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Species are hypotheses: avoid basing connectivity assessments on pillars of sand.

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1 Species are hypotheses: avoid basing connectivity assessments on pillars of sand

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38 **Abstract.**

39

40 Connectivity among populations determines the dynamics and evolution of marine populations, and its
41 assessment is essential in ecology in general and in conservation biology in particular. The robust basis of
42 any ecological study is the accurate delimitation of evolutionary units, such as populations, meta-
43 populations and species. Yet a disconnect still persists between the work of taxonomists describing species
44 as working hypotheses to be tested notably through molecular systematic studies, and the use of species
45 delimitation by molecular ecologists interested in describing patterns of gene flow. This problem is
46 particularly acute in the marine environment where the inventory of biodiversity is relatively delayed while
47 molecular studies since two decades have shown a high prevalence of cryptic species. In this review we
48 illustrate, based on a review of the marine population connectivity literature and case studies, how the
49 failure to recognize evolutionary-relevant unit boundaries leads to heavily biased estimates of connectivity.
50 We review the conceptual framework within which species delimitation can be formalized as falsifiable
51 hypotheses, and show how connectivity studies can feed integrative taxonomic work, and *vice versa*. Finally,
52 we suggest strategies for spatial, temporal and phylogenetic sampling to reduce the probability of
53 inadequately delimiting evolutionary units when engaging in connectivity studies.

54

55

56

57 **I. Context and problems.**

58

59 Population connectivity, involving the demographic and/or evolutionary interdependence of populations
60 through individual movements across their species range, has been subject to an increasing number of
61 studies for the last two decades (Scopus search for “population connectivity”, accessed on August 23th,
62 2014: 2,060 documents published since 1993, including 1,700 during the last 5 years). Connectivity studies
63 have bearing on both applied and theoretical research. For instance, assessing the extent of connectivity
64 among populations has become a key aspect in diverse areas of conservation biology. Applications of
65 connectivity studies include: 1) identifying genetically isolated populations of protected or endangered
66 species that should be monitored as separate conservation units (Palsbøll *et al.* 2007), 2) optimizing the size
67 and location of protected areas to create well-connected reserve networks (Kritzer & Sale 2004; Jones *et al.*
68 2007), 3) identifying stocks for fisheries management (Fogarty & Botsford 2007; Waples *et al.* 2008; Reiss *et*
69 *al.* 2009), and 4) evaluating the potential impacts of resource exploitation on population dynamics of local
70 communities (Bors *et al.* 2012; Plouviez *et al.* 2013). Besides these important applications to management
71 and conservation, connectivity studies can also help address long-standing questions. In ecology for
72 example they allow understanding to what extent marine populations are closed or open systems (Cowen *et*
73 *al.* 2000), in microevolution they can be used to identify factors that create and maintain genetic
74 differentiation (e.g. Bilton *et al.* 2002; Shank 2010), or to understand how local adaptation can occur in high
75 gene flow species (e.g. Nielsen *et al.* 2009).

76 In this review, we underline the crucial importance for connectivity studies of the state of taxonomic
77 knowledge and correlated aspects on the available background knowledge on the biology of organisms.
78 These factors are sometimes overlooked despite the fact that they should condition the sampling design
79 and the inferences made from genetic data. One of the main considerations we develop is that properly
80 estimating connectivity at the population level requires assessing the robustness of available taxonomic
81 hypotheses. This is not trivial as for a large part of biodiversity, taxonomic knowledge is inadequate or even
82 lacking. Indeed, a great portion of the world’s biodiversity remains to be described, mainly in under-
83 explored, difficult-to-access habitats such as tropical rainforests and the marine environment (e.g.
84 Appeltans *et al.* 2012). In addition, in these habitats, species descriptions are often based on few specimens
85 and species distribution ranges and ecological requirements are poorly known. The development of DNA-
86 sequencing techniques has considerably accelerated the rate of discovery and the documentation of species
87 distributions. This is notably true in the marine realm, especially for poorly studied eukaryotic phyla for
88 which few or no other characters were available as reference for taxonomic delimitations (e.g. cryptic
89 lineages of macro- and micro-algae that are subject to convergent evolution towards reduced
90 morphologies; see for review Leliaert *et al.* 2014). However, turning these DNA-based discoveries (generally

91 only based on data such as those gathered from the Barcode of Life program), into robustly and formally
92 named taxonomic entities remains a long and difficult process (Satler *et al.* 2013). Consequently, the
93 literature includes a large number of undescribed and un-named cryptic or pseudo-cryptic species for which
94 identification from one study to another is challenging in the absence of formal naming and description. As
95 recently underlined by Fontaine *et al.* (2012), the time lapse between specimen collection and formal
96 species description is extremely long (average “shelf life” of 21 years). A perhaps less appreciated issue is
97 that even for abundant, commercially-important or ecologically well-studied taxa, nominal species were
98 often described long ago and are not necessarily adequately delineated. This situation is particularly critical
99 in the marine environment (e.g. Uthicke *et al.* 2010; Jaafar *et al.* 2012; Mantelatto *et al.* 2014; Thomas *et al.*
100 2014). For instance *Ciona intestinalis* (Linnaeus, 1767), is a model organism in evolutionary developmental
101 biology and phylogeny and was among the first animals to have its genome fully sequenced (Dehal *et al.*
102 2002). However this taxon was only recently revealed as a complex of four cryptic species (Suzuki *et al.*
103 2005; Caputi *et al.* 2007; Nydam & Harrison 2007; Zhan *et al.* 2010), including two widespread species that
104 diverged ca. 3-4 my ago (Roux *et al.* 2013) but that are not yet formally named.

105 Beyond the discovery of recently diverged species, examples of taxonomic confusions at higher ranks are
106 also not rare. For instance in 2009, Johnson *et al.* revealed that three families of deep-sea fishes artificially
107 separated juveniles, males and females. This confusion was resolved by examining the morphology of more
108 specimens and by using molecular data. This taxonomic revision, in which the three families are
109 synonymized (the Cetomimidae being the valid family name), greatly changes the understanding of
110 connectivity in terms of life history traits and ecology but should also affect the strategy for sampling
111 populations.

112 The main objective of this review is thus to stress the fact that, when conducting connectivity assessments,
113 the taxonomic status of the studied organisms should be critically reassessed, and revised if necessary, in
114 light of newly acquired biological data. This point is particularly crucial in marine systems, which are the
115 focus of this review, both because the marine biota accumulates the greatest number of deep evolutionary
116 lineages (i.e. phyla), but also because it remains vastly under-sampled (see Costello *et al.* 2010 for
117 multicellular organisms and Not *et al.* 2012 for the phytoplankton). Additionally, we outline that the type of
118 data collected for estimating connectivity using population genetic methods may in turn be used to refine
119 the taxonomic knowledge at the species rank. Actually, cryptic species are typically revealed by population
120 genetic studies or by phylogeographic analyses. In fact, the patterns observed in connectivity studies and
121 species delimitations belong to the same divergence continuum because both result from the same
122 ecological and micro-evolutionary processes. Furthermore, the lack of knowledge for marine organisms is so
123 important (e.g. Knowlton 2000 or Webb *et al.* 2010) that the question of species delimitations does not only
124 concern the limits between recently separated lineages (i.e. the ‘emergent phylogenetic systems’ as defined

125 in Carstens *et al.* 2013) but also in the definition of higher-rank classifications as illustrated with the above
126 example of Cetomimidae. We here first illustrate the importance of accurate taxonomic evaluation in the
127 analysis of connectivity by examining one specific question (i.e. the correlation between pelagic duration
128 and genetic structure estimates). We then highlight the main issues in relation with taxonomic assignment
129 and propose a framework (guidelines) for preventing errors due to faulty taxonomy when conducting
130 connectivity studies.

131

132 **An illustration of the problem: pelagic larval duration, genetic structure, and obscure species boundaries.**

133 The issue of poor species delimitations extends beyond individual studies when we consider meta-analyses
134 or reviews investigating ecological trends such as the relationship between life-history traits and
135 connectivity levels. A first, fundamental, step before conducting such comparative studies should be to
136 ensure that comparisons are made across equivalent evolutionary units. Note that these evolutionary units
137 may or may not be named. For instance, comparative studies including measures of genetic structure such
138 as F_{ST} may produce biased conclusions if taxonomically-unresolved taxa are included. For instance in *C.*
139 *intestinalis* the value of F_{ST} computed with microsatellites is six times higher between two populations, each
140 made of the different cryptic species attributed to this name, compared to the values computed between
141 populations including only one of these cryptic species (Zhan *et al.* 2010). If cryptic species are not treated
142 as distinct entities, an analysis of genetic diversity among different sampling locations could lead to
143 confound extrinsic barriers to dispersal with intrinsic reproductive barriers. We illustrate this issue by
144 critically examining some of the source data used in a series of meta-analyses comparing measures of
145 genetic differentiation across multiple species (Weersing & Toonen 2009, Selkoe & Toonen 2011). These
146 meta-analyses address the question of whether early life-history traits influence connectivity and if this
147 influence may be evidenced by the analysis of neutral genetic markers. This question is debated in marine
148 molecular ecology because for most marine multi-cellular organisms, early life-history traits greatly differ
149 from adult life-history traits (especially in terms of dispersal ability), and thus putatively affect connectivity
150 among populations (especially if the habitat is fragmented). Different reviews and comparative studies have
151 reached conflicting conclusions (e.g. Kinlan & Gaines 2003; Siegel *et al.* 2003; Shanks 2009; Weersing &
152 Toonen 2009; Riginos *et al.* 2011; Selkoe & Toonen 2011). Weersing & Toonen (2009) examined how well
153 pelagic larval duration (PLD) correlates with genetic estimates of connectivity. Their meta-analysis was
154 based on a pseudo-random selection of papers from hits with selected keywords on electronic searches.
155 The analyzed dataset included 130 species from 87 papers. Contrasting with the general expectation, the
156 authors found only a weak correlation between PLD and genetic structure as estimated by the overall F_{ST} . As
157 emphasized by Selkoe & Toonen (2011), this result may be explained by various factors, including the
158 metrics used for estimating effective dispersal (i.e. gene flow), the type of markers and the sampling design.

159 In addition, we suggest that one possible explanation for such a weak correlation may be that unrecognized
160 cryptic species within organisms with high PLD have been included in the dataset. Indeed, if two cryptic
161 species are combined under a same name, F_{ST} estimates might be high not because the sampled
162 populations are geographically structured (i.e. limited gene flow) but simply because these populations do
163 not belong to the same species. At least some of the selected papers include questionable taxonomic
164 entities. For example, the reef fishes attributed to five different *Halichoeres* Rüppell, 1835 species had
165 among the highest F_{ST} values and unexpectedly the highest PLD. Examination of the available data from the
166 literature for these species shows that different taxonomic treatments have been applied for similar
167 divergence patterns in this genus (Rocha *et al.* 2005; Rocha *et al.* 2008). For example the name *Halichoeres*
168 *bivittatus* (Bloch, 1791) covers two divergent mitochondrial lineages whereas the names *Halichoeres*
169 *brasiliensis* (Bloch, 1791) and *Halichoeres radiatus* (Linnaeus, 1758) are attributed to two mitochondrial
170 lineages separated by a level of divergence similar to that of the lineages within *H. bivittatus*. The two
171 mitochondrial lineages of *H. bivittatus* are allopatric, except in Bermuda where both are present, although
172 not in the same environments. The high overall F_{ST} within *H. bivittatus* might thus result from endogenous
173 reproductive isolation barriers rather than dispersal barriers within a metapopulation. The case of the fish
174 *Elacatinus evelynae* (Böhlke & Robins, 1968) is another example. Several mitochondrial and nuclear genetic
175 markers, and an adequate sampling of closely related species revealed three distinct and well-supported
176 phylogenetic lineages (Taylor & Helberg 2005) but the very high F_{ST} value used in the meta-analysis by
177 Weersing & Toonen (2009) was estimated over a sample that combines these three lineages (Taylor &
178 Heldberg 2003). Whether the three lineages evidenced by the phylogenetic approach need to be described
179 as distinct species is an open question but an important criterion would be to determine if the divergence is
180 definitive (i.e. definitive cessation of reproductive exchanges whatever the causes). In the case of
181 reproductive isolation, the correlation between F_{ST} estimates and PLD should be evaluated only within each
182 lineage. These examples illustrate that depending on how the available results are interpreted in terms of
183 taxonomic status, conclusions of comparative studies on patterns of connectivity can vary widely.

184

185 **Theoretical framework for delimiting species: a primer.**

186 This critical analysis underlines a crucial aspect of taxonomy: described taxa are not facts but testable
187 hypotheses about the structure of biodiversity. This is true not only for higher taxonomic ranks, generally
188 considered as arbitrary ranks, but also for the species rank, which is the only taxonomic category for which a
189 “biological reality” is recognized by most scientists (see for example in Mishler 2009). The question is then
190 how such hypotheses are formulated in reference to a theoretical background and how they are then
191 revised in light of new empirical data. The literature about the species concept is very large and the debate
192 has often been considered as unsolvable (often referred as “the species problem”). In this debate, de

193 Queiroz (1998) was among the first to identify that two questions of different nature are mingled. He
194 indeed underlined that the question of the definition of the species category has to be distinguished from
195 that of the question of the adequacy of the criteria used in practice in the definition of the taxa ranked as
196 species. The definition of the species category is an ontological question that may be addressed based on
197 the theoretical ground offered by the Evolutionary Theory (as discussed in Samadi & Barberousse 2006).
198 How each species-taxon should be delineated is an epistemological question that requires evaluating how
199 well empirical data and analysis methods allow the taxonomists to propose sets of organisms that fit the
200 chosen definition of the species category. Following this line of thinking, species-taxa are viewed as
201 scientific hypotheses (species-hypotheses) engaged in a process of falsification based on the acquisition of
202 new evidence. Although this analysis was only recently accepted among biologists, it closely corresponds to
203 the working process of taxonomic revisions, which is one of the major activities of taxonomists (see
204 discussion in Barberousse & Samadi, 2010). This analysis is also the conceptual background of the presently
205 active field of integrative taxonomy. In the integrative approach to taxonomy, various attributes
206 (morphology, DNA, geographical range, habitat, behavior, etc...) and criteria (i.e. the so-called "species
207 concepts" such as the biological, the phylogenetic or the phenetic criteria), analyzed using various methods,
208 are taken into account to propose robust species-hypotheses, reproducible and testable (see Camargo &
209 Sites 2013 for a review on the integration of many characters and methods to delimit species). As
210 underlined by Yeates *et al.* (2011), among others, the way hypotheses are validated should not be primarily
211 the intersection among the evidence from different datasets but rather the prioritization of alternative
212 species-hypotheses based on their explanatory power as defined within the framework of Evolutionary
213 Theory. In this process the quality or robustness of available species-hypotheses greatly depends on the
214 state of knowledge about the concerned compartment of the tree of life. For instance the state of art of
215 species delimitations in birds is far more stable than that of most marine organisms. In fact uncertainties
216 about available species-hypotheses may have two different causes. First, the state of art of taxonomy may
217 be so preliminary that different life-stages of a same organism might have been erroneously classified in
218 different species or even families (see above in Johnson *et al.* 2009). Second, the evidence that two
219 evolutionary lineages are engaged in definitive divergence may not be easy to evaluate when the time
220 elapsed since the split between the emergent lineages is recent (the "grey zone" of de Queiroz, 1998). If the
221 relevance of taxonomic knowledge needs to be evaluated before conducting any biological study on a given
222 organism we also should underline that the results of connectivity studies, if carefully analyzed, can in turn
223 provide evidence about the state of divergence among emergent lineages.

224

225 **Where do we go from there?**

226 Four main sources of difficulty may strongly influence measures of connectivity and taxonomic hypotheses:
227 (1) the state of knowledge about the biology of the studied organisms, (2) the state of taxonomic
228 treatments of the studied organisms, (3) the spatial and temporal scales of sampling, and (4) the characters
229 used to infer connectivity patterns. In the following section we review these four points and give some
230 guidelines through examples taken from diverse marine model systems. We also offer some guidelines for
231 designing connectivity studies when little background knowledge is available.

232
233

234 **II. Methodological considerations and guidelines for connectivity studies.**

235

236 **1. Background knowledge on the biology of organisms.**

237 The availability of adequate background knowledge on the biology of organisms is crucial both for a robust
238 taxonomic framework and for the study of connectivity. As discussed in the introduction, one of the life
239 history traits commonly examined when studying connectivity data for marine organisms is pelagic larval
240 duration (e.g. Cowen & Sponaugle 2009; Shanks 2009; Weersing & Toonen 2009; Selkoe & Toonen 2011).
241 This biological character is also crucial for taxonomists when dealing with species delimitation (see an
242 example in gastropods in box 5). However, other biological attributes such as reproductive strategies, sexual
243 dimorphism or ecological specificities (i.e. habitat specialization, biological interactions, phenology, etc ...) can
244 significantly influence both interpretations of connectivity data and taxonomic decisions. For example,
245 advances in understanding of the migratory behavior of some focal marine organisms, such as eels, salmon
246 or whales (e.g. Bottom *et al.* 2005; Aarestrup *et al.* 2009; Baker *et al.* 2013) revealed the existence of
247 seasonal migrations among breeding and feeding sites, or discrete breeding sites, that have bearing on
248 connectivity estimates. For instance, sex-specific philopatry affects the pattern of genetic structure in sperm
249 whales (Engelhaupt *et al.* 2009) and humpback whales (Baker *et al.* 2013). In both cases, patterns of genetic
250 differentiation are interpreted as a consequence of sex-biased philopatry, because it was possible to
251 determine the gender of each individual analyzed, which is not possible for many poorly known organisms.
252 The importance of sexual dimorphism, season and location of sampling can greatly affect the observed
253 patterns of genetic differentiation and has consequence in the evaluation of species diversity (illustrated by
254 the example of *Lessonia* in box 2). While the migratory behavior of some well-studied marine species has
255 been characterized, the existence of seasonal migrations or discrete breeding sites is poorly known in most
256 marine groups. Thus, the unreported occurrence of seasonal migrations may bias interpretations obtained
257 when samples are collected from a single season. In extreme cases, different life history stages or sexes can
258 wrongly be attributed to different taxa (e.g. Johnson *et al.* 2009), confounding estimates of connectivity.
259 When the different sexes or stages inhabit different environments (such as different depths, e.g. the case of

260 the Cetomimidae in Johnson et al 2009), estimating dispersal from a single life-stage or sex can bias
261 interpretation of connectivity patterns but also taxonomic interpretations. Other attributes, such as
262 clonality or alternation of generations, can also complicate connectivity studies. In the case of clonal species
263 such as seagrasses or some algae, neglecting such attributes would lead to a severe underestimation of
264 gene flow or to overlook the presence of cryptic species (see in Box 4 and also Adjeroud *et al.* 2014 for
265 another case in corals). Thus, the sampling design and interpretation of genetic data in connectivity studies
266 and taxonomic revisions should take into account life history and ecological traits that could be relevant to
267 estimating dispersal. If, as in many cases, such knowledge is limited or inexistent for the species of interest,
268 one could examine information available for closely related taxa and/or incorporate this uncertainty in the
269 sampling design (by sampling at different seasons, or different depths for example).

270

271 **2. Taxonomic knowledge.**

272 Connectivity studies generally consider the described species (i.e. available species hypotheses) as facts
273 upon which the sampling scheme is established. In other words, the attributes associated with a species-
274 hypothesis (either extracted from the original description or from other biological studies based on this
275 species hypothesis) are considered as objective facts. Notably, the species range and habitat specificity are
276 among the important attributes extracted from those hypotheses that are used in the design of a
277 connectivity study. However, an important source of uncertainty in taxonomic hypotheses is the sampling of
278 organisms and/or of characters upon which the species-hypotheses were initially drawn. Thus, data
279 provided by a connectivity study are potentially a source of falsification of accepted taxonomic hypotheses.
280 Consequently, for organisms for which no recent taxonomic revision is available, the sampling scheme
281 should both follow the requirements of population biology but also that of phylogenetic systematics. For
282 example, only a widespread sampling of a supposedly cosmopolitan species may reveal several diverging
283 lineages restricted to isolated areas or distinct habitats as exemplified by the study of the supposedly
284 ubiquitous and widespread amphipod *Eurythenes gryllus* (Lichtenstein in Mandt, 1822) (Havermans *et al.*
285 2013) or the copepods *Pleuromamma piseki* Farran, 1929 and *Pleuromamma gracilis* Claus, 1863 (Halbert *et*
286 *al.* 2013). Collecting in under-sampled or unexplored areas may also reveal new divergent lineages, like in
287 the cosmopolitan red algae *Asparagopsis armata* Harvey, 1855 and *Asparagopsis taxiformis* (Delile) Trevisan
288 de Saint-Léon, 1845, for which human-mediated transports increase the uncertainty about taxon history
289 and systematics (Dijoux *et al.* 2014). Sampling is also an issue for species described as geographically
290 restricted or ecologically specialized: looking for (and sampling) the targeted species or a priori closely
291 related species in other localities or habitats may help rejecting hypotheses of endemism and/or
292 specialization. For example, a deep-sea chemosynthetic mussel attributed to a new *Idas* Jeffreys, 1876
293 species by Ritt *et al.* (2012), sampled in a deep cold seep site in the eastern Mediterranean sea, is included

294 in the same *cox1* cluster as the specimens attributed to *Idas simpsoni* (J. T. Marshall, 1900) and sampled at
295 about 150 m depth in the north Atlantic on vertebrate bones (Thubaut *et al.* 2013). In this example the new
296 species was considered as restricted to a given habitat that is highly fragmented and consequently other
297 specimens sampled in a distant and distinct habitat were falsely assigned to a distinct species. Two sets of
298 populations, identified as different species each associated with different habitats, may actually be
299 populations of a single species connected through (even if rare) gene flow. Similar observations were also
300 recently reported among chemosynthetic shrimps that were considered as distinct in cold seeps and vents,
301 while molecular analysis revealed the occurrence of a single species with high gene flow across the Atlantic
302 (Box 3; Teixeira *et al.* 2013). These examples illustrate the fact that the definition of the targeted group
303 (ingroup) may be biased due to a lack of samples from specimens a priori identified as belonging to a
304 different (related) species. Such sampling biases correspond to higher rank classification errors and may for
305 example exclude from genetic analyses a subset of populations despite its crucial role in shaping
306 connectivity patterns. As underlined below, the risk of inadequacy of available species-hypotheses should
307 not be ignored and sampling cannot be limited to the intra-specific level. Because of the poor state of
308 taxonomic knowledge for most marine organisms, taxonomic hypotheses (i.e. accepted classifications)
309 should thus be carefully evaluated prior to assessing connectivity.

310

311 **3. Guidelines for spatial and temporal sampling of organisms.**

312 Sampling should be guided by two objectives: adequately covering (1) both the putative geographic and
313 ecological range of the species of interest (coverage at the population level), and (2) potentially closely-
314 related species over a wide geographic and ecological range (taxonomic coverage). Coverage at the
315 population level should also take into account potential temporal variability in the geographic location of
316 individuals (e.g. different location and time of breeding and feeding) and in habitats (e.g. pelagic *versus*
317 benthic).

318 As explained above, this two-level sampling (developed in the case study presented in Box 5) should ensure
319 a proper estimation of the upper bound of intra-specific diversity and the lower bound of inter-specific
320 diversity as to determine their potential overlap. The genetic distance between conspecifics and individuals
321 from different species, known as the “barcoding gap” (Meyer & Paulay 2005), is often used as a threshold to
322 delimit species (at least in a primary step – Puillandre *et al.* 2012a) but its detection and meaning highly
323 depend on the sampling scheme. These sampling guidelines are of course difficult to follow in all situations,
324 especially in the marine realm: rarity, endemism, habitat fragmentation, human-mediated transports (i.e.
325 biological introduction), lack of knowledge on sister-species relationships for most taxa, technical difficulties
326 in sampling in the deep sea, among others factors, make the estimation of intra- and inter-specific diversity
327 difficult. Such strategy may point to new species-hypotheses and thus to the need of taxonomic revisions.

328 However, such revisions (i.e. providing named species-hypotheses) are possible only if the requirements of
329 the codes of biological nomenclature are taken into account in the sampling scheme. Indeed, an important
330 requirement for attributing species names to the redefined species hypotheses is the inclusion of type-
331 material linked to available species-names (see for example Puillandre *et al.* 2011). This last step is often
332 lacking in molecular studies and the literature is spoiled by arbitrarily-labeled species hypotheses (e.g.
333 OTUs, MOTUs, n. sp etc ...) that are difficult to compare from one study to another. As most name-bearing
334 specimens (holotypes) cannot be integrated in molecular studies (due to their age, their poor conservation
335 state, or simply because they were lost), sampling in type localities (as defined for example for animals in
336 the article 76 of the International Code of Zoological Nomenclature) is a minimum requirement for naming
337 species-hypotheses. This will both allow the corroboration (or the rejection) of available species-hypotheses
338 and an adequate analysis of the pattern of connectivity within the targeted species. Implementation of the
339 aforementioned sampling strategy and interpretation of the results should be guided as far as possible by
340 this two-level method, especially when assessing the reliability of the inferred patterns of connectivity. In
341 summary, these guidelines aim at identifying the adequate spatial and evolutionary scale for connectivity
342 studies.

343

344 **4. Guidelines for the sampling of characters.**

345 Classical markers widely used for marine organisms include genes from the phylogenetic literature and
346 notably those mobilized in the DNA barcoding initiatives (typically, the *cox1* gene for metazoans, Bucklin *et*
347 *al.* 2011; *cox1* or *RbcL* genes for macro-algae, Leliaert *et al.* 2014), but also other mitochondrial and nuclear
348 genes such as ribosomal genes or “universal” introns (e.g. Jarman *et al.* 2002; Chenuil *et al.* 2010; Gérard *et*
349 *al.* 2013). By using a set of unlinked genetic markers, it is possible both to refine the phylogenetic
350 relationships (phylogenetic criterion) and to test whether specimens from the same species are able to
351 recombine and, conversely, whether specimens from different species cannot (biological criterion). This can
352 be achieved by sequencing, for each specimen, at least two unlinked markers, such as one from the
353 mitochondrial genome (or chloroplast genome in plants and algae) and one from the nuclear genome. It is
354 now widely accepted that the use of a single marker, especially mitochondrial, is risky when delimiting
355 species, and should be used only in a first step to quickly propose primary species-hypotheses. As
356 underlined in the review of Carstens *et al.* (2013), multi-locus approaches are required especially when
357 considering recently diverging lineages, notably because of the genome wide heterogeneity of introgression
358 rates observed between such genomes (e.g. between the two tunicates *C. intestinalis* type A and type B,
359 Roux *et al.* 2013; or between the two sibling species of red algae *Gracilaria gracilis* (Stackhouse)
360 M.Steentoft, L.M.Irvine & W.F.Farnham, 1995 and *Gracilaria dura* (C.Agardh) J.Agardh, 1842, Destombe *et*
361 *al.* 2010). Furthermore, in some taxa (e.g. cnidarians, sponges, plants, algae...), classical markers sometimes

362 lack the necessary resolution to distinguish closely related species and to infer connectivity (e.g. Calderón *et*
363 *al.* 2006; Shearer & Coffroth 2008). Examples can be found in metazoan groups with remarkably low levels
364 of mitochondrial evolution such as corals, when this sole genetic marker is considered (e.g. Shearer &
365 Coffroth 2008; McFadden *et al.* 2011). However, for many poorly studied organisms, identifying a nuclear
366 marker variable at the species level in addition to, for example, the *cox1* gene can be tricky (e.g. McFadden
367 *et al.* 2010). In such taxa, genetic similarity among individuals may reflect above-species evolutionary
368 relationships rather than demographic connectivity (e.g. Miller *et al.* 2010). Clearly, these technical
369 difficulties should be overcome thanks to the new methods based on Next Generation Sequencing, such as
370 the RAD-sequencing (Baird *et al.* 2008). In a single run, many independent loci covering a large part of the
371 genome are sequenced. They are thus particularly appropriate to test for recombination among individuals
372 and to unravel diagnostic molecular characters among taxa (e.g. Pante *et al.* accepted).

373 Even if DNA characters often provide reliable data for species delimitation, either using a single locus to
374 propose primary species-hypotheses (e.g. “species discovery” by DNA barcoding) or multiple loci to test for
375 recombination among individuals within a species, they may not always provide decisive answers. When
376 genetic data are not fully conclusive, morphological, geographical and ecological data can help turn the
377 scales toward alternative hypotheses. For example Kekkonen & Hebert (2014) use a geographic criterion
378 (*i.e.* genetic units are or not in sympatry) to conclude about the status of the detected genetic units. Our
379 goal is not here to list all the characters and methods that can be included in the integrative taxonomy loop
380 (as illustrated in Figure 1), but just to emphasize the need to discuss alternate species hypotheses, referred
381 as (M)OTUs ((Molecular) Operational Taxonomy Units), ESU (Evolutionary Significant Units), PSH (Primary
382 Species Hypotheses) or candidate species, depending on the method and characters, using all the available
383 data to converge to robust, reproducible and testable hypotheses.

384

385 **5. Prioritization and workflow in multi-marker approaches.**

386 Integrative taxonomy has been proposed as a solution to overcome apparent contradictions in species-
387 hypotheses resulting from the use of different criteria or characters (Dayrat 2005, Will *et al.* 2005). This
388 approach is now widely accepted, although not always implemented in a fully integrated way. This results
389 from the fact that the different methodologies cannot be applied in a similar way to all taxonomic groups,
390 since each of them has its own limitations. For example, ecological data, commonly used in species
391 delimitation of terrestrial and shallow water organisms, are more difficult to gather when dealing with
392 deep-sea species. However, some general rules have been set up, and now, the debate does not primarily
393 concern the definition of the species concept or the suitability of an integrative approach, but rather the
394 relative weight and priority of the different characters and criteria used for species delimitation (DeSalle *et*
395 *al.* 2005; Padial *et al.* 2010; Yeates *et al.* 2011; Camargo & Sites 2013).

396 The principal source of difficulty for integrative taxonomy comes from cases of recent divergence
397 ('emergent phylogenetic systems' as defined in Carstens *et al.* 2013). For example, under the hypothesis of
398 a single species, high F_{ST} estimates are interpreted as population structure (low connectivity). If this species-
399 hypothesis covers in fact two recently separated lineages, depending on the level of shared polymorphism
400 among them, the F_{ST} estimates would be more or less close to one. Thus when taxonomy is uncertain, the
401 two alternative hypotheses explaining such high F_{ST} estimates should be examined. Wang *et al.* (2008)
402 illustrate this case by discussing how to discriminate between these two alternative hypotheses in their
403 investigation of two morphotypes of finless porpoise (genus *Neophocaena*, Cetacea) that occur in sympatry
404 in the strait of Taiwan. In their study, they apply population genetics methods to distinguish between a case
405 of recent speciation and a case of low but recurrent gene flow. When simulating two populations with no
406 gene flow during the last 18,000 years, simulated and observed F_{ST} and migration rate estimates were
407 similar. Thus, the simulated scenario of a recent speciation event matched empirical data, leading the
408 authors to conclude that the two morphotypes constitute two recent species that are currently in secondary
409 contact. In this study the two species are not reciprocally monophyletic (phylogenetic criterion) but the
410 population genetic approach supports the hypothesis of a complete breakage of gene flow (biological
411 criterion) between the two morphotypes. Reciprocal monophyly is a criterion frequently used for species
412 delimitation using molecular methods (e.g. Funk & Omland 2003) because it is much easier to evaluate than
413 the biological criterion (inter-fecundity within species and inter-sterility among species). When the
414 separation among studied species is ancient, the criterion of reciprocal monophyly is an operational method
415 for detecting species. Conversely, among recently diverged species (i.e. 'emergent phylogenetic systems'),
416 we recommend to give advantage to population genetic methods allowing the detection of lasting breakage
417 of gene flow over the criterion of reciprocal monophyly. New methods of analysis, mostly based on
418 coalescent theory, allow combining both criteria and should be as much as possible used in this context (e.g.
419 Yang and Rannala 2010, Leaché *et al.* 2014, Hsieh *et al.* 2014). These methodologies coupled with new
420 genotyping techniques allow the use, in a single analysis, of both the phylogenetic criterion (*i.e.* two sets of
421 organisms have distinct evolutionary trajectories) and the biological criterion (*i.e.* the two sets of organisms
422 are reproductively isolated) but were used only in a few pioneering studies (e.g. Leaché *et al.* 2014). Their
423 applicability on poorly known organisms of the marine realm has yet to be explored but the robustness of
424 such analyses will greatly depend on the quality of the sampling scheme relative to the state of taxonomic
425 (and biological) knowledge.

426

427 **III Conclusion and implications.**

428

429 In many marine species, particularly those with high fecundity and large population sizes, population
430 genetic studies have often revealed weak genetic structure, preventing accurate analyses of connectivity
431 patterns. However, strong genetic structure patterns have also been revealed in marine systems. In this
432 review, we argued that, in some cases, such observed patterns may actually be the outcome of incorrect
433 sampling and/or incorrect taxonomic assignment.

434 We showed that unreliable definition of evolutionary units can lead to either overestimation or
435 underestimation of connectivity. This situation is analogous to type I and type II statistical errors in a
436 hypothesis-testing framework where full connectivity is the null hypothesis. In this review we illustrated two
437 general cases, in which both types of errors may be encountered. These two cases correspond to the
438 number of evolutionary units being (1) larger or (2) smaller than the number of species acknowledged in the
439 study. In the first case, *i.e.* over-merging of evolutionary units, high genetic structure is equivalent to false
440 positives and may emerge when populations from different, unrecognized or cryptic species are lumped
441 together (e.g. Box 2 on the kelp genus *Lessonia* Bory de Saint-Vincent, 1825). This would lead to an increase
442 in type I statistical error, in which the null hypothesis of a lack of barrier to gene flow among populations
443 would be falsely rejected because species, and therefore populations, were not properly delimited. One
444 simple example documented above is when falsely inflated F_{ST} -values due to mixing of specimens from
445 different evolutionary entities are interpreted as picturing limited dispersal and increased genetic drift. The
446 issue lies in the interpretation of the data rather than in the data themselves, and some examples were
447 discussed above (e.g. Rocha *et al.* 2005; Taylor & Helberg 2005; Rocha *et al.* 2008). The second case, *i.e.*
448 over-splitting of evolutionary units, results in the flawed *a priori* assumption of a complete lack of
449 connectivity among taxa described as distinct species (see the example of the Atlantic shrimp named,
450 depending on its habitat, *Alvinocaris muricola* Williams, 1988 or *Alvinocaris markensis* Williams, 1988 as
451 detailed in Box 3), preventing their recognition as potentially interdependent entities and the will to engage
452 in connectivity studies among their populations.

453
454 As illustrated by the examples discussed in this paper, the issue of unreliable delineations of evolutionary
455 units has been exacerbated in the past two decades with the rise of molecular data, particularly in the
456 framework of phylogeographic studies documenting an ever-increasing number of cryptic species in the
457 marine realm (Appeltans *et al.* 2012). Species are hypotheses that need to be revised if necessary in light of
458 new information. To anticipate the possibility of false taxonomic hypotheses, we highlighted several
459 important aspects. First, the sampling design should encompass as much as possible 1) the entire known
460 distribution range of the targeted organism and 2) closely related taxa, or even some distantly related taxa
461 in taxonomic groups where the phylogeny is poorly known (e.g. deep-sea organisms). The latest point is
462 rarely considered in connectivity studies although it is an efficient way to ensure the correct definition of

463 the focal (i.e. ingroup) species. Second, any connectivity study should be careful in its design regarding life-
464 history traits: pelagic larval duration, reproductive system characteristics (sex-ratio, clonal reproduction,
465 ...etc) and social organization, which all have important consequences on connectivity patterns but also on
466 species delimitations. Third, the types of markers used should be diversified; in particular the sole use of
467 mitochondrial genes should be avoided. Altogether, the three points mentioned above are all relevant to
468 the framework of integrative taxonomy. This approach has proven to be helpful in species delineation and
469 meaningful when coupled with or used for connectivity studies.

470 Importantly, connectivity studies can reciprocally provide new data that can help defining new species in
471 previously recognized nominal species. This is particularly well illustrated in the case of introduced or
472 invasive marine species (see box 6). Human-driven dispersal adds to natural dispersal, thereby broadening
473 species ranges, sometimes at a worldwide scale. Connectivity and phylogeography studies of marine
474 invaders have shown that important and cosmopolitan invaders are actually species complexes (e.g. in
475 colonial tunicates *Botryllus schlosseri* (Pallas, 1766), Bock *et al.* 2012; in solitary tunicates *C. intestinalis* Zhan
476 *et al.* 2010; in the red alga *A. taxiformis* Dijoux *et al.* 2014). Cryptogenic, pseudo-indigenous species and
477 cryptic species, as defined by Carlton (2009) are new issues deserving integrative taxonomy and barcoding
478 studies, not only to verify the accuracy of invaders reports (e.g. McGlashan *et al.* 2008) or to quickly report
479 newly introduced species (Bishop *et al.* 2013), but also to examine gene flow between cryptic species not
480 fully reproductively isolated, in particular when invaders are morphologically similar to native species (e.g.
481 Nydam & Harrison 2011). More generally, delineating species can be particularly challenging considering
482 how rapidly species distribution can shift due to either introduction processes or global change. These
483 issues increase the complexity of taxonomic assignment and biogeography studies.

484 Finally, poor species delimitation and the use of inadequate taxonomy assignment are not restricted to
485 connectivity studies and have consequences on other fields of ecology and evolutionary studies, in
486 particular experimental ecology, medicine, evolutionary developmental biology, speciation and
487 phylogenetic studies, invasion biology research and biodiversity inventories (Bortolus 2008; Carlton 2009;
488 Collins & Cruickshank 2013). Recent methodological and conceptual advances will contribute to avoid the
489 pitfalls listed in this work, by taking advantage of the emergence of integrative taxonomy and the
490 development of new molecular techniques such as NGS-based methods, to provide a more accurate
491 exploration and understanding of the building-up of evolutionary units.

492

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503 **Boxes.**

504 **Box 1. Species delineation blurred by hybridization and suboptimal sampling design: the case of *Fucus***
505 **Linnaeus, 1753.**

506

507 The brown algae genus *Fucus* Linnaeus, 1753 is a well-known, textbook model in marine ecology,
508 exemplifying the distribution of different morphological species in well-defined zones along a vertical
509 intertidal gradient in the North Atlantic, from high- to low-water marks (Wahl *et al.* 2011). However, the
510 recent use of molecular markers has deeply modified our understanding of both taxonomy and connectivity
511 in this species complex.

512 The two closely related brown algae morphological species: *Fucus spiralis* Linnaeus, 1753 and *Fucus*
513 *vesiculosus* Linnaeus, 1753 are found in sympatry along eastern Atlantic rocky shores, from northern
514 Norway to Morocco. They differ mainly by their morphology, their vertical distribution on the shore and
515 their reproductive system. *F. spiralis* is hermaphrodite and is found in the higher intertidal, whereas *F.*
516 *vesiculosus* is dioecious and is located just lower on the shore. Molecular analyses of internal transcribed
517 spacer of nuclear ribosomal DNA (ITS) were not efficient to separate these two taxa (Serrão *et al.* 1999) and
518 their taxonomic status as two separate species was questioned. The use of five microsatellite loci on six
519 populations of *F. spiralis* and ten of *F. vesiculosus*, separated by tens to hundreds of kilometers, showed that
520 within each species individuals share diagnostic alleles whatever their geographic origin (Billard *et al.* 2005).
521 Consequently, the phylogeographic approach of Billard *et al.* (2005) demonstrated that these two
522 morphological species correspond to two different genetic entities. Using gene admixture detection
523 methods, intermediate individuals between the two species were detected along shores where the
524 distribution of the two species overlaps at the scale of few hundred meters (Engel *et al.* 2005). The authors
525 suggested a recent divergence of the two taxa with retention of ancestral polymorphism or introgressive
526 hybridization with varying levels of admixture.

527 Interestingly, using a more refined sampling (combining transects across the zonation of the shore and
528 quadrats at the different tidal levels), a new genetic entity previously overlooked by Engel *et al.* (2005) was
529 detected within the *F. spiralis* morphotype (Billard *et al.* 2010). These two entities were first labeled *F.*
530 *spiralis* Low and *F. spiralis* High according to their vertical distribution on the shore. Based on an integrative
531 approach combining molecular, physiological and morphological analyses, Zardi *et al.* (2011) proposed to
532 elevate the new genetic entity, *F. spiralis* Low, to the species level using the name *F. guiryi* G.I.Zardi,
533 K.R.Nicastro, E.S.Serrão & G.A.Pearson, 2011. The range distribution of *F. guiryi* was shown to occur from
534 Brittany to Morocco (Coyer *et al.* 2011). Common garden experiments performed on the three sympatric
535 genetic taxa: *F. spiralis*, *F. guiryi* and *F. vesiculosus* showed that physiological response to desiccation stress
536 differed between species and was consistent with their respective vertical distribution on steep

537 environmental clines in exposure time (Zardi *et al.* 2011). The opportunity of hybridization between the
538 three species was confirmed to be higher when they are physically close to each other on the shore while in
539 southern Portugal, where *F. vesiculosus* is only found in estuaries and *F. guiryi* on the open coast, the two
540 species never hybridize (Zardi *et al.* 2011). In addition to their ecology, it was proposed that selfing as well
541 as limited gamete dispersal contribute to species integrity in the hermaphroditic species *F. spiralis* and *F.*
542 *guiryi* (Engel *et al.* 2005; Billard *et al.* 2010; Zardi *et al.* 2011). Finally, a high level of asymmetric
543 introgression was reported between these different species using a combination of nuclear and
544 mitochondrial DNA markers (Coyer *et al.* 2011). A scenario of organelle capture via hybridization and spread
545 of neutral nuclear alleles during range expansion was suggested to explain the common occurrence of
546 introgression in this genus. This example clearly showed that organellar loci as barcoding tools should be
547 used with care because of introgression.

548 **Box 2. The integrative taxonomy loop: study case of two cryptic species in the kelp *Lessonia nigrescens***
549 **Bory de Saint-Vincent, 1826 complex.**

550 The discovery of cryptic species does not always lead to a taxonomic reexamination and in many cases,
551 cryptic species remain without taxonomic characterization (and therefore with no corresponding Latin
552 name) long after their detection. Here we describe the case of the kelp *Lessonia nigrescens* Bory de Saint-
553 Vincent, 1826 complex that was recently identified as two separate cryptic species thanks to a considerable
554 sampling effort, combining phylogenetic, population genetics and ecological approaches. This kelp forms
555 forests along the intertidal South Eastern Pacific coasts and was long considered as a unique species from
556 Cape Horn in southern Chile (ca. 55° S) to central Peru (ca. 12° S; Santelices 1989). Two cryptic species were
557 recently identified through a multigene phylogeographic study (using a combination of four markers located
558 in the three genomic compartments - chloroplast, mitochondrion and nucleus - with 1,000 individuals
559 covering more than 2,500 km of coastline) (Tellier *et al.* 2009). The two divergent genetic lineages show a
560 parapatric latitudinal distribution: one species extends from southern Peru (17°S) to central Chile (30° S),
561 and the other extends from central Chile (29° S) to Chiloe Island (42°S). Between 29° S and 30° S there is a
562 narrow area where both lineages spatially overlap in discrete patches where individuals belong to either the
563 northern or southern species. This contact zone between 29° S and 30° S has been described as a mosaic of
564 sites occupied by one species or the other (Tellier *et al.* 2009), rather than a true gradual transition zone,
565 with one species replacing the other. In addition, a detailed population genetic study of gene flow in the
566 transition zone confirmed a total absence of hybrids and mixed populations (Tellier *et al.* 2011). Ecological
567 differentiation between the two cryptic species was investigated, using different approaches (e.g. controlled
568 response to different thermal and desiccation stresses in the lab, Oppliger *et al.* 2012; López-Cristoffanini *et*
569 *al.* 2013). Both microscopic (haploid) and macroscopic (diploid) phases of the haploid-diploid kelp life cycle
570 were studied. Results demonstrated adaptive divergence for both phases that could explain the geographic
571 segregation of these two cryptic species. In addition, contrasted demographic processes could be observed
572 at various stages of their life cycle: female gametophytes from the Northern species develop rapidly but
573 give very few oogonia per gametophyte, whereas female gametophytes from the Southern species delay
574 their maturation, grow vegetatively and then produce numerous oogonia per individual. These results
575 strongly advocate that, even though the origin of the genetic differentiation of the two species is ancient,
576 ecological and intrinsic differences are probably still important for the maintenance of reproductive
577 isolation.

578 These two species were finally distinguished by subtle morphological differences leading to a formal
579 description of two pseudo-cryptic species as *Lessonia berteroa* Montagne, 1842 and *Lessonia spicata*
580 (Suhr) Santelices (Gonzalez *et al.* 2012).

581

582 **Box 3: Taxonomic inflation results in the flawed *a priori* assumption of a complete lack of connectivity**
583 **among morphologically distinct populations: an example with shrimps in the deep-sea environment.**

584 As reported in this manuscript, most examples of taxonomic bias leading to distorted connectivity
585 inferences are linked to the existence of hidden cryptic species or lineages. We here illustrate the opposite
586 case in which the polymorphism of morphological characters led to taxonomic inflation and thus to a false
587 inference of the connectivity pattern. This situation was found in the Alvinocaridae, a family of caridid
588 shrimps specialized in deep chemiosynthetic ecosystems. In the Atlantic, two species were described in
589 1988 by Williams: *A. muricola* associated with cold seeps from Western Atlantic and *A. markensis* from
590 hydrothermal vents of the Mid Atlantic Ridge. The two species were distinguished based on a slight
591 difference in an abdominal segment being entire or obscurely serrate. This distinction was made on juvenile
592 specimens and later confirmed on adults (Komai & Segonzac 2005), although on the basis of a distinct
593 morphological criterion that was the more inflated and convex nature of anterior part of the branchial
594 region in *A. muricola* compared to *A. markensis*. In 2005, Komai & Segonzac cautiously identified as *A. aff.*
595 *muricola* shrimps sampled along the continental margins of Eastern Africa. These authors extensively
596 discussed the pan Atlantic distribution of *A. muricola* given the puzzling absence of geographically
597 intermediate population. These authors hypothesized either the occurrence of large-scale dispersal
598 capacities or the existence of unknown suitable sites in the intermediate area. A barcoding approach based
599 on the Cytochrome Oxidase I gene (*cox1*), initially aiming at retracing the phylogenetic history of the
600 alvinocarid shrimp, casted doubt on the taxonomic status of those two species, showing entire sequence
601 match among haplotypes of both species (Teixeira *et al.* 2013). Indeed, using both mitochondrial and
602 nuclear sequences (*cox1* and 18S) together with a set of microsatellite loci, Teixeira *et al.* (2013) showed
603 that all populations respectively attributed to these two names share a unique set of alleles at both types of
604 genetic markers and display extremely weak genetic differentiation. This result eventually confirmed the
605 hypothesis of intermediate populations raised by Komai & Segonzac (2005) while more importantly
606 demonstrating that *A. markensis* and *A. muricola* should not be considered as two distinct species.
607 Following this interpretation the two species have to be synonymized in a single species that has a wide
608 geographic distribution but also a diversified range of suitable habitats in the deep sea. This example
609 supports the recommendation of investigating the phylogenetic framework of taxa studied including
610 closely-related taxa before engaging into comprehensive connectivity studies.
611 The same barcoding analysis also revealed that classification at both genus and species level should be
612 revised within the Alvinocaridae with possible cases of synonymy of species described as distinct genera
613 (e.g. *Rimicaris hybissae* Nye, Copley & Plouviez, 2012 reported in Eastern Atlantic seeps and *Chorocaris*
614 *chacei* (Williams & Rona, 1986) and Mid-Atlantic vents).

615 **Box 4. The implications of working on organisms with clonal populations and alternation of generations:**
616 **the case of seagrass meadows and algae.**

617

618 Seagrass meadows were initially expected to exhibit large-scale panmixia due to the positive buoyancy of
619 their seeds and rhizomatic fragments. In studying connectivity in those partially clonal organism, the first
620 issue was the ability to recognize clonal lineages on the basis of their multi locus genotypes (MLG) in order
621 to discard from the datasets the sampling units that are replicates of the same clonal lineage that would
622 potentially artificially inflate structure estimates such as F_{ST} among meadows (Arnaud-Haond *et al.* 2005;
623 Arnaud-Haond *et al.* 2007a). Even avoiding this potential bias, however, population genetic analysis across
624 the range area of several seagrass populations suggested a systematic structure and limitation to gene flow
625 across all geographical scales among almost all discrete meadows analyzed (Coyer 2004; Olsen, 2004,
626 Arnaud-Haond *et al.* 2007b; Alberto *et al.* 2008). Secondly however, fine grained analysis of the
627 distribution of clonal lineages and genetic polymorphism in *Zostera marina* Linnaeus, 1753 meadows
628 revealed a large influence of clonality, leading to a mosaic pattern of genetic patchiness largely driven by
629 patchy events of recolonization followed by the occupation of space through clonal spread, despite the
630 likely occurrence of much larger scale effective migration (Becheler *et al.* 2010; Becheler *et al.* 2014). These
631 results are casting doubts on the accuracy of the genetic concept of populations in those partially clonal
632 organisms, suggesting the ecological concept of distribution continuity (therefore the meadow scale) may
633 be more accurate as a framework for connectivity analysis (Becheler *et al.* 2010) and genetic differentiation
634 may not systematically reflect lack of connectivity. Additionally, the general lack of isolation by distance
635 patterns reported across seagrass species was interpreted as a clue for scarce but significant events of large-
636 scale dispersal influencing the dynamics, resilience and geographic distribution of genetic diversity in
637 seagrasses (Kendrick *et al.* 2012). Whereas this may hold for short-lived species with a regular turn over and
638 propagule recruitment, the decomposition of the genetic distance spectrum (Rozenfeld *et al.* 2007) among
639 meadows of the long-lived seagrass *Posidonia oceanica* (Linnaeus) Delile, 1813 (Arnaud-Haond *et al.* 2012)
640 also brings additional warning for the interpretation of genetic structure patterns in terms of contemporary
641 gene flow. This approach of genetic data performed at the level of clonal lineages indeed showed that at the
642 range scale, the pattern of genetic structure was reflecting ancient events of (re)colonization followed by
643 long term clonal spread and accumulation of divergent mutations within meadows at the regional scale,
644 rather than contemporary patterns of connectivity (Arnaud-Haond *et al.* 2014). Interestingly, in haploid-
645 diploid species like many seaweeds, there is an alternation of haploid gametophytes and diploid
646 sporophytes to complete the sexual life cycle. In such life cycle, dominance of clonal propagation can led to
647 an uncoupling of haploid and diploid phases and thus to differences in allele frequency between
648 gametophytes and tetrasporophytes (Sosa *et al.* 1998). This has been observed in species of red alga

649 reproducing mainly asexually (in *Gelidium arbusculum* Bory de Saint-Vincent ex Børgesen, 1927; Sosa *et al.*
650 1998; *Gracilaria chilensis* C.J.Bird, McLachlan & E.C.Oliveira, 1986; Guillemin *et al.* 2008) while the
651 occurrence of sex, even occasionally, in other species, explained the lack of significant differences between
652 haploid and diploid subpopulations (*Gelidium canariensis* (Grunow) Seoane Camba in Sosa & Garcia-Reina
653 1993; *Gracilaria gracilis* (Stackhouse) M.Steentoft, L.M.Irvine & W.F.Farnham 1995 in Engel *et al.* 2004;
654 *Cladophoropsis membranacea* (Hofman Bang ex C.Agardh) Børgesen, 1905 in van der Strate *et al.* 2002 and
655 *Chondrus crispus* Stackhouse, 1797 in Krueger *et al.* 2011). Genotyping both haploid and diploid individuals
656 in these species is necessary in order to detect the clonality and its effect on population structure.

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657 **Box 5. Large taxonomic and geographic sampling in the deep-sea gastropod *Bursa* Röding, 1798 helps**
658 **delineating species before testing hypotheses on modes of speciation.**

659 Deep-sea seamounts offer a multi-scale fragmented habitat for benthic organisms. For gastropods
660 inhabiting these environments, connectivity patterns have been shown to widely depend on the type of
661 larval development (Castelin *et al.* 2010; Castelin *et al.* 2012). Notably, the three deep-sea Bursidae species
662 (*Bursa latitudo* Garrard, 1961, *Bursa fijiensis* (Watson, 1881) and *Bursa quirihorai* Beu, 1987) share a largely
663 overlapping geographic range on the seamounts of the Norfolk and the Lord Howe ridges, and the
664 continental slopes of New Caledonia, in the Southwest Pacific, and are not genetically structured over this
665 area.

666 The prevalence of allopatric versus sympatric modes of speciation, were studied following Coyne & Orr
667 (2004)'s biogeographic approach that involves studying current distribution patterns and phylogenetics of
668 recently-separated sister species. In this approach, one of the first requirements is to show that the species
669 pairs have a sister relationship and that reproductive isolation is complete (i.e. absence of hybridization
670 between the sister-species). All Bursinae species have a long-lived planktonic-feeding larva and most of
671 them have a wide distribution range. Moreover, most are geographically rare and the distribution of
672 populations within the species range is usually sparse and highly fragmented. This leads to confusions in
673 Bursidae taxonomy with some geographically-isolated populations attributed to distinct species names, or
674 slightly distinct morphotypes collected at distant but not geographically isolated sites attributed to the
675 same species name. The sampling strategy was thus designed to meet a three-fold objective: extending the
676 taxonomic sampling without any *a priori* hypotheses about the systematics of the group, at both the genus
677 and the species level (e.g. including all available individuals, populations, species and genera of the family
678 Bursidae); extending the geographic sampling toward other oceans (Atlantic, Indian); and extending the
679 ecological sampling by adding species from both shallow-water and deep-sea environments.

680 Starting from this large taxon sampling, "Primary Species Hypotheses" (PSH) were drawn from the shell
681 morphology using the taxonomic literature and dry material of the Malacology collection at the French
682 National Museum of Natural History (MNHN). ESUs were then defined using both the GMYC (Pons *et al.*
683 2006) and ABGD methods (Puillandre *et al.* 2012b) applied on *cox1* sequences and then strengthened with
684 the analysis of a second mitochondrial gene (16S rRNA) and the nuclear 28S rRNA gene fragment. All
685 sources of evidence generated from these four steps were integrated to propose "Retained Secondary
686 Species Hypotheses" (RSSHs).

687 Over the sampled geographical range, the three morphologically diagnosable entities, *B. latitudo*, *B.*
688 *quirihorai* and *B. fijiensis*, are reciprocally monophyletic based on all tested loci. The dataset also revealed
689 cryptic lineages emphasizing that species diversity in the family Bursidae is underestimated and suggesting
690 that a taxonomic revision of the family should be performed. Among New Caledonian species, *B. fijiensis*

691 and *B. quirihorai* are reciprocally monophyletic. These two species are the two most closely related species
692 in the inferred phylogeny. The current biogeographic ranges of these two species and the estimated time of
693 divergence make the scenario of sympatric speciation the most likely.

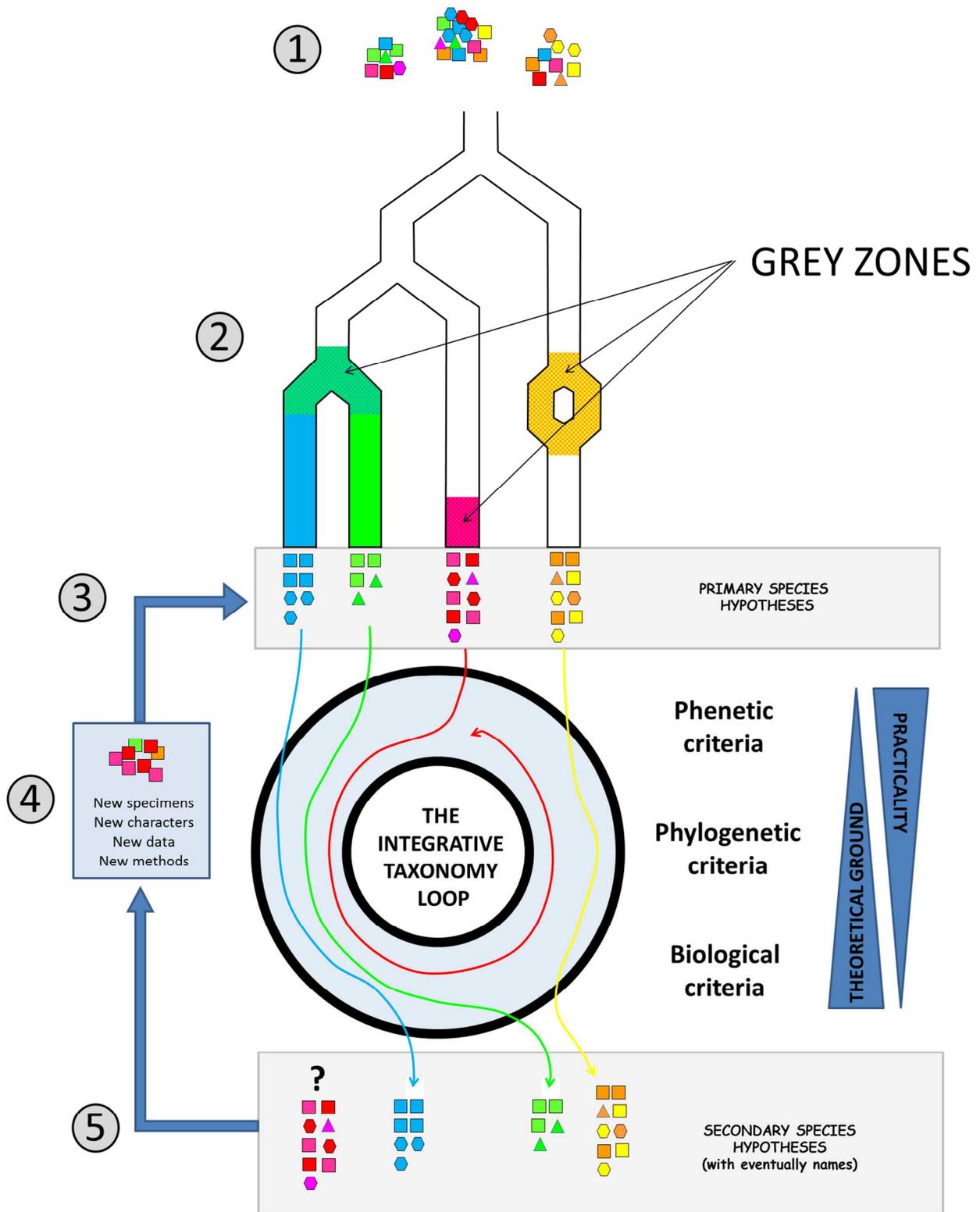
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694 **Box 6: Human-mediated transport may further increase complexity: from cryptogenic to cryptic species in**
695 **marine invaders - delineating species in a changing world.**

696 Besides the natural processes of expansion and contraction of the range colonized by a species over
697 evolutionary times, sudden species-range expansions -often through human-mediated transport- have been
698 observed at an increasing rate since the end of the 19th century. Since the original work by Elton published
699 in 1958 (Elton 1958), biological invasions have received a growing set of attention from the scientific
700 community (e.g. Blackburn *et al.* 2011 and Simberloff *et al.* 2013 for recent reviews and references herein).
701 Tightly linked to human activities particularly through shipping and aquaculture, biological invasions
702 severely alter biodiversity of marine ecosystems (Rilov & Crooks 2009). One of the important outcomes of
703 marine biological invasion studies has been to document biotic homogenization at a global scale with the
704 establishment of 'cosmopolitan' species and cryptogenic species (i.e. species that are neither demonstrably
705 native nor demonstrably introduced as defined by Carlton (1996; Haydar 2012). Interestingly, many of these
706 cosmopolitan invasive species actually appear to be composed of morphologically close taxa forming a
707 species complex. This was recently exemplified in two model ascidians, the colonial tunicate *Botryllus*
708 *schlosseri* (Bock *et al.* 2012) and the vase tunicate *Ciona intestinalis* (Nydam *et al.* 2011; Zhan *et al.* 2010).
709 Both taxa are distributed worldwide. Phylogeography, phylogeny, population genetics and genomic studies
710 showed for both taxa that they were actually characterized by marked genetic subdivision and large
711 genealogical divergence (up to 4-8 million years for type A and type B in *C. intestinalis*, Roux *et al.* 2013).
712 These two taxa are composed of (partially) reproductively isolated species, some of them came recently
713 into secondary contact because of their introduction with human activities. This is notably the case for two
714 members of the *Ciona intestinalis* species complex called type A and type B, which are living in sympatry
715 after the presumably recent introduction of Type A in the Northern Atlantic; the two species are
716 incompletely reproductively isolated in the English Channel where they are hybridizing (Nydam & Harrison
717 2011). The outcome of such secondary contact is unknown but may promote either adaptive introgression
718 or speciation (Abbott *et al.* 2013). Bock *et al.* (2012) also pointed out that the different cryptic species of the
719 *Botryllus schlosseri* complex are not characterized by similar invasive abilities questioning the reasons for
720 such differential abilities. Integrative taxonomy and molecular approaches are thus critical research to carry
721 out in biological invasions. They may help identify new cryptic introduced species with different invasion
722 abilities, understand the evolutionary history of cryptogenic species (e.g. the tunicate *Molgula manhattensis*
723 *manhattensis* (De Kay, 1843) (Haydar *et al.* 2011) and the fate of secondary contact between previously
724 isolated taxa. For management purpose, they are also helpful to identify new invaders (Bishop *et al.* 2013)
725 and verify the accuracy of published records of invasive species (e.g. McGlashan *et al.* 2008) - especially for
726 morphologically cryptic cosmopolitan taxa.

727 **Figure1: the integrative taxonomy loop**

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731 **Box figure 1.**
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735 **Box figure 2.**

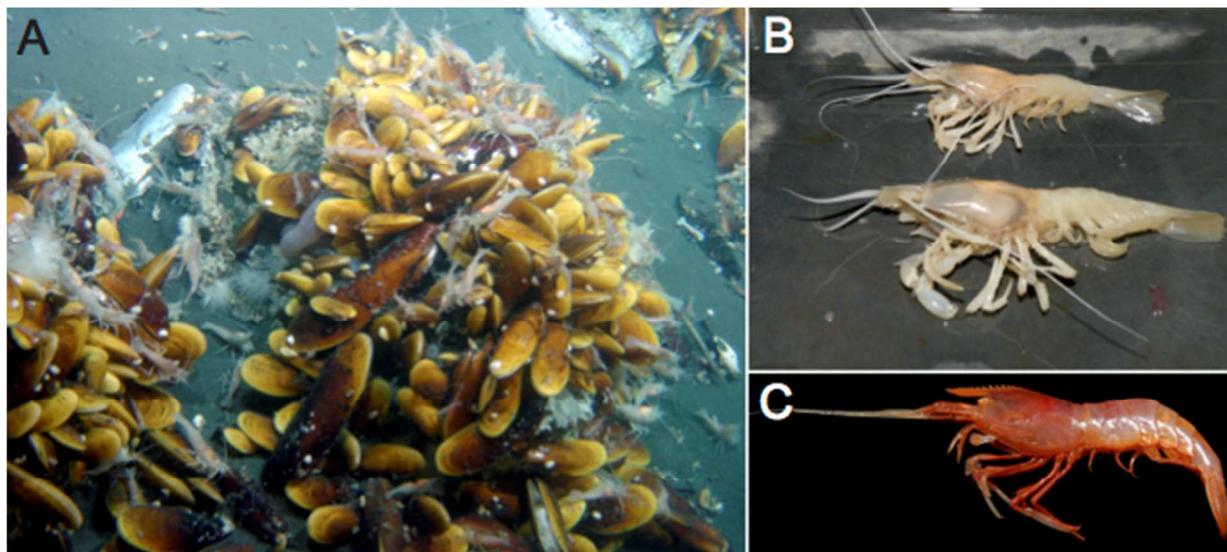
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739 **Box figure 3.**

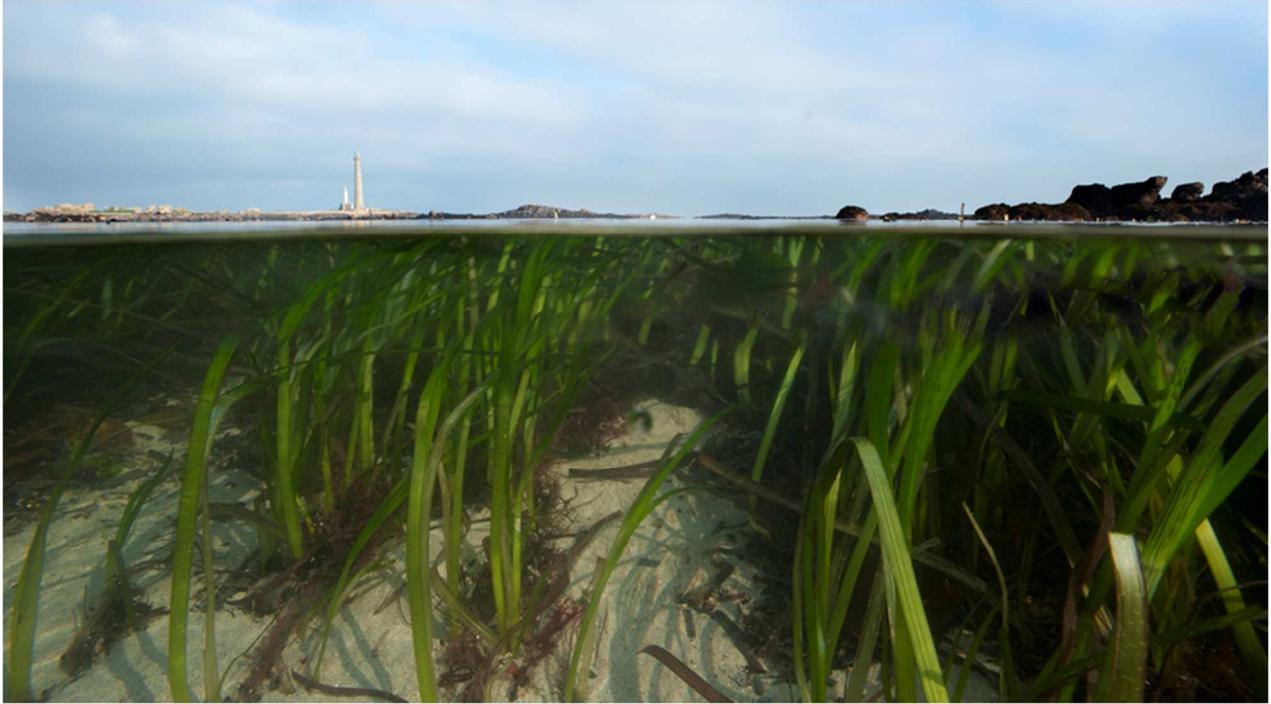
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743 **Box figure 4.**
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747 **Box figure 5.**
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751 **Box figure 6.**
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755 **Figure captions**

756

757 **Figure 1: The integrative taxonomy loop.** Integrative taxonomy consists in analyzing different characters,
 758 with different methods, and applying different criteria of species delimitation to propose species
 759 hypotheses that are as robust as possible. The different families of criteria (as discussed in Samadi &
 760 Barberousse 2014) are listed right of the loop, with the more theoretically grounded on the bottom, and the
 761 more operational (i.e. easy to test practically) on the top. Within each family of criteria different kind of
 762 characters (i.e. morphology, ethology, ecology, biochemical, genetic, etc...) and methods (i.e. distances,
 763 maximum parsimony, maximum likelihood, population genetics inferences, crossing experiments,
 764 observations, etc...) may be applied. The different steps are as follows. 1. Populations (putatively within-
 765 species) and phylogenetic (putative sister-species) sampling. 2. Sampled species may be highly
 766 differentiated (blue and green), recently diverged species that are still in a “grey zone” (see Glossary) with
 767 most characters undifferentiated (pink and red), or a single species that went through a temporal split into
 768 several temporary lineages (yellow/orange). 3. Primary Species Hypotheses (PSH) are proposed. 4. PSH are
 769 engaged in the integrative taxonomy loop and are evaluated, possibly with the addition of new material,
 770 using different criteria for species delimitation (the more theoretically-grounded biological criteria being
 771 tested directly using cross experiments or indirectly with unlinked markers, and complemented with more
 772 operational (i.e. practical) criteria). 5. When possible, taxonomic decisions are taken (PSH are turned in
 773 Secondary Species Hypotheses - SSH) and are named. Some lineages (i.e. the pink/red lineage) may stay in
 774 the loop, needing more conclusive data before being turned into SSHs. Most of the literature and methods
 775 for species delimitation focus on species that are currently in the grey zone (cf. Carstens *et al.* 2013), even
 776 though most delimitation cases fall outside of this range.

777

778 **Box figure 1. Species delineation in *Fucus*.** A: Vertical distribution of furoid species on rocky shore from
 779 high to low-water mark. Site: Viana do Castelo (GPS coordinates: 41° 41' 51''N, 8° 51' 07'' W, northern
 780 Portugal), photograph: Christophe Destombe. B: Detail of reproductive structure of *Fucus guiryi* showing a
 781 typical receptacle sterile rim (arrow). Photograph: Christophe Destombe.

782

783 **Box figure 2. The pseudocryptic kelp species *Lessonia spicata*.** A: This species (on the left of the picture) is
 784 found in central-south Chile (29°-41°S) in areas exposed to wave actions. Sporophyte individuals can be
 785 more than 2 meters long and are anchored to the rock by a holdfast. Site Los Molles (GPS coordinates:
 786 32°14'S-71°31'W, central Chile), photograph: Pablo Balzo. B: The gametophytes of this species are
 787 microscopic and show a sexual dimorphism with males being smaller and more branched than females.
 788 Photograph: Valeria Oppliger.

789

790 **Box figure 3. The deep-sea shrimp *Alvinocaris muricola* and *A. markensis*.** *A. Alvinocaris muricola* in situ,
791 on a mussel bed in in Western African cold seeps. (c) Ifremer, ROV Victor 6000, cruise Biozaïre. B-C.
792 specimens of *A. muricola* (B) and *A. markensis* (c) sampled respectively on Western African cold seeps
793 (cruise WACS, photograph Dominique Cowart) and Mid Atlantic Ridge hydrothermal vents (cruise BICOSE,
794 photograph Laure Corbari) (c) Ifremer.

795

796 **Box figure 4. *Zostera marina* meadows in Brittany.** Individual shoots (ramets) are easily distinguished while
797 rhizomatic connections linking ramets belonging to the same clonal lineage (genet) are hidden below the
798 sandy sédiment (photograph: Ifremer/Olivier Dugornay).

799

800 **Box figure 5. The deep-sea gastropod *Bursa*.** A specimen of *Bursa sp.* sampled in New Caledonia using a
801 dredge. The other organisms sampled in the same dredge are typical of hard bottoms (e.g. stylasterids,
802 deep-water solitary stony coral, etc...). Photograph MNHN/Pierre Lozouet.

803

804 **Box figure 6. The ascidian *Ciona*.** *C. intestinalis* type A is established in the English Channel, where it shares
805 habitats with its congener *C. intestinalis* type B. Photograph: Wilfried Thomas.

806

807

808

809 **Glossary.**

810 **Biological nomenclature:** set of rules that determine how names are given to taxa. In the binominal system,
811 when a new species is described based on a given set of biological criteria applied over a set of
812 specimens, one of them is designated as the holotype (the name-bearing specimen). When the
813 taxonomy is revised using new material (additional specimens) and/or new biological criteria the
814 position of the name-bearing specimens corresponding to the available names for the studied taxa
815 is used to name the new taxonomic hypotheses. When it is not possible to study the holotype for
816 the new character being proposed (e.g. molecular data), other categories of type material might be
817 used (paratypes, syntypes, etc...)

818 **Cryptic species: (see also pseudo-cryptic species):** two or more species lacking (obvious) morphological
819 diagnostic features that are classified as a single nominal species; it is the opposite situation as
820 compared to synonyms for which different scientific names are given to the same species.

821 **DNA barcoding:** identification of an unknown specimen by comparison of a standard DNA fragment (e.g.
822 Barcode fragment of the *cox1* gene for metazoans) to a database of sequences linked to identified
823 vouchers. By extension, use of a standard DNA fragment for biodiversity studies (e.g. to discover
824 new species; see Collins & Cruickshank (2013) for a distinction between “specimen identification”
825 and “species discovery and delimitation”).

826 **Divergence continuum:** Refers to the observation that differentiation between two emerging lineages
827 occurs through a series of changes that accumulate during the speciation process rather than
828 abrupt genetic changes. For a recent genomic example, see Lexer *et al.* (2014).

829 **Evolutionary significant unit (ESU):** defined in 1994 by Moritz to explicitly take into account the genetic
830 component of biodiversity in conservation, ESU is defining historically isolated groups of
831 populations showing reciprocal monophyly and significant genetic divergence at nuclear loci.

832 **Grey zone:** temporal zone of the genealogical network during which two lineages are definitively diverging,
833 but the criteria used for identifying divergence might not be applicable or in agreement (see in de
834 Queiroz 1998). During this period of time, characters might differentiate at a different pace, leading
835 to incompatible species hypotheses and justifying the need for integrative taxonomy.

836 **Integrative taxonomy:** species delimitation process in which species-hypotheses are proposed based on the
837 integration of several lines of evidence (characters, criteria, methods) in a reproducible and
838 falsifiable framework.

839 **Phylogeography:** the spatial distribution of phylogenetic lineages, particularly within and among closely
840 related species (Avice 2009).

841 **Population connectivity:** Demographic- and evolutionary-dependence among isolated patches of
842 individuals from a given species produced by the movement of individuals among patches.

843 **Pseudocryptic species:** morphologically overlooked species distinguished using other lines of evidence (e.g.
844 DNA) that proved to be finally distinguished by subtle morphological differences (e.g. Box 2 on the
845 kelp *Lessonia*).

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