Marine sponges as a powerful tool for trace elements biomonitoring studies in coastal environment

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In this work, we performed a comparative study on six marine sponge species collected along the French Mediterranean and Irish coasts for their TEs accumulation. Intra and inter-species variabilities were examined. Among the Mediterranean species, *Cymbaxinella damicornis* accumulates significantly more As and Cu than others sponge species; *Chondrilla nucula* more Ni and Mo and *Acanthella acuta* more Ag. Among Irish samples, *Hymeniacidon perlevis* showed higher accumulation properties for most of TEs in comparison to *Halichondria panicea*. Bioconcentration Factors were > 1 in all species for most of TEs. This study suggests that TEs bioaccumulation is most likely associated to differences in morphological features and/or to specific bacterial communities associated to different species. The determination of Pb isotope ratios revealed mainly natural Pb sources for Mediterranean and Kilkieran Bay's samples, and rather anthropogenic influence for Belfast samples. This study confirms that sponges represent a powerful tool for biomonitoring studies.

1. Introduction

The occurrence of trace elements (TEs) in aquatic environments is related to natural sources, but also to anthropogenic inputs from industrial activities. TE contaminations in the marine environment occur worldwide and the assessment of the pollution status is required, leading to the publication of several studies on this subject (Li et al., 2000; Caccia et al., 2003; Fang et al., 2014; Parra et al., 2015; Cabrita et al., 2017; Ashraf et al., 2017). Environmental surveys are often performed on water samples but, in marine ecosystems, the very low TEs concentrations and the high salinity of seawater cause severe limitations in the measurements (Søndergaard et al., 2015). Monitoring studies on TEs contamination are often focused on sediments and biota. Indeed, sediments act as sink and reservoirs of contaminants thus displaying higher TEs contents and not presenting problems related to short residence time such as for water (Varol, 2011; Liu et al., 2015). Marine organisms find also application as tools for characterizing the state of a marine ecosystem as some of them are recognized to accumulate TEs. They represent a good alternative especially in rocky shores where sediments are not easily found. Sessile and active filter feeding invertebrates are often used to determine temporal and spatial variation of TE in aquatic systems (Weis and Weis, 1999; Denton et al., 2006; Dahms et al., 2014). An ideal biomonitor should be a natural

accumulator of the element of interest and possibly with high concentration factor, in order to highlight possible difference between times or sites (Rainbow, 2002).

Historically, bivalves are considered as ideal biomonitor organisms as exemplified by the “mussel watch program” (Cantillo, 1998) also for their importance as widespread consumed seafood. More recently, sponges gained attention as possible biomonitors of TEs contamination as they are widespread, abundant in some ecosystems, long-living, easily sampled, sessile filter-feeders and highly tolerant to several pollutants (de Mestre et al., 2012; Batista et al., 2014). Additionally, sponge communities can remain stable for long periods, making them suitable model organisms for monitoring studies. Sponges pump large quantities of seawater ($100\text{--}1200\text{ mL h}^{-1}\text{ g}^{-1}$), more than any other marine invertebrates (Olesen and Weeks, 1994). These organisms have a high capacity for TEs accumulation (Patel et al., 1985; Padovan et al., 2012), exhibiting higher filtering capacity than bivalves (Negri et al., 2006; Gentric et al., 2016). They are considered as “biological particle traps” due to their ability to concentrate a wide range of chemicals from both the suspended and dissolved phases of the water, with usually high bioconcentration factors (BCF) (Perez et al., 2005; Cebrian et al., 2003). The detection of large quantities of TEs in sponges suggests either a high tolerance to diverse type of chemicals or the existence of detoxification systems (Aly et al., 2013). These characteristics make them

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suitable as a model organisms applied in TEs monitoring projects, and their use has been proposed in several studies (Negri et al., 2006; Venkateswara Rao et al., 2009; Illuminati et al., 2016). Sponges have also been reported as efficient environmental bioremediators of different chemicals or even microbes such as the organochlorine pesticide lindane (Aresta et al., 2015) and the bacterium *Escherichia coli* (Milanese et al., 2003). These characteristics, combined with a demonstrated interest for sponges as a source of novel pharmaceuticals (Anjum et al., 2016), open the way for self-financing sponge remediation programs (Gifford et al., 2007).

The main objective of this work was the assessment of TEs pollution in several sponge species of different morphologies and collected in different areas. A comparative study on TEs bioaccumulation was performed on six sponge species collected on the Southern French Mediterranean coast and in Ireland. These areas have been selected because of their differences in terms of TEs biogeochemical background and anthropogenic inputs. The bioaccumulation potential of these sponges was examined for a broad range of TEs, comparing with values measured in sediment samples collected in the same area. Additionally, measurements of Pb isotope ratios were performed for the first time in sponges and allowed us to give insights on the principal sources of Pb contamination in the studied zones.

2. Materials and methods

2.1. Sampling

Sponge and sediment samples were collected in three sampling sites: the Mediterranean coast, in the Grotte du Lido at Villefranche sur Mer, France (N 43° 41' 59.0", E 07° 19' 31.5") and two coastal sites in Ireland, Greenisland in the Belfast Lough (N54°41'24.383", O 5° 51' 36.633") and Kilkieran Bay (N 53° 21' 22.625", O 9° 42' 17.153") in Galway county (Fig. 1). Sediment samples were collected during each sampling session, using an acid pre-cleaned polyethylene tube. Sponges were collected using a plastic knife and placed in separate plastic bag. Different sponge species were sampled: *Acanthella acuta*, *Cymbaxinella*

damicornis, *Chondrilla nucula* and *Haliclona fulva* in the Mediterranean coast; *Halichondria panicea* and *Hymeniacidon perlevis* in Ireland. A list of samples with details on morphology and known bacterial associated population is presented in Table 1. These sponge species were described in details in a recently published paper (Orani et al., 2018). At least three specimens of similar size were randomly selected for each sponge species and during each sampling. Sediments and four species of Mediterranean sponges were collected by SCUBA diving at 20 m depth on February and September 2016 (winter and summer). During the first sampling, three specimens of *C. damicornis* were collected deeper in the cave while the remaining two specimens were from the entrance. During the second sampling, three specimens of *C. nucula* were collected at 5 m depth while three other specimens at 20 m. In Ireland, sediments and sponges were hand-sampled in intertidal zones. Samples from Kilkieran Bay and from Belfast were collected in September 2016 and in October 2016, respectively.

2.2. Sample preparation

All solutions were prepared with double deionized water obtained from Millipore water purification system (Elix & Synergy) (resistivity of $18.2 \text{ M}\Omega \text{ cm}^{-1}$, Total Organic Carbon $< 5 \mu\text{g L}^{-1}$ and microorganisms $< 0.1 \text{ UFC mL}^{-1}$). All PTFE and Teflon containers used for sample preparation and analyses were pre-cleaned using a procedure consisting of 24 h bath in 10% HNO_3 (prepared from proper dilution of 65% analytical grade HNO_3 , Fisher Chemicals) and careful rinsing three times with Milli-Q water. Sediment and sponge samples were freeze-dried, ground in agate mortar, and kept in closed PTFE tubes placed in a desiccator.

For trace elements analysis by ICP-MS, a microwave digestion was carried out into a closed microwave system (Ethos One, Milestone). Teflon reactors were filled with an aliquot of about 50 mg of sponge sample (or 100 mg of bulk sediment sample), with 5 mL of HNO_3 (Trace Metal Grade, 67 to 70% w/w, Fisher Chemical) and 2 mL of HF (Ultra Trace Elemental Analysis 47–51% w/w, Optima, Fisher Chemical). Temperature program used for acid digestion consisted of a 20-min

Fig. 1. Maps showing the sampling sites (indicated with black stars) located a) in the French Mediterranean coast (http://d-maps.com/carte.php?num_car=110470&lang=en) and b) in two locations along the Irish coast (http://d-maps.com/carte.php?num_car=14642&lang=en). Copyright d-maps.com

Table 1

List of specimens collected and analyzed in the present study with details on different morphological characteristics and microbial abundances. LMA stands for Low Microbial Abundance and HMA for High Microbial Abundance (Vacelet and Donadey, 1977).

Sponge specie	Shape	Skeleton	Microbial abundance	Number of specimens	Code	Sampling site
<i>Acanthella acuta</i>	Branch erected	Spongin and dense spicules	LMA	6	AC.A	Villefranche-sur-Mer
<i>Cymbaxinella damicornis</i>	Branch erected	Spicules	LMA	7	CY.D	Villefranche-sur-Mer
<i>Haliclona fulva</i>	Thick encrusting	Spicules	LMA	6	HA.F	Villefranche-sur-Mer
<i>Chondrilla nucula</i>	Thick encrusting	Dense collagen and spicules	HMA (cyanobacteria)	9	CH.N	Villefranche-sur-Mer
<i>Halichondria panicea</i>	Massive	Dense spicules, scarce spongin	LMA	4	HA.P	Kilkieran Bay
<i>Hymeniacidon perlevis</i>	Thick encrusting	Spicules, scarce spongin	HMA (cyanobacteria)	6	HA.P Bel	Belfast Lough
				3	HY-P	Kilkieran Bay
				7	HY-P Bel	Belfast Lough

Table 2

Comparison of measured concentrations and certified values in 2 CRMs: TORT-2 (lobster hepatopancreas) and MESS-2 (estuarine sediments).

		Measured value (mg kg ⁻¹)	Relative Standard Deviation (%)	Recovery (%)	Certified value (mg kg ⁻¹)
TORT-2 (n = 1-0)	V	2.2 ± 0.56	25	135	1.64 ± 0.15
	Ni	2.7 ± 0.7	25	106	2.5 ± 0.19
	Cu	97 ± 6	6	91	106 ± 10
	Zn	205 ± 36	17	114	180 ± 6
	As	19.8 ± 3.5	18	92	21.6 ± 1.8
	Mo	0.94 ± 0.21	22	99	0.95 ± 0.1
	Ag	6.1 ± 0.4	6		
	Cd	25.7 ± 1.7	6	96	26.7 ± 0.6
	Ba	2.6 ± 0.8	30		
	Pb	0.35 ± 0.20	57	100	0.35 ± 0.13
MESS-2 (n = 4)	V	234 ± 15	6	93	252 ± 10
	Ni	46 ± 2	4	94	49.3 ± 1.4
	Cu	37 ± 3	8	95	39.3 ± 2.0
	Zn	155 ± 12	8	90	172 ± 16
	As	19 ± 1	8	92	20.7 ± 0.8
	Mo	2.9 ± 0.8	27	101	2.85 ± 0.12
	Ba	794 ± 183	23		
	Pb	20 ± 4	23	91	21.9 ± 1.2

ramp temperature up to 180 °C, followed by an isothermal phase of 30 min. The mineralized solutions were then transferred to polyethylene pre-cleaned tubes and gravimetrically diluted with Milli-Q water, up to a final weight of about 50 g. These solutions were further diluted 15 times in Milli-Q water prior to ICP-MS analyses. The same sample preparation and analyses were performed on two Certified Reference Materials (CRMs) to ensure quality control of the entire analytical procedure. The used materials were TORT-2 (Lobster hepatopancreas, NRC, Canada) and MESS-2 (Estuarine sediment, NRC, Canada); 50 and 100 mg of these CRMs were used respectively. At least, one procedural blank and one CRM were included in each digestion run and analyzed with the rest of the samples. A summary of results obtained for CRMs analyzed in this study is presented in Table 2; and in all cases a good agreement with certified values was found. Cd and Ag in MESS-2 were below the limits of quantification, therefore they are not listed in the table with other analytes.

One specimen of each sponge species and one sediment sample for each location were selected for Pb isotope ratios analysis. An aliquot of about 100 mg of sponge or sediment sample was decomposed in 5 mL ultrapure HNO₃ (Fisher scientific, Hampton, USA) and 2 mL of ultrapure HF (Fisher scientific, Hampton, USA) using the Mars X-press Microwave (CEM Mars X-press) equipped with a 12 vessels carousel. After the digestion, the solutions were placed on a ceramic heating plate, under laminar hood, and concentrated to near dryness. The residues were taken up in 2% HNO₃, transferred to 50 mL PE tubes and stored at 4 °C. Solutions for Pb isotope ratios determination were diluted (at least 50 times) with 2% HNO₃ in class 100 clean room.

Table 3

(Q) ICP-MS settings used for the determination of trace metals in sediments and sponges and limits of detection.

Parameter	Value
RF power	1550 W
Plasma gas flow	15 L min ⁻¹
Auxiliary gas flow	1.1 mL min ⁻¹
Carrier gas flow	0.9 L min ⁻¹
Nebulizer	Quartz concentric
Spray chamber	Cyclonic
Interface	Pt sampling and skimmer cones
Oxide formation ¹⁴⁰ Ce ¹⁶ O ⁺ / ¹⁴⁰ Ce ⁺	< 3%
Double charged species	< 2%
Measured isotopes	⁵¹ V, ⁶⁰ Ni, ⁶³ Cu, ⁶⁶ Zn, ⁷⁵ As, ⁹⁸ Mo, ¹⁰⁷ Ag, ¹¹¹ Cd, ¹³⁸ Ba, ²⁰⁸ Pb
Mass for interference assessment	⁴⁰ Ar ³⁷ Cl, ⁸² Se, ⁹⁵ Mo ¹⁶ O, ³⁷ Cl ¹⁶ O
Limits of detection (µg L ⁻¹)	
V = 0.02	Zn = 0.3
Ni = 0.03	Cu = 0.008
As = 0.03	Mo = 0.002
Ag = 0.002	Cd = 0.007
Ba = 0.006	Pb = 0.002

2.3. Analyses and data treatments

A quadrupole Inductively Coupled Plasma Mass Spectrometer ((Q) ICP-MS; Elan DRCII, Perkin Elmer) was used for the determination of Ag, As, Cd, Cu, Mo, Ni, Pb, Ba and Zn in sponge and sediment samples. ICP-MS operating parameters are summarized in Table 3. Analytes quantification was performed by means of external calibration, using daily prepared standards obtained by proper dilution of multi-elemental standard for ICP-MS (ICP-MS standard N°3 Perkin Elmer® and ICPMS standard N°2 from SCP SCIENCE®). An internal standard solution, containing 10 and 1 µg L⁻¹ of Ge and Tb respectively, was prepared by dilution of mono-elemental standards of Ge and Tb (PlasmaCal, SCP SCIENCE®) and used during the ICP-MS analysis to correct for possible matrix effects. Daily analyses of the certified reference natural river water SLRS-5 or SLRS-6 (National Research Council, Canada) were carried out to check measurement accuracy (Barats et al., 2014). All results were corrected for instrumental blanks. The ongoing instrument performance was monitored with the repeated analysis of standards. Detection limits for analytes are also reported in Table 3.

Determination of Pb isotope ratios was carried out by means of High Resolution Sector Field (HR-SF) ICP-MS (NuAttom, NuInstruments Ltd., Wrexham, UK). The Standard Reference Material SRM 981 from the National Institute of Standards and Technology (NIST, USA) with certified Pb isotopic composition was used for correction of the instrumental mass discrimination. The bracketing approach based on linear law was used for correction (Vanhaecke et al., 2009). The instrument parameters for Pb isotopic ratios measurements with (HR-SF) ICP-MS are presented in Table 4. The ²⁰⁴Hg isobaric interference on ²⁰⁴Pb isotope was corrected by monitoring the mass ²⁰²Hg and applying a correction equation based on natural abundancy of isotopes ²⁰²Hg, ²⁰⁴Hg and ²⁰⁴Pb.

Table 4

(HR-SF) ICP-MS parameters optimized for the determination of Pb isotope ratios in sponges and sediment samples.

Parameter	Value
Coolant gas flow	13.5 L.min ⁻¹
Auxiliary gas flow	0.8 L.min ⁻¹
Nebulizer gas flow	29.6 psi
Plasma power	1300 W
Sensitivity for 1 ng g ⁻¹ (²⁰⁸ Pb)	> 800,000 cps
Background on mass 220	< 2 cps
Acquisition Mode	Peak Jumping
Number of sweeps	500
Number of cycles	25
Dwell time per isotope (ms)	1000
Measured isotopes	²⁰² Hg, ²⁰⁴ Pb, ²⁰⁶ Pb, ²⁰⁷ Pb, ²⁰⁸ Pb
Mass resolution	300
Oxide Formation ¹⁴⁰ Ce ¹⁶ O ⁺ / ¹⁴⁰ Ce ⁺	< 1%

Statistical data treatments were performed using XLSTAT (version 2014.05.5, Addinsoft, Paris, France). Principal component analysis (PCA) was applied to TE concentrations measured in sponges in order to highlight common trends between different species. Data were standardized using the biased standard deviation, to avoid biases due to differences in data dimensionality. Lead was not included in the PCA due to its high intra-species variability and it will be treated in a separated paragraph.

2.4. The use of lead isotope ratios

Lead isotope ratios measurement represents a very powerful analytical tool applied to several fields of study. Pb has a partially-radioactive isotopic composition, since three of its stable isotopes (²⁰⁶Pb, ²⁰⁷Pb and ²⁰⁸Pb) are the final product of the radioactive decay of ²³⁵U, ²³²Th and ²³⁸U respectively (Vanhaecke and Degryse, 2012). ²⁰⁴Pb is the only natural isotope and its abundance has remained constant since the formation of the solar system (Komárek et al., 2008). Thanks to this characteristic, Pb isotope composition is governed by the original composition of the parent isotopes, and thus is typical and characteristic of a given area/sample. Pb isotope ratios are a powerful tool often used for provenance studies in several fields, such as forensic science and cultural heritage investigations (Sjåstad et al., 2011; Drivelos and Georgiou, 2012; Ortega et al., 2012). The application of Pb isotope ratios measurement in environmental science is of great interest since it allows to distinguish between different Pb sources, whether they are natural or anthropogenic (Hinrichs et al., 2002; Larsen et al., 2012).

3. Results and discussion

3.1. TE concentrations in sponges

Average TE concentrations according to sampling site, date and sponge species are summarized in Tables 1S and 2S (supplementary information). Accounting for the intra-species variability, it was possible to evaluate the differences in TE bioaccumulation for different sampling conditions (depth, light, and seasons) or between different species of the same sampling site. Additionally, two species *H. panicea* and *H. perlevis* were collected in two different sampling sites around Ireland, making possible a direct comparison of these species bioaccumulation trends with respect to TE availability in the surrounding environment.

3.1.1. Intra-species variability

The variability of TE bioaccumulation within a same sponge species can be evaluated with the relative standard deviation (RSD%) calculated from different specimens of the same sponge species. In Mediterranean sponges, a variability of TE concentrations was noticed

depending on the target TE and sponge species, with an average RSD of 30%. Among TEs, Mo, As, Cd and Cu showed the lowest variabilities (averaging 21, 15, 16, 17 and 21%, respectively). The intra-species variability can also be very low for some sponge species e.g. for Ba in *C. damicornis* (1%) or for As in *C. nucula* at 20 m depth (3%) during the second sampling. The average RSD values for V, Ni, Zn, Ag and Ba was 33%. For all species collected in the Mediterranean sites, Pb showed by far the highest intra-specific variability, showing an average RSD of 69% and reaching values up to 121% (e.g. for *H. fulva* for the second sampling).

In Irish sponges, the intra-specific variability ranges from 2 to 55% for all TEs (even for Pb), with average around 33%, such as in Mediterranean samples. The maximum RSD value found for Irish samples is 55% for Ba measured in *H. panicea* from Kilkieran Bay. Arsenic showed the lowest intra-species variability for *H. panicea* from Belfast (14%) and for *H. perlevis* from Kilkieran (8%). Mo and Zn presented the lowest RSD values in *H. panicea* from Kilkieran (2%) and *H. perlevis* from Belfast (7%), respectively. Pb did not show the same behavior as in the Mediterranean samples, since its intra-specific variability for all studied species was similar to values obtained for other TE.

In this study, the natural variability within the same sponge species seems to be dependent both on the considered analyte and the sponge species itself. The factor most likely governing this intra-specific variability can be related to possible differences in organism age. Besides, it seems that some elements like Pb are more affected by this natural internal variability than other TE such as Cd, Mo, and V. For the Atlantic and Mediterranean sponge samples, the natural intra-specific variability observed in this study had average values around 30–33%, which agrees with literature data on TEs in sponges. Müller et al. (1998) reported RSD up to 33% for Cd and Cu in *Suberites domuncula*. Perez et al. (2005) showed RSD up to 70% and 50% for Mn and Pb in *Spongia officinalis*, respectively. These previous results are in the same range of RSD found for Pb in Mediterranean sponges analyzed in the present study. A natural variability has thus to be expected, even for samples analyzed in pristine environment such as the Antarctic (Grotti et al., 2008). This natural variability is not always taken into consideration in biomonitoring studies and should be considered when interpreting results.

3.1.2. Influence of sampling site on the TE concentrations in Mediterranean sponges

During the same sampling date, specimens of *C. damicornis* and *C. nucula* were collected at different depths and light conditions to evaluate the influence of physical parameters on the TE bioaccumulation. Results are shown in Fig. 2 and Table 1S. Differences between TE concentrations were examined taking into consideration the intra-specific variability.

- Influence of the light: During the sample collection carried out in February 2016 (first sampling), three specimens of *C. damicornis* were collected at the entrance of a cave and two others inside the cave. Considering the intra-species variability, TE concentrations were similar in the two cave sites. Slight differences were found on the mean Ni and As concentrations, but not so significant when considering respective RSD. Ni average is $3 \pm 1 \text{ mg kg}^{-1}$ for sponges collected in the cave entrance and $5 \pm 1 \text{ mg kg}^{-1}$ for those collected inside the cave. Unlikely, As was found in higher concentration in samples from the cave entrance: 85 ± 15 vs $67 \pm 4 \text{ mg kg}^{-1}$.
- Influence of the depth: During the second sample collection, three samples of *C. nucula* were collected at 5 m depth and other three specimens at 20 m depth. TE concentrations were generally similar for the two sampling depths. Variability in Mo was borderline (2.1 ± 0.3 and $1.52 \pm 0.08 \text{ mg kg}^{-1}$, for 5 and 20 m depth respectively), and V and Cd concentrations were slightly higher in samples collected at a depth of 20 m than at 5 m. Slight differences

Fig. 2. Graphic representation of inter-species variability of trace element mass fractions for sponges collected in Mediterranean coast during the two sampling campaigns. Comparisons for *C. damicornis* collected at the entrance (E) and center (C) of the cave as well as for *C. nucula* collected at two different depths (5 and 20 m), are also presented. Error bars stand for standard deviations on mean TE values obtained for different specimens of same sponge specie ($n \geq 3$).

occur also on the mean Cu and As sponge contents, but not significant considering the intra-specific variability.

Overall, small changes in the sampling location and/or depth do not have a significant impact on TEs accumulation in sponges. These organisms can thus be considered representative of a rather spread “zone” of environment, showing consistent and homogeneous TEs distribution within the same sampling period.

- Influence of the sampling season

Mediterranean sponges were collected at the same site during two seasons to estimate the influence of temporal variations in TEs bioaccumulation. The average values obtained for each specie in each season were compared, and differences were evaluated taking into consideration the intra-specific natural variability observed for each pool of specimens. Results are shown in Table 1S.

Considering the intra-specific variability, TEs bioconcentrations were overall similar for the two sampling seasons, especially for *C. nucula* and *A. acuta*. However, for some of them, concentrations were slightly higher during the first sampling carried out in September 2016 (summer): for As in *A. acuta* sponges, for Mo and Pb in *C. nucula* sponges, for Ba, As, Mo and Ag in *C. damicornis*, and for V, Ni, Ag, Ba

and Pb in *H. fulva* sponges. Overall, most of TEs presented none or negligible seasonal fluctuations considering RSD values. If a common trend of TEs enrichment in all sponge species were observed for a sampling period, this could signify an external increased input (e.g. from anthropogenic activities). But this behavior was not observed in sediment samples, as presented in one of the following paragraph. These fluctuations are most likely associated with natural processes: e.g. linked to changes in biological activities of associated bacteria, physio-chemical changes in the water column affecting the uptake and elimination of TEs.

3.1.3. Inter-specific variability

- Mediterranean sponges

The differences in TEs contents between different species were first evaluated for Mediterranean sponges. Since four different species were collected for two different sampling campaigns, it was possible to carry out a comparative study on the different accumulation properties of these species (Fig. 2).

The accumulation of TEs within sponges depends on the considered element. For instance, As, Cu and Zn presented the highest contents in all species. Additionally, the accumulation of TEs strongly depends on

Fig. 3. Biplot representation of PCA performed on all mass fractions found in Mediterranean sponge specimens analyzed in this study.

Fig. 4. Graphic comparison between TE concentrations on *H. panicea* and *H. perlevis* collected in two sampling locations in Ireland.

the sponge species. *C. damicornis* showed about two times higher As and Cu concentrations than other species: between 56 and 61 and 55–85 mg kg^{-1} , respectively. The highest content of Ni and Mo were found in *C. nucula*, with values twice (or more) higher than for other species. Zn showed also higher content in *C. nucula*, but only for the first sampling. The lowest Cd and V concentrations were revealed in *C. nucula* sponges. In this study, Ba and Pb accumulation in sponges was not shown to be specific to a sponge species. This result can be also due to high intra-species variations and seasonal variations previously described. A peculiar behavior was observed for Ag in *A. acuta*, which showed content up to 100 times higher than those found in the other three species. It is worth noticing that *H. fulva* is the only sponge species which did not exhibit a specific affinity for any TE comparing to the other sponge species.

A statistical analysis was performed to better describe the inter-specific variations. The biplot obtained by Principal Component Analyses (PCA) is presented in Fig. 3. The first two components (F1 and F2) explained 59.6% of the total variance. F1 accounted for 42.2% of

total variance and 17.49% for F2. High positive loadings of V, Cu, As, Cd and Ba and high negative loadings of Ni and Mo characterized the F1 axis. The F2 axis was mainly characterized by a Ag positive loading. These observations and the analysis of Fig. 3 led to the conclusions that a positive correlation ($p < 0.05$) was found for V, Cu, As, Cd, Ba and for Mo and Ni; these two groups of elements being negatively correlated between them. Ag was the only element that did not present any significant correlation with other analytes and thus it is the only one with high positive loadings on F2. The biplot graph shows four distinct groups, each of them including all specimens of the same species. These statistical results confirm the first observations on TEs concentrations and that sponge species can be distinguished based on their TEs bioaccumulation. This biplot highlights: (1) that *C. damicornis* accumulates preferentially As and Cu; (2) *C. nucula* Mo and Ni; (3) and an exceptional affinity of *A. acuta* for Ag; (4) no specificity for *H. fulva* was highlighted. Since Mediterranean sponges were collected in the same site and at the same date, we can assume that they were exposed to the same amount of TEs. The great differences in TEs accumulation in these

Fig. 5. Biplot representation of PCA performed on mass fractions found in Irish sponges collected in a) Kilkieran bay and b) Belfast lough.

sponges it is thus related to particular characteristics of a sponge species.

- Irish sponges

The inter-species variability of TEs concentrations were then evaluated for two sponge species collected in two Irish sampling sites. The results are shown in Fig. 4 and Table 2S. Principal Component Analysis was also performed separately for each sampling site in Ireland to highlight unique properties of each species, not considering differences on element availability in the surrounding seawater. Results are presented in two separated biplots (Fig. 5).

In the quite pristine area of Kilkieran bay, *H. perlevis* sponges revealed higher TEs concentrations than those measured in *H. panicea*, except for As and Mo with similar contents. The PCA results gave two components (F1 and F2) with eigenvalues higher than one, which

explained the 81.6% of variance (Fig. 5 a). F1 was characterized by high positive loadings of all analytes except As, the only element presenting negative contribution to this component. The F2 presented the highest positive loading of Mo and the highest one of Cd, while others TE show low positive or negative contribution. The separation of sponge species in the biplot is evident, as *H. panicea* specimens are grouped in the negative part of F1 while *H. perlevis* are grouped on the opposite side. Even though the As contents in these two species were similar, this metalloid seems to unequivocally distinguish the two sponge species in the PCA. Additionally, it can be noted that Cu, Ag, Zn and Cd form a first group whereas Pb, Ni, Ba and V another one. TEs forming the first group close to the *H. perlevis* samples agrees with the observation of higher Cu, Ag, Zn and Cd contents in *H. perlevis* sponges and thus specific affinity for this sponge species.

At the more contaminated site of Belfast Lough, *H. perlevis* sponges

Table 5

Summary of TE concentrations found in all sediments samples analyzed in this study. Standard deviations were calculated considering different samples collected in the same sampling site. The n refers to the number of specimens analyzed for each sponge specie.

Concentrations in	Sampling stations		
sediments (mg kg ⁻¹)	Villefranche (n = 6)	Killkieran (n = 3)	Belfast (n = 3)
V	9.5 ± 0.3	47 ± 1	59 ± 9
Ni	4.9 ± 0.2	10.9 ± 0.7	22 ± 3
Cu	3 ± 0.7	9 ± 5	7 ± 1
Zn	21 ± 2	49 ± 11	63 ± 9
As	7 ± 1	7 ± 2	2.9 ± 0.3
Mo	0.3 ± 0.1	0.35 ± 0.07	0.12 ± 0.02
Ag	0.17 ± 0.01	0.14 ± 0.08	0.10 ± 0.06
Cd	0.5 ± 0.4	0.08 ± 0.03	0.26 ± 0.03
Ba	10 ± 2	151 ± 24	142 ± 13
Pb	13 ± 4	8.7 ± 0.7	5.6 ± 0.3

revealed also significant higher concentrations for Cu, Cd, Ag and Zn, similar contents of As and Mo and lower concentrations for V, Ni and Ba. The PCA results gave two components (F1 and F2) with eigenvalues higher than one, which accounted for 87.1% of the total variance. The F1 was characterized by high positive values of V, Ni, Mo, Ba and Pb and by high negative loadings of Cu, Zn, Ag and Cd. The second component F2 shows high positive contribution from As, Pb, Ag and Cu and while in this case none of the factors gave negative contribution. The biplot shows again a clear separation between the two species, even if the samples appear to be quite scattered in the graph (Fig. 5b). Specifically, specimens of *H. perlevis* are grouped in the negative part of F1, close to Zn/Cd or Cu/Ag loadings but additionally characterized by negative scores in both F1 and F2. Specimens of *H. panicea*, on the other hand, are distinguishable as they show factors always positive in F1 and almost always negative in F2 (except for specimens HP Bel 3 and 4). While *H. perlevis* is characterized by its Cd and Zn accumulation, *H. panicea* separation is most likely related to Ba, Mo, Ni and V.

In conclusion, this survey on two sampling sites demonstrated that: (1) *H. perlevis* is characterized by higher affinity for Cu, Cd, Ag and Zn than *H. panicea* sponges; and (2) the remaining TE are not specifically accumulated by one of these two sponge species. The difference on Zn concentrations between both sponge species is considerable (at least 10 times more). Similarly, to Ag in *A. acuta*, *H. perlevis* present an outstanding feature to accumulate Zn.

3.1.4. Inter-site variability

For Irish sponges, the difference on TE bioaccumulation according to TE availability in the surrounding environment was investigated. Same species of Irish sponges were collected in two different sampling sites: a relatively uncontaminated area (Killkieran Bay) and in a more contaminated environment (Belfast Lough). Within the intra-specific variability, the comparison of TE concentrations in sponges highlights significant differences according to the sampling site (Fig. 4).

Except for As and Cd, *H. panicea* from Belfast showed higher TE concentrations than those in Killkieran: 2–3 times higher for Zn, Ag and Mo; 6–10 times higher for Ni, Zn, Cu and Pb; and > 20 times higher for Ba. On the contrary, As and Cd presented higher concentrations in *H. panicea* from Killkieran. For *H. perlevis* sponges, Ni, Cu, Pb and Ag revealed significant higher concentrations in Belfast than those in Killkieran; whereas Zn, Ba and Cd revealed significant higher concentrations in Killkieran. Considering the intra-specific variability, these results led to a first conclusion. Belfast is likely to be more contaminated than Killkieran for Ni, Cu, Pb and Ag, whereas Killkieran would be more contaminated in Cd. The inter-site variability being opposite or similar within the sponge species for the other TE (Zn, V, As, Ba, Mo), nothing can be concluded on the other TE bioavailability in the surrounding environment.

The great differences in TE accumulation in sponges and other

organisms can have different explanations: TE bioavailability and specific chemistry, geochemical influence on the TE availability in the environment as well species-specific physiological characteristics (Luoma and Rainbow, 2005).

3.2. TE bioaccumulation in sponges

A common way to interpret the accumulation of TE in organisms' tissue is the Bio-Concentration Factors (BCF). This parameter has been found useful in the evaluation of the sponge ability to accumulate the analyte with respect to its availability in the environment. According to the definition given by Rand et al. (1995), the bioconcentration is defined as the uptake of a chemical by an organism from the abiotic environment (non-living chemical and physical components of the environment that affect the living organisms, for instance sunlight, soil, water and pollution). Usually BCF for sponges are calculated as the ratio between the chemical concentrations measured in the organism and the one measured in local sediment (Negri et al., 2006; Mayzel et al., 2014; Pan et al., 2011). Value of 1 was set as threshold for all analytes, with results equal or above 1 being considered indicative of bioconcentration. When TE concentration in seawater is included in the study, the BAF are also calculated by dividing the TE sponge's content by the one found in seawater (Padovan et al., 2012; Venkateswara Rao et al., 2009; Batista et al., 2014).

3.2.1. Bio-concentration factors (BCF)

TEs concentration found in sediments from the three sampling sites are shown in Table 5. Overall, the values are typical of uncontaminated areas, based on the principal environmental regulations (US EPA, 2006; Split and Delta, 2010). We must emphasize that the grain size distribution appeared to be quite inhomogeneous, and the appearance was typical of sandy samples. These characteristics might explain the relatively low values and the high standard deviation for some elements among replicates collected in the same station (especially for Irish samples). Higher concentrations of V, Ni, Cu, Zn, and Ba were observed for sediments collected at Irish sites than at the French one. Similar Ag contents were measured in sediments regardless of the sampling site. Cd and Pb presented higher concentrations in the French site. Finally, the French site and Killkieran sampling site presented similar Mo and As contents in sediments, higher than those from the Belfast site. Comparing the two Irish sites, sediments revealed similar concentrations for most TE, except Ni, Cd, As and Mo. As and Mo revealed higher concentrations in sediments from Killkieran, whereas Ni and Cd exhibited higher concentrations in Belfast's samples.

In Belfast samples the higher Ni concentration in sediments is in accordance with higher Ni sponge contents. The bioavailability of Ni is thus suggested to be traced by Ni sponge content. Cd and Mo were found in higher concentration in sponges from Killkieran and Belfast, respectively; which is the opposite in sediments. For As, even if Irish sediments present significant differences on their concentrations, sponge contents were similar.

The BCF for Mediterranean and Irish sponges are presented in Table 6. BCF obtained for V, Pb, Ba and Ni were lower than 1 (or close) regardless of the sponge species, leading to the conclusion that these elements are more accumulated in sediments than in sponge' tissue. Ba was demonstrated to be toxic for some sponges (Ostrom and Simpson, 1978; Tompkins-MacDonald et al., 2009), and its BCF were already reported lower than 1 in several sponges species (Mayzel et al., 2014), thus implying its non-essential role in sponges' biological activities. In the same manner, a similar assumption can be made for V, which showed the same behavior in other sponge species, as reported by Mayzel et al. (2014). BCF for Pb were however close or higher than one, only in sponges collected in Belfast. BCF for Ni were found to be higher than one only for *H. fulva* and *C. nucula*. This behavior in *C. nucula* can be explained by a high concentration of dense collagen in this species (Pronzato, 2004). This intertwined network of proteins should have

Table 6

Average bioconcentration factors calculated for the sponge species analyzed in this study.

Sponge species (site)	Average BCF									
	V	Ni	Cu	Zn	As	Mo	Ag	Cd	Ba	Pb
<i>H. fulva</i> (Villefranche)	0.4	1.2	4.8	1.5	3.8	2.6	0.3	3.9	0.6	0.3
<i>C. damicornis</i> (Villefranche)	0.6	0.9	18.1	1.6	9.0	2.4	1.0	3.9	0.6	0.4
<i>A. acuta</i> (Villefranche)	0.5	0.6	5.7	2.0	5.4	2.3	41.1	3.7	0.6	0.3
<i>C. nucula</i> (Villefranche)	0.3	2.8	4.4	1.8	4.1	6.3	1.3	1.3	0.2	0.2
<i>H. panicea</i> (Kilkieran)	0.1	0.1	0.6	0.8	1.2	1.8	2.1	21.2	0.02	0.2
<i>H. panicea</i> (Belfast)	0.6	0.6	4.3	1.3	2.1	9.3	8.4	1.7	0.4	1.8
<i>H. perlevis</i> (Kilkieran)	0.2	0.2	2.5	27.8	1.0	1.9	5.4	31.5	0.4	0.3
<i>H. perlevis</i> (Belfast)	0.2	0.3	7.2	15.1	3.5	4.4	19.7	3.2	0.1	1.2

strong affinity with these pure transition metals, maybe due to the presence of basic amino acids acting as ligands for these transition metals. *C. nucula* is also a HMA (high microbial abundance, Vacelet and Donadey, 1977) and the associated cyanobacteria can be responsible for the observed behavior.

For all the other TEs (Cu, Zn, As, Mo, Ag, Cd), BCF were consistently higher than one. Cd and Cu were found to be accumulated by several sponges species collected in the Red and Mediterranean seas (Pan et al., 2011; Mayzel et al., 2014; Cebrian et al., 2007), highlighting their essential role in sponges' metabolism. The high Zn bioaccumulation is likely to be associated not only to its geochemical availability, but also to its crucial role in multiple biological processes as well as to its higher uptake rate for species of this family (Rainbow, 2002). This element is also present in all six classes of enzymes (Broadley et al., 2007), it is thus not surprising that Zn is actively accumulated by sponges. The specific bioconcentration of these TEs could also be associated to some adsorption phenomena related to particular biomolecules such as collagen, as already proposed for *C. nucula*.

The inter-site and inter-species variations of BCF are shown in Fig. 6. Except for Cd and Zn, all the BCFs calculated for Belfast samples were higher than those determined in Kilkieran sponges, despite the similar (V, Cu, Ag, Ba) or lower (As, Mo, Pb) content in sediments. This is particularly evident for Cu, As, Mo, Ag and Pb. Additionally the BCF found for *H. perlevis* were generally higher than those calculated for *H.*

panicea, sampling site being equal. Even if the massive shape of *H. panicea* should yield to higher water pumping rate, the bioaccumulation is higher for *H. perlevis*, thus other factors must be involved in the bioaccumulation processes. *H. perlevis* is a HMA sponge with several associated cyanobacteria described in this species (Alex et al., 2012) which can influence the bioaccumulation. Additionally, a tyrosine derivative has recently been found in this species, which can have a role in the TEs complexing (Mahajan et al., 2017). *C. damicornis* showed the highest BCF for Cu and As compared to other species (BCF = 18 and 9, respectively). Like *H. fulva* and *A. acuta*, this species is characterized by a low microbial abundance (LMA) (Vacelet and Donadey, 1977) with original bacterial community. The branch erected shape and in general the specie's shape cannot be responsible for a higher TEs accumulation as no correlation was found between the accumulation and the morphology. Instead, two reasons can explain the high concentration of As and Cu in *C. damicornis*: first a unique associated microbiota can be selective of these two TEs and the second option would be that high concentrations of secondary metabolites can complex these TEs. It is indeed known that pyrrole aminoimidazole metabolites are highly present in this species (Aiello et al., 2005). They can complex specifically Cu due to the presence of basic nitrogens, however for As associated bacteria may be a better explanation. Arsenic was previously shown to be highly accumulated by sponges, reaching concentration values above 100 mg kg⁻¹ (Pan et al., 2011; Yamaoka et al., 2006; Araújo et al., 2003). Arsenic accumulation was also demonstrated to be higher in demosponges than calcareous sponges (Yamaoka et al., 2001), and additionally quite variable among demosponges as demonstrated in the present study with a particular affinity for *C. damicornis*. Arsenic bioaccumulation and speciation were recently discussed in a published paper and the study underlines the potential role of this metalloid (and more specifically of arsenobetaine) in the protection of sponges against osmolytic stress (Orani et al., 2018). A new polyarsenic compound was also found in a sponge specie from New Caledonia (Mancini et al., 2006), opening the debate on a clear role of this element in sponges' metabolism.

As already observed in Zn concentrations results, Zn BCF were the highest in *H. perlevis* sponges (BCF = 15–28) regardless of the sampling site, showing a clear affinity of the species for this element. The highest BCF was found in *A. acuta* sponges for Ag (BCF = 41.1), as expected from the higher concentrations measured in this sponge specie. Similarly to results found in this study, the propensity for Ag accumulation in *A. acuta* sponges was already observed, also revealing a much reduced Ag decontamination process, compared to other species

Fig. 6. Graphical representation of some of the Bio-concentration factors calculated for all sponge species.

(Genta-Jouve et al., 2012). The reason for this particular behavior resides in the role of Ag in the secondary metabolism of this sponge specie as the sponge is known for containing high concentration of nitrile and isonitrile secondary metabolites which have strong affinity for silver ions. High Ag BCF values were also observed for Irish sponges, reaching 19.7 for *H. perlevis* in Belfast. Concerning Mo, its BCF is significantly higher for *C. nucula* among the in Mediterranean sponges (BCF = 6.3), and in *H. panicea* and *H. perlevis* in Belfast (BCF = 9.3 and 4.4, respectively). It is worth noticing that Mo bioaccumulation is higher in sponges from Belfast (BCF = 21.2–31.5) whereas Mo sediment content was significantly lower in Belfast than in Kilkieran. Mo was seldom included in TEs accumulation studies performed on sponges. Gentric et al. (2016) reported Mo sponge concentrations between 0.2 and 1.2 mg kg⁻¹ (similar to those measured in this study), showing a special affinity of this element for the specie *H. perlevis* comparing to the *Raspailia ramosa* species. Cd presented the same opposite behavior between sponge and sediment results. Cd BCF in both Irish sponge species were surprisingly high in Kilkieran Bay (21–31) whereas the Cd content in sediments was significantly lower in this sampling site comparing to Belfast. The discrepancy between Cd contamination profiles and accumulation in organisms was already observed in barnacles and mussels (Phillips and Rainbow, 1988) indicating the existence of subtle differences in bioavailability of this element. In other words, Cd bioavailability and thus its uptake is not necessarily proportional to its concentration in sediments or generally in the surrounding environment.

Padovan et al. (2012) reported BCF for the sponge *Spheciospongia vagabunda* up to 375, 48 and 1850 for Ni, Zn and Cd respectively, which is higher than BCF calculated in our study. However, sponges and sediments from this previous study were sampled close to a sewage outfall proving thus large anthropogenic TEs inputs. Similar BCF values were reported by Cebrian et al. (2007) in Northwestern Mediterranean sponges for Cu and Pb: i.e. up to 17.5 and up to 0.23, respectively for *Dysidea avara* and *Chondrosia reniformis*, values comparable with maximum BCF for Cu of 18 in *C. damicornis*, and similar than usual BCF for Pb (except in Belfast). Pan et al. (2011) measured TE in different sponges and sediments from Red Sea, showing maximum BCF of 74, 22, 3, 56 and 5 for Cd, Zn, Ag, Cu and As respectively. These results were higher than those reported in the present study for Cd and Cu, lower for Ag and overall comparable for Zn and As. Mayzel et al. (2014) analyzed 16 sponge species and reported BCF consistently lower than 1 for Ba (except for one specie) and V; additionally, the study shows BCF > 1 for As, Cd and Zn. A significant As BCF of 477 was previously measured in only one sponge specie, *Theonella swinhoei*, which was explained by the presence of a particular bacterium (Mayzel et al., 2014). It is worth noticing that *T. swinhoei* is a slow growing and long-lived specie. So As concentration could be linked to the age of the sponge and thus responsible for the elevated BCF. Negri et al. (2006) reported also BCF comparable with the present study for Cu, Pb and Zn (not considering *H. perlevis*), but much higher BCF were reported for As (up to 19) and Cd (up to 690). Results from our study as well as literature data thus support the evidence that the variability in TEs accumulation (and bioconcentration) in sponges occurs at many scales including within and between species, within and between locations, and related to the targeted TE.

3.2.2. Sponges: a powerful tool for biomonitoring studies

To better appreciate the extent of TEs bio-concentration in sponges, the results can be compared with values determined in other organisms, potentially used as monitors of TEs in coastal/marine environment. The great advantages of sponges comparing to other “common” biomonitors like bivalves is that they are simple organisms without complex organs and tissues. It is thus reasonable to think that sponge tissues show low intra-individual variation in TE concentrations (de Mestre et al., 2012).

In a baseline study conducted in Guam harbors the Indo-Pacific Ocean, sponges were shown to accumulate consistently higher amounts

of Ag, As, Cd, Cu, Ni, Pb and Zn in comparison to seaweeds, but not in comparison to oysters or other bivalves (Denton et al., 2006). Aly et al. (2013) found that most of TEs in sponge specimens of *H. oculata* from Poole Harbour (UK) were accumulated much more efficiently than in species of bivalve *Cerastoderma edule*, a common cockle. The latter showed TEs concentrations lower than sediment for almost all analytes while the sponge exhibited very high BCF. A study published by Perez et al. (2004) also showed that sponges from Mediterranean coasts concentrate more TEs than mussels (*Mytilus galloprovincialis*), exception made for Zn. Depending on the targeted TE, concentrations measured in sponges were from 6 to 145 times higher than those measured in mussels. Gentric et al. (2016) reported also higher concentration values of Mo, Ni, and Pb in *H. perlevis* and *R. ramosa* collected in Northern France, in comparison with *Crassostrea gigas* (oyster). On the other hand, this oyster specie showed much higher content of Cu and Zn in comparison with all the sponges species analyzed in the mentioned study. Cu and Zn concentrations in oysters were shown to reach > 10,000 mg kg⁻¹ (Weng and Wang, 2014; Birch et al., 2014). Other benthic organisms such as macroalgae, gastropods and nemertean were shown to accumulate TEs in extent more or less comparable to sponges. Majer et al. (2014) reported BCF values up to 2, 8, 2, 0.4, 1.6 and 5.6 for As, Cd, Cu, Ni, Pb and Zn respectively, for different benthic organisms collected in Antarctica. Corals also shown interesting TEs accumulation properties, with BCF reaching 22 for Cd, 7 for Pb, 24 for Cu and 66 for Zn (Esslemont, 2000).

In conclusion, elements such as Ag, As, Mo and Cd seem to have more affinity for sponges in comparison to other organisms. Even if Cu and Zn are often better accumulated by other organisms in comparison to sponges, these TEs are also significantly bioconcentrated in sponges. This and previous studies demonstrate that sponges are a good tool for TEs monitoring studies, especially since they are widespread in coastal ecosystems.

3.3. Source of lead

In the Mediterranean site, as already mentioned, Pb showed an unusual high variability within the species. The initial hypothesis was that the sponge samples were exposed to different Pb sources in different extent, thus leading to very high differences even among specimens of the same species. To assess this hypothesis, the analysis of Pb isotope ratios was performed in sponge and sediment samples. Results are presented in Fig. 7 and clearly show that all samples have been exposed to the same Pb source, as the isotopic compositions are practically identical. Pb isotope ratios were found to be more radiogenic than those reported for gasoline in France (Monna et al., 1995) or average French industrial Pb (Véron et al., 1999) but closer to values reported for natural Pb sources i.e. pre-industrial sediments discharged by the major French rivers (²⁰⁶Pb/²⁰⁷Pb = 1.197–1.206, Elbaz-Poulichet et al., 1986) and Miocene siltstone and Jurassic limestone analyzed by Monna et al. (1995). (See Fig. 8.)

Regarding Irish samples, the Pb concentrations found in sediments were higher in Kilkieran than in Belfast (8.7 vs 5.6 mg kg⁻¹) whereas it was the opposite in sponges with higher Pb contents in Belfast. Three-isotope graphs showed a clear linear trend for Irish samples ($r^2 = 0.9595$, Fig. 7) suggesting that at least two different Pb sources influenced these samples' isotopic composition. Samples collected in Belfast show lower ²⁰⁶Pb/²⁰⁷Pb as well as lower ²⁰⁶Pb/²⁰⁴Pb values, close to ²⁰⁶Pb/²⁰⁷Pb values reported by Fletcher et al. (1993) for Welsh ore and by Charlesworth et al. (2006) for Dublin surface sediments, influenced by the industrial activities of the area. Charlesworth et al. (2006) also reported ²⁰⁶Pb/²⁰⁷Pb equal to 1.191 for bottom sediment cores collected in the Irish Sea (between Dublin and Belfast). This value is close to our results on sponges and sediment collected in Kilkieran Bay, confirming that the Pb found in both, sponges and sediment from Kilkieran, is most likely associated to a natural source. These results thus show how the mere determination of Pb concentration is often not

Fig. 7. Three isotopes graph of Pb isotope ratios results obtained for Mediterranean sample plotted together with some literature data of common anthropogenic sources and typical natural composition of the area. Data from Elbaz-Poulichet et al. (1986), Monna et al. (1995), Véron et al. (1999).

sufficient for the assessment of contamination of a given place, especially when sediments of proper grain size are not available. The Pb isotopic composition can give relevant additional information and the ultimate confirmation of the presence of Pb from anthropogenic source.

4. Conclusion

In conclusion, the present work shows the great capabilities of sponges as TEs bioaccumulators and their suitability as environmental biomonitors. The six analyzed species show generally different accumulation properties depending on the considered element, sampling station and sponge species. Some elements such as Ba, V and in some cases Ni, were poorly accumulated by sponges, meaning that these elements are not involved in biological processes. Statistical data treatment by PCA and BCF calculations shows how it is possible to distinguish different species based on their bioaccumulation properties. Specifically, Cu and As are demonstrated to be highly accumulated in *C. damicornis*, Zn in *H. perlevis* and, as already shown in previous studies, Ag, Mo and Ni in *A. acuta*. Except for Ba, V and Pb, BCF were higher than one, highlighting the bioaccumulation properties of sponges. Our results show that the bioaccumulation of TEs is most likely associated to specific morphological features and/or to the specific bacterial community associated to different species rather than different pumping

rate. This study confirms that sponges are efficient bioindicators of TEs pollution state of coastal environments. Finally, the determination of Pb isotope ratios, offered the opportunity to evaluate the source of Pb in the studied area, revealing mostly a natural origin for samples collected in the Mediterranean area (even though the Pb concentrations were higher than both Irish sites) and Kilkieran bay, while an anthropogenic signature was mainly found in samples collected from Belfast Lough.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.marpolbul.2018.04.073>.

Fig. 8. Three isotopes graphs showing Pb isotope ratios results for sponges and sediments collected in Ireland. Data from Fletcher et al. (1993) and Charlesworth et al. (2006).

References

- Aiello, A., D'Esposito, M., Fattorusso, E., Menna, M., Müller, W.E.G., Perović-Ottstadt, S., Tsuruta, H., Gulder, T.A.M., Bringmann, G., 2005. Daminin, a bioactive pyrrole alkaloid from the Mediterranean sponge *Axinella damicornis*. *Tetrahedron* 61, 7266–7270. <http://dx.doi.org/10.1016/j.tet.2005.05.025>.
- Alex, A., Vasconcelos, V., Tamagnini, P., Santos, A., Antunes, A., 2012. Unusual symbiotic cyanobacteria association in the genetically diverse intertidal marine sponge *Hymeniacidon perlevis* (Demospongiae, Halichondrida). *PLoS One* 7, 1–12. <http://dx.doi.org/10.1371/journal.pone.0051834>.
- Aly, W., Williams, I.D., Hudson, M.D., 2013. Metal contamination in water, sediment and biota from a semi-enclosed coastal area. *Environ. Monit. Assess.* 185, 3879–3895. <http://dx.doi.org/10.1007/s10661-012-2837-0>.
- Anjum, K., Abbas, S.Q., Shah, S.A., Akhter, N., Batool, S., Hassan, S.S., 2016. Marine sponges as a drug treasure. *Biomol. Ther.* 24, 347–362.
- Araújo, M.F., Conceição, A., Barbosa, T., Lopes, M.T., Humanes, M., 2003. Elemental composition of marine sponges from the Berlengas Natural Park, western Portuguese coast. *X-Ray Spectrom.* 32, 428–433. <http://dx.doi.org/10.1002/xrs.660>.
- Aresta, A., Marzano, C.N., Lopane, C., Corriero, G., Longo, C., Zamboni, C., Stabili, L., 2015. Analytical investigations on the lindane bioremediation capability of the demosponge *Hymeniacidon perlevis*. *Mar. Pollut. Bull.* 90, 143–149. <http://dx.doi.org/10.1016/j.marpolbul.2014.11.003>.
- Ashraf, A., Saion, E., Gharibshahi, E., Kamari, H.M., Yap, C.K., Hamzah, M.S., Elias, M.S., 2017. Distribution of trace elements in Core marine sediments of coastal East Malaysia by instrumental neutron activation analysis. *Appl. Radiat. Isot.* 122, 96–105. <http://dx.doi.org/10.1016/j.apradiso.2017.01.006>.
- Barats, A., Féraud, G., Potot, C., Philippini, V., Travi, Y., Durrieu, G., Dubar, M., Simler, R., 2014. Naturally dissolved arsenic concentrations in the Alpine/Mediterranean Var River watershed (France). *Sci. Total Environ.* 473–474, 422–436. <http://dx.doi.org/10.1016/j.scitotenv.2013.12.007>.
- Batista, D., Muricy, G., Rocha, R.C., Miekeley, N.F., 2014. Marine sponges with contrasting life histories can be complementary biomonitors of heavy metal pollution in coastal ecosystems. *Environ. Sci. Pollut. Res.* 21, 5785–5794. <http://dx.doi.org/10.1007/s11356-014-2530-7>.
- Birch, G.F., Melwani, A., Lee, J.H., Apostolatos, C., 2014. The discrepancy in concentration of metals (Cu, Pb and Zn) in oyster tissue (*Saccostrea glomerata*) and ambient bottom sediment (Sydney estuary, Australia). *Mar. Pollut. Bull.* 80, 263–274. <http://dx.doi.org/10.1016/j.marpolbul.2013.12.005>.
- Broadley, M.R., White, P.J., Hammond, J.P., Zelko, I., Lux, A., 2007. Zinc in plants. *New Phytol.* 173, 677–702. <http://dx.doi.org/10.1111/j.1469-8137.2007.01996.x>.
- Cabrita, M.T., Padeiro, A., Amaro, E., dos Santos, M.C., Leppe, M., Verkulich, S., Hughes, K.A., Peter, H.-U., Canário, J., 2017. Evaluating trace element bioavailability and potential transfer into marine food chains using immobilised diatom model species *Phaeodactylum tricornutum*, on King George Island, Antarctica. *Mar. Pollut. Bull.* 121, 192–200. <http://dx.doi.org/10.1016/j.marpolbul.2017.05.059>.
- Caccia, V.G., Millero, F.J., Palanques, A., 2003. The distribution of trace metals in Florida Bay sediments. *Mar. Pollut. Bull.* 46, 1420–1433. [http://dx.doi.org/10.1016/S0025-326X\(03\)00288-1](http://dx.doi.org/10.1016/S0025-326X(03)00288-1).
- Cantillo, A.Y., 1998. Comparison of results of mussel watch programs of the United States and France with worldwide mussel watch studies. *Mar. Pollut. Bull.* 36, 712–717. [http://dx.doi.org/10.1016/S0025-326X\(98\)00049-6](http://dx.doi.org/10.1016/S0025-326X(98)00049-6).
- Cebrian, E., Martí, R., Uriz, J.M., Turon, X., 2003. Sublethal effects of contamination on the Mediterranean sponge *Crambe crambe*: metal accumulation and biological responses. *Mar. Pollut. Bull.* 46, 1273–1284. [http://dx.doi.org/10.1016/S0025-326X\(03\)00190-5](http://dx.doi.org/10.1016/S0025-326X(03)00190-5).
- Cebrian, E., Uriz, M.-J., Turon, X., 2007. Sponges as biomonitors of heavy metals in spatial and temporal surveys in northwestern Mediterranean: multispecies comparison. *Environ. Toxicol. Chem.* 26, 2430–2439. <http://dx.doi.org/10.1897/07-292.1>.
- Charlesworth, M.E., Chenery, S., Mellor, A., Service, M., 2006. Isotopic composition and concentration of Pb in suspended particulate matter of the Irish Sea reveals distribution and sources. *Mar. Pollut. Bull.* 52, 81–88. <http://dx.doi.org/10.1016/j.marpolbul.2005.08.009>.
- Dahms, S., van der Bank, F.H., Greenfield, R., 2014. A baseline study of metal contamination along the Namibian coastline for *Perna perna* and *Choromytilus meridionalis*. *Mar. Pollut. Bull.* 85, 297–305. <http://dx.doi.org/10.1016/j.marpolbul.2014.05.037>.
- Denton, G.R.W., Concepcion, L.P., Wood, H.R., Morrison, R.J., 2006. Trace metals in marine organisms from four harbours in Guam. *Mar. Pollut. Bull.* 52, 1784–1804. <http://dx.doi.org/10.1016/j.marpolbul.2006.09.010>.
- Drivelos, S.A., Georgiou, C.A., 2012. Multi-element and multi-isotope-ratio analysis to determine the geographical origin of foods in the European Union. *TrAC Trends Anal. Chem.* 40, 38–51. <http://dx.doi.org/10.1016/j.trac.2012.08.003>.
- Elbaz-Poulichet, F., Holliger, P., Martin, J.M., Petit, D., 1986. Stable lead isotopes ratios in major French rivers and estuaries. *Sci. Total Environ.* 54, 61–76. [http://dx.doi.org/10.1016/0048-9697\(86\)90256-1](http://dx.doi.org/10.1016/0048-9697(86)90256-1).
- Esslemont, G., 2000. Heavy metals in seawater, marine sediments and corals from the Townsville section, Great Barrier Reef Marine Park, Queensland. *Mar. Chem.* 71, 215–231. [http://dx.doi.org/10.1016/S0304-4203\(00\)00050-5](http://dx.doi.org/10.1016/S0304-4203(00)00050-5).
- Fang, T.-H., Hsiao, S.-H., Nan, F.-H., 2014. Nineteen trace elements in marine copepods collected from the coastal waters off northeastern Taiwan. *Cont. Shelf Res.* 91, 70–81. <http://dx.doi.org/10.1016/j.csr.2014.09.003>.
- Fletcher, C.J.N., Swainbank, I.G., Colman, T.B., 1993. Metallogenic evolution in Wales: constraints from lead isotope modelling. *J. Geol. Soc. Lond.* 150, 77–82. <http://dx.doi.org/10.1144/gsjgs.150.1.0076>.
- Genta-Jouve, G., Cachet, N., Oberhänsli, F., Noyer, C., Teyssié, J.L., Thomas, O.P., Lacoue-Labarthe, T., 2012. Comparative bioaccumulation kinetics of trace elements in Mediterranean marine sponges. *Chemosphere* 89, 340–349. <http://dx.doi.org/10.1016/j.chemosphere.2012.04.052>.
- Genric, C., Rehel, K., Dufour, A., Sauleau, P., 2016. Bioaccumulation of metallic trace elements and organic pollutants in marine sponges from the South Brittany Coast, France. *J. Environ. Sci. Health A* 51, 213–219. <http://dx.doi.org/10.1080/10934529.2015.1094327>.
- Gifford, S., Dunstan, R.H., O'Connor, W., Koller, C.E., MacFarlane, G.R., 2007. Aquatic zooremediation: deploying animals to remediate contaminated aquatic environments. *Trends Biotechnol.* 25, 60–65. <http://dx.doi.org/10.1016/j.tibtech.2006.12.002>.
- Grotti, M., Soggia, F., Lagomarsino, C., Riva, S.D., Goessler, W., Francesconi, K.A., 2008. Natural variability and distribution of trace elements in marine organisms from Antarctic coastal environments. *Antarct. Sci.* 20, 39–52. <http://dx.doi.org/10.1017/S0954102007000831>.
- Hinrichs, J., Dellwig, O., Brumsack, H.-J., 2002. Lead in sediments and suspended particulate matter of the German Bight: natural versus anthropogenic origin. *Appl. Geochem.* 17, 621–632. [http://dx.doi.org/10.1016/S0883-2927\(01\)00124-X](http://dx.doi.org/10.1016/S0883-2927(01)00124-X).
- Illuminati, S., Annibaldi, A., Truzzi, C., Scarponi, G., 2016. Heavy metal distribution in organic and siliceous marine sponge tissues measured by square wave anodic stripping voltammetry. *Mar. Pollut. Bull.* 111, 476–482. <http://dx.doi.org/10.1016/j.marpolbul.2016.06.098>.
- Komárek, M., Ettler, V., Chrastný, V., Mihaljevič, M., 2008. Lead isotopes in environmental sciences: a review. *Environ. Int.* 34, 562–577. <http://dx.doi.org/10.1016/j.envint.2007.10.005>.
- Larsen, M.M., Blustajn, J.S., Andersen, O., Dahlöf, I., 2012. Lead isotopes in marine surface sediments reveal historical use of leaded fuel. *J. Environ. Monit.* 14, 2893. <http://dx.doi.org/10.1039/c2em30579h>.
- Li, X., Wai, O.W.H., Li, Y.S., Coles, B.J., Ramsey, M.H., Thornton, I., 2000. Heavy metal distribution in sediment profiles of the Pearl River estuary, South China. *Appl. Geochem.* 15, 567–581. [http://dx.doi.org/10.1016/S0883-2927\(99\)00072-4](http://dx.doi.org/10.1016/S0883-2927(99)00072-4).
- Liu, L., Wang, Z., Ju, F., Zhang, T., 2015. Co-occurrence correlations of heavy metals in sediments revealed using network analysis. *Chemosphere* 119, 1305–1313. <http://dx.doi.org/10.1016/j.chemosphere.2014.01.068>.
- Luoma, S.N., Rainbow, P.S., 2005. Why is metal bioaccumulation so variable? Biodynamics as a unifying concept. *Environ. Sci. Technol.* 39, 1921–1931. <http://dx.doi.org/10.1021/es048947e>.
- Mahajan, N., Calabro, K., Morrow, C., Thomas, O., 2017. N-Dihydroxytyrosine identified from the Northeastern Atlantic marine sponge *Hymeniacidon perlevis* after chemical screening. *Nat. Prod. Commun.* 12, 945–946.
- Majer, A.P., Petti, M.A.V., Corbisier, T.N., Ribeiro, A.P., Theophilo, C.Y.S., Ferreira, P.A. de L., Figueira, R.C.L., 2014. Bioaccumulation of potentially toxic trace elements in benthic organisms of Admiralty Bay (King George Island, Antarctica). *Mar. Pollut. Bull.* 79, 321–325. <http://dx.doi.org/10.1016/j.marpolbul.2013.12.015>.
- Mancini, I., Guella, G., Frostin, M., Hnawia, E., Laurent, D., Debitus, C., Pietra, F., 2006. On the polyarsenic organic compound from nature: arsenicin A from the new Caledonian marine sponge *Echinochalina bargibanti*. *Chem. Eur. J.* 12, 8989–8994. <http://dx.doi.org/10.1002/chem.200600783>.
- Mayzel, B., Aizenberg, J., Ilan, M., 2014. The elemental composition of demospongiae from the Red Sea, Gulf of Aqaba. *PLoS One* 9. <http://dx.doi.org/10.1371/journal.pone.0095775>.
- de Mestre, C., Maher, W., Roberts, D., Broad, A., Krikowa, F., Davis, A.R., 2012. Sponges as sentinels: patterns of spatial and intra-individual variation in trace metal concentration. *Mar. Pollut. Bull.* 64, 80–89. <http://dx.doi.org/10.1016/j.marpolbul.2011.10.020>.
- Milanesi, M., Cellosi, E., Manconi, R., Sarà, A., Sidri, M., Pronzato, R., 2003. The marine sponge *Chondrilla nucula* Schmidt, 1862 as an elective candidate for bioremediation in integrated aquaculture. *Biomol. Eng.* 20, 363–368. [http://dx.doi.org/10.1016/S1389-0344\(03\)00052-2](http://dx.doi.org/10.1016/S1389-0344(03)00052-2).
- Monna, F., Ben Othman, D., Luck, J.M., 1995. Pb isotopes and Pb, Zn and Cd concentrations in the rivers feeding a coastal pond (Thau, southern France): constraints on the origin(s) and flux(es) of metals. *Sci. Total Environ.* 166, 19–34. [http://dx.doi.org/10.1016/0048-9697\(95\)04514-2](http://dx.doi.org/10.1016/0048-9697(95)04514-2).
- Müller, W.E.G., Batel, R., Lacorn, M., Steinhart, H., Simat, T., Lauenroth, S., Hassanein, H., Schröder, H.C., 1998. Accumulation of cadmium and zinc in the marine sponge *Suberites domuncula* and its potential consequences on single-strand breaks and on expression of heat-shock protein: a natural field study. *Mar. Ecol. Prog. Ser.* 167, 127–135. <http://dx.doi.org/10.3354/meps167127>.
- Negri, A., Burns, K., Boyle, S., Brinkman, D., Webster, N., 2006. Contamination in sediments, bivalves and sponges of McMurdo Sound, Antarctica. *Environ. Pollut.* 143, 456–467. <http://dx.doi.org/10.1016/j.envpol.2005.12.005>.
- Olesen, T.M.E., Weeks, J.M., 1994. Accumulation of Cd by the marine sponge *Halichondria panicea pallas*: effects upon filtration rate and its relevance for biomonitors. *Bull. Environ. Contam. Toxicol.* 52, 722–728. <http://dx.doi.org/10.1007/BF00195494>.
- Orani, A.M., Barats, A., Zitte, W., Morrow, C., Thomas, O.P., 2018. Comparative study on the bioaccumulation and biotransformation of arsenic by some northeastern Atlantic and northwestern Mediterranean sponges. *Chemosphere* 201, 826–839. <http://dx.doi.org/10.1016/j.chemosphere.2018.03.078>.
- Ortega, G.S., Pêchevran, C., Béail, S., Donard, O.F.X., 2012. A fit-for-purpose procedure for lead isotopic ratio determination in crude oil, asphaltene and kerogen samples by MC-ICPMS. *J. Anal. At. Spectrom.* 27, 1447. <http://dx.doi.org/10.1039/c2ja30143a>.
- Ostrom, K.M., Simpson, T.L., 1978. Calcium and the release from dormancy of freshwater sponge gemmules. *Dev. Biol.* 64, 332–338. [http://dx.doi.org/10.1016/0012-1606\(78\)90083-0](http://dx.doi.org/10.1016/0012-1606(78)90083-0).
- Padovan, A., Munksgaard, N., Alvarez, B., McGuinness, K., Parry, D., Gibb, K., 2012.

- Trace metal concentrations in the tropical sponge *Spheciospongia vagabunda* at a sewage outfall: synchrotron X-ray imaging reveals the micron-scale distribution of accumulated metals. *Hydrobiologia* 687, 275–288. <http://dx.doi.org/10.1007/s10750-011-0916-9>.
- Pan, K., Lee, O.O., Qian, P.Y., Wang, W.X., 2011. Sponges and sediments as monitoring tools of metal contamination in the eastern coast of the Red Sea, Saudi Arabia. *Mar. Pollut. Bull.* 62, 1140–1146. <http://dx.doi.org/10.1016/j.marpolbul.2011.02.043>.
- Parra, S., Bravo, M.A., Quiroz, W., Querol, X., Paipa, C., 2015. Distribution and pollution assessment of trace elements in marine sediments in the Quintero Bay (Chile). *Mar. Pollut. Bull.* 99, 256–263. <http://dx.doi.org/10.1016/j.marpolbul.2015.07.066>.
- Patel, B., Balani, M., Patel, S., 1985. Sponge “sentinel” of heavy metals. *Sci. Total Environ.* 41, 143–152.
- Perez, T., Vacelet, J., Rebouillon, P., 2004. In situ comparative study of several Mediterranean sponges as potential biomonitors for heavy metals. *Boll. Mus. Ist. Biol. Univ. Genova* 68, 517–525.
- Perez, T., Longet, D., Schembri, T., Rebouillon, P., Vacelet, J., 2005. Effects of 12 years' operation of a sewage treatment plant on trace metal occurrence within a Mediterranean commercial sponge (*Spongia officinalis*, Demospongiae). *Mar. Pollut. Bull.* 50, 301–309. <http://dx.doi.org/10.1016/j.marpolbul.2004.11.001>.
- Phillips, D., Rainbow, P., 1988. Barnacles and mussels as biomonitors of trace elements: a comparative study. *Mar. Ecol. Prog. Ser.* 49, 83–93. <http://dx.doi.org/10.3354/meps049083>.
- Pronzato, R., 2004. A climber sponge. *Boll. Mus. Ist. Biol. Univ. Genova* 68, 549–552.
- Rainbow, P., 2002. Trace metal concentrations in aquatic invertebrates: why and so what? *Environ. Pollut.* 120, 497–507. [http://dx.doi.org/10.1016/S0269-7491\(02\)00238-5](http://dx.doi.org/10.1016/S0269-7491(02)00238-5).
- Rand, G.M., Wells, P.G., McCarthy, L.S., 1995. Introduction to aquatic ecology. In: Rand, G.M. (Ed.), *Fundamentals of Aquatic Toxicology*. Taylor and Francis, London, pp. 3–53.
- Sjåstad, K.-E., Simonsen, S.L., Andersen, T., 2011. Use of lead isotopic ratios to discriminate glass samples in forensic science. *J. Anal. At. Spectrom.* 26, 325. <http://dx.doi.org/10.1039/c0ja00054j>.
- Søndergaard, J., Asmund, G., Larsen, M.M., 2015. Trace elements determination in seawater by ICP-MS with on-line pre-concentration on a Chelex-100 column using a “standard” instrument setup. *MethodsX* 2, 323–330. <http://dx.doi.org/10.1016/j.mex.2015.06.003>.
- Split, P.M., Delta, A., 2010. *Sediment Quality Guidelines for the Protection of Aquatic Life*. Canadian Council of Ministers of the Environment.
- Tompkins-MacDonald, G.J., Gallin, W.J., Sakarya, O., Degnan, B., Leys, S.P., Boland, L.M., 2009. Expression of a poriferan potassium channel: insights into the evolution of ion channels in metazoans. *J. Exp. Biol.* 212, 761–767. <http://dx.doi.org/10.1242/jeb.026971>.
- US EPA, 2006. *Marine Sediment Screening Benchmarks. III, Region*.
- Vacelet, J., Donadey, C., 1977. Electron microscope study of the association between some sponges and bacteria. *J. Exp. Mar. Biol. Ecol.* 30, 301–314. [http://dx.doi.org/10.1016/0022-0981\(77\)90038-7](http://dx.doi.org/10.1016/0022-0981(77)90038-7).
- Vanhaecke, F., Degryse, P., 2012. The isotopic composition of the elements. In: *Isotopic Analysis: Fundamentals and Applications Using ICP-MS*. John Wiley and Sons, pp. 1–29. <http://dx.doi.org/10.1002/9783527650484>.
- Vanhaecke, F., Balcaen, L., Malinovsky, D., 2009. Use of single-collector and multi-collector ICP-mass spectrometry for isotopic analysis. *J. Anal. At. Spectrom.* 24, 863. <http://dx.doi.org/10.1039/b903887f>.
- Varol, M., 2011. Assessment of heavy metal contamination in sediments of the Tigris River (Turkey) using pollution indices and multivariate statistical techniques. *J. Hazard. Mater.* 195, 355–364. <http://dx.doi.org/10.1016/j.jhazmat.2011.08.051>.
- Venkateswara Rao, J., Srikanth, K., Pallela, R., Gnaneshwar Rao, T., 2009. The use of marine sponge, *Haliclona tenuiramosa* as bioindicator to monitor heavy metal pollution in the coasts of Gulf of Mannar, India. *Environ. Monit. Assess.* 156, 451–459. <http://dx.doi.org/10.1007/s10661-008-0497-x>.
- Véron, A., Flament, P., Bertho, M.L., Alleman, L., Flegat, R., Hamelin, B., 1999. Isotopic evidence of pollutant lead sources in northwestern France. *Atmos. Environ.* 33, 3377–3388. [http://dx.doi.org/10.1016/S1352-2310\(98\)00376-8](http://dx.doi.org/10.1016/S1352-2310(98)00376-8).
- Weis, P., Weis, J.S., 1999. Accumulation of metals in consumers associated with chromated copper arsenate-treated wood panels. *Mar. Environ. Res.* 48, 73–81. [http://dx.doi.org/10.1016/S0141-1136\(99\)00028-8](http://dx.doi.org/10.1016/S0141-1136(99)00028-8).
- Weng, N., Wang, W.X., 2014. Variations of trace metals in two estuarine environments with contrasting pollution histories. *Sci. Total Environ.* 485–486, 604–614. <http://dx.doi.org/10.1016/j.scitotenv.2014.03.110>.
- Yamaoka, Y., Carmona, M.L., Oclarit, J.M., Jin, K., Shibata, Y., 2001. Arsenic compounds in marine sponge (*Haliclona permolis*, *Halichondria japonica*, *Halichondria okadai* and *Haliclona* sp. white) from Seto Inland Sea, Japan. *Appl. Organomet. Chem.* 15, 261–265. <http://dx.doi.org/10.1002/aoc.135>.
- Yamaoka, Y., Carmona, M.L., Oclarit, J.M., Jin, K., Shibata, Y., 2006. Characterization of water-soluble organoarsenic compounds in marine sponges. *Appl. Organomet. Chem.* 20, 545–548. <http://dx.doi.org/10.1002/aoc>.