Nitrate supply to grapevine rootstocks – new genome-wide findings

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Understanding the plant response to nitrate availability is crucial for sustainable agriculture. In viticulture, there is an additional element to consider: the choice of scion–rootstock couple, which allows the management of environmental cues (including nitrate availability) and productivity. Using the two rootstocks 1103 Paulsen and Riparia Gloire de Montpellier, known to confer different vigour to grafted Cabernet Sauvignon scions, Cochetel et al. (2017) have now performed the first genome-wide transcriptome study indicating the genetic basis of the response to heterogeneous nitrate supply in this situation.

Some of the first written evidence of the use of the grapevine (Vitis vinifera) cultivation is an ancient Sumeric text, interpreted by historians as referring to a technique for coping with an environmental stress (Mudge et al., 2009). From its ancient discovery through development in the 19th century, today grafting is applied to more than 70 woody perennial crop species cultivated for their fruits (Warschefsky et al., 2016). As reported by Warschefsky et al. (2016), among the wide list of primary targets for rootstock selection is a capacity to confer tolerance to biotic and abiotic stresses (such as nutrient scarcity), and in parallel to assure the best possible productivity of the scion. Nitrogen availability and nitrogen-use efficiency (NUE) are now high-priority research topics in the context of sustainable agricultural development (Zhang et al., 2015) and, during recent decades, the molecular mechanisms of nitrogen sensing in fluctuating environments have been deciphered (reviewed by O’Brien et al., 2016).

Different rootstocks can differently modulate the growth of a given grapevine scion. For example, a Cabernet Sauvignon (CS) scion grafted onto a Riparia Gloire de Montpellier (CS/RGM) rootstock is less vigorous than the same CS scion grafted onto a 1103 Paulsen (1103P) rootstock (Lecourt et al., 2015). The Cochetel et al. paper is the first report on the transcriptional reprogramming which occurs in these rootstocks (RGM and 1103P) when subjected to a nitrate-varying environment. The experimental set-up allowed the identification of modules of genes that were: (i) specifically linked to a rootstock genotype, (ii) common to the grapevine root response to nitrate and (iii) specific to the nitrate response of one variety of rootstock. The large amount of information obtained in this work on rootstock-specific gene modules will be a valuable source of molecular markers for selecting top-performing rootstock–scion combinations.

Hubs for nitrate responsiveness of rootstocks

Studying gene regulatory networks allows the identification of hubs, which are potentially key regulators of a specific condition (genotype and/or treatment). This approach was used to identify key factors that regulate nitrogen-responsive networks in Arabidopsis (Gutiérrez et al., 2008; Canales et al., 2014). Even though phylogenetically distant, Cochetel et al. have now demonstrated that grapevine and Arabidopsis share a common set of genes regulated by nitrate availability. For example, they show that VvNRT2.4a, a member of the high-affinity nitrate transporter family, is modulated by N supply in the two rootstocks.
Searching for N-specific/rootstock-specific hubs, they found two candidates: a BTB-like protein-coding gene and a TCP20 homologue. The BTB-like coding gene belongs to the modules correlating with high nitrate supply in RGM. In Arabidopsis and rice BT2, a BTB-like protein, is involved in the repression of high-affinity nitrate transporters in low nitrate conditions (Araus et al., 2016). In RGM roots, Cochetel et al. found higher VvNRT2.4a expression levels and nitrate content. In spite of this difference between the two species, BTB proteins were found to modulate nitrogen-use efficiency and to have an impact on leaf development (Araus et al., 2016), suggesting a role of BTB-like proteins in the control of CS scion growth by RGM. In parallel to this, the second hub found by Cochetel et al., the TCP20 homologue, which is RGM specific and not N-responsive, could be a key element for understanding RGM adaptation to N-poor soils. Very recently it was found that AtTCP20 interacts with NLP proteins and controls root development in N-deficient conditions (Guan et al., 2017). If this TCP20 homologue in RGM determines its aptitude to low-N adaptation, responding very rapidly to high nitrate supply, it might open new possibilities for rootstock selection.

An N-responsive, tri-partite system

Ruffel et al. (2011) identified sets of genes responding to the local or systemic response to nitrate availability, and the split-root system used by Cochetel et al. is an extremely useful tool for exploring this area. It is now clear that the root system response involves some molecular actors expressed in the foliar part of the plant (Ruffel et al., 2011, 2016; Ohkubo et al., 2017). The scion–rootstock combination effect arises from a bi-directional interaction, in which signals coming from the roots affect the scion physiology and vice versa. Adding the nitrate-heterogeneous compartments of the split root, we obtain a tri-partite system (the scion and the two parts of the split-root system). In a heterogeneous soil, a nutrient signal coming from one side of the root system should reach the distant parts of the root passing through the scion. Following this idea, it would be remarkable if the different responses to nitrate supply observed in CS/1103P and CS/RGM grafting combinations were due to a scion element that differentially interacts with the two rootstocks. Cochetel et al. point out the strong responsiveness of N-related genes (e.g. NRT2.4a, NPF6.3) under high nitrate treatment in RGM compared to 1103P. It is an open question as to whether this high N-responsiveness is explained by the fact that CS also integrates (in an additive way) the low nitrate information coming from the other side of the RGM roots, but not from the 1103P roots. However, completion of the transcriptomic data with CS/RGM and CS/1103P under split homogeneous conditions (high and low nitrate) will provide the answer.

New directions: not just nitrate

High-throughput methodologies are powerful tools for understanding the complex response to nutrient availability. Thinking and focusing on a single response or pathway is too restrictive – the system is more complex and able to integrate different environmental signals from the roots, which often share points of cross-talk (Kellermeier et al., 2014; Briat et al., 2015). Interestingly, the GARP transcription factor function is among the gene categories enriched for 1103P and RGM rootstocks under heterogeneous N supply. Recently it was demonstrated that GARP transcription factors are at the convergence between nitrate and phosphate signals in Arabidopsis (Medici et al., 2015; Nagarajan et al., 2016). In addition, the authors found that strigolactone biosynthesis genes were among the most tightly regulated in low nitrate conditions, in a rootstock-dependent manner. Strigolactones are important hormonal regulators of the phosphate starvation response (Sun et al., 2014; Kumar et al., 2015). Since N and P are fundamental macronutrients for plant biomass determination, the findings of Cochetel et al. suggest that a fine tuning of the three steps of acquisition, assimilation and allocation of N and P mineral forms takes place in grapevine rootstocks and is linked to the control of the scion growth. The close relationship between N and P levels in grafted grapevines was already studied in a root ionome analysis on the same varieties, which showed that N and P are finely modulated and influenced by the rootstock (Lecourt et al., 2015). Two particular unanswered questions remain. First, which molecular elements are responsible for the communication between rootstock and scion? And second, how is the N/P balance maintained even in a two-species assembly? Different mobile molecules such as hormones, miRNAs, proteins or small peptides are already targets of investigation as mobile elements connecting leaves and roots in non-grafted plants. These might also be good targets for understanding scion–rootstock communication. More specifically, SSPs (Small Secretory Peptides) and in particular CLE and CEP peptides, which are known to be regulated by multiple nutrient deficiencies and involved in the autoregulation of mycorrhization in different species (de Bang et al., 2017), are emerging as an important long-distance signalling molecules and will certainly receive increasing attention in this research area.

Key words: Grafting, grapevine, nitrate, RNA-seq, rootstocks, scion–rootstock couple, transcriptome, viticulture.


References


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