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Monazite, rhabdophane, xenotime & churchite: vibrational spectroscopy of gadolinium phosphate polymorphs

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Abstract:

Rare-earth phosphates with the general formula REEPO$_4$·nH$_2$O belong to four distinct structural types: monazite, rhabdophane, churchite, and xenotime. We report herein the first direct comparison between vibrational spectra of these compounds for the same metal cation *i.e.* gadolinium. The four GdPO$_4$·nH$_2$O samples were prepared through wet chemistry methods and first characterized by X-Ray Diffraction. Three distinct spectral domains, associated to the deformation and stretching modes of phosphate tetrahedra (PO$_4$) and to water molecules vibrations were then analyzed from FTIR and Raman data, and discussed regarding the structural characteristics of each sample. The most obvious differences between the spectra were associated to $\delta$(H$_2$O) and $\delta$(PO$_4$) modes and led to propose a simple method to rapidly and unambiguously discriminate the four polymorphs.

Keywords: phosphate, rare earth elements, monazite, FTIR, Raman
1. Introduction

Rare-earth phosphate minerals with the general formula REEPO₄·nH₂O belong to four distinct structural types, mainly depending on their hydration content and the atomic weight of the rare-earth elements incorporated (i.e. light or heavy REE) [1]. Among them, monazite LnPO₄ (hexagonal, P2₁/n), typically crystallizing with Ln = La-Tb [2], is probably the most widespread in nature and is therefore widely documented. Monazite constitutes one of the main ores exploited for lanthanide extraction, and naturally bears high contents in uranium and thorium (up to 15 wt.%), mainly through coupled substitutions with divalent elements such as calcium [3]. For heavy rare-earth elements, i.e. Ln = Gd-Lu, as well as for yttrium and scandium, anhydrous LnPO₄ phase adopts the zircon structure type (tetragonal, I₄₁/amd) [4], leading to the formation of xenotime. This latter frequently coexists with monazite within granitic and metamorphic rocks [5].

In comparison, related hydrated counterparts are far less present in nature. For light rare-earth elements, rhabdophane LnPO₄·0.667H₂O (monoclinic, C2) [6], is frequently restricted to the uppermost portion of the crust due to its metastable character regarding to monazite [7]. Hence, rhabdophane is one of the main LREE carriers in bauxite, laterites and hydrothermal systems, while monazite dominates in migmatic and metamorphic silicic rocks [8]. Finally, churchite is most often reported with the ideal formulae YPO₄·2H₂O (monoclinic, isostructural to gypsum) and also contains variable amounts of heavy rare-earth elements such as dysprosium and erbium. It was found in association with limonite deposits, but also in laterites [9]. One must note that this mineral is still frequently called as weinschenkite even if this name was early discarded by the Commission on New Minerals and Mineral Names [10].

Owing to their numerous potential applications in several fields of research, such as phosphor light emitters [11], geochronometers [12], thermal barrier coatings [13] or ceramics for the storage [14, 15] or the transmutation [16] of nuclear waste, REEPO₄·nH₂O compounds were investigated through a wide range of physico-chemical techniques of characterization. Among them, vibrational spectroscopy was frequently considered, due to its easy operation and to the intense signal provided by the vibration modes of the PO₄ tetrahedra. As a matter of fact, Raman spectra were used to validate the formation of monazite-type BkPO₄, CfPO₄, and EsPO₄, which remain up to now among the few samples prepared at a weighable scale with these heavy transuranic elements [17]. Raman spectroscopy was also demonstrated for long as a powerful tool in the analysis of scarce natural samples, such as REEPO₄·2H₂O churchite [18].
Along with the studies devoted to minerals, numerous works were also undertaken on synthetic samples series. Once again, monazite-type compounds were widely investigated, most often through comparisons along the light lanthanides series (Ln = La-Gd) [19-21] which allowed to point out a shift of the vibration modes towards the highest wavenumbers with the atomic number of the cation. Isostructural cheralite samples incorporating tetravalent actinides also present similar spectroscopic features [22]. In comparison, the studies concerning the xenotime series, obtained for heavy rare earth elements (Ln = Tb – Lu) [20, 23], appear to be more ancient and scarce. However, recent works renewed the interest for spectroscopy in this field, with some works dealing with the calculations of vibrational dynamics [24] and to the behavior of xenotime samples under pressure [25]. Quite similarly, only the Assaoudi’s group undertook systematic spectroscopic investigations of the hydrated rare earth phosphates, rhabdophane and churchite [26, 27].

However, if the evolution of the vibrational spectra was studied for each REEPO$_4$·nH$_2$O structural type as a function of the metal center, no study compared FTIR and Raman spectra of the four polymorphs for the same cation. This lack probably arises from the difficulty to stabilize the different phases for one given lanthanide element, gadolinium being the only one possible. Indeed, if GdPO$_4$ monazite as well as GdPO$_4$·0.667H$_2$O rhabdophane are well described [28, 29], a very limited number of synthesis protocols is available for xenotime and churchite [30]. The present study then reports for the first time the direct comparison of both Raman and FTIR spectra of the four gadolinium phosphate structures and establishes connections between their crystal structure, including symmetry of PO$_4$ groups and hydrogen bond network formed by water molecules, and their spectroscopic behavior.
2. Experimental

2.1 Preparation and characterization of the samples

All the samples were synthesized from GdCl$_3$·6H$_2$O (99%) as well as concentrated H$_3$PO$_4$ (85%) and HCl, all supplied by Sigma-Aldrich. In order to avoid any weighing bias due to the hygroscopic character of the gadolinium salt, this latter was first dissolved in 1M HCl. Gadolinium concentration was finally accurately determined by Inductively Coupled Plasma Atomic Emission Spectroscopy (ICP-AES) and was always in the 0.5-1M range.

GdPO$_4$·0.667H$_2$O rhabdophane [6] was obtained following the protocol reported in our previous work [7]. Hydrochloric solution containing gadolinium was mixed with 5M H$_3$PO$_4$ in a PTFE-lined vessel, an excess of 2 mol.% being considered to ensure the quantitative precipitation of the cations. The container was then transferred into an oven and left at 90°C for two weeks. The precipitate formed afterwards was recovered by centrifugation at 14000 rpm, then successively washed twice with deionized water and once with ethanol, and finally dried overnight in air at 90°C. The GdPO$_4$ monazite counterpart was obtained by heating the former compound at 1100°C for 6 hours in air. These firing conditions were chosen in order to fully convert rhabdophane into monazite, the transition being reported around 850°C [31].

GdPO$_4$·2H$_2$O churchite was obtained through a similar protocol than that described for the preparation of rhabdophane. However, in this case, the mixture of reactants was placed in a fridge at 4°C for several months. Separation of the precipitate, as well as washing and drying steps were repeated as described above. Finally, GdPO$_4$ xenotime was prepared after firing the churchite precursor at 1000°C for 6 hours in air [30].

The nature of the various GdPO$_4$·nH$_2$O phases prepared was first checked by the means of powder X-ray diffraction (PXRD) using a Bruker D8 advance diffractometer equipped with a lynxeye detector and having copper radiation (λ=1.54184Å). The collected patterns confirm the formation of four different polymorphs (Figure 1), in good agreement with the structures reported in the literature for Gd-based monazite (PDF file #01-083-0657 [4]), xenotime (#00-062-824 [32]), rhabdophane (#00-039-0232) and churchite (#00-021-0337 [33]). Associated unit cell parameters obtained from Rietveld refinement are gathered in Table 1.
Figure 1. XRD patterns obtained for the four gadolinium phosphate samples.

Table 1. Unit cell parameters determined for GdPO₄·nH₂O samples.

<table>
<thead>
<tr>
<th></th>
<th>a (Å)</th>
<th>b (Å)</th>
<th>c (Å)</th>
<th>β (°)</th>
<th>V (Å³)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rhabdophane</td>
<td>28.0273(2)</td>
<td>6.9262(1)</td>
<td>11.9914(1)</td>
<td>115.17(1)</td>
<td>2106.67(4)</td>
</tr>
<tr>
<td>Monazite</td>
<td>6.6515(1)</td>
<td>6.8446(1)</td>
<td>6.3340(2)</td>
<td>104.02(1)</td>
<td>279.77(1)</td>
</tr>
<tr>
<td>Churchite</td>
<td>6.2102(1)</td>
<td>15.1104(2)</td>
<td>5.6162(1)</td>
<td>114.95(1)</td>
<td>477.81(2)</td>
</tr>
<tr>
<td>Xenotime</td>
<td>6.9612(7)</td>
<td>6.1026(10)</td>
<td></td>
<td></td>
<td>295.73(9)</td>
</tr>
</tbody>
</table>

As previously stated, Gd-monazite adopts a P₂₁/n monoclinic structure. This latter is based on the widely-described nine-fold coordination of the cation which can be described as a pentagon interpenetrated by a distorted tetrahedron. These REEO₉ polyhedra then form chains along the [001] direction [2]. Within the xenotime structure, the PO₄ tetrahedra share corners and edges with REEO₈ polyhedra, which themselves share edges together to form chains parallel to the [100] direction. These chains are cross-linked with PO₄ units by sharing corners. The connections between monazite and xenotime structures, already described by Ni et al. [4],
consists in a shift of the (100) planes along with a slight rotation of the PO4 tetrahedra around [001].

Described for long as an hexagonal structure, GdPO4·0.667H2O rhabdophane was only recently reported to crystallize in the monoclinic system in the C2 space group. The structural arrangement consists in infinite channels oriented along the [101] direction and formed by the connection of six chains. One must note that the channels contain two types of chains, depending on the coordination of the cation which can be 8 or 9 [6].

Finally, the churchite crystallizes in the gypsum structure type in the C2/c space group of the monoclinic system [34]. Each Gd atom is coordinated to 8 oxygen atoms, six being provided by the phosphate groups and two by water molecules, to form distorted square antiprisms. These polyhedra are separated by phosphate groups and form infinite 2D layers stacked down the b axis.

Figure 2. View of the structure of: a) GdPO4·0.667 rhabdophane (along the [101] direction; b) GdPO4 monazite ([100] direction); c) GdPO4·2H2O churchite ([001] direction); d) GdPO4 xenotime ([001] direction).
2.2. Raman spectroscopy

Raman spectra were recorded by the means of a Horiba - Jobin Yvon Aramis apparatus equipped with an edge filter and using a Nd:YAG laser (532 nm). In order to avoid any laser-induced degradation of the compound, the power was turned down by the means of optical filters to about 1-4 mW depending on the sample analyzed. The laser beam was then focused on a small fraction of powder simply deposited on a glass lamella using an Olympus BX 41 microscope. A ×100 objective with a numerical aperture of 0.9, resulting in a spot size of about 1 µm² was used. The scattered Raman light was collected in a 180° backscattering geometry and dispersed by a grating of 1800 grooves/mm after having passed a 150 µm entrance slit, resulting in a spectral resolution lower than 1 cm⁻¹. For each spectrum, a dwell time of 90 to 180 seconds was considered with an average of 3 scans. Before analysis, the apparatus was calibrated with a silicon wafer, using the first-order Si line at 520.7 cm⁻¹.

2.3. FTIR spectroscopy

FTIR spectra were recorded in the 380-4000 cm⁻¹ range thanks to a Perkin-Elmer FTIR Spectrum 100 device. Powdered samples were deposited at the surface of an ATR crystal without any prior preparation. The spectra collected in such operating conditions exhibited a resolution lower than 2 cm⁻¹.
3. Results and discussion

3.1 General overview

The FTIR and Raman spectra recorded for the four gadolinium phosphate polymorphs are gathered in Figure 3. Band component analysis of the different data sets was carried out by the means of the Jandel Peakfit software, using pseudo-Voigt functions (Gaussian-Lorenzian ratio systematically higher than 0.7) with the minimum number of components. Correlation coefficients $R^2$ greater than 0.992 were usually obtained for the results reported in Table 2.

![Figure 3](image_url)

**Figure 3.** FTIR and Raman spectra recorded for the four gadolinium phosphate polymorphs: rhabdophane, monazite, churchite and monazite.
Table 2. Assignment of the vibration bands observed on the FTIR and Raman spectra recorded for the four gadolinium phosphate polymorphs: rhabdophane, monazite, churchite and monazite. Bands indicated into parenthesis are shoulders.

<table>
<thead>
<tr>
<th>Band assignment</th>
<th>Rhabdophane FTIR</th>
<th>Rhabdophane Raman</th>
<th>Monazite FTIR</th>
<th>Monazite Raman</th>
<th>Churchite FTIR</th>
<th>Churchite Raman</th>
<th>Xenotime FTIR</th>
<th>Xenotime Raman</th>
</tr>
</thead>
<tbody>
<tr>
<td>ν(H_2O)</td>
<td>3535 3475 3350</td>
<td>3540 3470 (3330)</td>
<td></td>
<td></td>
<td>3365</td>
<td>3380</td>
<td></td>
<td></td>
</tr>
<tr>
<td>δ(H_2O)</td>
<td>(1630) 1615</td>
<td></td>
<td>1705</td>
<td>(1690)</td>
<td>1630</td>
<td>(1610)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ν_3(PO_4)</td>
<td>1070 1027 1001</td>
<td>1106 (1070) 1022</td>
<td>1092</td>
<td>1071</td>
<td>1052</td>
<td>1058</td>
<td>1117</td>
<td>1057</td>
</tr>
<tr>
<td>ν_2(PO_4)</td>
<td>972 930 927</td>
<td>994 (933)</td>
<td>987 (943)</td>
<td>977 (878)</td>
<td>995</td>
<td>977</td>
<td>988</td>
<td>991</td>
</tr>
<tr>
<td>δ_3(PO_4)</td>
<td>620 576 538</td>
<td>634 (642)</td>
<td>632 (649)</td>
<td>(634)</td>
<td>636</td>
<td>646</td>
<td></td>
<td></td>
</tr>
<tr>
<td>δ_2(PO_4)</td>
<td>(506) 506 492</td>
<td>471 492</td>
<td>(493)</td>
<td>497</td>
<td></td>
<td></td>
<td>481</td>
<td></td>
</tr>
<tr>
<td>δ_s(PO_4)</td>
<td>(646) 538 538</td>
<td>429 539</td>
<td>(458)</td>
<td>(483)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lattice modes</td>
<td>404</td>
<td></td>
<td>362</td>
<td></td>
<td>364</td>
<td></td>
<td>346</td>
<td></td>
</tr>
</tbody>
</table>

All the spectra recorded for the four polymorphs could be split in three distinct zones of interest. The first one, comprised between 300 and 800 cm⁻¹, corresponds to the deformation
modes of the PO₄ tetrahedron, while stretching modes led to characteristic bands in the 900-1200 cm⁻¹ range. If these regions are similar for the various compounds studied herein, the number of vibration bands assigned to PO₄ groups differs depending on the crystal symmetry of the samples. The subsequent factor group analysis was reported by many authors for monazite [19-21], including recent papers bringing ab initio simulations to the state of the art [35]. Meanwhile, some data also exists in the literature for xenotime [20] and churchite [27]. However, it has to be reconsidered for rhabdophane due to the recent description of its structure as monoclinic (C2 space group) instead of the hexagonal P6₃22 [6]. Up to date, factor group analysis is then presented in Table 3. In parallel, the vibration bands assigned to the water molecules can be found between 1500 and 1700 cm⁻¹ (bending) and around 3600 cm⁻¹ (stretching). The results corresponding to these three different regions, i.e. to stretching and bending modes of phosphate entities, then to structural water vibrations, will be discussed in the following sections.
Table 3. Correlation of free PO$_4$ ion and the corresponding factor groups of gadolinium phosphate polymorphs.

<table>
<thead>
<tr>
<th>Free ion</th>
<th>Site symmetry</th>
<th>Factor group symmetry</th>
</tr>
</thead>
<tbody>
<tr>
<td>Td</td>
<td>C$_1$</td>
<td>C$_{2h}$</td>
</tr>
<tr>
<td>Monazite</td>
<td></td>
<td></td>
</tr>
<tr>
<td>A$_1$</td>
<td></td>
<td></td>
</tr>
<tr>
<td>E</td>
<td>9 A</td>
<td>9 Ag</td>
</tr>
<tr>
<td>2 F$_2$</td>
<td></td>
<td>9 Bg</td>
</tr>
<tr>
<td></td>
<td></td>
<td>9 Au</td>
</tr>
<tr>
<td></td>
<td></td>
<td>9 Bu</td>
</tr>
<tr>
<td>Td</td>
<td>D$_{2d}$</td>
<td>D$_{4h}$</td>
</tr>
<tr>
<td>Xenotime</td>
<td></td>
<td></td>
</tr>
<tr>
<td>A$_1$</td>
<td>2 A$_1$</td>
<td>2 A$_{1g}$</td>
</tr>
<tr>
<td>E</td>
<td>B$_1$</td>
<td>1 A$_{1u}$</td>
</tr>
<tr>
<td>2 F$_2$</td>
<td>2 B$_2$</td>
<td>2 B$_{1g}$</td>
</tr>
<tr>
<td></td>
<td>2 E</td>
<td>B$_2g$</td>
</tr>
<tr>
<td></td>
<td></td>
<td>2 B$_{2u}$</td>
</tr>
<tr>
<td></td>
<td></td>
<td>2 E$_g$</td>
</tr>
<tr>
<td></td>
<td></td>
<td>2 E$_u$</td>
</tr>
<tr>
<td>Td</td>
<td>C$_1$</td>
<td>C$_2$</td>
</tr>
<tr>
<td>Rhabd.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>A$_1$</td>
<td></td>
<td>5 A</td>
</tr>
<tr>
<td>E</td>
<td>9 A</td>
<td>4 B</td>
</tr>
<tr>
<td>2 F$_2$</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Td</td>
<td>C$_2$</td>
<td>C$_{2h}$</td>
</tr>
<tr>
<td>Churchite</td>
<td></td>
<td></td>
</tr>
<tr>
<td>A$_1$</td>
<td>5 A</td>
<td>5 Ag</td>
</tr>
<tr>
<td>E</td>
<td>4 B</td>
<td>4 Bg</td>
</tr>
<tr>
<td>2 F$_2$</td>
<td></td>
<td>5 Au</td>
</tr>
<tr>
<td></td>
<td></td>
<td>4 Bu</td>
</tr>
</tbody>
</table>
3.3 Phosphate stretching modes

The stretching modes of the PO$_4$ groups are observed in the Raman spectra gathered in Figure 4. In this zone (900-1200 cm$^{-1}$), the symmetric vibration, frequently labelled as $v_1$, appears to be located on a very small wavenumber range for the four crystal structures considered, i.e. at 977, 984, 987 and 994 cm$^{-1}$ for churchite, xenotime, monazite and rhabdophane, respectively. On this basis, the wavenumber assigned to the $v_1$(PO$_4$) vibrations can be hardly used to accurately discriminate the various polymorphs even if some differences can be underlined concerning the number of band components. Indeed, monazite and churchite led to a single vibration band, while shoulders can be observed for xenotime (991 cm$^{-1}$, i.e. at higher wavenumber than the main vibration band) and rhabdophane (985 cm$^{-1}$, i.e. at lower wavenumber).

On the other hand, the vibrations associated to the antisymmetric stretching mode ($v_3$) located above 1040 cm$^{-1}$ are generally of lower intensity. Nevertheless, they present significant differences depending on the sample considered, particularly concerning the position of the main vibration band. Indeed, the wavenumber of the band with the maximal intensity varies from 1048 cm$^{-1}$ for xenotime to 1058, 1071 and 1108 cm$^{-1}$ for churchite, monazite and rhabdophane, respectively. Even if no clear correlation can be established between the P-O distance in the different structures and the $v_3$ wavenumber, the lowest wavenumbers were measured for xenotime and churchite. In both these structural type, gadolinium is eight-fold coordinated, while its coordination is 9 for rhabdophane and monazite. This lower coordination state results in shorter Gd-O distances, then in longest P-O lengths, in good agreement with the position of the PO$_4$ stretching bands in the spectra. One must also note that if only one antisymmetric vibration band is observed for rhabdophane and churchite, several smaller vibration bands were detected for monazite and xenotime. Particularly, the existence of three bands of weak intensity at around 1000, 1040, and 1090 cm$^{-1}$ for GdPO$_4$ monazite, that was already evidenced by Begun et al. and also observed for the other members of the LnPO$_4$ series [20], can be considered as a characteristic feature for this structural type.
Figure 4. Raman spectra recorded in the 900-1200 cm$^{-1}$ domain for gadolinium-bearing rhabdophane, monazite, churchite and monazite.

Compared to Raman ones, the FTIR spectra (Figure 5) recorded for the four samples appear much more convoluted. Indeed, a wide massif was systematically recorded between 800 and 1200 cm$^{-1}$, which makes it difficult to unambiguously discriminate the components assigned to symmetric and antisymmetric stretching vibration modes. Nevertheless, a sharp band can be observed for rhabdophane (at 972 cm$^{-1}$) and monazite (at 960 cm$^{-1}$). It could be assigned to the $\nu_1$ symmetrical stretching mode. Also, these values are in good agreement with that recently reported by Heuser et al. [21]. For xenotime and churchite, the resolution of the spectra collected was probably lowered by the crystallization state. Also, it might arise from the presence of bands related to forbidden vibration modes. Nevertheless, a simple first-basis approach could be considered in this case by assigning all the vibration bands above 1000 cm$^{-1}$ to antisymmetric stretching vibration modes ($\nu_3$). Finally, one must note that a specific vibration mode related to water molecules can be observed for churchite at 732 cm$^{-1}$. It will be discussed in the dedicated section.
Figure 5. FTIR spectra recorded in the 700-1300 cm\(^{-1}\) range for gadolinium bearing rhabdophane, monazite, churchite and monazite.

3.4 Phosphate deformation modes

The Raman spectral zone corresponding to the bending modes of the PO\(_4\) groups is mainly characterized by the presence of an intense band around 470-480 cm\(^{-1}\) (Figure 6) and assigned to the symmetric vibration (frequently labelled as \(v_2\)). As already noted by Silva et al., this mode occurs systematically as a broad band, regardless of any splitting of the degeneracy [19]. The position of this vibration mode appears to be weakly dependent on the structural type considered, with monazite, rhabdophane and xenotime features occurring on a 10 cm\(^{-1}\) range. More specifically, churchite is characterized by the very low intensity of the \(v_2\) mode. For this sample, the most intense band occurs at around 360 cm\(^{-1}\), i.e. just above the wavenumber domain usually assigned to external vibration modes. Nevertheless, it was systematically correlated to phosphate internal modes in the literature [26, 27]. On the FTIR spectra, the \(v_2\) vibration mode of the PO\(_4\) units was observed for monazite, rhabdophane and churchite, even if it appears frequently as a shoulder. For the two hydrated compounds, the existence of the \(v_2\) band confirms the C\(_2\) site symmetry proposed from crystal structure. On the contrary, the D\(_{2d}\)
site symmetry of the phosphate groups in the xenotime structure leads the symmetrical bending vibration to be absent on the FTIR spectra [21].

Figure 6. Raman spectra recorded in the 400-800 cm\(^{-1}\) range for gadolinium based rhabdophane and monazite and between 300 and 700 cm\(^{-1}\) for gadolinium based churchite and monazite.

At higher wavenumber, i.e. typically between 500 and 700 cm\(^{-1}\), the antisymmetric bending modes (\(\nu_4\)) mostly generate intense vibration bands in the FTIR spectra, which result in some large massifs. For rhabdophane, churchite, and xenotime, two main bands were detected at around 520-540 cm\(^{-1}\) and 620-640 cm\(^{-1}\), while monazite exhibited more complex features between 500 and 600 cm\(^{-1}\), which were sometimes depicted as the succession of two doublets (538/570 then 585/623 cm\(^{-1}\)) [36]. On the other hand, Raman spectra often present a collection of several low-intensity bands, which wavenumbers match with that of FTIR vibrations. These latter then do not follow the geometry selection rules and probably come from the presence of solid defects. However, the structure of these additional bands can be considered as characteristic of a given structural type. Particularly, the various publications dealing with monazite-type samples systematically report bands at about 540, 580 and 600 cm\(^{-1}\) [19]. The same applies for the quadruplet recorded between 557 and 586 cm\(^{-1}\) for xenotime.
Figure 7. FTIR spectra recorded in the 400-700 cm\(^{-1}\) range for GdPO\(_4\)·nH\(_2\)O rhabdophane, monazite, churchite and monazite.

3.5 Water molecules

The spectral zones associated to the vibrations of water molecules, \(i.e.\) bending modes typically between 1500 and 1600 cm\(^{-1}\) and stretching modes above 3000 cm\(^{-1}\), were particularly probed for GdPO\(_4\)·0.667H\(_2\)O rhabdophane and GdPO\(_4\)·2H\(_2\)O churchite samples, which are well known to be hydrated compounds (Figure 8). One must note that a characteristic water rocking mode can also be observed for churchite on the FTIR spectra as a broad band at around 730 cm\(^{-1}\) [27]. Also, even if xenotime sometimes showed some weak intensity bands in the regions mentioned above, it was more likely correlated to absorbed water due to the powder’s small grain size (thus to an important number of surface adsorption sites), and the existence of structural water molecule in the structure was ruled out.

Whether regarding FTIR or Raman spectra, the characteristic features of water molecules appear to be strongly different between churchite and rhabdophane. For the FTIR-active bending modes, churchite presents two distinct bands at 1630 and 1705 cm\(^{-1}\) with small shoulders at lower wavenumbers (\(i.e.\) 1610 and 1690 cm\(^{-1}\)) which can be assigned to forbidden
vibration modes, while rhabdophane exhibits only strong vibration band at 1615 cm\(^{-1}\) with a shoulder at 1630 cm\(^{-1}\). A similar behavior was observed for stretching modes, since more complex signals were systematically recorded for churchite. As a matter of fact, rhabdophane only presented one main vibration at around 3500 cm\(^{-1}\) both in Raman and FTIR spectra, which is accompanied by smaller shoulders. On the other hand, churchite is characterized by large massifs in the 3000-4000 cm\(^{-1}\) region, which are systematically composed by three components or more.

**Figure 8.** Raman spectra in the 2500-4000 cm\(^{-1}\) range and FTIR spectra in the 1500-1800 cm\(^{-1}\) and 2500-4000 cm\(^{-1}\) regions, recorded for gadolinium rhabdophane, monazite, churchite and monazite.

Such differences can be easily correlated to the crystal structure of the two compounds. Indeed, churchite bears two non-equivalent water molecules in its structure, one being strongly attached to the PO\(_4\) network and the other more loosely bound in the interlayer space [34, 37]. On this basis, the bending band observed at higher wavenumber (\(i.e.\) around 1700 cm\(^{-1}\)) should correspond to the longest bond distance, then to the less strongly coordinated water molecule. A similar assignment was already made for isostructural CaSO\(_4\)·2H\(_2\)O gypsum, for which the second band (1685 cm\(^{-1}\)) was found to progressively disappear upon dehydration while the first one (1620 cm\(^{-1}\)) remained present in the final CaSO\(_4\)·0.5H\(_2\)O spectra [38]. Moreover, the wavenumber of the stretching modes observed on the FTIR spectra can be used to evaluate the d(H···O) bond lengths in the hydrogen bond network thanks to the empirical relation established by Libowitzky from mineral samples (including silicates and phosphates) [39]. Based on the three band components at 2870, 3132 and 3365 cm\(^{-1}\), distances of about 1.67,
1.76, and 1.89 Å, respectively, were obtained, which are in the same range of magnitude than those reported by Frost et al. for natural samples of Y-churchite [40].

In the case of rhabdophane, the recently revised structure includes four non-equivalent water molecules [6]. Nevertheless, all of them take part to the coordination of the metal center, resulting in a close set of M-O₆ bond distances. As a consequence, both Raman and FTIR spectra present strongly overlapping bands in the 3000-4000 cm⁻¹ range. The narrowing of the stretching vibration bands can also be associated to the decrease of the hydration content of the GdPO₄ sample compared to churchite. Indeed, it led to the weakening of the hydrogen bonding which generally tends to broaden the signal in this region [41]. Similarly, the existence of only one bending mode at 1615 cm⁻¹ could be correlated to the quasi-equivalent nature of the water molecules in the rhabdophane structure. Also, as in the case of churchite, a parallel can be drawn with hydrated forms of gypsum. Indeed, Mesbah et al. already stressed out the isostructural character of CaSO₄·0.625H₂O and rhabdophane, as well as that of CaSO₄·0.5H₂O with the hexagonal LnPO₄·0.5H₂O [42]. Nevertheless, if numerous studies were undertaken concerning the vibrational characterization of dehydrate and hemihydrate gypsum, no reference spectra seems to exist, to our knowledge, for CaSO₄·0.625H₂O. Only Fritz et al. reported a Raman spectra for the selenite counterpart CaSeO₄·0.625H₂O [43] but without investigating the typical wavenumber range of water molecules vibration. In these conditions, the FTIR and Raman data collected in this study for GdPO₄·0.667H₂O rhabdophane can be considered as a potential first-basis approach to forecast the spectral characteristics of water in these compounds.

3.6 Discussion

The comparison of the vibrational spectra of LnPO₄·nH₂O lanthanide phosphate polymorphs was undertaken for the first time using a single metal cation, i.e. gadolinium. Even if the assignment of the various vibration modes was reported for long in the literature for several monazite, rhabdophane, churchite and xenotime type samples, this direct comparison can be used to propose some guidelines to quickly discriminate the different structural types. First, whereas the symmetrical stretching vibrations of the phosphate tetrahedra usually led to the more intense vibration bands, they appeared to have very close wavenumbers. Also, their resolution is strongly dependent on the crystallization state of the samples, which can bias their analysis, particularly for natural samples. On this basis, their examination should be discarded to unambiguously identify the sample.
Conversely, the identification of water vibration modes seems to be the more rapid way to identify the crystal structure of any LnPO$_4$·nH$_2$O sample. Indeed, the existence of well-defined vibration bands in the 1500-1600 cm$^{-1}$ range of the FTIR spectra, related to the $\delta$(H$_2$O) modes, attests of the existence of structural water in the sample. Here, bending modes are to be preferred, as the spectral region associated to stretching vibration modes could also show some large bands in the presence of adsorbed water. Thereafter, the number of $\delta$(H$_2$O) modes easily lead to identify the structure, churchite being associated to two bands, while rhabdophane only exhibits one single signal in this region.

For anhydrous samples adopting the xenotime or monazite structures, the main difference in the spectral signature probably comes from the vibrations associated to the phosphate bending modes. In the FTIR spectra, the $\delta$(PO$_4$) mode ($\nu_2$) is inactive for the tetragonal xenotime, while it led to one vibration band for the monoclinic monazite. Nevertheless, the corresponding signal is frequently merged into a large massif gathering both symmetric and anti-symmetric modes, which can hampers its identification. Then, we suggest to use the Raman data in this case. Indeed, xenotime presents only one intense $\nu_2$ band, while two signals are detected for monazite, the second band located at about 430 cm$^{-1}$ being almost as intense.

Based on these recommendations, a schematic methodology is proposed in Figure 9 to rapidly discriminate the four gadolinium phosphate polymorphs. Once again, it demonstrates the complementarity of FTIR and Raman spectroscopies, which have to be used together to unambiguously assign one of the four polymorphs. Also, as the method is mainly based on the number of vibration bands observed, it could be easily applied to any chemical system, including pure parent compounds or solid solutions involving several metal cations. In these conditions, it appears suitable for field investigations, particularly in the domain of geochemistry where monazite, rhabdophane, churchite and xenotime have already dragged a significant attention.
4. Conclusion

Gadolinium phosphates with monazite, rhabdophane, churchite, and xenotime structures were analyzed by vibrational spectroscopy (FTIR and Raman), leading to the first direct comparison of LnPO$_4$·nH$_2$O polymorphs for a single metal cation. General overview led to split the spectra into three zones, corresponding to the bending and the stretching vibration modes of phosphate tetrahedra and to the vibration modes of water molecules. For all the samples studied, the wavenumbers associated to $\nu_s$(PO$_4$) bands were found to be very close, and can be hardly used to discriminate the various structural type, despite of their important intensities. Conversely, phosphate bending modes were found to be more informative. Particularly, the number of bands observed on the Raman spectra can differentiate monazite from xenotime, provided that the presence of water molecules was previously ruled out. For hydrated samples, i.e. rhabdophane and churchite, the $\delta$(H$_2$O) modes were found to be well-defined in FTIR spectra, and attested of the structural character of the water molecules. Moreover, the number of bands observed can once again be used to identify the structure of the GdPO$_4$·nH$_2$O sample.

The comparison of the vibrational spectroscopy data led to propose a simple methodology to rapidly discriminate the four LnPO$_4$·nH$_2$O polymorphs, which could be applied during field investigations, for example in the domain of geochemistry.
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