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1 **Changes of sewage sludge digestate-derived biochar properties after chemical treatments**
2 **and the influence on As(III and V) and Cd(II) sorption**

3
4 Suchanya Wongrod^{a,b,c}, Stéphane Simon^{b,*}, Eric D. van Hullebusch^{a,c,d}, Piet N.L. Lens^c, Gilles
5 Guibaud^b

6
7 ^a Université Paris-Est, Laboratoire Géomatériaux et Environnement (EA 4508), UPEM, 77454,
8 Marne-la-Vallée, France

9 ^b Université de Limoges, PEIRENE, Equipe Développement d'indicateurs ou prévision de la
10 qualité des eaux, URA IRSTEA, 123 avenue Albert Thomas, 87060 Limoges, France

11 ^c UNESCO-IHE Institute for Water Education, P.O. Box 3015, 2601 DA Delft, The Netherlands

12 ^d Institut de Physique du Globe de Paris, Sorbonne Paris Cité, Université Paris Diderot, UMR
13 7154, CNRS, F-75005 Paris, France

* Corresponding author. Université de Limoges, PEIRENE, Equipe Développement d'indicateurs ou prévision de la qualité des eaux, URA IRSTEA, 123 avenue Albert Thomas, 87060 Limoges, France
Email: stephane.simon@unilim.fr

14 **Abstract**

15 This work seeks to extend the knowledge on the effect of chemical treatments of sewage
16 sludge digestate (SSD)-derived biochar for As(III and V) and Cd(II) sorption ability using
17 potassium hydroxide (KOH) or hydrogen peroxide (H₂O₂). Results showed the increases of pH of
18 point of zero charge, the Brunauer-Emmett-Teller (BET) surface area and cation exchange
19 capacity (CEC) after chemical treatments of biochar. The sorption ability was enhanced from 1.6
20 $\mu\text{mol g}^{-1}$ (As(V)) and 16.1 $\mu\text{mol g}^{-1}$ (Cd(II)) on raw biochar to 8.5 $\mu\text{mol g}^{-1}$ (As(V)) and 318.5
21 $\mu\text{mol g}^{-1}$ (Cd(II)) on KOH-modified biochar. Furthermore, arsenic redox distribution showed a
22 large oxidation (70%) of As(III) to As(V) in KOH-biochar with batch washing, while a partial
23 oxidation (7%) was observed in KOH-biochar with batch and subsequently column washing. The
24 washing procedures after KOH treatment play an important role on arsenic sorption, due to the
25 release of phosphate (PO_4^{3-}) as well as organic matter from the biochar that may subsequently
26 lead to the oxidation of As(III) to As(V). Our findings highlight the potential influence of biochar
27 on the redox transformation of As(III) to As(V) and therefore require a careful assessment while
28 investigating the fate of As in aquatic environments.

29
30 Keywords: Sewage sludge digestate derived biochar, chemical activation, As(III), As(V), Cd(II),
31 biochar washing

32

33 1. Introduction

34 Arsenic (As) and cadmium (Cd) contamination in water creates a pressing toxicity
35 towards aquatic organisms. Arsenic is mostly present as two inorganic species, arsenite (As(III))
36 and arsenate (As(V)), whereas a divalent cadmium ion (Cd(II)) dominantly prevails in water. Due
37 to the toxicity of As and Cd, numerous treatments have been dedicated to removing these
38 elements from polluted water. Among them, sorption is a cost-effective and simple treatment for
39 metal(loid) removal from water (Ding et al., 2016; Ofomaja et al., 2014; Peng et al., 2018; Wang
40 et al., 2016; Zhou et al., 2017). In recent years, biochar, a black solid char obtained from
41 pyrolysis (> 250 °C) of biomass under oxygen-limiting conditions (Ahmad et al., 2014), has been
42 considered as a potential alternative to activate charcoal in sorption treatments. Sewage sludge
43 digestate (SSD) obtained from wastewater treatment plants is currently considered as a promising
44 material to produce a low-cost biochar (Agrafioti et al., 2013; Rajapaksha et al., 2014; Yuan et
45 al., 2015; Zielińska and Oleszczuk, 2015).

46 During the pyrolysis step of biochar preparation, the organic matter from biomass
47 undergoes structural modifications such as oxidation and conversion of aliphatic forms to
48 aromatic forms (Zama et al., 2017). The resulting biochar generally harbors negative and
49 positive charges on its surface depending on the neighboring pH. Consequently, at circumneutral
50 pH, where negative charges are predominant, it has been applied to sorb cationic metals like lead
51 (Pb), zinc (Zn) and cadmium (Cd) from polluted water (Ding et al., 2016; Ho et al., 2017; Park et
52 al., 2017; Wongrod et al., 2018). Unfortunately, the sorption ability of SSD biochar for
53 metal(loid)s, particularly As oxyanions, is relatively low compared to activated carbon or paper
54 mill sludge biochar (Table S1, supplementary information). Therefore, biochar modifications like
55 physical activation and chemical treatment have been applied to improve its sorption ability (Li
56 et al., 2017; Rajapaksha et al., 2016; Sizmur et al., 2017). Even though there are many studies on
57 the adsorption capacity of raw and modified biochars (Liu et al., 2017; Zuo et al., 2017), an
58 evaluation of the modification of the biochar properties is yet to be done. Indeed, the biochar
59 surface properties are important parameters that influence the behavior of a sorbent to sorb
60 anionic and cationic pollutants.

61 The present work aimed to study the consequences of chemical treatments of SSD biochar
62 using KOH or H₂O₂ on its arsenic and cadmium sorption capacities. Batch experiments were
63 performed to evaluate the sorption kinetics and capacity of raw and modified biochars for As(III,
64 V) and Cd(II). Several surface properties, such as the pH of point of zero charge (pH_{PZC}), the
65 cation exchange capacity (CEC) and the Brunauer-Emmett-Teller (BET) surface area were
66 determined to access information on, respectively, the net surface charge, cation exchange
67 capacity and available specific surface area of the SSD biochars. These parameters can be used to
68 discuss the sorption characteristics of the biochar. The influence of chemical modifications of the
69 biochar onto sorption of redox-sensitive elements was also investigated through the determination
70 of arsenic redox transformation (*i.e.* As(III) and As(V)) during sorption experiments. Finally, the
71 consequences of biochar washing on all these parameters were considered by comparing the
72 results obtained with a KOH-modified biochar submitted either to only a batch washing or to a
73 subsequent continuous column washing.

74

75 2. Materials and methods

76 2.1 Chemical reagents

77 Cadmium ($\text{Cd}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$, 99% w/w, Merck), arsenate ($\text{Na}_2\text{HAsO}_4 \cdot 7\text{H}_2\text{O}$, 98% w/w,
78 Sigma-Aldrich) and arsenite (AsNaO_2 , 98% w/w, Merck) were used to prepare 10 mM of Cd(II),
79 As(V) and As(III) stock solutions, respectively. The ultrapure water (18.2 M Ω , MilliQ Gradient
80 A10, Millipore SAS 67120, Molsheim, France) was used to wash biochars, as well as to prepare
81 and dilute all solutions and samples.

82

83 2.2 Biochar preparation and chemical treatment

84 Pellets of domestic sewage sludge digestate (SSD) were obtained from a wastewater
85 treatment plant (WWTP) located in Limoges, France (size: 285,000 equivalent population). The
86 main treatment processes of the WWTP include activated sludges with anoxic treatment and
87 addition of iron(III) chloride (FeCl_3) to, respectively, remove nitrogen(N) and phosphorus (P).
88 Particularly, the SSD sample was collected in the storage tank after dewatering and drying
89 processes.

90 The sample was heated at 350 °C for 15 min under slow pyrolysis technology to produce
91 SSD biochar and more information was described in our previous work (Wongrod et al., 2018).
92 The SSD biochar was chemically treated for 2 h with a 10% H_2O_2 solution (100 g: 1 l) or with 2
93 M KOH solution (100 g: 2.5 l) (Wongrod et al., 2018).

94 The raw and chemically-modified biochars were subsequently washed as follows: a triple
95 batch washing in a row followed by a continuous column washing (Wongrod et al., 2018). These
96 washing steps were performed to remove competitive ions (*e.g.* PO_4^{3-} , $\text{HCO}_3^-/\text{CO}_3^{2-}$, Ca^{2+} and
97 Mg^{2+}) and organic matter released by the biochar during the chemical treatment. These released
98 compounds can react with elements and thus interfere with the biochar-metal(loid) sorption
99 system. In addition, clogging of the biochar pore sites by small particles can be eliminated
100 through these washing steps. To determine if the washing procedure influence the metal(loid)
101 sorption, a KOH-modified biochar was washed only with a batch procedure. The raw SSD
102 biochar, its H_2O_2 and KOH-modified SSD biochar are denoted as, respectively, BSS, BH_2O_2 and
103 BKOH. The KOH-modified biochar washed with only the batch procedure is labeled as
104 BKOH^{bat} .

105

106 2.3 Biochar characterization

107 X-ray powder diffraction (XRD) and Fourier transform infrared spectroscopy (FTIR) of
108 the SSD and its derived biochar were analyzed to observe the changes of, respectively, crystalline
109 structures and surface functional groups after biochar production. The total amount of metal and
110 metalloid in the SSD and its derived biochar was determined using acid digestion from the EPA
111 Method 3050B (EPA, 1996). The details of the digestion procedures are described by Wongrod et
112 al. (2018).

113 The pH (pH-meter LPH 330T, Tacussel, France) and electrical conductivity (EC) (CDM
114 210 conductivity-meter, Radiometer, Denmark) of the biochar were measured in water. The

115 surface area of biochar was measured using N₂ sorption by the Brunauer-Emmett-Teller (BET)
116 method at 77 K (3Flex, Micromeritics, USA). Pretreatment of biochar was performed by drying
117 at 105 °C for 5 h prior to the BET surface area (S_{BET}) analysis.

118 The pH of point of zero charge (pH_{PZC}) of the biochar was determined from zetametry by
119 determining the zeta potential at different pH ranges (Mahmood et al., 2011). For the
120 determination of the cation exchange capacity (CEC), 2 g of biochar (dry weight) were mixed
121 with 40 ml of 0.05 N cobalt hexamine trichloride ([Co(NH₃)₆]Cl₃) solution (99% w/w, Sigma-
122 Aldrich) in polypropylene tubes (Aran et al., 2008). The details of the biochar preparation for
123 pH_{PZC} and CEC measurement are provided in the supplementary information.

124

125 *2.4 Sorption experiments*

126 In sorption kinetic experiments, 10 µM As(III), As(V) and Cd(II) were prepared
127 separately from stock solutions of 1 mM As(III), As(V) and Cd(II) (as AsNaO₂, NaHAsO₄.7H₂O
128 and Cd(NO₃)₂, respectively). Raw and modified biochars (600 mg) were added to 150 ml of
129 metal(loid) solution. The initial pH was adjusted to 5 using HNO₃ and NaOH. The solutions were
130 agitated at (20 ± 2) °C at 180 rpm for 24 h using an orbital shaker (KS 501 digital, IKA™, USA).
131 The sorption experiments were performed in duplicate. Samples of solution were collected over
132 time. The total collected volume was less than 5% of the initial volume to prevent any alteration
133 of the sorption equilibrium. Sampled solutions were filtered through Lab Logistics Group (LLG)
134 cellulose acetate syringe filters (0.45 µm) and their As and Cd concentrations were determined by
135 graphite furnace atomic absorption spectrometry (240Z, Agilent Technologies, USA) at λ 193.7
136 nm and λ 228.8 nm for As and Cd, respectively. The pseudo-first-order (PFO) and the pseudo-
137 second-order (PSO) kinetics models were used to describe the experimental data. The
138 intraparticle diffusion model was also studied to access information on the mechanism of the
139 sorption processes (Zhao et al., 2015a).

140 Adsorption isotherms were performed with initial concentrations ranging from 10 to 300
141 µM for As(III, V) and from 10 to 4000 µM for Cd(II). After 24 h of agitation, the residual
142 concentrations of As were determined by graphite furnace atomic absorption spectrometry as
143 detailed previously, whereas the residual concentrations of Cd were determined by Microwave
144 Plasma Atomic Emission Spectroscopy (MP-AES) (Agilent 4100, USA) at λ 228.8 nm. Two
145 common adsorption isotherm models (Langmuir and Freundlich) were used to fit the
146 experimental data. Arsenic speciation was determined by High-Performance Liquid
147 Chromatography (HPLC) coupled to Atomic Fluorescence Spectrometry (AFS) with Hydride
148 Generation (HG) (PS Analytical Millennium Excalibur, P S Analytical, UK). For the
149 chromatographic conditions, a Hamilton PRP-X100 column was used with a phosphate buffer
150 solution as the mobile phase at a rate of 1 ml min⁻¹. Detailed information can be found in Wan et
151 al. (2014).

152

153 *2.5 Statistical analysis*

154 Results are reported as means ± standard deviation (n=3) unless otherwise stated.
155 Uncertainties of sorption parameters were obtained by non-linear regression using Statistica

156 software (v6.1, StatSoft). The t-test with two-tailed distribution at a statistical significance level
157 of $P \leq 0.01$ were used for data comparison.

158

159 3. Results and discussion

160 3.1 Influence of chemical treatment on biochar characteristics

161 The XRD analysis showed that quartz was present in both SSD and BSS with
162 insignificant modification of the surface properties after biochar pyrolysis (Fig. S1). FTIR spectra
163 demonstrated similar binding between atoms of both the SSD and BSS (Fig. S2). The absorbance
164 peaks of hydroxyl ($-\text{OH}$) groups (3425 cm^{-1}), $-\text{CH}_3$ stretching of aliphatic groups (2924 cm^{-1}),
165 $\text{C}=\text{C}$ stretching vibrations (1600 cm^{-1}), and $\text{C}-\text{C}$ skeleton and $\text{C}-\text{O}$ stretching (1033 cm^{-1}) were
166 mainly detected on the SSD and BSS (Fig. S2). These functional groups can be involved in the
167 sorption for metal(loid)s (Pituello et al., 2014).

168 The pH values of the suspensions of the raw and chemically-modified biochars are given
169 in Table 1. The KOH treatment induced a change from 6.4 (± 0.1) for BSS to 8.4 (± 0.1) for
170 BKOH and 10.1 (± 0.1) for BKOH^{bat} , whereas no significant difference ($P \leq 0.01$) was observed
171 after H_2O_2 treatment (Table 1). Results also showed no significant changes of the electrical
172 conductivity (EC) among each biochar suspension (*i.e.* BSS, BH_2O_2 and BKOH), except for
173 BKOH^{bat} where a 50-fold higher value ($324 \pm 2 \mu\text{S cm}^{-1}$) was observed. The high EC value of
174 BKOH^{bat} is probably due to a significant release of weakly sorbed ions (*e.g.* PO_4^{3-} , HCO_3^- , CO_3^{2-} ,
175 Ca^{2+} and Mg^{2+}) from BKOH^{bat} as previously reported by Wongrod et al. (2018). A previous study
176 published by Yuan et al. (2015) is in agreement with our finding, as the authors have reported a
177 very high EC of $7750 \mu\text{S cm}^{-1}$ in the sewage sludge biochar that was not washed with ultrapure
178 water after pyrolysis (Table 1).

179 Fig. 1 shows the evolution of the zeta potential according to pH values for BSS, BH_2O_2 ,
180 BKOH and BKOH^{bat} . The corresponding pH_{PZC} are given in Table 1. Results demonstrated
181 similar pH_{PZC} values for both BSS (~ 2.7) and BH_2O_2 (~ 2.9) biochars, whereas a slightly higher
182 value was obtained for BKOH (~ 3.4). Such limited variations of pH_{PZC} (2.7 – 3.4) are not prone
183 to significantly modify the pH on the As and Cd sorption, except if the manipulations are
184 performed at a pH lower than 5 (*i.e.* pH at sorption experiment, see section 2.4). In contrast, the
185 more negative values of zeta potential observed for BH_2O_2 and BKOH^{bat} (Fig. 1) could indicate
186 higher amounts of negative charges at the surfaces of these two modified biochars, resulting in
187 higher sorption capacities towards cationic metals.

188 The pH_{PZC} results showed that all biochars carried net negative charges at neutral pH (Fig.
189 1). At present, the pH_{PZC} of biochar has been rarely reported in the literature and there is a lack of
190 information to understand its variability among different chemical treatments. However, our
191 results are in agreement with a straw-derived biochar with a $\text{pH}_{\text{PZC}} \sim 1.9$ (Qiu et al., 2009) and a
192 paper mill sludge with pH_{PZC} ranging from 4.0 – 6.0 (Guo et al., 2018).

193 The BET surface area (S_{BET}) of the raw and modified biochars is given in Table 1. A
194 significant increase of the S_{BET} was observed for BH_2O_2 and BKOH, from $0.4 (\pm 0.1)$ to $5.7 (\pm$
195 $0.1)$ and $7.9 (\pm 0.1) \text{ m}^2 \text{ g}^{-1}$, respectively. This increasing S_{BET} is an indicator of higher porosity on
196 the modified biochar and thus a growing chance for metal(loid)s to sorb onto the biochar surface
197 (*e.g.* by complexation) (Mohan et al., 2014). Our findings are similar to those determined for

198 sewage sludge biochars ($4.0 - 14.3 \text{ m}^2 \text{ g}^{-1}$) (Agrafioti et al., 2013; Yuan et al., 2015) and
199 pineapple-peel-derived biochars ($0.7 - 2.1 \text{ m}^2 \text{ g}^{-1}$) (Wang et al., 2016). Nevertheless, these values
200 are still far lower than those reported for activated carbons ($1215.0 - 1316.0 \text{ m}^2 \text{ g}^{-1}$) (Gong et al.,
201 2015).

202 The CEC values determined for the different biochars are presented in Table 1. Results
203 showed a slight increase of CEC after treating the biochar with H_2O_2 from $1.9 (\pm 0.1)$ to $3.0 (\pm$
204 $0.1) \text{ cmol}^+ \text{ kg}^{-1}$, whereas a significantly increase (6 times) after the KOH treatment ($P \leq 0.01$)
205 was reported. This higher CEC of BKOH could induce an enhanced sorption ability towards
206 cationic metals. Previously published data showed variable high ranges of the CEC of the
207 biochars from $45.7 \text{ cmol}^+ \text{ kg}^{-1}$ to $483.4 \text{ cmol}^+ \text{ kg}^{-1}$ on wood and straw-based biochars,
208 respectively (Ding et al., 2016; Jiang et al., 2014).

209 The variations in the S_{BET} and CEC after chemical treatments of the biochars suggest that
210 H_2O_2 and KOH-modified biochars provided higher amounts of active pore sites, surface
211 functional groups and exchangeable cations that could enhance the sorption of metallic elements
212 on the biochar surface. For anion sorption, the increase of S_{BET} could influence the arsenic
213 sorption capacity (Table 1).

214 Concerning the effect of the washing procedures after KOH treatment, the results showed
215 significantly higher values of both pH and conductivity on BKOH^{bat} than BKOH (Table 1),
216 which were attributed to a release of ions such as PO_4^{3-} and $\text{HCO}_3^{2-}/\text{CO}_3^{2-}$ from the biochar
217 (Wongrod et al., 2018). As discussed previously (Wongrod et al., 2018), the insufficient batch
218 washing after biochar modification led to the release of ions (*e.g.* PO_4^{3-} , $\text{HCO}_3^-/\text{CO}_3^{2-}$, Ca^{2+} and
219 Mg^{2+}) and organic compounds from the biochar which interacted with metals and modified the
220 sorption behavior. In the case of As(V), PO_4^{3-} is expected to be a strong competitor, reducing its
221 sorption onto the biochar. The pH_{PZC} , S_{BET} and CEC values were also significantly affected by
222 the incomplete batch washing, resulting in an alteration of the biochar surface properties.

223

224 3.2 Effect of chemical treatment and washing on Cd(II) sorption

225 The results for the kinetic study of the Cd(II) sorption fitted better to the pseudo-second-
226 order (PSO) (Fig. 2a) than intraparticle diffusion (Fig. 2b) models. All values for the kinetic
227 model-fitting can be found in Table 2 and Table S2 (supplementary information). Results showed
228 that the sorption of Cd(II) by all biochars were well-described by the PSO kinetic model (Table 2
229 and Fig. 2a), literally suggesting that the sorption mainly occurred *via* simple chemical reaction
230 (Nethaji et al., 2013). Both the KOH and H_2O_2 -treated SSD biochars showed faster removal
231 kinetics than the raw biochar, reaching the equilibrium in less than 5 h for BH_2O_2 and BKOH,
232 and after 10 h for BSS (Fig. 2a). These results are consistent with a previous study with faster
233 kinetics on the KOH-modified switchgrass biochar than the raw biochar (Regmi et al., 2012). Fig.
234 2b shows that the plots did not pass through the origin as the C values are not equal to zero
235 (Table 2). This indicates some degree of boundary layer and the intraparticle diffusion was not
236 the only rate controlling step, but other mechanisms like film diffusion and external mass transfer
237 may also play important roles in the sorption (Nethaji et al., 2013).

238 Sorption isotherms are shown in Fig. 3, and the corresponding parameters can be found in
239 Table 3 and Table S3 (supplementary information). Cd sorption was better described by the

240 Langmuir model (Table S3), suggesting the possibility of a monolayer sorption of Cd onto the
241 homogeneous biochar surfaces (Zhao et al., 2015b). The Langmuir maximal sorption capacity,
242 Q_m , increased by a 15-fold to 20-fold factor after treatment with H_2O_2 and KOH, respectively
243 (Table 3). This can be linked to the previous observation that the chemical treatments not only
244 increase the S_{BET} , but also the CEC, resulting in an increase of the sorption sites and
245 exchangeable cations. The higher Q_m value of BKOH compared to BH_2O_2 is consistent with the
246 more pronounced improvement of both S_{BET} and CEC with the KOH treatment (Table 1).

247 The effect of the washing procedure applied after KOH treatment on the adsorption
248 kinetics and isotherms are shown in Fig. 2 and 3 and Tables 2 and 3. The sorption kinetic
249 constant for biochar with the KOH treatment was lower without the column washing step, but the
250 value was still higher than that for the raw biochar. This finding could result from the reduction
251 of S_{BET} for the insufficient batch washed BKOH^{bat}. A slightly higher Q_m was observed for
252 BKOH^{bat} compared to BKOH, meaning that the sorption of this cationic metal was not strongly
253 affected by the washing procedure. The release of ions and organic compounds from the
254 BKOH^{bat} did not appear to modify the Cd(II) behavior, in contrast to what was observed in a
255 previous study with Pb(II) (Wongrod et al., 2018). This could be due to a higher affinity of Cd(II)
256 for biochar sites (*i.e.* higher K_L values than Pb(II)) and/or weaker interactions between Cd(II) and
257 dissolved compounds in the solutions. For example in our findings, the K_L values of BKOH were
258 $0.032 \text{ l } \mu\text{mol}^{-1}$ and $0.007 \text{ l } \mu\text{mol}^{-1}$ for Cd(II) and Pb(II), respectively (Wongrod et al., 2018).

259

260 3.3 Arsenic sorption

261 3.3.1 Effect of chemical treatment of biochar on As(V) sorption

262 Fig. 4(a) shows the effect of chemical treatments on the adsorption kinetics for As(V)
263 onto BSS, BH_2O_2 and BKOH. All experimental data fitted with a pseudo-second-order kinetic
264 model ($R^2 \geq 0.98$, Table S2, supplementary information) and corresponding parameters are given
265 in Table 2. These results showed that the sorption kinetics were unchanged for BH_2O_2 , with a
266 sorption equilibrium reached after 10 h, while it was reached after 24 h for BKOH.

267 The sorption isotherms for As(V) are shown in Fig. 4(b). Similar to Cd, the Langmuir
268 model provided the best fit for As(V) sorption (Table S3, supplementary information). The
269 corresponding sorption parameters are given in Table 3. No significant evolution was observed
270 for the values of K_L , indicating that the chemical treatments did not modify the affinity of the
271 biochar for As(V). In contrast, the sorption capacity was enhanced by both H_2O_2 and KOH-
272 treated biochars (except BKOH^{bat}), and the Q_m values being, respectively, about 4 times and 5
273 times higher than the raw biochar.

274 These results indicate that H_2O_2 treatment only improved the sorption capacity, whereas
275 KOH treatment improved the sorption capacity but induced slower sorption kinetics for As(V)
276 (Tables 2 and 3). In both cases, the enhanced sorption ability for As(V) by the chemically-
277 modified biochars could be linked to the increase of the S_{BET} , which was slightly higher for
278 BKOH than BH_2O_2 (Table 1). These results are in agreement with previous studies (Ding et al.,
279 2016; Wu et al., 2017), who showed that the surface area plays an important role in the sorption
280 of elements onto biochar. The slightly increased pH_{pzc} of BKOH (Table 1) may result in a little

281 increase of the positive charges at the surface of the biochar, and thus enhance the sorption ability
282 for the anionic As(V).

283

284 3.3.2 Comparison of biochar washing

285 The effect of the washing procedure applied after KOH treatment on the adsorption
286 kinetics and isotherms for arsenic are presented in Fig. 5(a-d). The corresponding sorption
287 parameters are given in Tables 2 and 3. The experiments were performed for As(V) and As(III) to
288 determine if the washing procedure affects similarly both arsenic species, taking into account
289 that, in the experimental conditions, As(III) is neutral whereas As(V) is negatively-charged.

290 It appeared that the washing procedures of KOH-biochars had a low impact on the
291 sorption kinetics for As(III), k_2 values increasing from $0.20 (\pm 0.05) \text{ h}^{-1}$ to $0.38 (\pm 0.07) \text{ h}^{-1}$ with
292 the addition of the subsequent column washing step (Fig. 5a). In contrast, the addition of this step
293 strongly affected the sorption kinetics for As(V) (Fig. 5b), inducing a 3-fold decrease of k_2 (Table
294 2). The sorption isotherms showed an opposite trend: the column washing step only marginally
295 modified the Q_m of the biochar for As(III), but induced an almost 10-fold improvement for
296 As(V). This strong difference of the Q_m values for As(V) can be attributed to the PO_4^{3-} released
297 from BKOH^{bat} , as highlighted in a previous study (Wongrod et al., 2018). Indeed, the PO_4^{3-} may
298 strongly compete with As(V) for sorption onto the biochar. It has been previously established that
299 excessive PO_4^{3-} concentrations could mobilize As(V) from the sorbent (Neupane et al., 2014).
300 This is likely due to the similar chemical properties between PO_4^{3-} and As(V) since both
301 chemicals are negatively-charged oxyanions. Moreover, the leaching of dissolved organic carbon
302 (DOC) from the biochar (Wongrod et al., 2018) could accelerate the mobilization of As, as
303 observed in a previous study in a biochar-soil system (Kim et al., 2018). Concerning the sorption
304 constant K_L , there was no significant difference ($P \leq 0.01$) according to the washing procedures
305 for As(V) and As(III), indicating that no evolution in the affinity of biochar for both arsenic
306 species was observed. When comparing K_L values between As(III) and As(V), it appears that the
307 latter has less affinity for BKOH and BKOH^{bat} (Table 3). It could be hypothesized from these
308 results that arsenic species interact differently with the biochar surface or with different types of
309 surface functions.

310

311 3.3.3 Evolution of arsenic redox speciation

312 The arsenic redox speciation in solution was determined during As(III) and As(V)
313 sorption experiments in order to determine whether the biochar was able to induce redox
314 conversions. During the sorption of As(III) onto BKOH^{bat} , up to 70% of arsenic in solution was
315 As(V), despite only As(III) was initially added. A partial oxidation (<10%) was also observed in
316 the presence of BKOH and no As(V) was detected in controls. In contrast, no reduction of As(V)
317 to As(III) was noticed during As(V) sorption experiments. These results show that biochar is able
318 to modify the arsenic redox state in solution and this could be strongly increased with an
319 insufficient washing. These findings are in agreement with a previous study (Fakour and Lin,
320 2014) that reported an increased oxidation of As(III) in the presence of organic matter in solution.

321

322 **4. Conclusions**

323 Chemical treatment of biochar is considered to enhance the sorption performance for both
324 oxyanions and cationic metals. The treatment with KOH or H₂O₂ strongly affects the sorption
325 kinetics for As(III and V) and Cd(II). For As(V), KOH treatment induced a slower kinetic
326 sorption compared to the raw biochar, while no significant variation was noticed for the H₂O₂-
327 modified biochar. In contrast, both treatments showed faster sorption for Cd(II). All treatments
328 induced a strong sorption capacity enhancement, particularly with alkali treatment. This is
329 consistent with the observed evolutions of biochar properties such as CEC, S_{BET} and, to a lesser
330 extent, pH_{ZPC}. Unfortunately, as previously highlighted, without the implementation of a careful
331 washing procedure, this treatment also generated a strong release of inorganic and organic
332 compounds from the biochar that can alter the evaluation of its sorption properties. This issue
333 clearly affected As(V) sorption due to the competition with the released PO₄³⁻.

334 When working with a redox-sensitive element, such as arsenic, it was observed that the
335 biochar may also play an important role in the oxidation of As(III) to As(V). This phenomenon
336 was enhanced by the insufficient washing process. The potential chemical transformation of
337 arsenic should be carefully considered in future research in order to better understand the role of
338 chemically-modified biochar on the removal of such compounds from contaminated water.

339

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346 **References**

- 347 Agrafioti, E., Bouras, G., Kalderis, D., Diamadopoulos, E., 2013. Biochar production by sewage
348 sludge pyrolysis. *J. Anal. Appl. Pyrolysis* 101, 72–78.
- 349 Ahmad, M., Rajapaksha, A.U., Lim, J.E., Zhang, M., Bolan, N., Mohan, D., Vithanage, M., Lee,
350 S.S., Ok, Y.S., 2014. Biochar as a sorbent for contaminant management in soil and water: A
351 review. *Chemosphere* 99, 19–23.
- 352 Aran, D., Maul, A., Masfaraud, J.F., 2008. A spectrophotometric measurement of soil cation
353 exchange capacity based on cobaltihexamine chloride absorbance. *Comptes. Rendus.*
354 *Geosci.* 340, 865–871.
- 355 Ding, Z., Hu, X., Wan, Y., Wang, S., Gao, B., 2016. Removal of lead, copper, cadmium, zinc,
356 and nickel from aqueous solutions by alkali-modified biochar: Batch and column tests. *J.*
357 *Ind. Eng. Chem.* 33, 239–245.
- 358 EPA, 1996. Method 3050B: Acid digestion of sediments, sludges, and soils. revision, 2, 1–12.
- 359 Fakour, H., Lin, T.F., 2014. Effect of humic acid on As redox transformation and kinetic
360 adsorption onto iron oxide based adsorbent (IBA). *Int. J. Environ. Res. Public Health* 11,
361 10710–10736.
- 362 Gong, X.J., Li, W.G., Zhang, D.Y., Fan, W.B., Zhang, X.R., 2015. Adsorption of arsenic from
363 micro-polluted water by an innovative coal-based mesoporous activated carbon in the
364 presence of co-existing ions. *Int. Biodeterior. Biodegrad.* 102, 256–264.
- 365 Guo, K., Gao, B., Yue, Q., Xu, X., Li, R., Shen, X., 2018. Characterization and performance of a
366 novel lignin-based flocculant for the treatment of dye wastewater. *Int. Biodeterior.*
367 *Biodegrad.* 133, 99–107.
- 368 Ho, S.H., Chen, Y., Yang, Z., Nagarajan, D., Chang, J.S., Ren, N., 2017. High-efficiency removal
369 of lead from wastewater by biochar derived from anaerobic digestion sludge. *Bioresour.*
370 *Technol.* 246, 142–149.
- 371 Jiang, T., Xu, R., Gu, T., Jiang, J., 2014. Effect of crop-straw derived biochars on Pb(II)
372 adsorption in two variable charge soils. *J. Integr. Agric.* 13, 507–516.
- 373 Kim, H. Bin, Kim, S.H., Jeon, E.K., Kim, D., Tsang, D.C.W., Alessi, D.S., Kwon, E.E., Baek, K.,
374 2018. Effect of dissolved organic carbon from sludge, rice straw and spent coffee ground
375 biochar on the mobility of arsenic in soil. *Sci. Total Environ.* 636, 1241–1248.
- 376 Li, H., Dong, X., da Silva, E.B., de Oliveira, L.M., Chen, Y., Ma, L.Q., 2017. Mechanisms of
377 metal sorption by biochars: Biochar characteristics and modifications. *Chemosphere* 178,
378 466–478.
- 379 Liu, S., Huang, B., Chai, L., Liu, Y., Zeng, G., Wang, X., Zeng, W., Shang, M., Deng, J., Zhou,
380 Z., 2017. Enhancement of As(V) adsorption from aqueous solution by a magnetic
381 chitosan/biochar composite. *RSC Adv.* 7, 10891–10900.
- 382 Mahmood, T., Saddique, M.T., Naeem, A., Westerho, P., Mustafa, S., 2011. Comparison of
383 different methods for the point of zero charge determination of NiO. *Ind. Eng. Chem. Res.*
384 50, 10017–10023.
- 385 Mohan, D., Sarswat, A., Ok, Y.S., Pittman, C.U., 2014. Organic and inorganic contaminants
386 removal from water with biochar, a renewable, low cost and sustainable adsorbent – A
387 critical review. *Bioresour. Technol.* 160, 191–202.
- 388 Nethaji, S., Sivasamy, A., Mandal, A.B., 2013. Adsorption isotherms, kinetics and mechanism
389 for the adsorption of cationic and anionic dyes onto carbonaceous particles prepared from
390 *Juglans regia* shell biomass. *Int. J. Environ. Sci. Technol.* 10, 231–242.
- 391 Neupane, G., Donahoe, R.J., Arai, Y., 2014. Kinetics of competitive adsorption/desorption of

392 arsenate and phosphate at the ferrihydrite-water interface. *Chem. Geol.* 368, 31–38.

393 Ofomaja, A.E., Pholosi, A., Naidoo, E.B., 2014. Kinetics and competitive modeling of cesium
394 biosorption onto iron(III) hexacyanoferrate modified pine cone powder. *Int. Biodeterior.*
395 *Biodegrad.* 92, 71–78.

396 Park, J.H., Wang, J.J., Kim, S.H., Cho, J.S., Kang, S.W., Delaune, R.D., Han, K.J., Seo, D.C.,
397 2017. Recycling of rice straw through pyrolysis and its adsorption behaviors for Cu and Zn
398 ions in aqueous solution. *Colloids Surfaces A* 533, 330–337.

399 Peng, Q., Zhang, F., Zhou, Y., Zhang, J., Wei, J., Mao, Q., Huang, H., Chen, A., Chai, L., Luo,
400 L., 2018. Formation of composite sorbent by *P. chrysogenum* strain F1 and ferrihydrite in
401 water for arsenic removal. *Int. Biodeterior. Biodegrad.* 132, 208–215.

402 Pituello, C., Francioso, O., Simonetti, G., Pisi, A., Torreggiani, A., Berti, A., Morari, F., 2014.
403 Characterization of chemical–physical, structural and morphological properties of biochars
404 from biowastes produced at different temperatures. *J. Soils Sediments* 15, 792–804.

405 Qiu, Y., Zheng, Z., Zhou, Z., Sheng, G.D., 2009. Effectiveness and mechanisms of dye
406 adsorption on a straw-based biochar. *Bioresour. Technol.* 100, 5348–5351.

407 Rajapaksha, A.U., Chen, S.S., Tsang, D.C.W., Zhang, M., Vithanage, M., Mandal, S., Gao, B.,
408 Bolan, N.S., Ok, Y.S., 2016. Engineered/designer biochar for contaminant
409 removal/immobilization from soil and water: Potential and implication of biochar
410 modification. *Chemosphere* 148, 276–291.

411 Rajapaksha, A.U., Vithanage, M., Zhang, M., Ahmad, M., Mohan, D., Chang, S.X., Ok, Y.S.,
412 2014. Pyrolysis condition affected sulfamethazine sorption by tea waste biochars. *Bioresour.*
413 *Technol.* 166, 303–308.

414 Regmi, P., Garcia Moscoso, J.L., Kumar, S., Cao, X., Mao, J., Schafran, G., 2012. Removal of
415 copper and cadmium from aqueous solution using switchgrass biochar produced via
416 hydrothermal carbonization process. *J. Environ. Manage.* 109, 61–69.

417 Sizmur, T., Fresno, T., Akgül, G., Frost, H., Moreno-Jiménez, E., 2017. Biochar modification to
418 enhance sorption of inorganics from water. *Bioresour. Technol.* 246, 34–47.

419 Wan, J., Pressigout, J., Simon, S., Deluchat, V., 2014. Distribution of As trapping along a
420 ZVI/sand bed reactor. *Chem. Eng. J.* 246, 322–327.

421 Wang, C., Gu, L., Liu, X., Zhang, X., Cao, L., 2016. Sorption behavior of Cr(VI) on pineapple-
422 peel-derived biochar and the influence of coexisting pyrene. *Int. Biodeterior. Biodegradation*
423 111, 78–84.

424 Wongrod, S., Simon, S., Guibaud, G., Lens, P.N.L., Pechaud, Y., Huguenot, D., van Hullebusch,
425 E.D., 2018. Lead sorption by biochar produced from digestates: Consequences of chemical
426 modification and washing. *J. Environ. Manage.* 219, 277–284.

427 Wu, W., Li, J., Lan, T., Müller, K., Khan, N., Chen, X., Xu, S., Zheng, L., Chu, Y., Li, J., Yuan,
428 G., Wang, H., 2017. Unraveling sorption of lead in aqueous solutions by chemically
429 modified biochar derived from coconut fiber: A microscopic and spectroscopic
430 investigation. *Sci. Total Environ.* 576, 766–774.

431 Yuan, H., Lu, T., Huang, H., Zhao, D., Kobayashi, N., Chen, Y., 2015. Influence of pyrolysis
432 temperature on physical and chemical properties of biochar made from sewage sludge. *J.*
433 *Anal. Appl. Pyrolysis* 112, 284–289.

434 Zama, E.F., Zhu, Y.G., Reid, B.J., Sun, G.X., 2017. The role of biochar properties in influencing
435 the sorption and desorption of Pb(II), Cd(II) and As(III) in aqueous solution. *J. Clean. Prod.*
436 148, 127–136.

437 Zhao, L., Xue, F., Yu, B., Xie, J., Zhang, X., Wu, R., Wang, R., Hu, Z., Yang, S.T., Luo, J.,
438 2015a. TiO₂–graphene sponge for the removal of tetracycline. *J. Nanoparticle Res.* 17, 1–9.

- 439 Zhao, Z., Wu, M., Jiang, Q., Zhang, Y., Chang, X., Zhan, K., 2015b. Adsorption and desorption
440 studies of anthocyanins from black peanut skins on macroporous resins. *Int. J. Food Eng.* 11,
441 841–849.
- 442 Zhou, Z., Liu, Y., Liu, S., Liu, H., Zeng, G., Tan, X., Yang, C., Ding, Y., Yan, Z., Cai, X., 2017.
443 Sorption performance and mechanisms of arsenic(V) removal by magnetic gelatin-modified
444 biochar. *Chem. Eng. J.* 314, 223–231.
- 445 Zielińska, A., Oleszczuk, P., 2015. The conversion of sewage sludge into biochar reduces
446 polycyclic aromatic hydrocarbon content and ecotoxicity but increases trace metal content.
447 *Biomass Bioenergy* 75, 235–244.
- 448 Zuo, W.Q., Chen, C., Cui, H.J., Fu, M.L., 2017. Enhanced removal of Cd(II) from aqueous
449 solution using CaCO₃ nanoparticle modified sewage sludge biochar. *RSC Adv.* 7, 16238–
450 16243.

451 **Figures**

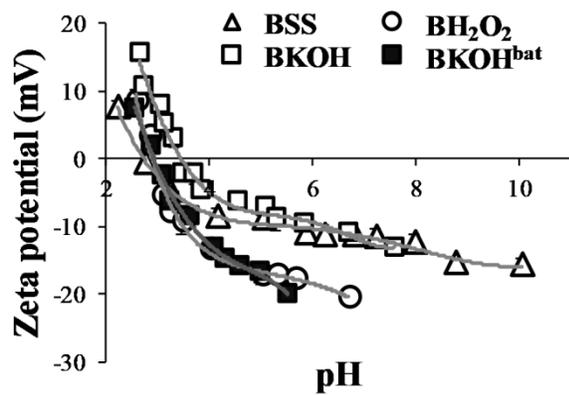


Fig. 1. Evolution of the zeta potential as a function of the pH values.

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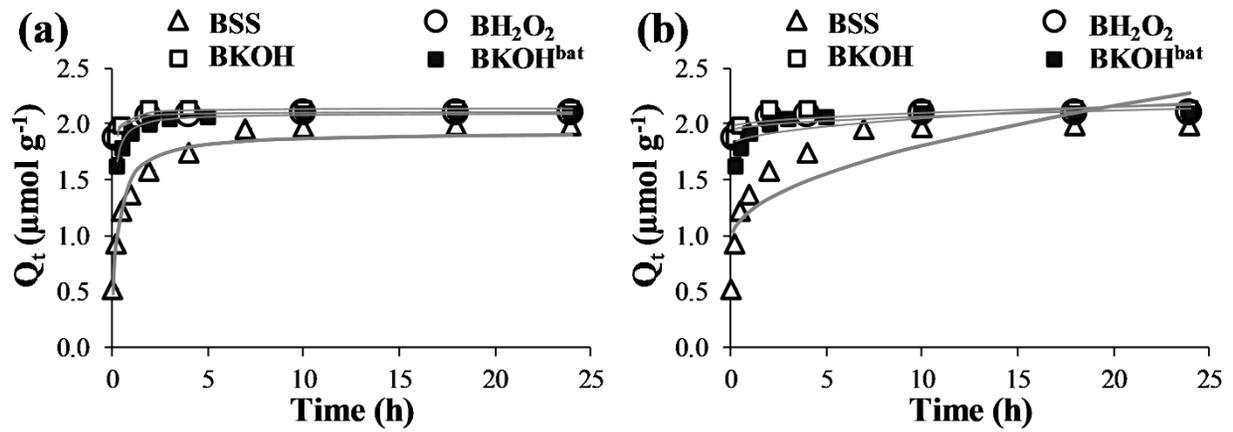


Fig. 2. Effect of chemical treatment and biochar washing on adsorption kinetics for Cd(II) by the raw and chemically-modified biochars fitted with the pseudo-second-order kinetic model (a), and with the intraparticle diffusion model (b) (initial pH=5, initial concentration: $10 \mu\text{mol l}^{-1}$).

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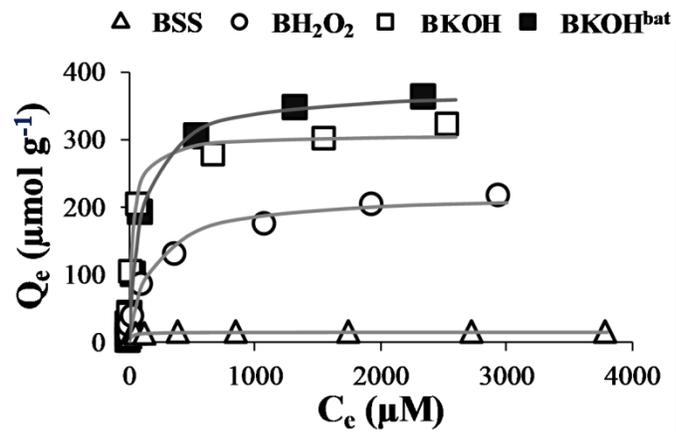


Fig. 3. Effect of chemical treatment and biochar washing on adsorption isotherms for Cd(II) by the raw and chemically-modified biochars (initial pH=5, 24 h, Langmuir model fitting).

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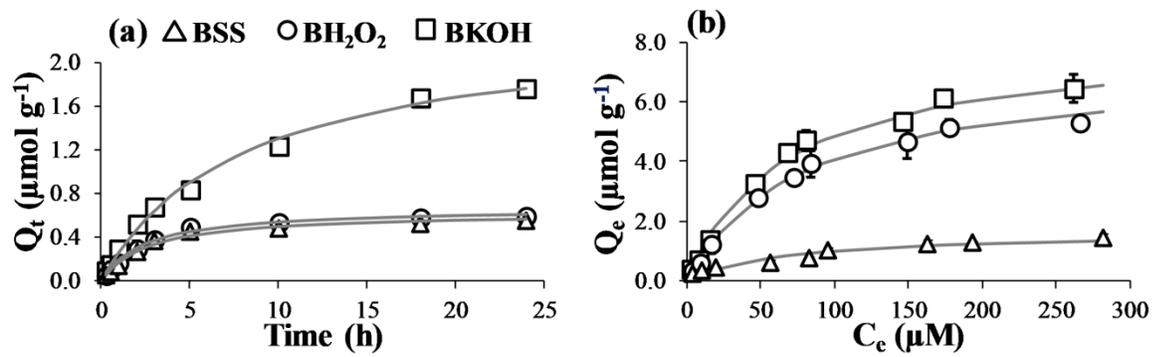


Fig. 4. Effect of chemical treatment on As(V) adsorption kinetics by biochars (a) (initial pH=5, initial concentration: $10 \mu\text{mol l}^{-1}$, pseudo-second-order kinetic model fitting), and As(V) adsorption isotherms by biochars (b) (initial pH=5, 24 h, Langmuir model fitting).

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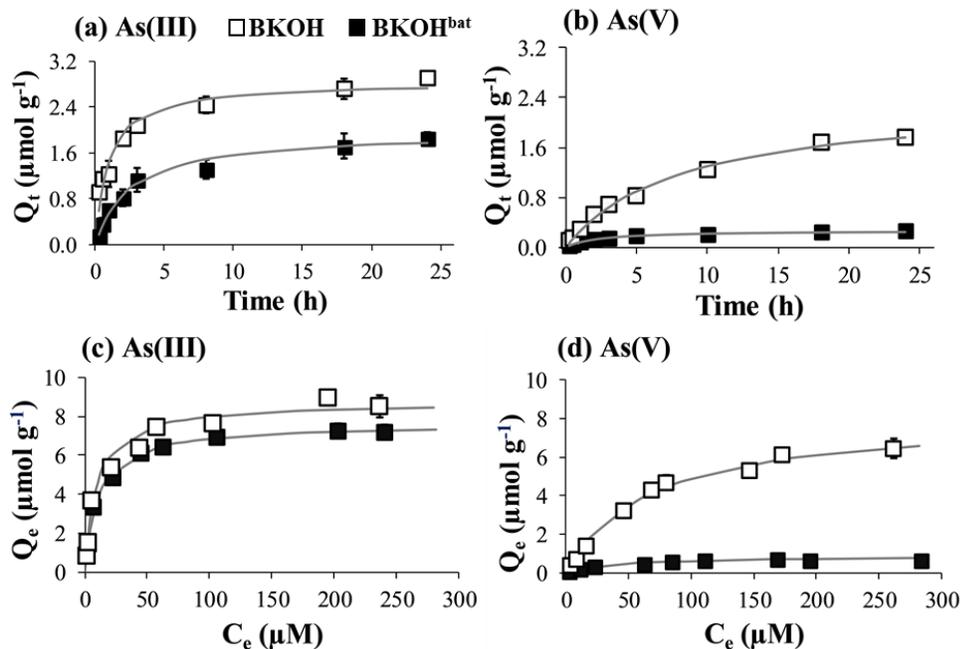


Fig. 5. Influence of biochar washing of KOH-modified biochars on adsorption kinetics for As(III) (a) and As(V) (b) (initial pH=5, initial concentration: $10 \mu\text{mol l}^{-1}$, pseudo-second-order kinetic model fitting), and on adsorption isotherms for As(III) (c) and As(V) (d) (initial pH=5, 24 h, Langmuir model fitting).

457 **Tables**458 **Table 1.** pH, electrical conductivity and surface area of the raw and chemically-modified
459 biochars.

Biochar	pH in water	pH _{pzc}	Electrical conductivity (EC) ($\mu\text{S cm}^{-1}$)	S _{BET} ^a ($\text{m}^2 \text{g}^{-1}$)	CEC ^b ($\text{cmol}^+ \text{kg}^{-1}$)	Reference
BSS ^c	6.4 ±0.1	2.7	4.0 ±0.1	0.4 ±0.1	2.0 ±0.1	This study
BH ₂ O ₂ ^c	6.5 ±0.1	2.9	4.1 ±0.3	5.7 ±0.1	2.9 ±0.1	This study
BKOH ^c	8.4 ±0.1	3.4	6.2 ±0.5	7.9 ±0.1	13.4 ±0.1	This study
BKOH ^{bat c}	10.0 ±0.1	2.9	324.0 ±2.4	3.0 ±0.1	20.8 ±0.1	This study
Sewage sludge (300 °C)	6.7 ±0.2	na ^d	7750.0 ±160.0	14.3 ±0.2	na	Yuan et al. (2015)
Sewage sludge (300 °C)	na	na	na	4.0	na	Agrafioti et al. (2013)

460 ^a S_{BET} refers to Brunauer-Emmett-Teller surface area of biochar.461 ^b CEC refers to cation exchange capacity of biochar.462 ^c BSS: biochar from sewage sludge digestate, BH₂O₂: H₂O₂-modified biochar, BKOH and BKOH^{bat}:
463 KOH-modified biochars, respectively, with batch and column washing, and with batch washing.464 ^d na refers to not available.

465 **Table 2.** Constants for adsorption kinetics of As(III), As(V) and Cd(III) by biochars with the
 466 pseudo-second-order and intraparticle diffusion models.

467 Pseudo-second-order model

Biochar	As(III)			As(V)			Cd(II)		
	k_2^a	Q_e^b	R^2	k_2	Q_e	R^2	k_2	Q_e	R^2
BSS ^c	-	-	-	0.630 ±0.098	0.62 ±0.03	0.985	1.724 ±0.204	1.93 ±0.04	0.983
BH ₂ O ₂ ^c	-	-	-	0.619 ±0.092	0.66 ±0.03	0.985	12.251 ±0.803	2.09 ±0.01	9.985
BKOH ^c	0.378 ±0.067	2.83 ±0.11	0.961	0.052 ±0.006	2.35 ±0.09	0.995	12.907 ±0.479	2.14 ±0.01	0.993
BKOH ^{bat c}	0.205 ±0.042	1.94 ±0.10	0.975	1.599 ±0.340	0.27 ±0.02	0.969	6.298 ±0.338	2.09 ±0.10	0.987

Intraparticle diffusion model

Biochar	As(III)			As(V)			Cd(II)		
	k_i^d	C	R^2	k_i	C	R^2	k_i	C	R^2
BSS	-	-	-	0.115 ±0.013	0.065 ±0.035	0.910	0.269 ±0.039	0.956 ±0.101	0.852
BH ₂ O ₂	-	-	-	0.121 ±0.014	0.083 ±0.036	0.913	0.042 ±0.010	1.932 ±0.029	0.787
BKOH	0.429 ±0.044	0.981 ±0.113	0.938	0.396 ±0.012	-0.074 ±0.030	0.993	0.041 ±0.013	1.973 ±0.036	0.699
BKOH ^{bat}	0.358 ±0.033	0.219 ±0.086	0.948	0.050 ±0.006	0.036 ±0.015	0.919	0.077 ±0.018	1.799 ±0.046	0.749

468 ^a k_2 unit: $\text{g}(\mu\text{mol h})^{-1}$.

469 ^b Q_e unit: in $\mu\text{mol g}^{-1}$.

470 ^c BSS: biochar from sewage sludge digestate, BH₂O₂: H₂O₂-modified biochar, BKOH and BKOH^{bat}:
 471 KOH-modified biochars, respectively, with batch and column washing, and with batch washing.

472 ^d k_i unit: $\mu\text{mol}(\text{g h}^{1/2})^{-1}$.

473 **Table 3.** Parameters determined from adsorption isotherms for As(III), As(V) and Cd(II) by
 474 biochars with the Langmuir model.

Biochar	As(III)			As(V)			Cd(II)		
	K_L^a	Q_m^b	R^2	K_L	Q_m	R^2	K_L	Q_m	R^2
BSS ^c	-	-	-	0.015 ±0.006	1.60 ±0.23	0.958	0.061 ±0.009	15.49 ±0.32	0.992
BH ₂ O ₂ ^c	-	-	-	0.012 ±0.003	7.22 ±0.67	0.989	0.005 ±0.001	218.73 ±6.91	0.990
BKOH ^c	0.103 ±0.018	8.74 ±0.33	0.976	0.014 ±0.003	8.11 ±0.53	0.994	0.032 ±0.004	306.09 ±6.07	0.994
BKOH ^{bat c}	0.088 ±0.007	7.60 ±0.12	0.996	0.024 ±0.008	0.85 ±0.09	0.965	0.009 ±0.001	372.49 ±6.71	0.996

475 ^a K_L unit: in l μmol^{-1} .

476 ^b Q_m unit: in $\mu\text{mol g}^{-1}$.

477 ^c BSS: biochar from sewage sludge digestate, BH₂O₂: H₂O₂-modified biochar, BKOH and BKOH^{bat}:
 478 KOH-modified biochars, respectively, with batch and column washing, and with batch washing.

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495 **Supplementary Information**

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497 **Changes of sewage sludge digestate-derived biochar properties after chemical treatments**

498 **and the influence on As(III and V) and Cd(II) sorption**

499

500 Suchanya Wongrod^{a,b,c}, Stéphane Simon^{b,†}, Eric D. van Hullebusch^{a,c,d}, Piet N.L. Lens^c, Gilles

501 Guibaud^b

502

503 ^a Université Paris-Est, Laboratoire Géomatériaux et Environnement (EA 4508), UPEM, 77454,

504 Marne-la-Vallée, France

505 ^b Université de Limoges, PEIRENE, Equipe Développement d'indicateurs ou prévision de la

506 qualité des eaux, URA IRSTEA, 123 avenue Albert Thomas, 87060 Limoges, France

507 ^c UNESCO-IHE Institute for Water Education, P.O. Box 3015, 2601 DA Delft, The Netherlands

508 ^d Institut de Physique du Globe de Paris, Sorbonne Paris Cité, Université Paris Diderot, UMR

509 7154, CNRS, F-75005 Paris, France

† Corresponding author. Université de Limoges, PEIRENE, Equipe Développement d'indicateurs ou prévision de la qualité des eaux, URA IRSTEA, 123 avenue Albert Thomas, 87060 Limoges, France
Email: stephane.simon@unilim.fr

510 **Measurement of pH of point of zero charge (pH_{PZC}) of biochar**

511 The pH_{PZC} of raw and chemically-modified biochars was determined from zetametry by
512 determining the zeta potential at different pH ranges (Mahmood et al., 2011). Firstly, 40 mL of
513 background electrolyte solution (0.01 M NaNO₃) containing 0.2 g of biochar was equilibrated
514 with continuous magnetic stirring at 20 (± 2) °C for 40 min. The pH of the mixture was adjusted
515 in the range of 2.0 – 10.0 by using HNO₃ (0.1, 0.5 and 1 M) and NaOH (0.1, 0.5 and 1 M). After
516 40 min of stirring, the pH was measured and readjusted to correct the pH value until each
517 solution reached a stable pH. The suspension was then filtered with a 0.45 µm cellulose acetate
518 syringe filter before determining the zeta potential (Zetasizer Nanoseries (Nano-ZS), Malvern,
519 UK). This filtration was performed to prevent the settlement of biochar particles in the
520 measurement cell of the zetameter and to avoid the clogging of big particles in the device. We
521 hypothesize that all biochar particles display the same behavior, neglecting the particle size.

522

523 **Measurement of the cation exchange capacity (CEC) of biochar**

524 For the determination of the CEC, 2 g of biochar (dry weight) were mixed with 40 mL of 0.05 N
525 **cobalt hexamine trichloride** ([Co(NH₃)₆]Cl₃) solution (99% w/w, Sigma-Aldrich) in
526 polypropylene tubes (Aran et al., 2008). The suspensions were shaken for 1 h using an orbital
527 shaker (KS 501 digital, IKATM, USA) at 60 rpm, then centrifuged at 7000 g for 10 min (Rotina
528 420, Hettich, Germany). The supernatant was filtered through Whatman polyethersulfone syringe
529 filters (0.2 µm). The pH (Präzisions-pH-meter E510, Metrohm AG, Switzerland) and absorbency
530 at 472 nm (UV–vis spectrophotometer, Lambda 365, PerkinElmer, USA) of each sample were
531 measured immediately. All experiments were performed in triplicate and the results are reported
532 as mean values. The CEC (meq 100 g⁻¹ or cmol⁺ kg⁻¹) can be calculated as follows using Eq. (1)
533 (Aran et al., 2008):

$$\text{CEC} = \left[\frac{\text{Abs}_{\text{Co}} - \text{Abs}_{\text{sample}}}{\text{Abs}_{\text{Co}}} \right] \times 50 \times \frac{V}{m} \times 100 \quad (1),$$

534

535 where Abs_{Co} and Abs_{sample} refer to the absorbency (at 472 nm) of 0.05 N [Co(NH₃)₆]Cl₃ and of
536 sample (supernatant after filtration), respectively. V and m correspond to solution volume in liter
537 and biochar dry mass in gram, respectively.

538

539 **X-ray powder diffraction (XRD) analysis**

540 X-ray diffractometry (XRD) was employed to identify the crystalline structures presented in raw
541 and modified biochars. The samples were grounded to less than 100 µm particle size and were
542 characterized using a powder diffractometer (AXS D8, Bruker, Germany) with Cu Kα radiation
543 at 1.54 Å wavelength over the 2θ range from 10° to 32°, for 10 seconds per step, at 40 kV voltage

544 and 40 mA current with a Solx (Si-Li) detector. The crystalline compounds present in biochar
545 were identified using the X'Pert HighScore software for data analysis with the reference from the
546 International Center for Diffraction Data (ICDD).

547

548 **Fourier-transform infrared spectroscopy (FTIR) analysis**

549 FTIR spectra were recorded to identify the functional groups present on biochar surfaces. Each
550 sample was mixed with KBr in a ratio of 1 mg sample (particle size 100 μm) per 200 mg KBr and
551 the pellet was prepared by compression under vacuum (Jouraiphy et al., 2005). The analysis was
552 performed using a Shimadzu IRAffinity-1 Spectrometer with a deuterated-triglycine sulfate
553 (DTGS) detector. The number of scans was 12 with a resolution of 16 cm^{-1} and a frequency in a
554 range from 4000 to 800 cm^{-1} .

555 XRD spectra of raw sewage sludge digestate and its derived biochar

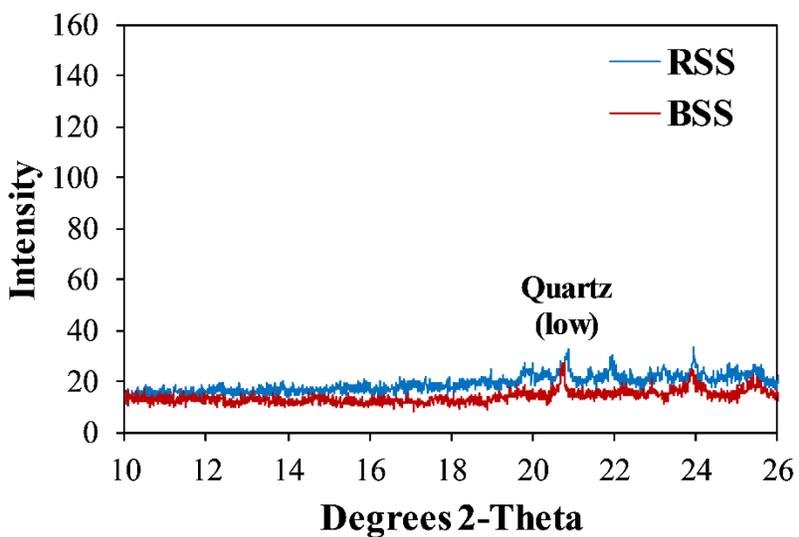


Fig. S1. XRD spectra of raw sewage sludge digestate (RSS) and biochar from sewage sludge digestate (BSS) produced at 350 °C under slow pyrolysis.

556

557 **FTIR spectra of raw sewage sludge digestate and its derived biochar**

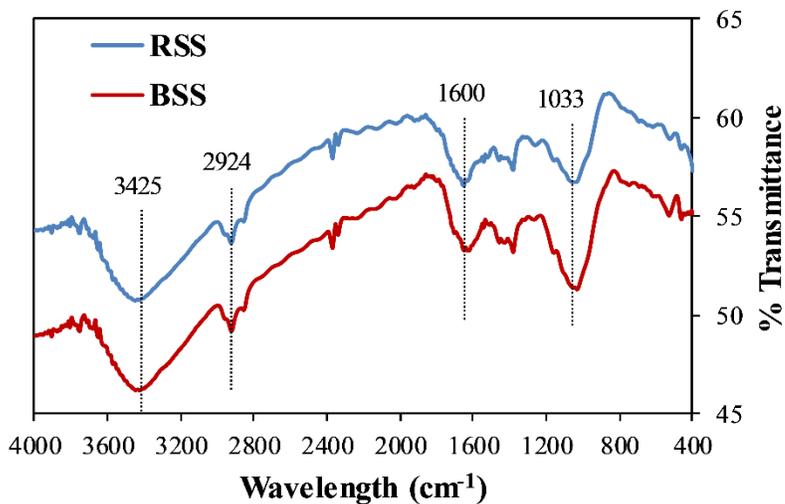


Fig. S2. FTIR spectra of raw sewage sludge digestate (RSS) and biochar from sewage sludge digestate (BSS) produced at 350 °C under slow pyrolysis.

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560 **Table S1.** Comparison of the sorption performance of As(V) and Cd(II) by different types of biochars.

Adsorbent	Element	Maximum adsorption capacity ($\mu\text{mol g}^{-1}$)	Concentration ranges (μM)	Solution pH	References
BSS ^a	As(V)	1.6	10 – 300	5.0	This study
BH ₂ O ₂ ^a	As(V)	7.1	10 – 300	5.0	This study
BKOH ^a	As(V)	8.5	10 – 300	5.0	This study
Fe(II)-loaded activated carbon	As(V)	27	7 – 113	3.0	Özge et al. (2013)
Paper mill sludge biochar	As(V)	303	280 – 2529	6.5	Yoon et al. (2017)
Magnetic carbonaceous tea waste	As(V)	507	10 – 1335	5.0	Wen et al. (2017)
Carbonaceous nanofibers	As(V)	670	100 – 900	5.0	Cheng et al. (2016)
BSS	Cd(II)	16	10 – 4000	5.0	This study
BH ₂ O ₂	Cd(II)	220	10 – 4000	5.0	This study
BKOH	Cd(II)	318	10 – 4000	5.0	This study
Paper mill sludge biochar	Cd(II)	369	187 – 2506	6.5	Yoon et al. (2017)
H ₂ O ₂ -treated yak manure biochar	Cd(II)	419	10 – 1779	-	Wang and Liu (2018)
Graphene oxide nanosheets	Cd(II)	945	40 – 450	6.0	Zhao et al. (2011)

561 ^a BSS, BH₂O₂ and BKOH refer to, respectively, biochar produced from sewage sludge digestate, and its derived biochar with H₂O₂ and
562 KOH treatments.

563 **Table S2.** Chemical characteristics of raw sewage sludge digestate (RSS) and its derived biochar
 564 produced at 350 °C under slow pyrolysis (BSS).

Parameters	RSS	BSS
pH in water	6.0 ±0.1	6.4 ±0.1
As (mg kg ⁻¹)	79 ±3 ^a	47 ±1
Cd (mg kg ⁻¹)	dl ^b	9 ±1
Cr (mg kg ⁻¹)	42 ±1	48 ±1
Cu (mg kg ⁻¹)	390 ±10	617 ±7
Ni (mg kg ⁻¹)	25 ±1	41 ±1
Pb (mg kg ⁻¹)	80 ±9	85 ±3
Zn (mg kg ⁻¹)	756 ±10	1017 ±13

565 ^a Values reported as means (n=3) on dry mass basis followed by standard deviation.

566 ^b dl refers to value below detection limit.

567 **Adsorption kinetics**

568 The sorption kinetic data for arsenic and cadmium by biochars was calculated using the pseudo-
569 first-order (PFO) (Eq. 2), pseudo-second-order (PSO) (Eq. 3) and intraparticle diffusion (Eq. 4)
570 kinetic models, which are illustrated as followed:

$$\log(Q_e - Q_t) = \log Q_e - \frac{k_1 t}{2.303} \quad (2),$$

571

$$\frac{t}{Q_t} = \frac{1}{k_2 Q_e^2} + \frac{t}{Q_e} \quad (3),$$

572

$$Q_t = k_i t^{1/2} + C \quad (4),$$

573

574 where Q_t and Q_e are arsenic or cadmium adsorption capacity ($\mu\text{mol g}^{-1}$) at time t (h) and at
575 equilibrium, k_1 (h^{-1}), k_2 ($\text{g}(\mu\text{mol h})^{-1}$) and k_i ($\mu\text{mol}(\text{g h}^{1/2})^{-1}$) are the rate constants for the PFO,
576 PSO and intraparticle diffusion kinetic models, respectively (Liu and Zhang, 2009; Zhao et al.,
577 2015). C is the intraparticle diffusion constant related to the boundary layer diffusion. All
578 uncertainties of sorption kinetic parameters were obtained by non-linear regression using
579 Statistica software (v6.1, StatSoft).

580

581 **Adsorption isotherms**

582 Langmuir and Freundlich models are presented, respectively, in Eqs. (4) and (5):

$$\frac{C_e}{Q_e} = \frac{1}{K_L Q_m} + \frac{C_e}{Q_m} \quad (5),$$

583

$$\ln Q_e = \ln K_F + \frac{1}{n} \ln C_e \quad (6),$$

584

585 where C_e is the equilibrium arsenic or cadmium solution concentration (μM). Q_e and Q_m are the
586 arsenic or cadmium adsorption capacity at equilibrium and at maximum capacity ($\mu\text{mol g}^{-1}$),
587 respectively. K_L and K_F are the adsorption constants for the Langmuir and Freundlich isotherm
588 models, and n is the Freundlich constant which indicates the favorability of adsorption,
589 respectively.

590 **Table S3.** Constants for the pseudo-first-order (PFO), pseudo-second-order (PSO) and intraparticle diffusion kinetic models for As(III,
 591 V) and Cd(II) sorption by different types of biochars (BSS: biochar from sewage sludge digestate, BH₂O₂: H₂O₂-modified biochar, BKOH and
 592 BKOH^{bat}: KOH-modified biochars, respectively, with batch and column washing and with batch washing).

Biochar	As(III)								
	PFO			PSO			Intraparticle diffusion		
	k_1^a	Q_e^b	R^2	k_2^a	Q_e	R^2	k_i	C	R^2
BSS	-	-	-	-	-	-	-	-	-
BH ₂ O ₂	-	-	-	-	-	-	-	-	-
BKOH	0.128	1.62	0.934	0.378	2.83	0.961	0.429	0.981	0.938
BKOH ^{bat}	0.134	1.44	0.967	0.205	1.94	0.975	0.358	0.219	0.948
Biochar	As(V)								
	PFO			PSO			Intraparticle diffusion		
	k_1	Q_e	R^2	k_2	Q_e	R^2	k_i	C	R^2
BSS	0.167	0.40	0.918	0.630	0.62	0.985	0.115	0.065	0.910
BH ₂ O ₂	0.188	0.44	0.948	0.619	0.66	0.985	0.121	0.083	0.913
BKOH	0.165	1.84	0.969	0.052	2.35	0.995	0.396	-0.074	0.993
BKOH ^{bat}	0.139	0.19	0.954	1.599	0.27	0.969	0.050	0.036	0.919
Biochar	Cd(II)								
	PFO			PSO			Intraparticle diffusion		
	k_1	Q_e	R^2	k_2	Q_e	R^2	k_i	C	R^2
BSS	0.184	0.94	0.923	1.724	1.93	0.983	0.269	0.956	0.852
BH ₂ O ₂	0.400	0.14	0.881	12.251	2.09	9.985	0.042	1.932	0.787
BKOH	1.217	0.19	0.763	12.907	2.14	0.993	0.041	1.973	0.699
BKOH ^{bat}	0.652	0.38	0.942	6.298	2.09	0.987	0.077	1.799	0.749

593 ^a k_1 and k_2 units: h⁻¹ and g(μmol h)⁻¹, respectively.

594 ^b Q_e unit: in μmol g⁻¹.

595

596 **Table S4.** Constants for the Langmuir and Freundlich isotherms for As(III), As(V) and Cd(II) sorption by different types of biochars
 597 (BSS: biochar from sewage sludge digestate, BH₂O₂: H₂O₂-modified biochar, BKOH and BKOH^{bat}: KOH-modified biochars, respectively, with
 598 batch and column washing and with batch washing).

Biochar	As(III)						As(V)						Cd(II)					
	Langmuir			Freundlich			Langmuir			Freundlich			Langmuir			Freundlich		
	K_L^a	Q_m^b	R^2	K_F^c	n	R^2	K_L	Q_m	R^2	K_F	n	R^2	K_L	Q_m	R^2	K_F	n	R^2
BSS	-	-	-	-	-	-	0.015	1.60	0.958	0.112	2.185	0.986	0.061	15.49	0.992	3.163	4.336	0.796
BH ₂ O ₂	-	-	-	-	-	-	0.012	7.22	0.989	0.156	1.456	0.966	0.005	218.73	0.990	5.893	2.008	0.942
BKOH	0.103	8.74	0.976	1.059	2.299	0.884	0.014	8.11	0.994	0.196	1.473	0.967	0.032	306.09	0.994	23.460	2.570	0.933
BKOH ^{bat}	0.088	7.60	0.996	1.007	2.420	0.880	0.024	0.85	0.965	0.044	1.867	0.906	0.009	372.49	0.996	9.013	1.849	0.882

599 ^a K_L unit: in $l \mu\text{mol}^{-1}$.600 ^b Q_m unit: in $\mu\text{mol g}^{-1}$.601 ^c K_F unit: in $(\mu\text{mol g}^{-1})(l \mu\text{mol}^{-1})^{1/n}$.

602 **References**

- 603 Aran, D., Maul, A., Masfaraud, J.F., 2008. A spectrophotometric measurement of soil cation
604 exchange capacity based on cobaltihexamine chloride absorbance. *Comptes. Rendus.*
605 *Geosci.* 340, 865–871.
- 606 Cheng, W., Ding, C., Wang, X., Wu, Z., Sun, Y., Yu, S., Hayat, T., Wang, X., 2016. Competitive
607 sorption of As(V) and Cr(VI) on carbonaceous nanofibers. *Chem. Eng. J.* 293, 311–318.
- 608 Jouraiphy, A., Amir, S., El Gharous, M., Revel, J.C., Hafidi, M., 2005. Chemical and
609 spectroscopic analysis of organic matter transformation during composting of sewage sludge
610 and green plant waste. *Int. Biodeterior. Biodegrad.* 56, 101–108.
- 611 Liu, Z., Zhang, F.S., 2009. Removal of lead from water using biochars prepared from
612 hydrothermal liquefaction of biomass. *J. Hazard. Mater.* 167, 933–939.
- 613 Mahmood, T., Saddique, M.T., Naeem, A., Westerho, P., Mustafa, S., 2011. Comparison of
614 different methods for the point of zero charge determination of NiO. *Ind. Eng. Chem. Res.*
615 50, 10017–10023.
- 616 Tuna, A.O.A., Ozdemir, E., Simsek, E.B., Beker, U., 2013. Removal of As(V) from aqueous
617 solution by activated carbon-based hybrid adsorbents: Impact of experimental conditions.
618 *Chem. Eng. J.* 223, 116–128.
- 619 Wang, Y., Liu, R., 2018. H₂O₂ treatment enhanced the heavy metals removal by manure biochar
620 in aqueous solutions. *Sci. Total Environ.* 628–629, 1139–1148.
- 621 Wen, T., Wang, J., Yu, S., Chen, Z.S., 2017. Magnetic porous carbonaceous material produced
622 from tea waste for efficient removal of As(V), Cr(VI), humic acid and dyes. *ACS Sustain.*
623 *Chem. Eng.* 5, 4371–4380.
- 624 Yoon, K., Cho, D.W., Tsang, D.C.W., Bolan, N., Rinklebe, J., Song, H., 2017. Fabrication of
625 engineered biochar from paper mill sludge and its application into removal of arsenic and
626 cadmium in acidic water. *Bioresour. Technol.* 246, 69–75.
- 627 Zhao, G., Li, J., Ren, X., Chen, C., Wang, X., 2011. Few-layered graphene oxide nanosheets as
628 superior sorbents for heavy metal ion pollution management. *Environ. Sci. Technol.* 45,
629 10454–10462.
- 630 Zhao, Z., Wu, M., Jiang, Q., Zhang, Y., Chang, X., Zhan, K., 2015. Adsorption and desorption
631 studies of anthocyanins from black peanut skins on macroporous resins. *Int. J. Food Eng.* 11,
632 841–849.
- 633