

# Heterogeneity of the chemical composition and thermal stability of particulate organic matter in French forest soils

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Heterogeneity of the chemical composition and thermal stability of particulate organic matter in French forest soils Laure Soucémarianadin<sup>1,\*</sup>, Lauric Cécillon<sup>1,2</sup>, Claire Chenu<sup>3</sup>, François Baudin<sup>4</sup>, Manuel Nicolas<sup>5</sup>, Cyril Girardin<sup>3</sup>, Amicie Delahaie<sup>1</sup> and Pierre Barré<sup>1</sup> <sup>1</sup> Laboratoire de Géologie, PSL Research University, CNRS-ENS UMR8538, Paris, France <sup>2</sup> Université de Normandie, UNIROUEN, IRSTEA, ECODIV, 76000 Rouen, France <sup>3</sup> UMR ECOSYS, INRA, AgroParisTech, Université Paris-Saclay, 78850 Thiverval-Grignon, France <sup>4</sup> Sorbonne Université/CNRS, UMR ISTeP, 4 place Jussieu 75005 Paris, France <sup>5</sup> Office National des Forêts, R&D, 77300 Fontainebleau, France \* Corresponding author: Laure Soucémarianadin, souce@geologie.ens.fr Laboratoire de Géologie (UMR 8538) Ecole Normale Supérieure, 24 Rue Lhomond 75231 Paris CEDEX 5, France; phone: +331 44 32 22 94; fax: +331 44 32 22 00 Type: Regular article 

#### Abstract

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25 In temperate forests, soils contain a significant part of the ecosystem carbon (C) stock that can be subjected to C losses upon global changes. In forest soils, particulate organic matter 26 27 (POM) is a major contributor to the labile C pool and its dynamics can significantly influence 28 the overall total soil organic carbon stock. However, POM has been overlooked in forest soils, 29 specifically in deep horizons. 30 We isolated the POM fraction of mineral soil samples collected in 52 French forest sites, 31 using a size- (> 50  $\mu$ m) and density- (< 1.6 g·cm<sup>-3</sup>) fractionation scheme. These soil samples 32 presented variability in terms of depth (0–10 cm; 40–80 cm), soil class (dystric Cambisol, 33 eutric Cambisol, entic Podzol) and vegetation type (deciduous, coniferous). First, we 34 determined the POM chemical composition and thermal stability using elemental analysis, mid infrared-attenuated total reflectance spectroscopy and Rock-Eval thermal analysis. Then, 35 we assessed how depth, soil class and vegetation type influenced POM chemistry and thermal 36 37 stability in these temperate forest soils. 38 Depth, soil class and vegetation type were all important factors influencing POM chemistry 39 and thermal stability. Variations in POM chemistry (higher C/N ratio, lower ether + alcohol 40 and carbonyl + carboxyl ratios and decrease in hydrogen-rich compounds) and increase in 41 thermal stability with depth suggested different POM input sources for the surface and deep 42 soil layers and an increased biogeochemical stability of POM in deep soil layers. Whatever 43 the vegetation, POM in eutric Cambisols had lower aliphatic and higher aromatic ratios than 44 POM in dystric Cambisols. POM in soils under deciduous trees had higher aliphatic and 45 carbonyl + carboxyl ratios and lower aromatic ratio, more hydrogen-rich and less oxygen-rich compounds than POM in soils under coniferous trees, reflecting the difference in litter 46 47 chemistry between the two vegetation types. POM from deciduous plots was also significantly 48 more thermally stable than from coniferous plots, suggesting a higher biogeochemical 49 stability for POM in deciduous forest soils. This study highlights the variations in POM chemistry and thermal stability existing within 50 51 and among soil profiles and the role of depth, soil class and vegetation type in these 52 variations. It appears that if POM can be regarded as a labile carbon fraction in soils, its 53 lability varies depending on the ecosystem (soil, vegetation) and depth considered. 54 55 **Keywords**: forest soils; soil organic carbon stability; particulate organic matter; Rock-Eval; mid-infrared spectroscopy; environmental drivers. 56 57 Abbreviations: Rock-Eval 6 (RE6), particulate organic matter (POM), mid-infrared-58 attenuated total reflectance (MIR-ATR) 59 60

#### 1. Introduction

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62 Soil organic carbon (SOC) is made up of very heterogeneous compounds (Trumbore, 1997; 63 Amundson, 2001) with turnover times ranging from a few days/weeks to several centuries. 64 The most labile SOC pools are very sensitive to environmental changes (Carter et al., 1998; 65 Zhang et al., 2007), which could result in rapid variations of SOC stocks (~years) and as such 66 they play an important role in global warming and its mitigation. Labile SOC is also central to 67 short-to medium-term nutrient availability and soil structural stability (Wander, 2004). The 68 particulate organic matter (POM) fraction represents a labile SOC pool constituted of partially 69 decomposed organic debris, i.e., plant and animal residues, (Spycher et al., 1983; Christensen, 70 1992) with a mean residence time that is usually considered to be < 20 years for temperate in-71 situ conditions (e.g., Trumbore et al., 1996; Balesdent, 1996). POM fractions are separated 72 and quantified as low-density and/or coarse-size fractions (often > 50–250 μm; Wander, 73 2004). POM is primarily a source of energy and C for microbial decomposers and soil fauna 74 but is also a source of N and other nutrients (Haynes, 2005). POM fractions are typically 75 enriched in carbohydrates relative to whole soils and heavy fractions (Baldock et al., 1992; 76 Baldock et al., 1997) with carbohydrates representing 5 to 25% of the C in the POM fraction 77 (Oades, 1972; Oades et al., 1987). The elemental composition of POM also plays a major role 78 in its dynamics and organic materials with higher N content (low C/N ratio) tend to 79 decompose faster than those with low N content (Swift et al., 1979; Aber and Melillo, 1980; 80 Melillo et al., 1982; Cotrufo et al., 1995). 81 Soils experiencing conditions that will limit decomposition (e.g., cold and dry climates or low 82 pH) and that are under permanent vegetation allowing for high inputs of aboveground and 83 belowground plant litter (e.g., forests and permanent grasslands) will tend to favour POM 84 accumulation (e.g., Whitehead et al., 1975; Balesdent et al., 1998; Leifeld et al., 2009). In 85 agricultural soils, the POM fraction contains 2-18% of total SOC and 1-16% of total soil N

(Gregorich and Janzen, 1996). In forest topsoils, the POM fraction represents 5–47% of total 86 87 SOC and 3–40% of total soil N (Christensen, 1992). Variations in SOC stocks after land-use 88 change have been largely associated to C gains or losses in the POM fraction (Poeplau and 89 Don, 2013). 90 Some studies have investigated the effects of environmental factors on POM properties. POM 91 quantity and chemistry have been shown to vary according to vegetation (Quideau et al., 92 2001; Laganière et al., 2011; Schrumpf et al., 2013) or parent material/soil type (Kölbl and 93 Kögel-Knabner, 2004; Angst et al, 2018). However, these studies often lack multi-site 94 observations. And despite their importance for SOC dynamics and soil quality, POM fractions 95 from surface and deep horizons of forest soils remain poorly documented. 96 Mid-infrared (MIR) spectroscopy can provide information related to soil chemical 97 composition as most chemical bounds found in soil minerals and organic compounds have 98 spectral absorptions at specific wavelengths of the IR domain (Nguyen et al., 1991). The 99 chemical characterization of soil organic matter (SOM) in bulk soils by MIR spectroscopy is 100 made difficult because of mineral interferences (Nguyen et al., 1991; Reeves, 2012; Cécillon 101 et al., 2012), while the characterization of pure SOM fractions does not suffer this caveat. The 102 MIR technique in the attenuated total reflectance mode (MIR-ATR) has been used to describe 103 the chemical composition of pure SOM in Histosols (Pengerud et al., 2013; Robroek et al., 104 2015) or to study the bulk chemistry of the POM fractions in mineral soils (Puissant et al., 105 2017). 106 Among thermal analyses used to characterize SOM, Rock-Eval (RE) analysis has provided 107 promising results showing that SOC thermal stability observed in RE thermograms results can 108 be related to SOC biogeochemical stability (Gregorich et al., 2015; Saenger et al., 2015; Barré 109 et al., 2016; Soucémarianadin et al., 2018a; Poeplau et al., 2019). RE was also shown to 110 provide information on organic matter evolutions in soils at various depth or during

composting (Hetényi et al., 2005; Hetényi et al., 2006; Sebag et al., 2006; Albrecht et al., 2015), and it has been specifically applied to characterize litters, bulk soil, organic layers and POM fractions in mineral soils (Disnar et al., 2003; Saenger et al., 2015; Sebag et al., 2016). The objectives of this study were thus to characterize POM chemistry and thermal stability in a set of French forest soil samples and then to assess the importance of various environmental factors, namely soil depth, soil class and vegetation type in controlling POM chemistry and thermal stability. To this purpose, we separated by size and density the POM fraction from a set of French forest soil samples that covers a large pedoclimatic variability as well as tree species diversity and includes deep samples (up to 0.8 m). We then used a set of complementary techniques (i.e., C and N elemental analysis, MIR-ATR spectroscopy and RE thermal analysis) to characterize POM chemistry and thermal stability. We hypothesized that 1/vegetation type (coniferous vs. deciduous dominant canopy trees) affects POM chemistry and thermal stability. Specifically we expected POM from coniferous trees to have more aromatic moieties because of their (likely) higher lignin content (Berg et al., 2013) but a lower thermal stability than POM from deciduous plots, at least in the surface layer. Deciduous forests have a more rapid plant-soil nutrient cycling than coniferous forests (Cole and Rapp, 1981) and litter is decomposed faster. In parallel, more decomposed organic matter has been shown to have higher thermal stability (Disnar et al. 2003); 2/ eutric Cambisols provide higher physical protection to POM than the two other soil classes due to their higher calcium (Rowley et al., 2018) and clay contents (Kölb and Kögel-Knabner, 2004). This could result in lower thermal stability for POM in eutric Cambisols; and 3/depth influences POM C/N ratio, increasing with depth as POM would be likely more decomposed in surface than in the deep layer due to more favourable environmental conditions and a greater contribution of partially decomposed leaf litter.

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#### 2. Material and methods

2.1. Sampling

Mineral soils samples were collected in 52 forest sites of the French national network for the long term monitoring of forest ecosystems ("RENECOFOR"). These permanent plots were established in 1992 (Ulrich, 1995) by the National Forest Service (ONF) and are part of the European network ICP-FORESTS level 2 (Fig. 1a). Details regarding these sites have been previously published (Soucémarianadin et al., 2018b). Briefly, they presented variability in terms of soil type with a class constituted of soils related to entic Podzols, another class constituted of eutric and epileptic Cambisols as well as a few Calcisols and a last class constituted of dystric and hyperdystric Cambisols (IUSS Working Group, 2015). Forest vegetation varied also among sites, with coniferous [silver fir (Abies alba Mill.); Norway spruce (Picea abies (L.) H. Karst.); European larch (Larix decidua Mill.); Scots pine (Pinus sylvestris L.)] and deciduous [beech (Fagus sylvatica L.); sessile (Quercus petraea (Matt.) Liebl.) and/or pedunculate oaks (Quercus robur L.)] stands. Finally, the sites also varied in terms of climate (continental influence, oceanic influence, mountainous influence; with mean annual precipitation = 703–1894 mm and mean annual temperature = 4.8–12.3 °C for the 1971–2000 period). Samples representing two mineral soil layers were obtained (0–10 cm = topsoil and 40–80 cm = subsoil; Fig. 1b) at each site. Samples of the topsoil layer were composite of  $5 \times 5$  sampling points and were collected by digging a 50 cm wide soil profile (Ponette et al., 1997; Jonard et al., 2017). Samples of the deep soil layer (subsoil) were composite from two soil pits (Brêthes et al., 1997). Samples storage and pooling have been detailed elsewhere (Soucémarianadin et al., 2018a). The pooled samples were sieved at 2 mm before analysis.

#### 2.2. Particle size and density SOC fractionation

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The protocol builds on that of Balesdent et al. (1991) combining both size and density separation. Because our protocol involved both density and size fractionation, we chose to refer to the isolated fraction as the particulate organic matter (POM) fraction (and not just light fraction). Details regarding the POM fraction isolation procedure have been previously published (Soucémarianadin et al., 2018a). Briefly 25 g of samples were shaken overnight in a 0.5% sodium hexametaphostate solution with ten 5 mm-diameter glass beads. Samples were thoroughly rinsed over a 50- $\mu$ m mesh with deionized water. The sand fraction (> 50  $\mu$ m) was then mixed with a sodium polytungstate (SPT) solution of density =  $1.6 \pm 0.03$  g·cm<sup>-3</sup> shaken and left to settle down overnight. We collected the floating material with a spatula and placed it over a 50-µm mesh sieve. If material was still found floating, SPT solution was added back to the flask and the previous step was repeated using centrifugation for 30 minutes (2750 rpm) to accelerate the separation, as many times as needed until nothing was left floating. As soon as the POM fraction was added to the sieve, it was thoroughly rinsed using deionized water, and this was repeated throughout the whole process. We relied on a visual check (after drying; magnification ×10) of the POM material and the sieve for residual traces of SPT. In case traces of salt were observed the sample was again thoroughly rinsed with deionized water, dried and checked one more time. The sieves and fractions were then placed in the oven at 50 °C for 24 h before being weighed. POM fractions were further ground with a ball mill (mixer mill MM 200, Retsch Gmbh) or a mortar and pestle when the sample mass was less than 0.05 g. A total of 102 samples were initially fractionated for POM. However, some samples still contained traces of SPT as shown by their low C content ( $\leq 30\%$ ) and/or MIR-ATR spectra (see section 2.4.) and because we were concerned with possible interactions altering the

thermal analysis signals, they were thus excluded from the statistical analysis. This resulted in the exclusion of 16 samples, (n = 6) for the 0–10 cm soil layer and (n = 10) for the 40–80 cm soil layer, leaving a total of 86 samples.

Organic carbon and total nitrogen concentrations of the ground POM fractions were determined by dry combustion with an elemental analyzer (CHN NA 1500, Carlo "Elba").

Using these concentrations we determined the C/N ratio of the POM samples.

#### 2.4. Mid-infrared spectroscopy

The chemical composition of the POM OC was further assessed by Fourier transform mid-infrared-attenuated total reflectance (MIR-ATR) spectroscopy. Prior to analysis, the ground samples were dried overnight at 40 °C to standardize their water content without altering organic matter chemistry.

MIR-ATR spectra were acquired with a Nicolet *iS10* infrared spectrometer (Thermo Fischer Scientific Inc., Madison, WI, USA) equipped with an ATR device (diamond crystal) over the spectral range 4000–650 cm<sup>-1</sup>, with a spectral resolution of 4 cm<sup>-1</sup> and 16 scans. All spectra were corrected for atmospheric interferences (H<sub>2</sub>O and CO<sub>2</sub>). Absorbance was calculated as the inverse logarithm of the measured reflectance.

Pre-processing of the spectra included a systematic offset-correction, which shifted individual spectrum to set the *y*-minimum to zero (Bruker, 2011), using the 4000–3950 cm<sup>-1</sup> region as a reference.

Four spectral regions, adapted from Puissant et al. (2017), corresponding to specific C functional groups were used to characterize POM chemistry: (1) the aliphatic (CH<sub>2</sub> and CH<sub>3</sub> stretch) region between 2950–2900 cm<sup>-1</sup>; (2) the carbonyl and carboxyl (C=O stretch) region

between 1750–1690 cm<sup>-1</sup>; (3) the aromatic C (C=C bond) region between 1605–1575 cm<sup>-1</sup> 211 212 and (4) the ether and alcohol (C-O stretch typically found in carbohydrates) region between 1180–1145 cm<sup>-1</sup> (Fig. A.1). Relative ratios for these four regions were then calculated using 213 214 Eq. 1: 215 Relative ratio (region i) = (area region i)/ $\sum$  (areas of 4 regions)) (Eq. 1) 216 The contribution of soil minerals (e.g. phyllosilicates) or SPT pollution to the intensities of 217 the four waveband-regions was negligible for the 86 selected POM fractions. The potential 218 pollution of POM fractions with SPT was tested by comparing all POM MIR-ATR spectra to 219 the spectrum of pure SPT powder and making sure that there were no over-average peaks, specifically for the two peaks centered around 720 cm<sup>-1</sup> and 860 cm<sup>-1</sup>. These two peaks are 220 221 high intensity and well-defined in the pure SPT powder spectrum but not apparent in most 222 POM spectra. Therefore the presence of peaks centered on these two wavelengths indicated the possibility of SPT contamination and these samples were thus excluded (see section 2.2.). 223 224 Signal processing of the MIR-ATR spectra was performed with the R environment software 225 v.3.3 (R Core Team, 2016) using the pracma (Borchers, 2015) and hyperSpec (Beleites and 226 Sergo, 2015) R packages. 227 228 2.5. Thermal analysis: Rock-Eval 6 229 Samples were also analysed with a Rock-Eval 6 turbo device (Vinci Technologies, France; 230 Behar et al., 2001). We modified the procedure developed for the analysis of SOM by Disnar 231 et al. (2003) as detailed in Soucémarianadin et al. (2018b). Briefly, about 10 mg of ground 232 sample mixed with 50 mg of ground Fontainebleau sand (previously muffle-furnaced at 233 850 °C for 15 minutes to ensure that it was C-free) were exposed to two consecutive thermal

treatments. Samples were first introduced in a pyrolysis oven (under N<sub>2</sub> atmosphere) then in a

combustion oven (under laboratory air atmosphere). The pyrolysis began with an isothermal

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step (at 200 °C) that lasted around 200 seconds and during which the free hydrocarbons (HC) were thermovaporized (S1 peak). A flame ionization detector allowed for the quantification of the pyrolysis effluents (mostly HC), while an infrared detector quantified CO and CO<sub>2</sub> during both the pyrolysis and oxidation stages (Fig. A.2). Two standard RE6 parameters were determined: the hydrogen index (HI; mg HC·g<sup>-1</sup> TOC) and the oxygen index (OI<sub>RE6</sub>; mg  $O_2 \cdot g^{-1}$  TOC). HI corresponds to the quantity of pyrolyzed relative to the total organic carbon content of the sample and describes the relative enrichment/depletion of POM in hydrogen-rich moieties. OI<sub>RE6</sub> corresponds to the oxygen yield as CO and CO<sub>2</sub> during the thermal pyrolysis divided by the total organic carbon content of the sample. It describes the relative oxidation status of POM. In this study, as POM does not contain inorganic C, we corrected the calculation of HI (HI corrected; expressed as mg HC⋅g<sup>-1</sup> OC) to consider all the C signals (TOC and MinC) as organic C. This did not change drastically the values (Table B.1). This corrected HI value is simply referred to as HI hereafter. We derived one additional RE6 parameter describing the thermal stability of POM: T<sub>50 CO2 PYR</sub>, the temperature at which 50% of the CO<sub>2</sub> resulting from organic matter pyrolysis had evolved. Because the signal was noisy at the beginning of the pyrolysis, we started the integration for T<sub>50 CO2 PYR</sub> right after the S1 peak. This T<sub>50</sub> temperature parameter and the HI index have been previously shown as good thermal indicators of SOM biogeochemical stability (Barré et al., 2016; Cécillon et al., 2018). While HI decreased with time since barefallow and with the proportion of persistent SOC in a soil sample, T<sub>50 CO2 PYR</sub> increased (Barré et al., 2016; Cécillon et al., 2018). The latter also appeared as the most important thermal parameter to predict the proportion of persistent SOC with a multivariate model (Cécillon et al., 2018). As such, T<sub>50 CO2 PYR</sub> could be considered as an indicator of C persistence in soils.

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Signal processing of the RE6 thermograms (signal integration and calculation of the  $T_{50}$  temperature) was performed with the R environment software v.3.3 (R Core Team, 2016) using the hyperSpec (Beleites and Sergo, 2015) and pracma (Borchers, 2015) R packages.

2.6. Calculations and statistical analyses

Although our design was not truly balanced, the distribution of our 86 samples, representing 52 plots (Fig. 1b), among the classes of the various environmental factors (depth, soil class and vegetation type) was not too biased (Table C.1).

The statistical analysis to determine the driving factors of POM chemistry was performed in a similar manner as previously published for the SOC stability of the bulk mineral soil (Soucémarianadin et al., 2018b). Briefly, we used multivariate models to assess the effects of the three environmental factors (depth, soil class and vegetation type) on the different POM chemistry and thermal stability parameters. The linear mixed models introduced a random intercept for each site (≈ to treat "site" as random effect) to take into account that the two layers constituted repeated measures (increasing depth within a same RENECOFOR site). To do so, we added the compound symmetry structure to a generalized least squares (gls) function. The latter fits a linear model using generalized least squares (Pinheiro et al. 2016).

do so, we added the compound symmetry structure to a generalized least squares (gls) function. The latter fits a linear model using generalized least squares (Pinheiro et al. 2016). Model selection was then implemented with a top-down strategy for optimal fit with the restricted maximum likelihood (REML) approach. The response variables were transformed, to the exception of T<sub>50\_CO2\_PYR</sub>, using a reciprocal transformation or the Box-Cox transformation technique, as they showed evidence of the variance increasing with the mean response. After transformation, all residuals followed a normal distribution. We relied on the Cook's distance to identify potential outliers.

All comparisons were considered significant at an alpha value ( $\alpha$ ) of 0.05.

Relationships between the parameters related to POM chemistry and parameters linked to thermal stability were estimated using Spearman rank correlation as the data did not meet the assumption of normality. All statistical analyses were performed using the R 3.3 statistical software (R Core Team, 2016) with the nlme (Pinheiro et al., 2016), lme4 (Bates et al., 2015) and car (Fox and Weisberg, 2011) packages.

#### 3. Results

The effects of climate were tested in a preliminary analysis but this factor (or its interactions with other factors) never had a significant influence on parameters of POM chemistry and thermal stability. We thus decided not to present the results here for concision sake.

#### 3.1. Heterogeneity of POM chemistry

#### 3.1.1. Effects of vegetation type

Vegetation type was highly significant for all parameters except for C/N ratio and MIR-ATR ether + alcohol ratio (Table 1). When comparing the three other MIR-ATR-derived ratios in our 52 sites, POM in deciduous plots had higher aliphatic and carbonyl + carboxyl ratios and lower aromatic ratio (0.191  $\pm$  0.016, 0.220  $\pm$  0.031 and 0.239  $\pm$  0.048, respectively) than the POM found in soils from coniferous plots (0.181  $\pm$  0.009, 0.207  $\pm$  0.021 and 0.268  $\pm$  0.030, respectively; Table 2).

#### 3.1.2. Effect of soil class

Soil class was highly significant for all parameters except for both the MIR-ATR aromatic ratio and ether + alcohol ratio (Table 1). In our sample set, when differences were observed among soil classes, the general trend was that a given parameter (C/N ratio, aliphatic and carbonyl + carboxyl ratios) was lower in POM from eutric Cambisols ( $34 \pm 7$ ;  $0.178 \pm 0.010$ ;

 $0.190 \pm 0.020$ , respectively) than in POM from dystric Cambisols ( $42 \pm 14$ ;  $0.192 \pm 0.016$ ; 311  $0.226 \pm 0.023$ , respectively; Table 2). Values of these parameters for the POM in entic 312 Podzols were lower than for the POM in dystric Cambisols but still significantly greater than 313 for the POM in eutric Cambisols (Table 2). 314 3.1.3. Effect of soil depth 315 The effect of depth was significant for all parameters of POM chemistry except the aliphatic 316 ratio (Table 1). The MIR-ATR ether + alcohol and carbonyl + carboxyl ratios significantly 317 decreased with depth  $(0.357 \pm 0.019 \text{ to } 0.336 \pm 0.017 \text{ and } 0.221 \pm 0.025 \text{ to } 0.204 \pm 0.026,$ 318 respectively), while the aromatic ratio significantly increased (0.236  $\pm$  0.038 to 0.274  $\pm$  0.038; 319 Table 2). The C/N ratio also increased from  $30 \pm 8$  in the surface layer to  $48 \pm 11$  in the deep 320 layer (Table 2). 321 3.1.4. Factor interactions 322 Depth × soil was the most common significant interaction (C/N ratio, aliphatic ratio and 323 aromatic ratio), followed by soil × vegetation (aromatic ratio and carbonyl + carboxyl ratio) 324 and depth  $\times$  vegetation (carbonyl + carboxyl ratio; Table 1). By looking closely at these 325 interactions, different patterns were observed. 326 Soil class was an important driver of POM chemistry through its interactions with both depth 327 and vegetation type, with the effects of these two environmental drivers being generally 328 buffered in eutric Cambisols. Specifically there was no effect of vegetation type on aromatic 329 ratio and carbonyl + carboxyl ratio in eutric Cambisols (Fig. D.1). Moreover, for these two 330 ratios, differences among soil classes were smaller under the coniferous vegetation (Fig. D.1). 331 Similarly, the differences between the surface and deep layers in C/N ratio, aliphatic ratio and 332 aromatic ratio were reduced in eutric Cambisols compared with the two other soil classes 333 (Fig. 2). If there was no overall effect of depth on the aliphatic ratio of POM, opposite trends

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were found in dystric Cambisol (decrease) and entic Podzol (increase) with depth (depth ×

soil interaction; Fig. 2). For the C/N ratio, it appeared that the soil effect was mainly due to differences in the deep layer (Fig. 2). Finally, for the carbonyl + carboxyl ratio, the depth

effect was not observed in the deciduous plots (Fig. 3).

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- 3.2. Heterogeneity of POM Rock-Eval thermal parameters
- 3.2.1. Effect of vegetation type
- Vegetation type significantly influenced all parameters of POM thermal parameters (Table 1).
- 342 HI and  $T_{50\_CO2\_PYR}$  were higher in POM of soils under deciduous trees (346 ± 88 mg HC·g<sup>-1</sup>
- OC and 358  $\pm$  10 °C, respectively) than coniferous trees (213  $\pm$  59 mg HC·g<sup>-1</sup> OC and 355  $\pm$
- 8 °C, respectively; Table 2), while  $OI_{RE6}$  was significantly lower (166 ± 42 mg  $O_2 \cdot g^{-1}$  TOC
- 345 vs.  $194 \pm 26 \text{ mg O}_2 \cdot \text{g}^{-1} \text{ TOC}$ ; Table 2).
- 3.2.2. Effect of soil class
- 347 HI and  $T_{50\_CO2\_PYR}$  were significantly lower in POM from eutric Cambisols (244  $\pm$  87 mg
- 348  $HC \cdot g^{-1}$  OC and 351  $\pm$  5 °C, respectively) than from dystric Cambisols (305  $\pm$  114 mg  $HC \cdot g^{-1}$
- OC and  $360 \pm 10$  °C) and entic Podzols (Table 2). Conversely, OI<sub>RE6</sub> was higher in POM
- from eutric Cambisols (214  $\pm$  30 mg O<sub>2</sub>·g<sup>-1</sup> TOC) than from entic Podzols and dystric
- Cambisols (171  $\pm$  27 and 158  $\pm$  29 mg O<sub>2</sub>· g<sup>-1</sup> TOC, respectively; Table 2).
- 352 3.2.3. Effect of soil depth
- 353 The effect of depth was significant for HI and T<sub>50\_CO2\_PYR</sub> but not for OI<sub>RE6</sub> (Table 1). It is
- worth noting that the variability of  $OI_{RE6}$  was however greater in the deep layer (sd = 45 mg
- 355  $O_2 \cdot g^{-1}$  TOC vs. sd = 29 mg  $O_2 \cdot g^{-1}$  TOC in the surface layer; Table 2) and overshadowed the
- 356 slight increase with depth.
- 357 HI decreased from  $301 \pm 86$  mg HC·g<sup>-1</sup> OC in the surface layer to  $248 \pm 108$  mg HC·g<sup>-1</sup> OC
- 358 in the deep layer (Table 2).  $T_{50\_CO2\_PYR}$  was higher in the deep layer (363 ± 9 °C) than in the

surface layer (351  $\pm$  5 °C; Table 2), indicating an increase in thermal stability of the POM fraction with depth.

3.2.4. Factor interactions

Depth  $\times$  soil ( $T_{50\_CO2\_PYR}$ ), soil  $\times$  vegetation ( $OI_{RE6}$ ) and depth  $\times$  vegetation (HI) were all significant interactions (Table 1). For  $T_{50\_CO2\_PYR}$ , the soil effect was mainly due to differences in the deep soil layer (Fig. 2). For  $OI_{RE6}$ , the vegetation effect was not significant in eutric Cambisols (Fig. D.1). For HI, the depth effect was not observed in the deciduous plots (Fig. 3).

#### 4. Discussion

4.1. Influence of vegetation type on POM chemistry and thermal stability

Results from the literature were generally consistent with our findings of POM of deciduous origin having more aliphatic and carbonyl/carboxyl and less aromatic C than POM of coniferous origin. Using <sup>13</sup>C CP/MAS NMR, Preston et al. (2000) found significantly more aromatic and phenolic C in foliar litters of coniferous species than deciduous species, supporting other studies that showed that deciduous species tend to have lower concentrations of lignin than coniferous species (Berg et al., 2013). With the same technique, Rumpel et al. (2002) observed a greater proportion of alkyl-C in beech than spruce roots sampled from deep layer (B horizons), and these authors suggested that suberin might be an important contributor of the alkyl-C. As the percentage of fine root biomass in the first 30 cm of soil is higher in deciduous temperate forests than their coniferous counterparts (Jackson et al., 1997), this could at least partially explain our higher MIR-ATR carbonyl + carboxyl ratio in the deciduous POM. Both the aliphatic and carbonyl + carboxyl ratios of deciduous POM had a variability that was much larger than for the coniferous POM and illustrate the fact that inter-

species differences are important when it comes to plant chemistry (Berg and McClaugherty, 2013). Further studies looking at species level are therefore warranted. In our study plots, HI and T<sub>50\_CO2\_PYR</sub> were significantly higher in POM of soils under deciduous trees than coniferous trees (Table 2) while OI<sub>RE6</sub> was significantly lower (Table 2). As coniferous litter is generally considered more difficult to decompose (e.g., Prescott, 2010), we would have expected a higher T<sub>50 CO2 PYR</sub> for the POM in coniferous plots. When looking more closely, the difference in T<sub>50 CO2 PYR</sub> between the two vegetation types originated mainly from the surface layer (Fig. 3). This difference in thermal stability of the POM between the two vegetation types might be related to a higher turnover in deciduous plots (Cole and Rapp, 1981; Quideau et al., 2001) and/or, in coniferous plots, to a protection from decomposition due to complexation of root tissues and litter with Al and Fe (Rasse et al., 2005). Previous study of the thermal stability of total SOC in these forest sites (Soucémarianadin et al., 2018b) showed that total SOC was also more thermally stable in the surface layer of deciduous plots than coniferous plots. Overall, these results only partially validated our first hypothesis, as we were not able to highlight that POM in soils under deciduous trees was in fact more decomposed. The absence of a vegetation effect on the POM C/N ratio could be explained by previous observations in soils under deciduous than in coniferous species. If, for our study species, the C/N ratio of fresh foliage found in the literature appear much lower in deciduous than in coniferous species (Cools et al., 2014), the C/N ratio of organic layers did not significantly differ between the deciduous (30  $\pm$  6) and coniferous plots (31  $\pm$  7) in our study sites (Brêthes et al., 1997). POM in topsoil is more likely to derive from the organic layers than directly from fresh leaves/needles.

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4.2. Influence of soil class on POM chemistry and thermal stability

In our study sites, POM from eutric Cambisols was significantly different from POM from dystric Cambisols and entic Podzols. Using <sup>13</sup>C NMR spectroscopy and elemental analysis, Kölbl and Kögel-Knabner (2004) reported a protection of POM by clay through higher aggregation, leading to a less decomposed POM (higher C/N and O-alkyl and lower aryl C) in a clayey soil compared with a sandy soil. Yeasmin et al. (2017) observed variations in the chemical composition of POM ( $d < 1.8 \text{ g} \cdot \text{cm}^{-3}$ ) among four soil types, using elemental analysis and MIR spectroscopy in the diffuse reflectance mode. Specifically, POM in their clayey soil had higher C/N ratio than in the other soil types and POM in the sandy soil had a large aliphatic C band compared with POM in the clayey soil. These results contradict our own observations as POM in eutric Cambisols (clay content =  $37 \pm 11\%$ ; Soucémarianadin et al. 2018b) had lower C/N ratio than POM from dystric Cambisols and entic Podzols (clay content =  $18 \pm 9\%$  and  $15 \pm 9\%$ , respectively). However, they agree with the lower aliphatic and carbonyl + carboxyl ratios we observed in eutric Cambisols. Yeasmin et al. (2017) also reported only small (non-significant) differences in the aromatic spectral region and no difference for the ether + alcohol band of POM among the four soil types, similar to our results. Finally, comparing soils with clay content varying between 12% and 72%, Golchin et al. (1994) did not either observe a clear trend on POM C/N ratio or chemical composition with soil texture. When combining the differences in POM chemistry we observed in eutric Cambisols compared with dystric Cambisols and entic Podzols, with the differences in thermal parameters (lower HI and higher OI<sub>RE6</sub> in POM from eutric Cambisols), our results suggest that POM in eutric Cambisols is more decomposed than in dystric Cambisols and entic Podzols. This contradicts the lower T<sub>50\_CO2\_PYR</sub> values observed in eutric Cambisols but it appears that the significant difference in thermal stability among soil classes are only present in the deep layer (Fig. 2), while differences in HI were mainly found in the surface layer.

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Overall, our results thus disprove our second hypothesis. One possible explanation could be that as eutric Cambisols have a greater water availability (Brêthes et al., 1997) and more nutrient exchange sites (cation exchange capacity = 24.4 ± 12.3 cmol(+)/kg) as well as a higher pH (Soucémarianadin et al., 2018b), this could promote decomposition. Moreover, there could be a threshold value for clay content to determine whether it promotes or reduces decomposition and clay mineralogy is also likely to affect decomposition (Fissore et al., 2016). Our result call for a more in-depth characterization of the chemical nature of the POM from the different soil classes (maybe using <sup>13</sup>C NMR) to further explain the variations observed among classes. Although as previously mentioned, our design was not overall too unbalanced, it should finally be noted that soil and vegetation appeared as confounded factors in our design. POM from coniferous plots was found preferentially on Podzols (70%), while POM from deciduous plots was preferentially associated with eutric Cambisols (59%). These unbalances could have affected our results and prevented us from properly discriminating the effects of vegetation *vs.* soil on POM characteristics in our study sites.

4.3. Contrasting POM chemistry and thermal stability at various soil depths suggests different C input sources

The increase in POM C/N ratio with depth was consistent with previous observations in forest ecosystems (Spycher et al., 1983; Schrumpf et al., 2013). The ether + alcohol and carbonyl + carboxyl ratios are considered as representative of labile C compounds (Baldock et al., 1992; Sarkhot et al., 2007; Ng et al., 2014) and their observed decrease with depth could correspond to the decomposition of POM from the surface to the deep layer (Baldock et al., 1997). Similarly, with depth, HI decreased while T<sub>50\_CO2\_PYR</sub> increased: these opposite trends match what is generally observed for bulk soil OC and organic material (Disnar et al., 2003; Sebag et al., 2016; Soucémarianadin et al., 2018a). These evolutions have been linked to an

increased decomposition and a decrease in labile C compounds, resulting in a more thermally and biogeochemically stable OC (Sebag et al., 2006; Albrecht et al., 2015; Barré et al., 2016; Cécillon et al., 2018). Our results for the thermal parameters could thus suggest a more decomposed POM in the deep layers. However, if POM in the deep layers were more decomposed we would have expected a decrease of the C/N ratio. The fact that the latter was significantly higher in the deep layers would hint at the fact that the differences in chemistry between POM from the surface and deep layers originated from variations in POM sources. The high C/N ratios observed in the deep layers matched a higher contribution of roots compared to leaf litter in the deep layers. C/N ratios of fine roots are closer to 50–60 (Hobbie et al., 2010) and even higher (Angst et al., 2016), while C/N ratios of fresh foliage or humus layers range between 15 and 45 (Brêthes et al., 1997; Disnar et al., 2003; Cools et al., 2014). Roots tend to have higher lignin content than leaves, which could explain the increase in the aromatic ratio (Natelhoffer and Fry, 1988; Berg and McClaugherty, 2013). This difference in main sources of POM could also explain the decrease of HI with depth. There are variations of HI values among plant organs, with "woody" materials tending to have lower HI. For instance, high HI values have been observed for needles or OL horizons of deciduous forest floors (≈ 300; Disnar et al., 2003; Sebag et al., 2006; Carrie et al., 2012), while bark HI was only 140 mg HC·g<sup>-1</sup> TOC (Carrie et al., 2012). The observed increase of T<sub>50 CO2 PYR</sub> in the POM fraction of the deep layer could also be related to the fact that the deeper POM comes from a different source that is more stable (i.e., woody roots vs. leaves and needles litter). This could also explain the increase in the aromatic ratio (Kögel-Knabner, 2002). These results thus confirmed our third hypothesis. The lack of correlation between HI and T<sub>50</sub> CO<sub>2</sub> PYR (Table E.1) supports this hypothesis of different POM inputs between the two layers rather than just a decomposition gradient. Soil class and

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vegetation type were also important factors, confirming combined effects of input quality and decomposition for POM chemistry and thermal stability.

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4.4. Some POM are more labile than others

The strong heterogeneity we observed in the POM samples of our 52 forest sites suggest that POM lability varies. All the chemical (higher C/N ratio and aromatic ratio, lower ether + alcohol and carbonyl + carboxyl ratios) and thermal (higher T<sub>50</sub> CO<sub>2</sub> PYR and lower HI) properties of deep layer POM point towards a more labile POM in the topsoil than subsoil. Aryl C is considered as relatively stable carbon forms (Ng et al., 2014), while carbonyl C is considered as more labile C (Sarkhot et al., 2007; Ng et al., 2014). The moderate to strong positive correlation between C/N ratio and T<sub>50 CO2 PYR</sub> (Table E.1) suggested that POM with higher C/N ratios were more thermally (and biogeochemically) stable. This could be linked to previous reports showing that litters with low C/N ratio are more easily decomposed (Cotrufo et al., 1995), thus connecting thermal and chemical indicators of biogeochemical stability. Free POM (light fraction) turnover has been shown to increase by an order of magnitude from the 0–5 cm layer (turnover < 10 years) to the 10–20 cm layer (turnover  $\approx 100$  years) (Schrumpf and Kaiser, 2015). In the mineral soil, an increased SOC decomposition and turnover (Balesdent et al., 2018; Cécillon et al., 2018) with depth, time since bare-fallow or increase in soil temperature has been linked to a decrease in HI (Barré et al., 2016; Sebag et al., 2016; Soucémarianadin et al., 2018b; Poeplau et al., 2019). Similarly to SOC, it would be expected that POM with lower HI is less energy-rich and would require more energy to further break down (Barré et al., 2016), resulting in a greater thermal and biogeochemical stability. However, our results suggest that differences in lability between topsoil and subsoil POM are less important in eutric Cambisols (a similar pattern was observed for total SOC;

Soucémarianadin et al. 2018b) and under deciduous vegetation. The chemical (higher aliphatic and carbonyl + carboxyl ratios; lower aromatic ratio) and thermal (higher T<sub>50 CO2 PYR</sub> and HI; lower OI<sub>RE6</sub>) characteristics of the POM from soils in deciduous dominant plots suggest that it is less labile than the POM fraction from soils in coniferous plots, despite a lower lignin content. Our results also suggest that POM of deciduous and coniferous plots are not as different in eutric Cambisols. Compared with the POM fraction in dystric Cambisols or entic Podzols, POM in eutric Cambisols presented chemical (lower C/N ratio and aliphatic and carbonyl + carboxyl ratios) and thermal (lower T<sub>50 CO2 PYR</sub> and HI; higher OI<sub>RE6</sub>) properties suggesting that it was more decomposed, yet less thermally stable. These differences were mostly observed in the deep layer. It could also be that the proportion of pyrogenic carbon in the deep layers of eutric Cambisols is less important than in the two other soil classes: as more pyrogenic carbon gets potentially stabilized in organo-mineral interactions stimulated by the finer texture of the eutric Cambisols, this results in a reduced vertical transfer in the soil profile for the latter soil class. POM is considered as part of labile SOC (Wander, 2004) and HI, OIRE6 and T50\_CO2\_PYR values of the POM fraction, when compared with values obtained for the bulk soil or the mineral-associated OM fraction (Saenger et al., 2015; Soucémarianadin et al., 2018b), reflected a lower thermal stability of the POM fraction. This lower thermal stability can be linked to the shorter persistence of the fraction of total SOC present as POM in the ecosystem. For instance, Baisden et al. (2002) showed that at least 90% of the free POM from topsoils of Californian grasslands (with secondary oaks) was turning over in less than 10 years. In a German beech forest, Schrumpf and Kaiser (2015) found a similar turnover for the free POM (light fraction) in the 0–5 cm layer. The potential contribution of recalcitrant pyrogenic carbon to the POM fraction should also be taken into consideration as an additional source of POM heterogeneity. The presence of this

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residue of the incomplete combustion of organic matter would create a strong heterogeneity in the chemical composition of the POM fraction. As pyrogenic carbon has a greater intrinsic recalcitrance due to higher condensation, which makes it less prone to decomposition (Schmidt et al., 2011), its presence could also affect POM thermal stability and overall dynamics. Hence, POM containing pyrogenic carbon would present lower OI<sub>RE6</sub> and HI values and increased thermal stability (*e.g.*, higher T<sub>50\_CO2\_PYR</sub>) as shown by Poot et al. (2009) for various pyrogenic carbon samples.

Taken together, our results suggest that caution should be applied when using POM quantity as an indicator of SOC lability as the quality of the POM, hence its lability, will be strongly dependent on the ecosystem (soil type, vegetation) and depth considered.

#### 5. Conclusions

The variable thermal stability of POM illustrates its heterogeneity as a labile SOC component and suggests variations in turnover of C in POM. Our results also suggested that various sources of POM inputs exist within a soil profile. Although the C in POM only corresponds to 10 to 20% of total SOC in our French forest soils, it is likely to significantly influence SOC persistence overall. While confirming that POM is part of the more labile SOC, our results showed that, in French temperate forests, depth, soil class and vegetation type were all important factors to explain the heterogeneity of POM chemistry and thermal stability. This suggests contrasting persistence of POM in soils as a function of these three factors.

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## Figures captions

Fig. 1. (a) Location of the 52 study sites from the French national network for the long term monitoring of forest ecosystems (RENECOFOR) and their repartition among the vegetation types and soil classes; (b) Number of samples by depths and analyses performed to

characterize the POM.

**Fig. 2.** Interactions depth  $\times$  soil class for C/N ratio,  $T_{50\_CO2\_PYR}$  and MIR-ATR derived aliphatic ratio and aromatic ratio. The horizontal black line shows the median. The bottom and top of the box show the first and third quartiles, respectively. Different letters indicate significant differences among the three soil classes in topsoil or subsoil for each parameter.

**Fig. 3.** Interactions depth  $\times$  vegetation type for the MIR-ATR derived carbonyl + carboxyl ratio and the RE6 Hydrogen Index (corrected) and  $T_{50\_CO2\_PYR}$ . The horizontal black line shows the median. The bottom and top of the box show the first and third quartiles, respectively. Different letters indicate significant differences between vegetation types in topsoil or subsoil for each parameter.

## **Tables**

**Table 1.** Details of models and their significant terms selected to explain variations in parameters from Rock-Eval 6 (HI corrected,  $OI_{RE6}$ , and  $T_{50\_CO2\_PYR}$ ) and elemental analysis (C/N ratio) and ratios derived from MIR-ATR spectroscopy (aliphatic, ether and alcohol, aromatic, carbonyl and carboxyl) for the POM fraction of the 52 study plots. All models used a gls function (see details in the *Calculations and statistical analyses* section § 2.6). Significance is indicated as follows: \*\*\*: p < 0.001; \*\*: p < 0.01; \*: p < 0.05. The number of samples considered in each model is provided; outliers were determined with Cook's distance.

Response variable	Transformation	Predictors in final model <sup>a</sup> and level of significance	n
HI corrected (HI)	sqrt (HI)	$depth^{***} + soil^{***} + veg^{***} + depth \times veg^{***}$	86
$\mathrm{OI}_{\mathrm{RE6}}$	$log_{10} (OI_{RE6})$	$soil^{***} + veg^{***} + soil \times veg^{**}$	86
$T_{50\_CO2\_PYR}$		$depth^{***} + soil^{***} + veg^{***} + depth \times soil^{***}$	84
C/N ratio	$log_{10}$ (C/N)	$depth^{***} + soil^{***} + depth \times soil^{***}$	85
aliphatic ratio (ali)	1/ali	soil*** + veg*** + depth × soil**	84
aromatic ratio (arom)	1/arom	depth*** + veg*** + depth × soil** + soil × veg***	84
ether + alcohol ratio (ether)	1/ether	depth***	84
carbonyl + carboxyl ratio (carbo)	sqrt (carbo)	depth*** + soil*** + veg*** + depth × veg** + soil × veg**	85

<sup>&</sup>lt;sup>a</sup> For all models, we used the compound symmetry structure [corCompSymm(form =~ 1|plot)], which is similar to the variance structure of a random-intercept-only model. In our case, it allowed to treat each site as random factor.

**Table 2.** Mean, standard deviation (sd), minimum (min) and maximum (max) values of the parameters derived from elemental analysis and Rock-Eval thermal analysis as well as the ratios of C functions derived from MIR-ATR spectroscopy for the POM fraction for each depth, soil class and vegetation type. For each parameter, different letters indicate significant differences among means of the classes of the three factors (depth, soil class and vegetation type). Differences were observed for transformed data as specified in Table 1.

 $\mathbf{OI}_{\mathbf{RE6}} (\mathbf{mg} \ \mathbf{O}_2 \cdot \mathbf{g}^{-1} \ \mathbf{TOC})$ 

 $T_{50\_CO2\_PYR}\,(^{\circ}C)$ 

**HI corrected** (mg HC·g<sup>-1</sup> OC)

C/N ratio

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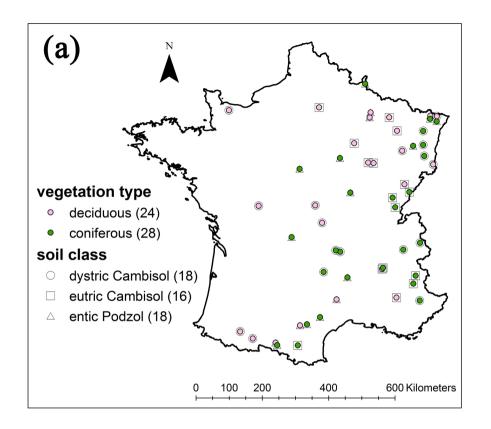
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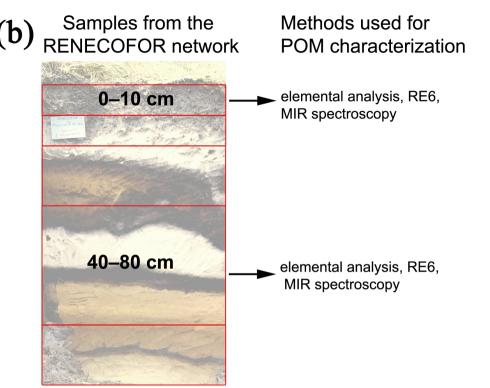
		0/11	- 444			(11)	0 0	00)	V = ,	TEO (1112	28 -0	<b>U</b> )		- 30_CO2_1	IK ( )	
	mean	sd	min	max	mean	sd	min	max	mean	sd	min	max	mean	sd	min	max
DEPTH																_
surface	30 <sup>a</sup>	8	20	71	301 <sup>a</sup>	86	187	504	177 <sup>a</sup>	29	107	247	351 <sup>a</sup>	5	341	363
deep	$48^{b}$	11	28	78	$248^{b}$	108	106	493	186ª	45	106	302	363 <sup>b</sup>	9	348	388
SOIL																
dystric Cambisol	42a	14	22	78	$305^{a}$	114	107	504	158a	29	106	222	$360^{a}$	10	346	388
eutric Cambisol	34 <sup>b</sup>	7	22	50	$244^{b}$	87	106	454	$214^{b}$	30	154	302	351 <sup>b</sup>	5	341	363
entic Podzol	$40^{ab}$	15	20	71	$280^{ab}$	89	132	436	171 <sup>a</sup>	27	125	221	$358^{a}$	9	345	375
<b>VEGETATION</b>																
coniferous	$40^{a}$	15	20	78	213a	59	106	332	194ª	26	134	259	355a	8	341	375
deciduous	37 <sup>a</sup>	11	22	71	$346^{b}$	88	186	504	166 <sup>b</sup>	42	106	302	358 <sup>b</sup>	10	345	388
ALL	39	12	22	71	277	90	106	461	181	33	120	265	356	8	344	377
	aliphatic ratio carbonyl + carboxyl ratio		atio		aromati	c ratio		eth	er + alc	ohol rat	io					
	mean	sd	min	max	mean	sd	min	max	mean	sd	min	max	mean	sd	min	max
DEPTH																
surface	$0.186^{a}$	0.014	0.160	0.223	$0.221^{a}$	0.025	0.164	0.278	$0.236^{a}$	0.038	0.172	0.329	$0.357^{a}$	0.019	0.330	0.412
deep	$0.186^{a}$	0.014	0.158	0.213	$0.204^{b}$	0.026	0.155	0.253	$0.274^{b}$	0.038	0.204	0.340	$0.336^{b}$	0.017	0.309	0.394
SOIL																
dystric Cambisol	$0.192^{a}$	0.016	0.161	0.223	$0.226^{a}$	0.023	0.176	0.278	$0.236^{a}$	0.037	0.172	0.305	$0.347^{a}$	0.017	0.314	0.386
eutric Cambisol	$0.178^{b}$	0.010	0.158	0.201	$0.190^{b}$	0.020	0.155	0.247	$0.285^{a}$	0.040	0.172	0.340	$0.348^{a}$	0.024	0.314	0.412
entic Podzol	$0.188^{a}$	0.011	0.170	0.212	$0.224^{a}$	0.020	0.177	0.255	$0.241^{a}$	0.031	0.191	0.300	$0.347^{a}$	0.022	0.309	0.412
<b>VEGETATION</b>																
coniferous	$0.181^{a}$	0.009	0.158	0.203	$0.207^{a}$	0.021	0.155	0.250	$0.268^{a}$	0.030	0.210	0.340	$0.344^{a}$	0.015	0.309	0.373
deciduous	$0.191^{b}$	0.016	0.160	0.223	$0.220^{b}$	0.031	0.162	0.278	$0.239^{b}$	0.048	0.172	0.330	$0.350^{a}$	0.025	0.314	0.412
ALL	0.186	0.013	0.161	0.214	0.213	0.024	0.163	0.262	0.254	0.038	0.185	0.326	0.347	0.020	0.314	0.400

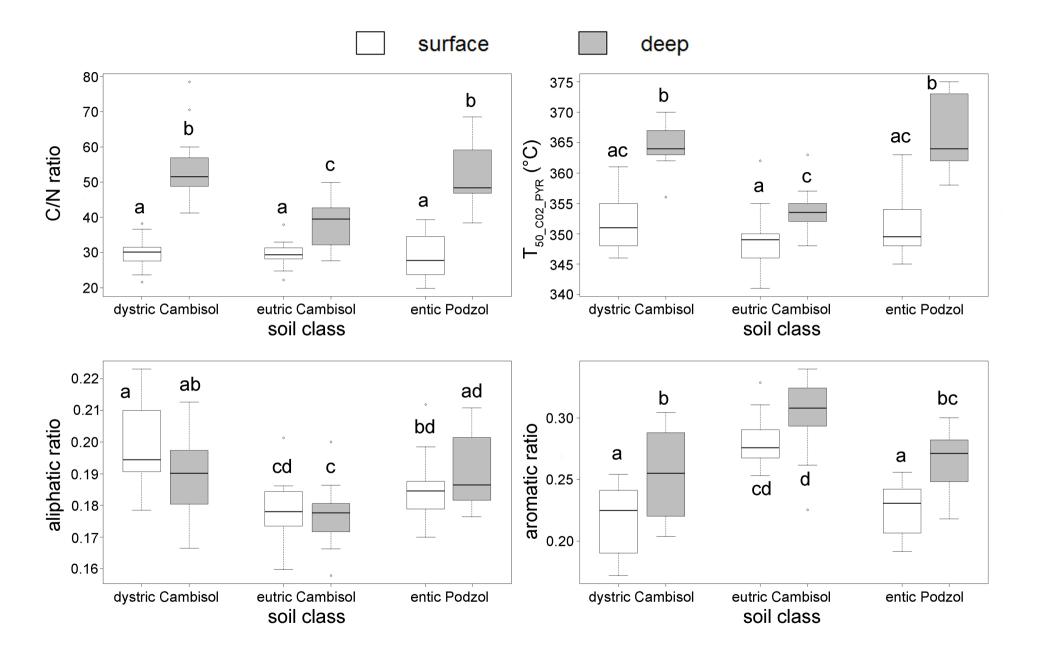
**Supplementary materials** 849 850 851 APPENDIX A – information related to the analysis of POM chemistry and thermal 852 stability 853 854 Fig. A.1. Absorbance spectra (with offset-correction) of the 86 POM samples considered in 855 this study. The four spectral regions of interest used for the calculation of the relative aliphatic  $(2950-2900 \text{ cm}^{-1})$ , carbonyl + carboxyl  $(1750-1690 \text{ cm}^{-1})$ , aromatic  $(1605-1575 \text{ cm}^{-1})$  and 856 ether + alcohol (1180–1145 cm<sup>-1</sup>) ratios are indicated. 857 858 859 Fig. A.2. Description of the Rock-Eval 6 thermal analysis (adapted from Baudin et al. 2017) 860 and calculation of the three RE6-derived parameters (Hydrogen Index; Oxygen Index and 861 T<sub>50\_CO2\_PYR</sub>, the temperature at which 50% of the SOM converted to CO<sub>2</sub> had evolved during 862 the pyrolysis phase). 863 Reference: Baudin, F., Tribovillard, N. and Trichet, J., 2017. Géologie De La Matière 864 Organique. 324 pp. EDP Sciences, Lille, France. 865 866 APPENDIX B – chemical and thermal characteristics of the 86 selected samples 867 868 **Table B.1**. Values of the parameters derived from elemental analysis and Rock-Eval thermal 869 analysis as well as the ratios of C functions derived from MIR-ATR spectroscopy for the 870 POM fraction for each of the 86 POM samples considered in this study. For comparison, HI 871 values without and with correction for the inorganic carbon are included. 872

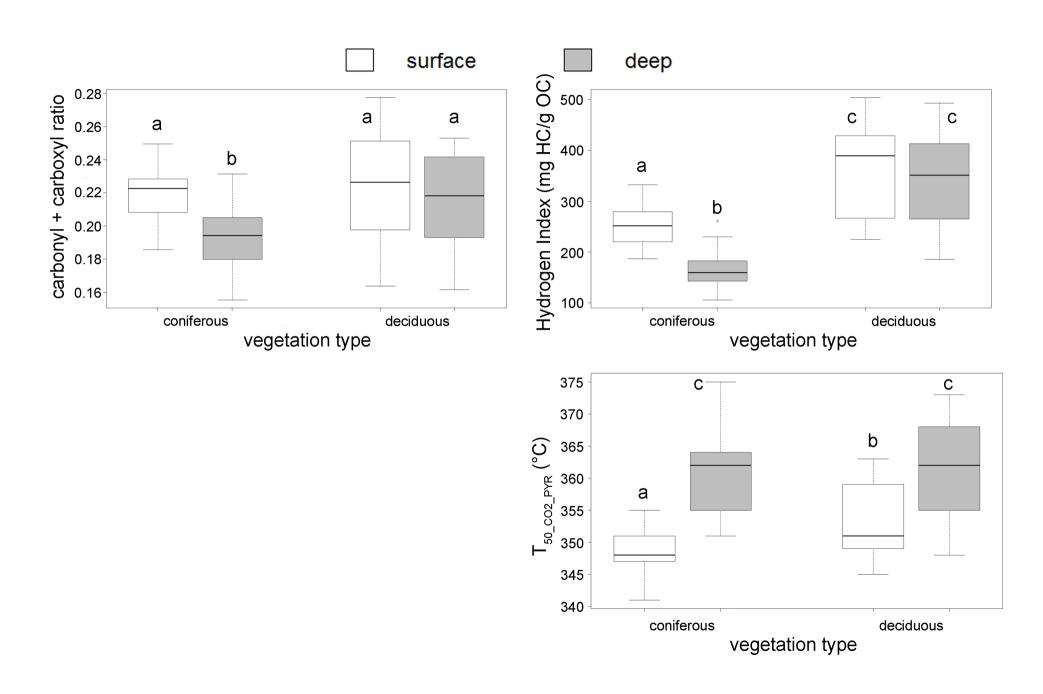
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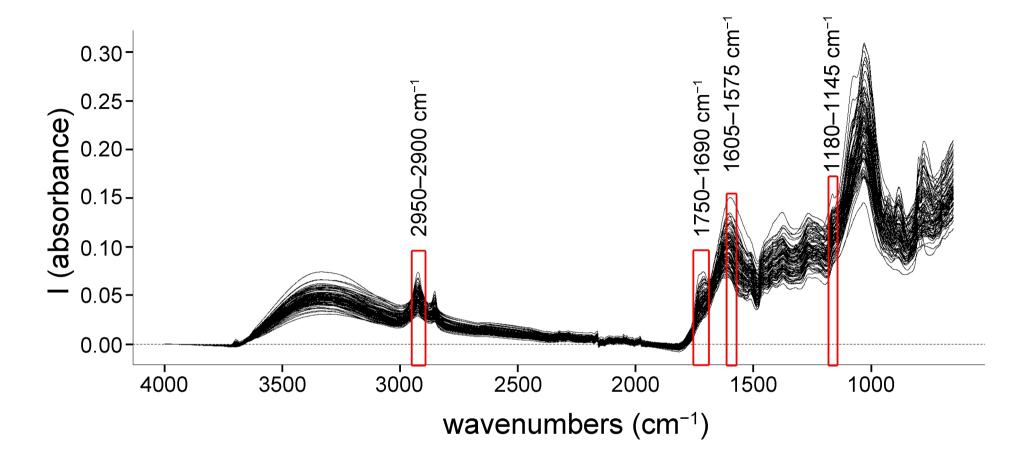
874	$ \label{eq:APPENDIX} \textbf{C}-\textbf{information related to the distribution of the 86 POM samples among} $
875	the different classes of environmental factors (depth; soil class; vegetation type)
876	
877	<b>Table C.1</b> . Distribution of the 86 POM samples considered in this study among the various
878	classes of depth (surface and deep layers; 0–10 cm and 40–80 cm), soil (dystric Cambisol,
879	eutric Cambisol, entic Podzol) and vegetation (coniferous and deciduous).
880	
881	
882	APPENDIX D – effects of interactions of the environmental factors on POM chemistry
883	and thermal parameters
884	
885	<b>Fig. D.1.</b> Interactions soil class $\times$ vegetation type for MIR-ATR derived aromatic ratio and
886	$carbonyl + carboxyl \ ratio \ and \ the \ OI_{RE6}. \ Different \ letters \ indicate \ significant \ differences$
887	among the three soil classes in coniferous or deciduous plots for each parameter.
888	
889	
890	APPENDIX E – correlations between parameters of POM chemistry and thermal
891	stability
892	
893	Table E.1. Table of correlations for all samples and for each layer individually between the
894	POM chemical properties (C/N ratio, ether + alcohol ratio, aromatic ratio, carbonyl +
895	carboxyl ratio, aliphatic ratio) and POM thermal stability (HI corrected, $OI_{RE6}$ , $T_{50\_CO2\_PYR}$ ).
896	Significance is indicated as follows: ***: $p < 0.001$ ; **: $p < 0.01$ ; *: $p < 0.05$ . The high (>
897	0.6) correlations between parameters derived from different methods are marked in bold. n =
898	86 for all layers; $n = 46$ for surface layer and $n = 40$ for deep layer, respectively.

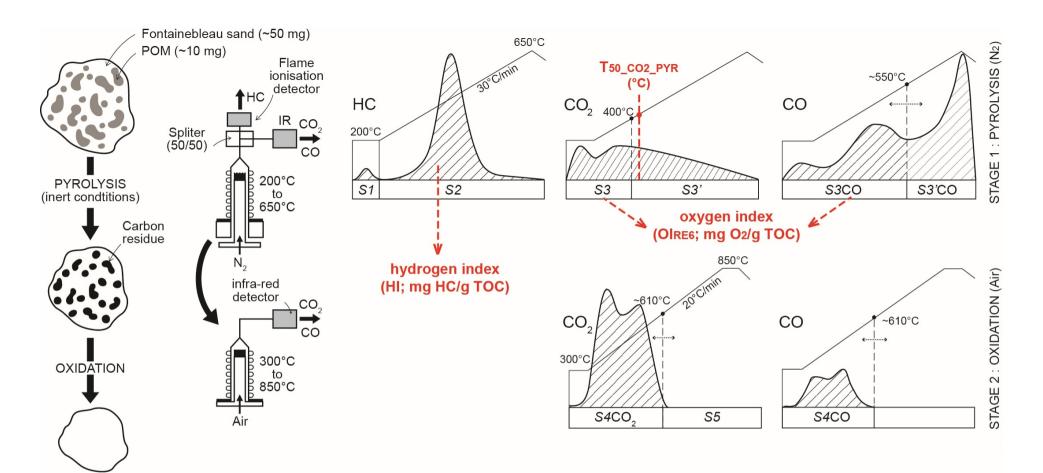












**Table B.1.** Values of the parameters derived from elemental analysis and Rock-Eval thermal analysis as well as the ratios of C functions derived from MIR-ATR spectroscopy for the POM fraction for each of the 86 POM samples considered in this study. For comparison, HI values without and with correction for the inorganic carbon are included.

samples	layer	vegetation type	soil class	C/N ratio	HI mg HC·g <sup>-1</sup> TOC)	HI corrected (mg HC·g <sup>-1</sup> OC)	$\begin{array}{c} OI_{RE6}(mg\\ O_2 \cdot g^{-1}\ TOC) \end{array}$	T <sub>50_CO2_PYR</sub> (°C)	ether + alcohol ratio (1180–1145 cm <sup>-1</sup> )	aromatic ratio (1605–1575 cm <sup>-1</sup> )	carbonyl + carboxyl ratio (1690– 1750 cm <sup>-1</sup> )	aliphatic ratio (2950–2900 cm <sup>-1</sup> )
CHP10_1	surface	deciduous	eutric Cambisol	25	467	454	177	347	0.412	0.172	0.232	0.184
CHP10_4	deep	deciduous	eutric Cambisol	31	433	433	302	363	0.393	0.225	0.197	0.184
CHP55_1	surface	deciduous	eutric Cambisol	32	450	436	166	349	0.366	0.186	0.247	0.201
CHP55_4	deep	deciduous	eutric Cambisol	28	416	399	187	354	0.314	0.262	0.225	0.200
CHP65_1	surface	deciduous	dystric Cambisol	31	417	404	169	349	0.381	0.199	0.226	0.194
CHP65_4	deep	deciduous	dystric Cambisol	71	345	330	106	388	0.353	0.217	0.241	0.189
CHS03_1	surface	deciduous	dystric Cambisol	29	471	446	134	350	0.358	0.181	0.251	0.210
CHS03_4	deep	deciduous	dystric Cambisol	51	506	493	136	362	0.362	0.208	0.218	0.213
CHS51_1	surface	deciduous	entic Podzol	28	401	363	148	349	0.412	0.192	0.217	0.179
CHS57b_1	surface	deciduous	entic Podzol	24	398	389	154	348	0.383	0.194	0.225	0.198
CHS68_1	surface	deciduous	dystric Cambisol	26	440	429	158	351	0.362	0.206	0.237	0.195
CHS86_1	surface	deciduous	dystric Cambisol	31	259	251	138	360	0.386	0.230	0.222	0.161
CHS86_4	deep	deciduous	dystric Cambisol	48	364	351	135	368	0.337	0.255	0.213	0.195
CPS67_1	surface	deciduous	dystric Cambisol	38	431	421	107	361	0.344	0.177	0.256	0.223
CPS67_4	deep	deciduous	dystric Cambisol	52	374	360	139	370	0.332	0.224	0.246	0.198
EPC08_1	surface	coniferous	entic Podzol	23	270	260	193	351	0.348	0.253	0.228	0.170
EPC34_1	surface	coniferous	entic Podzol	20	300	291	188	349	0.345	0.237	0.233	0.185
EPC34_4	deep	coniferous	entic Podzol	38	150	143	181	362	0.336	0.300	0.177	0.186
EPC39a_1	surface	coniferous	eutric Cambisol	30	237	228	209	346	0.367	0.270	0.189	0.174
EPC71_1	surface	coniferous	entic Podzol	32	281	272	177	351	0.353	0.242	0.225	0.180

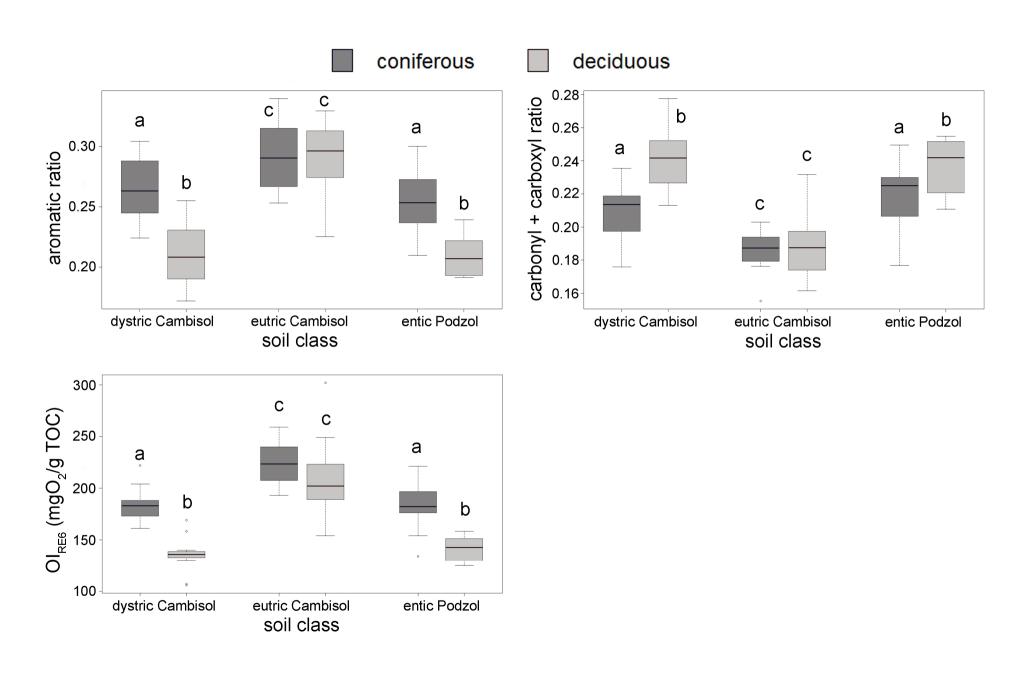
EPC73_1	surface	coniferous	dystric Cambisol	31	316	305	204	347	0.349	0.238	0.219	0.194
EPC73_4	deep	coniferous	dystric Cambisol	53	170	164	165	363	0.343	0.288	0.197	0.171
EPC81_1	surface	coniferous	entic Podzol	23	250	241	182	355	0.340	0.250	0.226	0.184
EPC81_4	deep	coniferous	entic Podzol	46	138	132	203	364	0.338	0.279	0.207	0.176
EPC87_1	surface	coniferous	entic Podzol	23	197	190	197	350	0.344	0.256	0.222	0.179
EPC88_1	surface	coniferous	dystric Cambisol	22	326	315	185	347	0.373	0.225	0.224	0.179
EPC88_4	deep	coniferous	dystric Cambisol	41	149	142	222	356	0.337	0.304	0.185	0.173
HET03_1	surface	deciduous	dystric Cambisol	35	523	504	137	357	0.334	0.172	0.278	0.217
HET03_4	deep	deciduous	dystric Cambisol	47	441	427	140	362	0.335	0.203	0.253	0.208
HET04_1	surface	deciduous	eutric Cambisol	29	257	242	215	345	0.384	0.280	0.176	0.160
HET04_4	deep	deciduous	eutric Cambisol	28	245	236	200	357	0.344	0.315	0.162	0.179
HET09_4	deep	deciduous	entic Podzol	47	402	390	133	369	0.323	0.226	0.248	0.203
HET14_1	surface	deciduous	dystric Cambisol	29	302	283	133	361	0.376	0.231	0.227	0.166
HET14_4	deep	deciduous	dystric Cambisol	49	366	354	138	369	0.333	0.235	0.236	0.197
HET21_1	surface	deciduous	eutric Cambisol	28	242	225	206	355	0.333	0.329	0.164	0.174
HET21_4	deep	deciduous	eutric Cambisol	40	259	248	224	356	0.317	0.330	0.175	0.179
HET25_1	surface	deciduous	eutric Cambisol	33	245	231	154	362	0.349	0.273	0.205	0.173
HET25_4	deep	deciduous	eutric Cambisol	43	233	223	197	353	0.327	0.310	0.184	0.179
HET26_1	surface	deciduous	eutric Cambisol	25	244	232	223	349	0.345	0.311	0.173	0.171
HET26_4	deep	deciduous	eutric Cambisol	32	198	186	249	348	0.331	0.324	0.168	0.177
HET30_1	surface	deciduous	entic Podzol	28	427	414	158	354	0.330	0.203	0.255	0.212
HET30_4	deep	deciduous	entic Podzol	47	314	302	145	373	0.357	0.239	0.211	0.193
HET52_1	surface	deciduous	eutric Cambisol	31	279	269	189	349	0.358	0.268	0.189	0.186
HET52_4	deep	deciduous	eutric Cambisol	43	284	269	229	353	0.318	0.299	0.197	0.186
HET54b_1	surface	deciduous	eutric Cambisol	30	277	267	192	351	0.340	0.276	0.198	0.186
HET55_1	surface	deciduous	entic Podzol	38	441	430	125	363	0.378	0.191	0.242	0.188
HET60_1	surface	deciduous	eutric Cambisol	22	300	288	212	346	0.338	0.296	0.186	0.180
HET60_4	deep	deciduous	eutric Cambisol	39	295	277	202	355	0.351	0.293	0.189	0.166
HET65_4	deep	deciduous	dystric Cambisol	60	277	261	132	383	0.314	0.249	0.242	0.194

HET81_1	surface	deciduous	entic Podzol	30	401	390	140	359	0.347	0.211	0.255	0.187
HET81_4	deep	deciduous	entic Podzol	50	460	436	127	364	0.330	0.218	0.241	0.211
HET88_1	surface	deciduous	dystric Cambisol	37	434	423	134	352	0.349	0.182	0.259	0.211
HET88_4	deep	deciduous	dystric Cambisol	50	449	432	130	367	0.328	0.209	0.251	0.212
MEL05_1	surface	coniferous	eutric Cambisol	29	204	196	247	341	0.366	0.253	0.203	0.178
MEL05_4	deep	coniferous	eutric Cambisol	35	166	156	212	353	0.347	0.328	0.155	0.170
PS04_4	deep	coniferous	dystric Cambisol	78	112	107	161	363	0.354	0.288	0.176	0.182
PS15_1	surface	coniferous	dystric Cambisol	28	239	231	180	353	0.339	0.254	0.219	0.188
PS41_1	surface	coniferous	entic Podzol	39	261	253	157	354	0.355	0.242	0.228	0.175
PS41_4	deep	coniferous	entic Podzol	69	206	193	221	360	0.343	0.274	0.200	0.183
PS63_1	surface	coniferous	dystric Cambisol	32	252	243	184	349	0.342	0.248	0.218	0.191
PS63_4	deep	coniferous	dystric Cambisol	55	173	173	173	363	0.348	0.279	0.194	0.178
PS67a_1	surface	coniferous	entic Podzol	26	292	283	177	348	0.360	0.230	0.234	0.176
PS67a_4	deep	coniferous	entic Podzol	53	161	153	154	375	0.331	0.298	0.188	0.182
PS67b_1	surface	coniferous	entic Podzol	71	284	276	162	348	0.360	0.231	0.227	0.182
PS88_1	surface	coniferous	entic Podzol	37	285	276	182	348	0.354	0.236	0.224	0.186
PS88_4	deep	coniferous	entic Podzol	67	223	214	134	374	0.323	0.264	0.210	0.203
PS89_1	surface	coniferous	entic Podzol	37	283	274	192	348	0.355	0.210	0.250	0.185
PS89_4	deep	coniferous	entic Podzol	66	188	178	212	362	0.370	0.257	0.194	0.179
SP05_4	deep	coniferous	eutric Cambisol	50	111	106	231	355	0.323	0.340	0.179	0.158
SP07_1	surface	coniferous	entic Podzol	26	343	332	201	345	0.338	0.225	0.240	0.197
SP07_4	deep	coniferous	entic Podzol	48	241	230	175	373	0.309	0.286	0.206	0.200
SP09_4	deep	coniferous	entic Podzol	47	285	261	196	358	0.316	0.271	0.231	0.181
SP11_1	surface	coniferous	eutric Cambisol	38	223	215	216	344	0.370	0.264	0.193	0.172
SP11_4	deep	coniferous	eutric Cambisol	42	166	157	259	351	0.327	0.325	0.176	0.172
SP25_1	surface	coniferous	eutric Cambisol	33	207	200	193	347	0.362	0.260	0.194	0.184
SP25_4	deep	coniferous	eutric Cambisol	43	170	160	251	354	0.324	0.306	0.196	0.174
SP26_1	surface	coniferous	eutric Cambisol	29	209	202	199	349	0.343	0.286	0.186	0.185
SP26_4	deep	coniferous	eutric Cambisol	42	197	183	233	352	0.356	0.291	0.180	0.174

SP38_1	surface	coniferous	dystric Cambisol	27	234	226	171	353	0.346	0.247	0.217	0.191
SP38_4	deep	coniferous	dystric Cambisol	57	166	151	177	366	0.323	0.291	0.201	0.184
SP39_1	surface	coniferous	eutric Cambisol	28	194	187	206	351	0.346	0.290	0.186	0.177
SP39_4	deep	coniferous	eutric Cambisol	36	181	163	233	352	0.332	0.296	0.191	0.181
SP57_4	deep	coniferous	dystric Cambisol	47	233	221	190	364	0.324	0.272	0.214	0.190
SP63_1	surface	coniferous	dystric Cambisol	24	334	324	182	346	0.341	0.224	0.235	0.199
SP68_1	surface	coniferous	dystric Cambisol	30	258	251	187	347	0.356	0.245	0.214	0.186
SP68_4	deep	coniferous	dystric Cambisol	57	139	133	188	367	0.338	0.291	0.205	0.166

**Table C.1**. Distribution of the 86 POM samples considered in this study among the various classes of depth (surface and deep layers; 0–10 cm and 40–80 cm), soil (dystric Cambisol, eutric Cambisol, entic Podzol) and vegetation (coniferous and deciduous).

	0–10 cm	40–80 cm	coniferous	deciduous
dystric Cambisol	15	15	14	16
eutric Cambisol	15	14	12	17
entic Podzol	16	11	19	8
total			45	41
coniferous	24	21		
deciduous	22	19		
total	46	40		



**Table E.1.** Table of correlations for all samples and for each layer individually between the POM chemical properties (C/N ratio, ether + alcohol ratio, aromatic ratio, carbonyl + carboxyl ratio, aliphatic ratio) and POM thermal stability (HI corrected,  $OI_{RE6}$ ,  $T_{50\_CO2\_PYR}$ ). Significance is indicated as follows: \*\*\*: p < 0.001; \*\*: p < 0.01; \*: p < 0.05. The high (> 0.6) correlations between parameters derived from different methods are marked in bold. n = 86 for all layers; n = 46 for surface layer and n = 40 for deep layer, respectively.

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ALL LAYERS	C/N ratio	ether + alcohol ratio	aromatic ratio	carbonyl + carboxyl ratio	aliphatic ratio	HI corrected	$\mathrm{OI}_{\mathrm{RE6}}$
ether + alcohol ratio	-0.35***						
aromatic ratio	0.19	-0.44***					
carbonyl + carboxyl ratio	-0.07	0.08	-0.88***				
aliphatic ratio	0.08	-0.21	-0.63***	0.64***			
HI corrected	-0.25*	0.25*	-0.83***	0.75***	0.65***		
$OI_{RE6}$	-0.21	-0.07	0.67***	-0.70***	-0.59***	-0.56***	
$T_{50\_CO2\_PYR}$	0.68***	-0.45***	0.07	0.09	0.20	-0.05	-0.46***
SURFACE LAYER							
	C/N ratio	ether + alcohol ratio	aromatic ratio	carbonyl + carboxyl ratio	aliphatic ratio	HI corrected	$\mathrm{OI}_{\mathrm{RE6}}$
ether + alcohol ratio	0.19						
aromatic ratio	-0.15	-0.29					
carbonyl + carboxyl ratio	0.13	-0.03	-0.87***				
aliphatic ratio	0.12	-0.31*	-0.62***	0.60***			
HI corrected	0.03	0.15	-0.87***	0.78***	0.62***		
$OI_{RE6}$	-0.28	-0.20	0.75***	-0.69***	-0.42**	-0.65***	
$T_{50\_CO2\_PYR}$	0.17	-0.25	-0.16	0.31*	0.10	0.14	-0.65***
DEEP LAYER							
	C/N ratio	ether + alcohol ratio	aromatic ratio	carbonyl + carboxyl ratio	aliphatic ratio	HI corrected	$\mathrm{OI}_{\mathrm{RE6}}$
ether + alcohol ratio	0.07						
aromatic ratio	-0.43**	-0.17					
carbonyl + carboxyl ratio	0.41**	-0.22	-0.87***				
aliphatic ratio	0.25	-0.24	-0.77***	0.74***			
HI corrected	-0.07	-0.05	-0.71***	0.65***	0.71***		
$OI_{RE6}$	-0.59***	0.06	0.70***	-0.70***	-0.71***	-0.46**	
$T_{50\_CO2\_PYR}$	0.65***	-0.02	-0.62***	0.63***	0.55***	0.24	-0.79***