Coxiella burnetii Multilevel Disk Space Infection, Epidural Abscess, and Vertebral Osteomyelitis Secondary to Contiguous Spread From Infected Abdominal Aortic Aneurysm or Graft: Report of 4 Cases Acquired in the US and Review of the Literature

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Coxiella burnetii Multilevel Disk Space Infection, Epidural Abscess, and Vertebral Osteomyelitis Secondary to Contiguous Spread From Infected Abdominal Aortic Aneurysm or Graft: Report of 4 Cases Acquired in the US and Review of the Literature

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Background. Chronic Coxiella burnetii infections such as vertebral osteomyelitis caused by contiguous spread from an infected abdominal aortic graft or aneurysm have been rarely reported and are associated with significant morbidity and mortality.

Methods. We present the first four reported US acquired cases of Coxiella burnetii vertebral osteomyelitis caused by contiguous spread from an infected abdominal aortic graft or aneurysm.

Results. Presenting symptoms included progressive back pain, malaise, and weight loss with recent or remote animal exposure. Typical imaging findings demonstrated a peri-aortic collection with extension to the paraspinal muscles and vertebrae. Antibiotic regimens included doxycycline with either hydroxychloroquine or a quinolone for at least 2 years or as chronic suppression.

Conclusions. C. burnetii vertebral osteomyelitis is rare and can occur by contiguous spread from an abdominal aneurysm or vascular graft infection. It should be suspected in patients where pre-antibiotic cultures are negative with animal/farming exposure.

Keywords. Coxiella burnetii; infected abdominal aortic aneurysm; Q fever; vascular graft; vertebral osteomyelitis.

Coxiella burnetii infections occur sporadically worldwide and are more common near farming communities or communities with animal exposure. Although most infections go unnoticed, chronic infections, such as infective endocarditis, cause significant morbidity. Other noncardiac endovascular involvement such as vascular aneurysm and/or graft infections have rarely been reported from other countries. We report 4 cases of C. burnetii multilevel disc space infection and vertebral osteomyelitis caused by contiguous spread from infected abdominal aortic grafts and/or mycotic aneurysm and review the literature of noncardiac endovascular and osteoarticular chronic infections.

METHODS

We conducted a retrospective review of all C. burnetii abdominal aortic aneurysm or aortic graft infections with associated vertebral osteomyelitis presenting at our institution from January 1990 to July 2017. A real-time polymerase chain reaction (PCR) assay, which rapidly detects C. burnetii DNA by targeting a sequence of the shikimate dehydrogenase gene (aroE) unique to C. burnetii, was used to confirm infection from tissue and blood specimens unless otherwise stated. Blinded clinical validation of this PCR assay demonstrated 100% sensitivity and specificity when used on cardiac valve tissue [1]. Indirect immunofluorescence testing for C. burnetii IgG and IgM antibodies was performed on a Focus Diagnostics assay (package inserts: Q fever IFA IgG Focus Diagnostics, Inc., Cypress, California; Q fever IFA IgM, Focus Diagnostics, Inc., Cypress, California).

A systematic literature search for additional reported cases of noncardiac endovascular and osteoarticular chronic C. burnetii infections was performed with the assistance of a specialist medical librarian, guided by the authors. The search was restricted to English-language publications from January 1966 to July 2017; findings from conference proceedings were excluded.

Case 1

A 74-year-old male farmer from Iowa with a history of hypertension and coronary artery disease underwent a bifurcated aorto-iliac Dacron graft repair of an abdominal aortic aneurysm (AAA) in May 2005. In early 2006, he was treated for a methicillin-resistant Staphylococcus aureus (MRSA) superficial surgical site infection of an abdominal wall hernia repair. There was no documented MRSA blood stream infection; additional details about this infection are not known. The
patient remained well until January 2007, when he developed progressive back pain. An MRI in mid-March 2007 (Figure 1) showed a left psoas muscle and T10-L3 epidural abscess with L1-L3 osteomyelitis. An open debridement of the lumbar vertebrae and epidural abscess was performed. All obtained surgical cultures were negative for bacterial (aerobic and anaerobic), fungal, and mycobacterial organisms at 6 weeks of incubation. The patient did not receive any antimicrobial therapy prior to culture ascertainment. A tuberculin skin test (TST), Brucella, and Bartonella serologies were also negative. He completed a 6-week course of empiric vancomycin without improvement in back pain. The patient denied fevers, chills, sweats, abdominal pain, or new neurologic symptoms. He denied overseas travel. Exposures included deer hunting, raising livestock, and drinking unpasteurized milk within the past year. Due to persistent symptoms, he was transferred to our institution on May 29, 2007. On examination at admission, his temperature was 36.9°C, BP was 131/94 mmHg, and pulse was 89/min. He was nontoxic-appearing and had a soft 1/6 systolic murmur and tenderness on lumbar spinal percussion. Laboratory tests showed a hemoglobin of 10 g/dL, WBC of 6.4 × 10⁹/L, platelets of 107 × 10⁹/L; serum Cr of 1.4 mg/dL, sedimentation rate of 53 mm/h, C-reactive protein (CRP) of 3.6 mg/dL (normal reference range < 0.8 mg/dL), and normal liver function tests.

Computerized tomography (CT) of the abdomen and pelvis and magnetic resonance imaging (MRI) of the thoracolumbar spine (Figure 2) in May 2007 showed diffuse enhancement of L1, L2, and L3 and the disk spaces of L5-S1 and L2-L3, an epidural abscess extending from T11 to L3, and an L2 fracture with destruction of the body. Paraspinal soft tissues and psoas muscles were thickened and enhanced bilaterally with a new right psoas muscle abscess. There was enhancement of the abdominal aortic graft from T10 to the level of proximal common iliac arteries and an area of aneurysmal dilatation above the graft that was worrisome for a mycotic aneurysm.

Additional serologic testing performed in May 2007 noted negative blood cultures obtained off antimicrobial therapy. Fungal, Francisella tularensis, Brucella, Bartonella serologies, TST, and tuberculosis interferon-gamma release assay (TB-IGRA) were all negative. The C. burnetii IgG phase 1 titer was 1:1024, and the

![Figure 1. Lumbar spine MRI with psoas muscle and T10 to L3 epidural abscess, L1, L2, and L3 osteomyelitis. Images: (A) sagittal T2, (B) sagittal T1, and (C) axial.](https://academic.oup.com/ofid/article-abstract/4/4/ofx192/4139802)

![Figure 2. Thoracolumbar spine MRI with diffuse enhancement of L1-L3, epidural abscess extending from T11 to L3, L2 fracture, paraspinal soft tissue and psoas muscles thickening and enhancement, and right psoas muscle abscess. Images: (A) sagittal T2, (B) sagittal T1, and (C) axial.](https://academic.oup.com/ofid/article-abstract/4/4/ofx192/4139802)
IgM phase 1 titer was ≥1:16. The phase 2 titers of IgG 1:256, and IgM <1:16 was consistent with a chronic infection. A transesophageal echocardiogram (TEE) showed no valvular abnormalities.

On June 1, 2007, the patient underwent a temporary axillo-femoral bypass; explantation of the infected graft and mycotic aneurysm; insertion of rifampin-soaked Dacron aorto-bi-iliac, celiac, superior mesenteric, and right renal grafts nested away from the infected field; and soft tissue and L1, L2 debridement (Figure 3). Histopathologic examination of explanted tissue revealed granulomas with neutrophilic inflammation. Aortic and vertebral tissues were sent for cultures and were negative for bacteria, fungi, and mycobacteria. *C. burnetii* DNA PCR assay using the *hspAB*-associated element was positive from aortic aneurysm tissue and vertebral body tissue; in addition, *C. burnetii* was able to be cultured from shell-vial cultures using human embryonic lung fibroblasts inoculated by aortic aneurysm tissue and vertebral body tissue (Figure 4) at a specialist research laboratory (Unité des Rickettsies, Université de la Méditerranée, Marseille, France) [2, 3]. The patient was started on doxycycline and ciprofloxacin; the latter was subsequently replaced with hydroxychloroquine. His course was complicated by an acute myocardial infarction, congestive heart failure, bowel perforation, and an intra-abdominal abscess with sepsis. After a prolonged hospitalization and discussions with family, life support was withdrawn 55 days after surgery.

**Case 2**

A 60-year-old female from North Dakota underwent an urgent repair for a leaking abdominal aneurysm using an aorto-bi-iliac polyester graft in July 2009. After her aneurysm repair, she developed progressive back pain, unintentional weight loss, and gait instability. Her medical history included hypertension, hyperlipidemia, chronic obstructive pulmonary disease, coronary artery disease, and pulmonary carcinoid status post–wedge resection. She lived on a farm with animal exposure 20 years ago; there was no other known animal exposure or international travel.

A CT of the lumbar spine showed erosive L2-L4 vertebral changes with complex L2-L3 fractures. Fluid and air were noted around the aortic graft, with a paraspinal phlegmon along the psoas muscle. A CT-guided aspiration and L3 vertebral body biopsy tissue were negative for bacterial, fungal, and mycobacterial growth (off antimicrobials).

Transthoracic echocardiography was negative for evidence of endocarditis. A 2-staged surgery was performed, the first for the spine debridement and stabilization with hardware fixation. Tissue and bone cultures remained negative. Pathology noted chronic inflammation, giant cell reaction, calcification, and focal necrosis. Five days later, she underwent resection of the infected aorto-bi-iliac graft, debridement of the aneurysm sac and peri-aortic tissues, and insertion of a rifampin-soaked Hemashield aorto-bi-iliac graft wrapped with omentum. Postoperation, she was placed on empiric vancomycin and meropenem. Operative cultures remained negative. Pathology showed necrotizing granulomatous inflammation with negative stains for bacteria, fungi, and mycobacteria. TB-IGRA, fungal and Bartonella serologies, and urine fungal antigen testing were negative. *C. burnetii* serologies were an IgG phase 1 titer of 1:1024 and an IgM phase 1 titer of <1:16. The phase 2 titers were an IgG of 1:256 and an IgM of <1:16. PCR for *C. burnetii* was positive from the aortic graft and bone. Antibiotics were changed to ciprofloxacin and doxycycline, which she continued to date with anticipated chronic indefinite suppression. Her serologies at 4.5 years postsurgery were an IgG phase 1 titer of 1:1024 and phase 2 IgG titers of 1:128. She was clinically stable from the infection perspective at last follow-up, 6.5 years after surgery.

**Case 3**

A 57-year-old female from South Dakota with a known 3.6 cm AAA was diagnosed with metastatic colon adenocarcinoma in March 2009, for which she underwent diverting ileostomy and bowel resection. She completed chemotherapy with fluorouracil, folinic acid, oxaliplatin, and bevacizumab in April 2010. In July 2010, a CT screen of the abdomen showed a fluid collection consistent with hematoma surrounding the distal aorta with stable aneurysm size. She had no intervention at that time. In January 2011, a repeat CT of the abdomen/pelvis showed

![Figure 3](https://academic.oup.com/ofid/article-abstract/4/4/ofx192/4139802/4139802)

Figure 3. (A) Vascular debridement and reconstruction with rifampin-soaked graft. (B) Debridement of the vertebrae.

*Coxiella burnetii* Noncardiac Endovascular Infection With Contiguous Vertebral Osteomyelitis • OFID • 3
a 4.3 cm × 4 cm AAA with considerable intraluminal thrombus, thickening of the anterior and posterolateral walls, and an irregularly shaped infiltrative-appearing structure between the aorta and left psoas muscle. Erosive changes of the anterior margins of multiple vertebral bodies were increased compared with July 2010. The patient underwent extensive retroperitoneal debridement, repair of the juxtarenal mycotic aneurysm with a rifampin-soaked InterGard Dacron aorto-bi-iliac graft, and debridement of L2-L4. Histopathology of tissue revealed dense fibrosis with chronic inflammation, with no evidence of fungal or mycobacterial organisms on special stains. Cultures for bacteria, fungi, and mycobacteria were negative. TB-IGRA, Brucella, and fungal serologies were negative. Cultures and PCR were positive from the lumbar spine and aorta. A TEE did not show evidence for endocarditis. She had spent time at a cattle and sheep ranch in Texas shortly before the onset of infection and lived in a rural community with surrounding livestock. She was started on ciprofloxacin and doxycycline; due to persistent nausea, her regimen was changed to hydroxychloroquine and delayed-release doxycycline. She completed 1 year of therapy with hydroxychloroquine and doxycycline and was transitioned to chronic suppressive doxycycline therapy. Colon cancer treatment was re-initiated in 2012 due to metastatic disease with infusional fluorouracil and bevacizumab therapy followed by folinic acid (leucovorin)–fluorouracil-irinotecan plus cetuximab therapy. She remained stable from the infection perspective until November 2016, when she passed away from metastatic colon cancer.

Case 4

A 67-year-old male from Minnesota underwent an endovascular aortic stent graft repair of an AAA in November 2014. The patient remained well until November 2015, when he developed right-sided hip pain and malaise. A CT angiogram demonstrated a large hyperdense collection consistent with an abscess extending from the posterior portion of the aneurysm sac to the psoas and iliopsoas muscles; no leak from the endoluminal graft was noted. CT-guided aspiration of this intra-abdominal collection yielded no growth on bacterial, fungal, and mycobacterial cultures. No antimicrobial therapy was initiated at that time. Over the next 6 months, he developed low back pain, sweats, and malaise. Repeat CT in June 2016 demonstrated an enlarging 4 cm × 2.1 cm × 5.5 cm collection extending from the posterior portion of the aneurysm sac, with L2-L3 vertebral osteomyelitis. Repeat CT-guided aspiration of this intra-abdominal collection again yielded negative cultures. He remained off antimicrobial therapy throughout.

Notable exposures included residing in a rural area with surrounding livestock, regularly drinking unpasteurized goat milk from a nearby farm for 12–18 months prior to onset of symptoms, hunting wild game, and spending his childhood on a dairy farm. He had never traveled overseas.

On examination in June 2016, his temperature was 36.3ºC, BP was 128/84 mmHg, and pulse was 97/min. He was nontoxic, appearing with unremarkable cardiac examination. No spinal tenderness was present; however, pain was elicited on right hip flexion. Laboratory tests showed a hemoglobin of 15.3 g/dL, WBC of 9.5 × 10^9/L, platelets of 227 × 10^9/L, serum Cr of 1.0 mg/dL, sedimentation rate of 20 mm/h, CRP of 27.9 mg/dL (normal reference range < 0.8 mg/dL), and normal liver function tests.

CT of the abdomen and pelvis and a lumbar spine MRI in June 2016 showed a multiloculated abscess extending from the aneurysm sac into the prevertebral space and along the left iliopsoas muscle, contiguous with erosions of L2-L3 vertebrae. Enhancement and edema through L2-L3 and bilateral psoas muscle thickening and enhancement were noted. Indium-111 tagged white blood cell scan noted no increased uptake. He underwent explantation of the aortic endograft with placement

Figure 4. (A) Direct immunofluorescence staining with anti-Coxiella polyclonal antiserum used to confirm growth of C. burnetii in human fibroblast cells inoculated with aortic aneurysm tissue. Green fluorescent areas indicate the presence of Coxiella burnetii. (B) Presence of C. burnetii by Gimenez staining in human fibroblast cells inoculated with aortic aneurysm tissue.
of an aorto-bi-iliac cryopreserved allograft; a copious amount of cheese-like material was debrided from the aneurysm sac and psoas muscle. The L2-L3 disc space was also debrided. Aortic tissue and abscess cultures were negative for bacterial, fungal, and mycobacterial growth. Operative pathology noted inflammation with negative stains for bacteria, fungi, and mycobacteria.

He completed a 6-week course of empiric vancomycin, cefepime, and metronidazole without significant clinical improvement. Repeat inflammatory markers were elevated, with a sedimentation rate of 73 mm/h and CRP 75.8 mg/dL. C. burnetii IgG phase 1 titer was 1:1634, and IgM phase 1 titer was <1:16; the phase 2 titers were IgG 1:8192 and IgM <1:16, consistent with a chronic infection. TEE showed no valvular abnormalities. He was started on doxycycline and hydroxychloroquine for an intended minimum 24-month course.

**DISCUSSION**

Q fever is a zoonotic infection that occurs worldwide. About 120–170 cases are reported annually in the United States, with an incidence of approximately 0.38 cases per million persons per year [4, 5]. C. burnetii is a fastidious obligate intracellular rickettsial microorganism [6, 7]. Its ability to form a hardy spore-like state allows it to resist desiccation, extreme temperatures, and common chemical disinfectants and enables it to persist in the environment for long periods of time [8, 9]. C. burnetii’s virulence, potential for aerosolization, and resistant environmental state have led it to be classified as a Category B bioterrorism organism by the US Centers for Disease Control [10, 11].

Transmission is primarily by inhalation of small droplets or dust containing aerosolized organisms or by ingestion of contaminated dairy products [7, 10]. Less commonly, human-to-human transmission has been reported through blood transfusion, specimen handling by laboratory workers, and sexual contact [12–15]. In addition, multiple tick species have been shown to be competent vectors of C. burnetii, with rare reported cases of possible human transmission [16–18]. It can form reservoirs in mammals, arthropods, and birds, with the primary reservoirs being cattle, sheep, and goats [5, 19]. Animals are usually asymptomatic but shed the bacteria in reproductive secretions, milk, urine, or feces [20]. Conventionally, C. burnetii was recognized as an infection affecting farmers, agricultural or abattoir workers, veterinarians, or those who have contact with parturient animals [10]. Most cases in the United States, however, had no reported exposures to sheep, cattle, or goats [5]. In Europe and North America, outbreaks and isolated cases have been reported without recognized exposure to animals or animal products. These are thought to be due to airborne spread of Coxiella spores from farmland to residential and urban areas [21–24].

C. burnetii infections may present acutely or as a chronic infection. Around 50% of acute infections are mild or asymptomatic. Symptomatic acute infections present as nonspecific febrile illness, pneumonia, hepatitis, or other organ involvement [19]. Chronic infections occur in less than 5% of acutely infected patients and may manifest more than 6 months after exposure. C. burnetii endocarditis accounts for 60–70% of all chronic infections. Risk factors for chronic infection include immunosuppression and pregnancy, with a mortality risk of 5–40% [6, 19].

**Coxiella burnetii Osteoarticular Infections**

Osteoarticular C. burnetii infections are rare, reported in 7 of 313 (2%) patients with chronic C. burnetii in 1 study [19]. These include osteomyelitis of the long bones, sternum, and vertebrae, or multifocal osteomyelitis [19, 25–31]. C. burnetii native joint infection has been reported in 6 cases, half of which were monoarticular, involving the hip or knee. Joint aspiration often shows a mononuclear predominance. A prosthetic knee joint infection was recently reported by our group [32].

**Coxiella burnetii Vascular Aneurysms or Vascular Graft Infections**

C. burnetii infections of vascular aneurysms or grafts have been reported from countries other than the United States [33–39]. A compilation of 30 cases [34, 40] of C. burnetii–infected aortic aneurysms (52.5%) or vascular grafts (47.5%) showed that most patients have some environmental exposure such as residing in rural areas, ingesting unpasteurized dairy products, and contact with livestock or domestic animals. The most common location of infection is the infrarenal portion of the aorta [41]. The most common symptoms were fever (82.5%), weight loss and abdominal pain (52.5% each), and fatigue (27.5%) [34].

**Coxiella burnetii Vascular Aneurysms or Graft Infections With Contiguous Spread to the Vertebrae**

Table 1 summarizes all reported cases of C. burnetii–infected vascular aneurysms or grafts with contiguous spread to the vertebrae. In the review by Botelho-Nevers et al., 23.3% (7/30) of patients with C. burnetii–infected vascular aneurysms or grafts had contiguous vertebral involvement [34]. Prior to our report, none have been reported as acquired in the United States. Of these 24 cases, 8 were infected abdominal aortic vascular grafts whereas the rest were aortic aneurysmal infections. The predominant location of these vascular infections was the infrarenal aorta while only 1 was a thoracic aortic aneurysm. Where information was available, surgical intervention was performed in all cases except for 1 [42]. There was no incident of contiguous vertebral osteomyelitis complicating any of the 122 cases of vascular aneurysm or graft C. burnetii infections in a recent report from the Netherlands [39].

Diagnosis of C. burnetii infection requires a high index of suspicion. It should be suspected when bacterial fungal and mycobacterial cultures are negative in the absence of prior antimicrobial exposure as this is a common cause of lack of growth in cultures. Histopathologic examination of infected tissue demonstrates noncaseating, occasionally necrotizing...
**Table 1. Cases of Reported Coxiella burnetii-Infected Vascular Aneurysm or Graft Infection With Contiguous Spread to Vertebrae**

<table>
<thead>
<tr>
<th>Case Number</th>
<th>Reference</th>
<th>Age/Sex/Location</th>
<th>Presentation</th>
<th>Exposures</th>
<th>Aneurysm/Graft</th>
<th>Vascular Surgery</th>
<th>Vertebal Debridement</th>
<th>Outcomes</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 Case 1</td>
<td>74/M/US</td>
<td>Back pain</td>
<td></td>
<td>Deer, livestock, unpasteurized milk</td>
<td>Infrarenal aortic graft</td>
<td>Yes</td>
<td>Yes</td>
<td>Died of secondary complications</td>
</tr>
<tr>
<td>2 Case 2</td>
<td>60/F/US</td>
<td>Back pain, weight loss</td>
<td></td>
<td>Remote farming exposure</td>
<td>Juxtarenal aortic graft</td>
<td>Yes</td>
<td>Yes</td>
<td>Well at 6.5 years postsurgery</td>
</tr>
<tr>
<td>3 Case 3</td>
<td>57/F/US</td>
<td>Asymptomatic—notted on staging CT for cancer</td>
<td>Cattle &amp; sheep farm exposure, rural residence</td>
<td>Infrarenal aortic aneurysm</td>
<td>Yes</td>
<td>Yes</td>
<td>Well at 6 years postsurgery, passed away due to malignancy</td>
<td></td>
</tr>
<tr>
<td>4 Case 4</td>
<td>67/M/US</td>
<td>Back pain, sweats, malaise</td>
<td>Unpasteurized goat milk, rural residence</td>
<td>Infrarenal aortic graft</td>
<td>Yes</td>
<td>Yes</td>
<td>Well 6 at months postsurgery</td>
<td></td>
</tr>
<tr>
<td>5 [46]</td>
<td>67/M/Canada</td>
<td>Back &amp; abdominal pain, increasing pseudoaneurysm size, positive serology, positive PCR from aorta</td>
<td>Unknown</td>
<td>Infrarenal aortic graft</td>
<td>Yes</td>
<td>No</td>
<td>Died at 18 months—PE, decreasing titers</td>
<td></td>
</tr>
<tr>
<td>6 [47]</td>
<td>64/M/France</td>
<td>Back pain, malaise, positive serology, positive PCR from psoas abscess</td>
<td>Unknown</td>
<td>Infrarenal aortic graft</td>
<td>Yes</td>
<td>No</td>
<td>Died at 3 months postsurgery</td>
<td></td>
</tr>
<tr>
<td>7 [48]</td>
<td>57/M/Jordan</td>
<td>Back pain, fever, positive serology, positive PCR on vertebrae</td>
<td>Remote unpasteurized milk</td>
<td>Infrarenal aortic aneurysm</td>
<td>Yes</td>
<td>No</td>
<td>Cured (20 months)</td>
<td></td>
</tr>
<tr>
<td>8 [49]</td>
<td>91/M/France</td>
<td>Fever, back pain, positive serology</td>
<td>Unknown</td>
<td>Infra-renal ruptured aortic aneurysm</td>
<td>Aortic allograft</td>
<td>No</td>
<td>Cured (20 months)</td>
<td></td>
</tr>
<tr>
<td>9 [50]</td>
<td>92/M/France</td>
<td>Back pain, positive serology</td>
<td>Unknown</td>
<td>Infrarenal aortic aneurysm</td>
<td>Aortic allograft</td>
<td>No</td>
<td>Lost to follow-up at 6 months</td>
<td></td>
</tr>
<tr>
<td>10 [51]</td>
<td>54/M/Israel</td>
<td>Back pain, positive serology</td>
<td>Unknown</td>
<td>Infrarenal aortic aneurysm</td>
<td>Dacron graft, extra-anatomic bypass + stent graft</td>
<td>Unknown</td>
<td></td>
<td></td>
</tr>
<tr>
<td>11 [52]</td>
<td>77/M/Netherlands</td>
<td>Increasing aneurysm size on surveillance, positive serology, positive PCR from blood and aortic tissue</td>
<td>Unknown</td>
<td>Infrarenal aortic aneurysm</td>
<td>Spiral vein reconstruction</td>
<td>Died postoperatively—ischemic colon</td>
<td></td>
<td></td>
</tr>
<tr>
<td>12 [31]</td>
<td>72/M/France</td>
<td>Back pain, weight loss, positive serology</td>
<td>Goat cheese</td>
<td>Aortobiiliac endograft</td>
<td>Graft explantation, extra-anatomic bypass</td>
<td>Unknown</td>
<td>Improved titers at 6 months</td>
<td></td>
</tr>
<tr>
<td>13 [42]</td>
<td>66/M/France</td>
<td>Lumbar pain, fever, positive serology, positive PCR from psoas abscess</td>
<td>Sporadic farm animal contact</td>
<td>Infrarenal aortic aneurysm</td>
<td>No</td>
<td>No</td>
<td>Doxycline + hydroxychloroquine for 3 years, cured</td>
<td></td>
</tr>
<tr>
<td>14 [42]</td>
<td>65/M/France</td>
<td>Weight loss, back pain, positive serology, positive PCR from psoas abscess</td>
<td>Unknown</td>
<td>Infrarenal aortic aneurysm</td>
<td>Yes</td>
<td>Yes</td>
<td>Cured</td>
<td></td>
</tr>
<tr>
<td>15–21 [34]</td>
<td>France</td>
<td>Unknown</td>
<td>Unknown</td>
<td>Infrarenal aneurysm (5), graft (1), thoracic aortic aneurysm (1)</td>
<td>Unknown</td>
<td>Unknown</td>
<td>Died (1), cured (7)</td>
<td></td>
</tr>
<tr>
<td>22 [40]</td>
<td>70/M/France</td>
<td>Fever, lumbar pain, aneurysm rupture, positive serology, positive culture &amp; PCR from aorta</td>
<td>Rural residence</td>
<td>Infrarenal aortic aneurysm</td>
<td>Dacron graft</td>
<td>Unknown</td>
<td>Died</td>
<td></td>
</tr>
<tr>
<td>23 [53]</td>
<td>70/M/France</td>
<td>Lumbar pain, aneurysm rupture, positive serology, positive culture from aorta</td>
<td>Unknown</td>
<td>Infrarenal aortic aneurysm</td>
<td>Dacron graft</td>
<td>Unknown</td>
<td>Cured</td>
<td></td>
</tr>
<tr>
<td>24 [54]</td>
<td>67/M/France</td>
<td>Lumbar pain, weight loss, positive serology, positive PCR &amp; culture from aorta/vertebrae</td>
<td>Rural residence</td>
<td>Infrarenal aortobifemoral Dacron graft</td>
<td>Graft explantation, extra-anatomic bypass</td>
<td>Yes</td>
<td>Cured</td>
<td></td>
</tr>
</tbody>
</table>

Abbreviations: CT, computerized tomography; PE, pulmonary embolism; PCR, polymerase chain reaction.
granulomas, often with characteristic doughnut-shaped ring granuloma surrounded by fibroblasts. Diagnosis of *C. burnetii* infection is usually confirmed with serologic testing and real-time *C. burnetii* DNA PCR on blood or serum. The serologic response is biphasic. Phase 1 positive titers (IgG, IgM) of ≥1:800 represent chronic infections while phase 2 (IgG, IgM) positive titers are seen with acute infections. Confirmation can also be made with the use of DNA amplification techniques and special cultures. DNA amplification from tissue is more sensitive than blood [43]. *C. burnetii* can be grown in a shell-vial of human embryonic lung fibroblast cells. Direct immunofluorescence can be used to confirm the presence of *C. burnetii* (Figure 4).

For 3 of our 4 patients, surgical tissue samples were sent to the French National Reference Center for Rickettsial Diseases in Marseille, France, for molecular testing and cultures. Management of infected vascular aneurysms or grafts with contiguous vertebral spread requires a combined surgical and medical approach. As outlined in Table 1, all patients for whom details are available except 1 underwent vascular and/or orthopedic debridement with or without reconstructive surgery. Antimicrobial treatment with doxycycline along with hydroxychloroquine or fluoroquinolones or rifampin is recommended for chronic *C. burnetii* infection. Measurement of doxycycline concentration in serum may be helpful in the management of severe infections [44]. Duration of antibiotic therapy for severe infections is at least 18–36 months; however, most cases involving prosthetic material require long-term suppressive therapy. Response to therapy should be assessed by measuring serial serological titers with the goal of phase 1 titers being less than ≤1:200 [45]. Persistence of high titers is associated with a high relapse rate. *C. burnetii* may survive for prolonged periods in monocytes and macrophages, which may in part explain their resistance to antimicrobial therapy.

Overall, of the 20 reported patients whose outcomes are known, 14 of 20 (70%) were cured or doing well and 6 (30%) died. In previous reports, an overall mortality of 25% have been known, 14 of 20 (70%) were cured or doing well and 6 (30%) resistance to antimicrobial therapy.

CONCLUSIONS

*C. burnetii* infections of abdominal aortic aneurysms or vascular grafts, with or without spread to contiguous structures, occur in the United States and other countries where *C. burnetii* is endemic. Although *C. burnetii* infections are uncommon in the United States, noncardiac endovascular infections do occur and may be under-recognized. Patients may present with nonspecific symptoms or symptoms related to vertebral extension. Diagnosis is dependent on elucidating the exposure history, recent or remote, and rigorously excluding other microbial etiologies prior to administering antimicrobial therapy. Withholding antimicrobial therapy prior to obtaining vertebral or vascular tissue cultures in cases where the diagnosis is unclear optimizes yield from cultures, allowing for identification of pathogens. Receiving antimicrobial therapy prior to cultures can confound the clinical picture as it is unclear if growth in a culture of common organisms has been compromised by the antimicrobial therapy or if the true pathogen is a fastidious organism. Therefore, whenever it is safe to do so, we advocate withholding antimicrobial therapy until microbiologic diagnosis can be established through culture, molecular, or serologic methods. In our cases, extensive deep surgical routine cultures were negative; the withholding of antimicrobial therapy was an important factor for the “true culture-negative” work-up, which ultimately yielded the diagnosis. Understandably, withholding antimicrobial therapy may not be feasible in critically ill patients. Without surgery and removal of infected foreign material, prognosis is likely poor. Surgical intervention requires a multidisciplinary approach with vascular and orthopedic surgical teams. These patients require initial multidrug therapy to control infection, followed by, usually, lifelong suppressive therapy together with close follow-up to monitor response.

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