times in a volume of 15 μ L. The quality of RNA (the integrity of the 18 and 28S ribosomal bands) for the largest sections was evaluated by electrophoresis on a 1% agarose-formaldehyde gel. First strand cDNAs were synthesized from 14 μ L of total RNA using the SuperScript® VILO™ cDNA Synthesis Kit (Invitrogen Ltd, Paisley, U. K.) and according to the supplier's instructions. The duration of the transcription phase was 90 min. Different lengths of *Rb* fragments were amplified in order to find the maximum length yielding a reliable sequencing resolution from microdissected samples. Following this, three overlapping parts of the coding sequence of the *Rb* cDNA: RbA1, RbA2 and RbB, containing the region of functional importance, A and B, were amplified. Primer pairs used to amplify the region comprised between 620 and 1942 bp of the *Rb* cDNA (Accession number: **AY973250**) are described in Table 1. Next, 2 or 4 μ L of the reverse transcribed product was used as a template for subsequent PCR in a 25 μ L final volume using 2.5 units of the Expand High FidelityPLUS enzyme (Roche Diagnostics Ltd, West Sussex, U. K.), primers at a final concentration of 1 μ M, and following the supplier's instructions. PCR reactions were performed according to the following programme: one cycle at 94°C for 2 min and 40 amplification cycles at 94°C for 30 s, 60°C (RbA1) or 65°C (RbA2 and RbB) for 30 s, and 72°C for 1 min. 10 μ L of each PCR product were then sequenced commercially (Macrogen, Amsterdam, Netherlands).

2.4. Suppression subtractive hybridisation (SSH)

The SSH method was performed to facilitate the identification of genes differentially expressed between normal and HCA samples. Total RNAs from 8 normal fish and 8 HCA fish samples were extracted using the High Pure RNA Tissue kit (Roche Diagnostics Ltd, West Sussex, UK) according to the supplier's instructions. RNA integrity was assessed by formaldehyde-agarose gel electrophoresis, and 8 samples from each treatment group (LCM-derived HCA and normal tissues respectively) were randomly selected and pooled together so two RNA pools were obtained with each sample having an equal concentration (312.5 ng). SMARTer PCR cDNA Synthesis Kit (Clontech, UK) reagents were used to generate cDNA and the Advantage 2 PCR Kit (Clontech, UK) was used for PCR reactions. The SSH procedure utilised the PCR-Select cDNA Subtraction Kit (Clontech, UK) with normal and HCA samples as the driver and tester respectively. The manufacturer's protocol was followed with the exception of phenol-chloroform extraction followed by purification using the NucleoSpin Gel and PCR Clean-up kit (Machery-Nagel, UK) as per manufacturer's instructions, which was substituted for column chromatography (to avoid the handling and use of phenol, a highly toxic chemical).

In order to generate sub-clones of varying sizes, the final PCR products resulting from SSH were run on a 1% TBE agarose gel, stained post-run with ethidium bromide (Invitrogen, Paisley, UK), to generate smears which were then cut from the gel in 4 size-related pieces, each of which was purified with the NucleoSpin Gel and PCR Clean-up reagents (Machery-Nagel, UK). Subcloning with blue/white screening was performed both directly with the PCR products and also with gel-purified products using either the Original TA Cloning Kit with the pCR2.1 vector (Life Technologies, UK) or the TOPO TA Cloning Kit For Sequencing with the pCR4-TOPO vector (Life Technologies, UK) according to the manufacturer's protocols. The chemically competent cells used were TOP10 *E. coli* (Life Technologies, UK) and MAX Efficiency *DH10B E.coli* (Life Technologies, UK). White colonies were used to inoculate overnight liquid cultures which were screened for an insert using PCR with M13 primers. Plasmids containing inserts were purified with

the Nucleospin Plasmid kit reagents (Machery-Nagel, UK) and sequenced (Macrogen Europe, Amsterdam, The Netherlands). Sequences were identified by protein and nucleotide BLAST searches (blastx and blastn) on the NCBI database with results giving E-value $> 10^{-5}$ considered a match.

2.5. Quantitative real-time PCR validation of SSH results

The following genes were selected for real-time PCR validation of the SSH results: *Ras-related C3 botulinum toxin substrate 1 (Rac1)*, proto-oncogene protein *c-fos*, and *cyclin-dependent kinase inhibitor 1B (CDKN1B)*. The same RNAs (n = 8 fish for each treatment) were used for RT-PCR as were used for the SSH experiment in addition to 4 extra available RNAs selected from the normal group. Total RNA concentrations were measured using a Qubit 1.0 Fluorometer (Life Technologies, UK) and reagents. cDNA synthesis was performed using 1 μ g total RNA and SuperScript® VILO cDNA Synthesis Kit reagents (Life Technologies, UK), followed by treatment with 0.5 μ L Ribonuclease H (RNase H) and its associated buffer (Thermo Fisher Scientific, Loughborough, UK) at 37 °C for 45 min.

RT-PCR reactions were performed on a CFX96 Real-Time PCR Detection System (BioRad, UK) in a total volume of 20 μ L consisting of 10 μ L FastStart Universal SYBR Green Master (Rox) (Roche, UK), 7 μ L molecular grade water (Fisher Scientific, UK), 2 μ L primer mix and 1 μ L template cDNA. Reactions were optimised using final primer concentrations of 100 nM for reference genes and 300 nM for genes of interest. Primer sequences and amplicon sizes are listed in Table 1. PCR products from each primer pair were sequenced to confirm target gene amplification. No-template cDNA controls were included with all runs to confirm the lack of amplification and melt peaks were also universally generated to confirm the absence of secondary products such as primer dimers and to demonstrate primer specificity. Primer efficiencies were calculated, according to the MIQE guidelines (Bustin et al., 2009), from at least four points over a 10X cDNA dilution range and were all within the 90% to 110% range (Table 1).

2.6. Statistical analyses

Any significant difference in biometric parameters (sex, body length, body weight, and FC) or *Rb* genetic profiles between the HCA and normal samples was assessed using generalised linear models (glm) in R (R Development Core Team 2013). The influence of the *Rb* genetic profiles on the phenotype was assessed using the glm function (binomial family, link logit) and excluding the non determined (nd) samples. The effects of the sex ratio on the biometric parameters was assessed using a non parametric Kruskal-Wallis test in R as the normality of the residuals was not respected.

As part of the SSH optimisation, two genes were considered for their suitability as reference genes for this dataset: *EF1* and *α TUB*. Expression data was generated for each gene and stability across normal and HCA was tested. An unpaired T Test was performed in both cases using GraphPad InStat v3 (GraphPad Software Inc., La Jolla, USA), and although mean expression levels were stable between treatments for *α TUB* ($p = 0.1055$), this was not the case for *EF1* ($p = 0.0127$). As such, the three target genes were normalised to *α TUB* alone using the $2^{-\Delta C_t}$ method (Livak and Schmittgen, 2001).

All subsequent data on the transcriptional response of the genes selected for the validation of the SSH results were tested for normality of the residuals and homogeneity of variance using GraphPad InStat v3 software. An unpaired t test was therefore performed for *cfos* in order to test for significance between normal and HCA samples, whereas the Mann-Whitney Test was performed for *Rac1* and *CDKN1B*. Statistical significance was accepted at the $p < 0.05$ level.

3. Results

3.1. Fish biometric data

There were no significant differences of sex ($p = 0.11$), length ($p = 0.198$), weight ($p = 0.348$) or FC ($p = 0.168$) between the (presumptively) normal fish and those with HCA (Table 2). In addition, no significant effect of the sex was found on the biometric parameters in the LCM-derived samples

(length: $p = 0.08$; weight: $p = 0.19$; FC: $p = 0.26$) and the SSH (length: $p = 0.10$; weight: $p = 0.10$; FC: $p = 0.74$).

3.2. *Rb* mutational profile characterisation

Rb genetic profiles were characterised in LCM-derived normal and HCA samples of North Sea dab. Four nucleotides were identified as changed in the *Rb* coding sequence at 996 bp (G to A), 1088 bp (T to C), 1514 bp (G to T) and 1592 bp (G to T) leading to several different genetic profiles (Table 3). All of these changes occurred within the *Rb* sequence encoding the functionally important and conserved A and B domains. However, the *Rb* genetic profiles from normal and HCA samples showed no significant difference ($p = 0.311$).

3.3. SSH analysis

A number of differentially expressed genes, both up-regulated and down-regulated in HCA samples, were identified from the SSH experiment based on sequence similarity with other NCBI database sequences (Table 4). All dab sequences obtained here were also submitted to the database and were given accession numbers. Sequenced clones that generated no named matches, or matches below the E-value 10^{-5} cut-off, were not included.

3.4. Validation of differentially expressed transcripts

Relative expression levels of the 3 selected genes used for qPCR validation of SSH are shown in Figure 3. No significant differences between normal and HCA samples were detected for *RAC1* ($p = 0.0691$), *c-fos* ($p = 0.4893$) or *CDKN1B* ($p = 0.3837$).

4. Discussion

Herein, we adopted two parallel approaches, *Rb* allele frequencies and global differential gene expression, to characterise molecular-level hallmarks of HCA development. The biometric

parameters used showed that no significant difference occurred between the apparently normal and HCA phenotype samples analysed. This is consistent with a previous comprehensive study, using 165 dabs and 6 sites, whereby the HSI was not significantly different between sites with high prevalence of tumour-bearing individuals and reference sites (Lerebours et al. 2014).

In terms of *Rb* genetic profiles observed in the LCM derived samples, four nucleotide positions were altered in a manner similar to that previously identified in dab displaying liver HCC and HCA lesions captured at sites of differing contaminant burdens in the North Sea (Lerebours et al., 2014). These four positions correspond to a region of functional importance of the *Rb* gene, leading to 8 genetic profiles previously identified (Lerebours et al., 2014) along with two new profiles both observed in samples dissected from fish captured at the North Dogger site (Table 3). In this study, albeit using a low sample number (n=8 fish), there is no apparent association between *Rb* profiles and HCA phenotype. This is in contrast to previous work (with a higher sample number of n=165 fish and control sampling sites in the English Channel), whereby *Rb* profiles were not randomly distributed and specific *Rb* profiles were associated with different phenotypes i.e. normal, pre-neoplastic and neoplastic (Lerebours et al., 2014). Interestingly, *Rb* alleles previously associated with pre-neoplastic lesions (allele profiles N and M, Lerebours et al., 2014) are observed within the ‘normal’ tissue samples analysed herein (Table 3). It is possible that a degree of phenotypic latency exists whereby molecular events take place before neoplastic alterations are observed. Conversely, profiles previously associated with normal histopathology (allele profile A, Lerebours et al., 2014), are observed within the ‘HCA’ samples analysed herein (Table 3). However, the dissection of the liver in the previous study may have included different populations of cells, normal and neoplastic. Similarly, the present study has shown several *Rb* allele genotypes obtained within a laser micro-dissected homogeneous population of cells. Additionally, other molecular events may have a role in the neoplastic development. Previous work conducted using dab samples has suggested the involvement of epigenetic mechanisms with a decrease in the global methylation in the

development of hepatocellular neoplasm (Mirbahai et al., 2011). A combination of genetic and epigenetic changes may thus be involved in the neoplastic transformation of cells.

Using the parallel SSH approach, we generated libraries enriched for genes that varied between normal and HCA samples. A total of 82 successfully ligated plasmids were cloned, isolated and sequenced. The success rate for unique sequences obtained from the SSH experiment was 30.5%, which is comparable to similar studies using non-model fish (8%) (Urbatzka et al., 2013); lower relative to studies using fish model organisms (45-80%) (Marchand et al., 2006; Luckenbach et al., 2008; Leguen et al., 2015); and an improvement compared with invertebrate species such as molluscs (6-22%) (Boutet et al., 2008; Craft et al., 2010; Ciocan et al., 2011). Duplicate sequences accounted for 27% of clones and the remaining 43% of clones could not be identified based on sequence similarities.

A number of transcripts highlighted by the SSH approach were isolated (Table 4) and a few were selected for validation by qPCR. No significant difference in mRNA transcript expression was detected for any of the three genes (*Rac1*, *c-fos*, and *CDKN1B*) selected (Figure 3). This may be a result of low sample numbers, or reflect inter-individual variability, or represent an actual lack of change in mRNA expression between the (presumptively) normal and HCA liver samples. As such, further analysis is required, using the sequences obtained herein, to facilitate further validation. The following transcripts highlighted and discussed should thus be interpreted accordingly.

mRNA transcripts up-regulated in dab liver adenoma cells relative to normal hepatocytes

In HCA samples (Figure 2), sequences associated with cell signalling, cell cycle, gene expression regulation, protein transport and protein degradation were isolated (Table 4). Several genes involved in cell signalling were up-regulated, according to the SSH technique, in adenoma samples. For example, *arrestin domain containing 3* (*arrdc3*) encodes a protein in mammals that plays a role in regulating cell surface expression of G protein-coupled and adrenergic receptors (Qi

et al., 2014). Relevantly, *arrdc3* up-regulation has been associated with the suppression of cancer cell proliferation, migration and invasion in breast tumour development (Draheim et al., 2010). Of the other cell signalling transcripts, *Rac-1* encodes a G protein belonging to the Rho family of GTPases, which is also implicated in regulation of cancer cell migration (Oppelt et al., 2014). Up-regulation of *Rac-1* has also previously been associated with progression of testicular and breast cancer in mammals (Kamai et al., 2004; Baugher et al., 2005). *Calmodulin-like* gene was also observed as up-regulated in HCA samples relative to the normal cell samples (Table 4). Calmodulin is a calcium binding messenger protein that is ubiquitous in the cytoplasm and mediates the control of several enzymes, ion channels, aquaporins and other proteins. Its up-regulation has been associated with parathyroid adenomas in mammals (Oldham et al., 1982), although there are no previous reports related to HCA development.

Tribbles homolog 1-like transcript has been identified as similarly up-regulated in the dab HCA samples using the SSH technique. *Tribbles* was originally identified as a cell cycle regulator in mammals but is also considered as an adaptor in signalling pathways involved in tumour development (Yokoyama and Nakamura, 2011; Sakai et al., 2016). Relevantly, the mechanism of tribbles-induced action is thought to involve the ubiquitin-proteasome system, of which a *26S proteasome non-ATPase regulatory subunit-like* transcript and a *ubiquitin-conjugating enzyme E2 G1-like transcript*, are also among the up-regulated transcripts in the same cells sampled (Table 4). Of the remaining transcripts identified as up-regulated using the SSH approach, *myosin light peptide* is worthy of highlighting due to its possible contributory role in proliferation of breast cancer cells (Cui et al., 2010).

mRNA transcripts down-regulated in dab liver adenoma cells relative to normal hepatocytes

In the HCA samples (Figure 2), further sequences associated with cell signalling and cell cycle control were highlighted as down-regulated using the SSH approach relative to control hepatocytes (Table 4). Of particular note, *Ankyrin (ANKS1B)* encodes a tyrosine kinase protein

involved signal transduction and apoptosis, and has been previously reported as down-regulated in smoking related clear cell renal carcinoma (Eckel-Passow et al., 2014). *ANKS1B* down-regulated expression has accordingly been suggested as a potential early warning detection marker of tumour development and prevention (Eckel-Passow et al., 2014). C6orf58 represents another signalling peptide which is involved in liver development and has relevantly been identified as mutated/dysfunctional in HCC (Li et al., 2011). Similarly, the *RIMS2-like* transcript encodes a protein that primarily tethers calcium channels to mediate synapsis in mammals (Kaesler et al., 2012), yet it has not previously been observed as having any role in tumour development.

Cell cycle control elements down-regulated in HCA cells compared with the normal hepatocytes includes: *cyclin T2-like*, *CDKN1B* and *RhoA* (Table 4). Cyclin T2 has been identified as interacting with cyclin dependent kinases (cdks) and the tumour suppressor gene retinoblastoma, each of which is involved in cell cycle progression (Simone et al., 2002). *CDKN1B* (also known as *p27/kip1*) expression has been reported as frequently down-regulated in many human cancers (Garcia-Fernandez et al., 2011). *RhoA-like* encodes a small GTPase protein in mammals and its activity is often lost in ras-mediated tumourigenesis (Zandvakili et al., 2015). Of the remaining down-regulated transcripts, *c-Fos* represents a transcription factor protein that also regulates cell signaling and proliferation in mammals but is usually up-regulated in tumourigenesis, including HCC (Yeun et al., 2001), in contrast to the findings here.

Several of the gene expressions observed in our dab HCA samples (using the SSH technique) are consistent with those reported previously in dab HCA tissue reported by Small et al. (2010). Using a flounder cDNA microarray approach with 6 individual dab, Small et al. (2010) identified multiple 60S ribosomal proteins, a complement component C8 protein, a Rag C (ras related) GTP binding protein and a liver metabolism related protein aldose reductase, as all being similarly differentially expressed in the HCA samples analysed. In contrast, vitellogenin was reported as highly up-regulated in (a) male dab samples displaying HCA or HCC and (b) female

dab displaying HCC but not in HCA. However, in this study (using mixed sex) a down-regulation was observed in HCA cell samples relative to microdissected normal hepatocytes.

This study on normal and neoplastic hepatocytes from dab has allowed (i) a comparison of *Rb* sequence profiles of cells displaying HCA versus normal ones and (ii) a comparison of the transcriptome. This study is the first to provide information specific to the neoplastic pathway of HCA in the flatfish dab and offers a number of differentially regulated genes for future analysis. The approach presented here adds to our understanding of the mechanisms of liver tumors in sentinel marine flatfish. This greater insight into the drivers of disease will further aid in the interpretation of field data where fish are increasingly being used as sentinels of wider environmental health (Stentiford et al., 2010; Lang et al., 2015; Lyons et al., 2015).

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Figure and Table Legends

Figure 1. Overview of the methods used to generate the samples used in the various analyses.

Figure 2. Image of a frozen section, collected on a glass slide, and showing a benign HCA. The arrow shows the edge of the HCA, which can be distinguished by architectural alterations compared to the surrounding parenchyma. The magnification used is x20, H and E stain.

Figure 3. Relative gene expression of normal and HCA samples for (A) *Rac1*, (B) *c-fos*, and (C) *CDKN1B* with mean data plotted (+SD); n=12 fish for normal and n=8 fish for HCA. **

Table 1. Primer pairs used for *Rb* mutational profile analysis and qPCR validation of SSH results.

Table 2. Biometric data for body length (L), body weight (BW), Fulton index Condition (FC)($FC = BW/L^3 \times 100$) separated for male and female. Data are expressed as mean (SD) for each sex group.

Table 3. *Rb* genetic profiles found in LCM-derived liver HCA cells and normal hepatocytes. **Rb* profile as previously reported (Lerebours et al., 2014).

Table 4. Dab partial *cDNA* sequences obtained from SSH which are either up-regulated or down-regulated in HCA samples relative to normal hepatocytes, and their identifications based on sequence similarity obtained by NCBI database BLAST searches.

Figure 1.

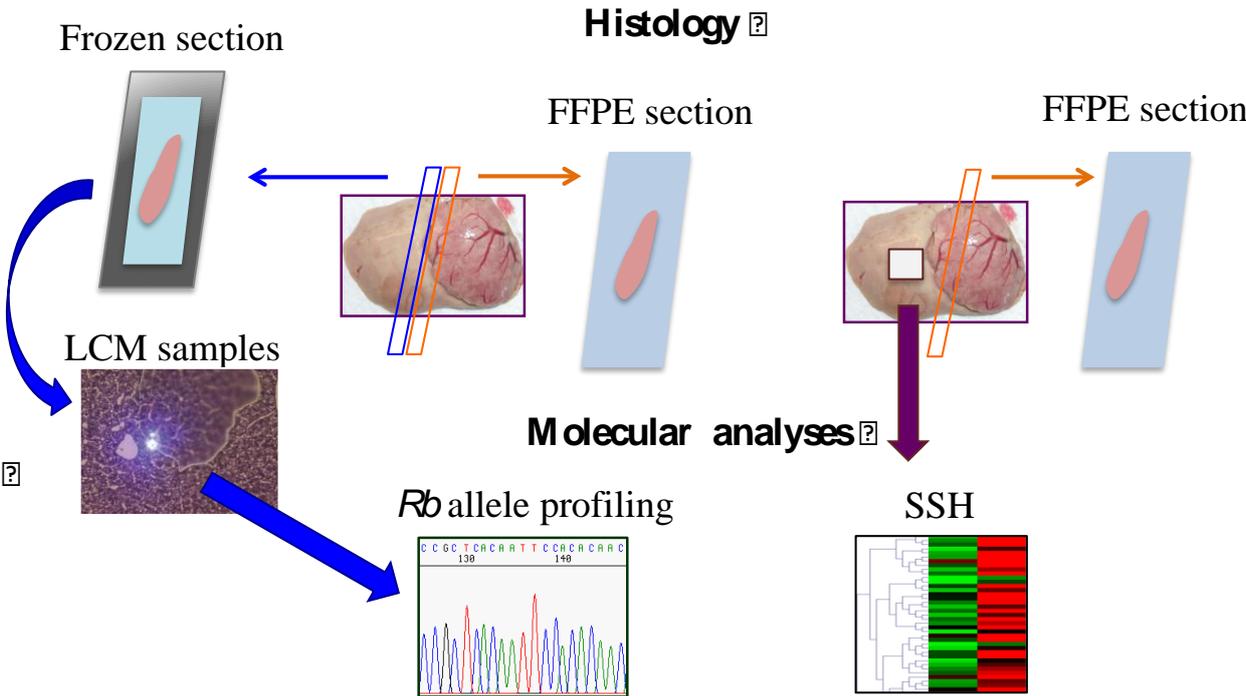


Figure 2.

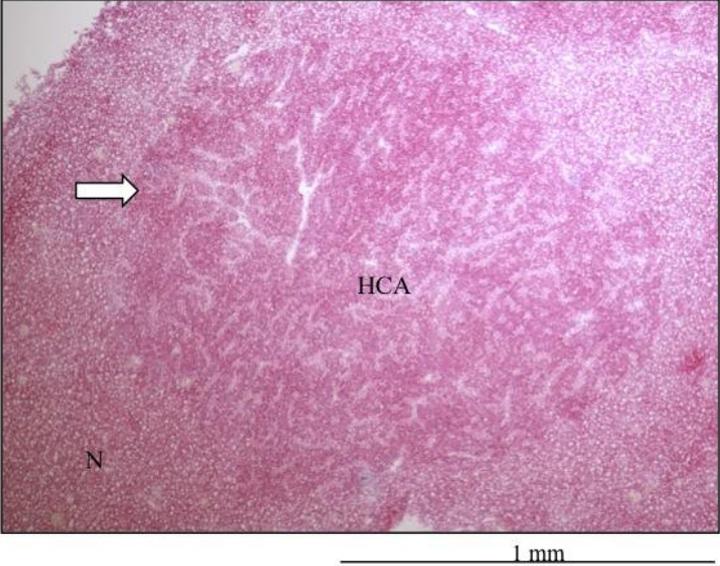


Figure 3.

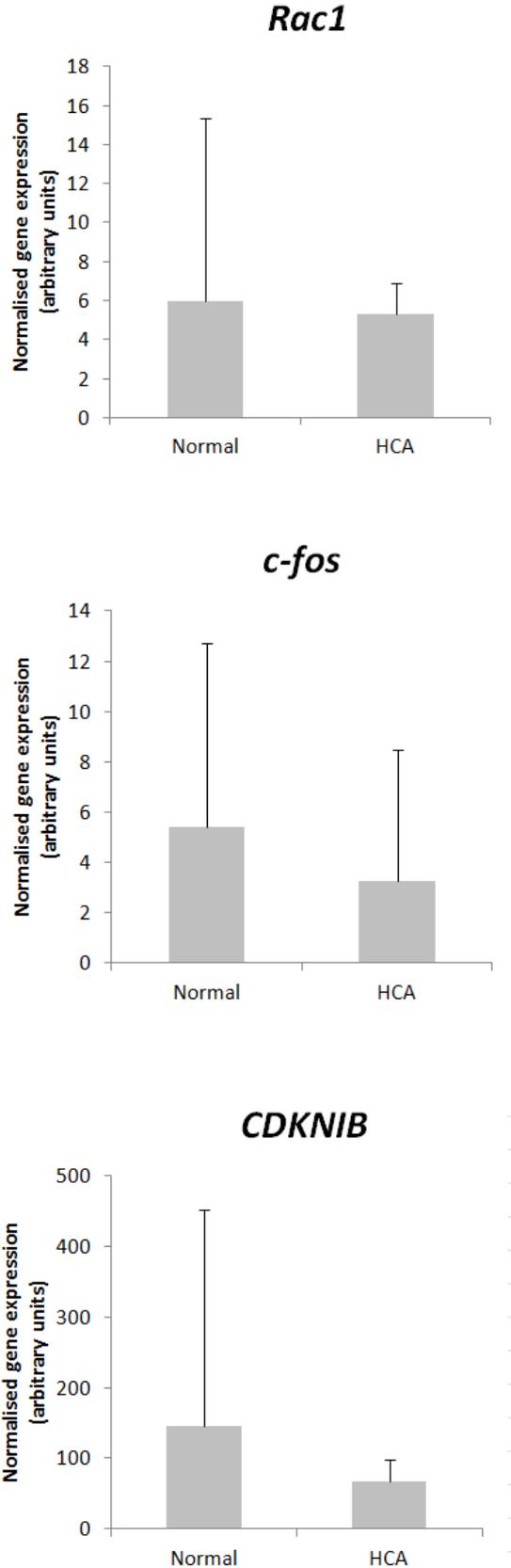


Table 1.

| Gene | Primer | Sequence 5'-3' | Amplicon size (bp) | Amplification efficiencies (%) |
|----------------------------|----------------|-------------------------|---------------------------|---------------------------------------|
| <i>EF1</i> | <i>EF1_F1</i> | AGTGCGGAGGAATCGACAAG | 286 | 103.3 |
| | <i>EF1_R1</i> | CCTCGAACTCACCAACACC | | |
| <i>αTUB</i> | <i>TUB_F1</i> | TCAATCACAGCCTCACTTCG | 197 | 94.2 |
| | <i>TUB_R1</i> | ATCTGATTGGCTGGCTCAAA | | |
| <i>c-fos</i> | <i>cFos_F1</i> | TCGATGTGTCTTCAGAGCAA | 93 | 90.4 |
| | <i>cFos_R1</i> | TTTCTTTTCAAAACAGCTCACA | | |
| <i>CDKN1B</i> | <i>CDKI_F3</i> | AGTCTCCTGAAGGACGAGGAG | 110 | 101.2 |
| | <i>CDKI_R3</i> | CACACTGCTACATTGACTGCTTT | | |
| <i>Rac1</i> | <i>Rac1_F3</i> | TGTATCACACTTCACTCACTGT | 102 | 102.2 |
| | <i>Rac1_R3</i> | GCTAGGCTTGCAGGGAGATT | | |
| <i>Rb (620 – 1070 bp)</i> | <i>RbA1_F</i> | AATCAGAGCTGCCATGACCT | 450 | |
| | <i>RbA1_R</i> | CCAGGGGAAACAAACATCTG | | |
| <i>Rb (983 – 1437 bp)</i> | <i>RbA2_F</i> | GGCAGCATATGGAGAGAGCGG | 454 | |
| | <i>RbA2_R</i> | GAGCAGGCGGCTGGGTTGG | | |
| <i>Rb (1350 – 1942 bp)</i> | <i>RbB_F</i> | CGTCCGGGCCATCGTGTCTT | 592 | |
| | <i>RbB_R</i> | ACGTTGTTGCTGCCAGGCACA | | |

Table 2.

| Technique | Phenotype | Sex (<i>n</i> fish) | Length (cm) | Weight (g) | FC |
|------------------|------------------|----------------------------|--------------------|-------------------|-----------|
| LCM | Normal | F (<i>n</i> =8) | 23 (1.3) | 112.3 (24.6) | 0.9 (0.1) |
| | | M (<i>n</i> =3) | 22.1 (0.7) | 91.3 (10.3) | 0.8 (0.0) |
| | HCA | F (<i>n</i> =5) | 24.8 (3.0) | 146.4 (87) | 1.0 (0.1) |
| | | M (<i>n</i> =2) | 21.8 (0.3) | 98.3 (16.7) | 0.9 (0.1) |
| SSH and qPCR | Normal | F (<i>n</i> =7) | 26.2 (1.9) | 167.3 (31.5) | 0.9 (0.1) |
| | | M (<i>n</i> =1) | 17.5 | 48.4 | 0.9 |
| | HCA | F (<i>n</i> =8) | 28.5 (4.2) | 231.5 (81.7) | 1.0 (0.2) |
| qPCR only | Normal | F (<i>n</i> =4) | 24.8 (2.2) | 132.1 (36.3) | 0.8 (0.1) |

The additional 4 normal samples that were used for qPCR validation but were not in the SSH can be added to the new row on the table above. The sample numbers are: 267, 269, 410 and 411. Done

Table 3.

| Sampling site (sex) | <i>Rb</i> genetic alterations | | | | Rb profile* |
|--|-------------------------------|--------|--------|--------|-------------|
| | 996bp | 1088bp | 1514bp | 1592bp | |
| <i>Hepatocellular adenoma (HCA) n=7 fish</i> | | | | | |
| North East Dogger (F) | G | T | G | G | A |
| Central Dogger (M) | G | T | G | G | A |
| North Dogger (M) | G | nd | G | G | nd |
| North Dogger (F) | G | T/C | G/T | G/T | B |
| North Dogger (F) | G | nd | T | T | nd |
| West Dogger (F) | G | T/C | T | T | O |
| West Dogger (F) | G | T/C | G/T | G/T | B |
| <i>Normal n=11 fish</i> | | | | | |
| North East Dogger (M) | G | T/C | G/T | G/T | B |
| North East Dogger (F) | G | C | G/T | G/T | P |
| North East Dogger (F) | G | T/C | G | G/T | N |
| North East Dogger (M) | G | T/C | G/T | G/T | B |
| North East Dogger (M) | G | T/C | G/T | G/T | B |
| North East Dogger (F) | G | T/C | G/T | G/T | B |
| Central Dogger (F) | G | T/C | G | G/T | N |
| Central Dogger (F) | G | T | G | G | A |
| North Dogger (F) | G | T/C | G/T | G/T | B |
| North Dogger (F) | G | C | G | G | M |
| West Dogger (F) | G | T | G | G | A |

1 **Table 4.**

| Clone accession number | Gene identity | Length (bp) | Species match | Accession number of match | e-value |
|--|--|-------------|---------------------------------|---------------------------|----------|
| Up-regulated in adenoma samples | | | | | |
| JZ714697 | Arrestin domain containing 3 (arrdc3) | 515 | <i>Oreochromis niloticus</i> | XM_005457484.1 | 1.00E-27 |
| JZ714698 | Calmodulin-like | 275 | <i>Pundamilia nyererei</i> | XM_005735127.1 | 2.00E-28 |
| JZ714699 | Ras-related C3 botulinum toxin substrate 1 (Rac1)* | 258 | <i>Psetta maxima</i> | FJ361904.1 | 4.00E-99 |
| JZ714701 | Coatomer subunit delta-like (ARCN1) | 210 | <i>O. niloticus</i> | XM_005462437.1 | 3.00E-39 |
| JZ714702 | Tribbles homolog 1-like | 295 | <i>O. niloticus</i> | XM_003444147.2 | 2.00E-31 |
| JZ714703 | Transcription factor AP-1-like | 132 | <i>Haplochromis burtoni</i> | XM_005923214.1 | 5.00E-35 |
| JZ714704 | Ubiquitin-conjugating enzyme E2 G1-like | 380 | <i>Lepisosteus oculatus</i> | XM_006640271.1 | 1.00E-45 |
| JZ714700 | 26S proteasome non-ATPase regulatory subunit 2-like | 200 | <i>Neolamprologus brichardi</i> | XM_006801164.1 | 1.00E-72 |
| JZ714705 | Myosin light polypeptide 6 putative mRNA | 180 | <i>Anoplopoma fimbria</i> | BT083308.1 | 8.00E-22 |
| Down-regulated in adenoma samples | | | | | |
| JZ714707 | Ankyrin repeat and sterile alpha motif domain-containing protein 1B-like (ANKS1B-like) | 351 | <i>H. burtoni</i> | XM_005947551.1 | 8.00E-63 |
| JZ714710 | UPF0762 protein C6orf58 homolog | 178 | <i>H. burtoni</i> | XM_005917613.1 | 3.00E-20 |
| JZ714711 | Regulating synaptic membrane exocytosis protein 2-like (RIMS2-like) | 218 | <i>O. niloticus</i> | XM_005472540.1 | 2.00E-51 |

| | | | | | |
|----------|--|------|---------------------------------|----------------|-----------|
| JZ714717 | Cytokine inducible SH2-containing protein (cish) | 227 | <i>Gasterosteus aculeatus</i> | NM_001267677.1 | 2.00E-47 |
| JZ714712 | Cyclin-T2-like | 217 | <i>O. niloticus</i> | XM_005450425.1 | 1.00E-21 |
| JZ714713 | Cyclin-dependent kinase inhibitor 1B (CDKN1B) putative mRNA* | 510 | <i>A. fimbria</i> | BT082547.1 | 1.00E-62 |
| JZ714721 | Transforming protein RhoA-like | 199 | <i>O. niloticus</i> | XM_003448207.2 | 8.00E-35 |
| JZ714708 | Proto-oncogene protein c-fos* | 266 | <i>Dicentrarchus labrax</i> | DQ838581.1 | 2.00E-78 |
| JZ714720 | U1 small nuclear ribonucleoprotein A-like | 202 | <i>Neolamprologus brichardi</i> | XM_006795364.1 | 3.00E-08 |
| JZ714714 | 60S ribosomal protein L13-like | 336 | <i>O. niloticus</i> | XM_003442274.2 | 7.00E-118 |
| JZ714715 | 60S ribosomal protein L19-like | 322 | <i>O. niloticus</i> | XM_003442012.2 | 1.00E-105 |
| JZ714706 | 60S ribosomal protein L5 (rpl5 gene) | 350 | <i>Platichthys flesus</i> | AJ843088.1 | 3.00E-146 |
| JZ714716 | Complement component C9 | 428 | <i>Paralichthys olivaceus</i> | AB020963.1 | 7.00E-53 |
| JZ714718 | Glucose-fructose oxidoreductase domain-containing protein 1-like | 1064 | <i>O. niloticus</i> | XM_005471621.1 | 2.00E-13 |
| JZ714709 | Vitellogenin | 271 | <i>Pleuronectes platessa</i> | AJ416328.1 | 3.00E-115 |
| JZ714719 | Vitellogenin C | 170 | <i>Thunnus thynnus</i> | GU217573.1 | 9.00E-54 |

