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miR-125b and miR-532-3p predict the efficiency of rituximab-mediated lymphodepletion in chronic lymphocytic leukemia patients. A French Innovative Leukemia Organization study

Anne-Laure Gagez, Isabelle Duroux-Richard, Stéphane Leprêtre, Frédérique Orsini-Piocelle, Rémi Letestu, Sophie De Guibert, Edouard Tuaillon, Véronique Leblond, Olfa Khalifa, Valérie Gouilleux-Gruart, Anne Banos, Olivier Tournilhac, Jehan Dupuis, Christian Jorgensen, Guillaume Cartron and Florence Apparailly

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Correspondence:
g-cartron@chu-montpellier.fr

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The underlying in vivo mechanisms of rituximab action remain incompletely understood in chronic lymphocytic leukemia. Recent data suggest that circulating micro-ribonucleic acids correlate with chronic lymphocytic leukemia progression and response to rituximab. Our study aimed at identifying circulating micro-ribonucleic acids that predict response to rituximab monotherapy in chronic lymphocytic leukemia patients. Using a hierarchical clustering of micro-ribonucleic acid expression profiles discriminating 10 untreated patients with low or high lymphocyte counts, we found 26 micro-ribonucleic acids significantly deregulated. Using individual real-time reverse transcription polymerase chain reaction, the expression levels of micro-ribonucleic acids representative of these two clusters were further validated in a larger cohort (n=61). MiR-125b and miR-532-3p were inversely correlated with rituximab-induced lymphodepletion (P=0.020 and P=0.001, respectively) and with the CD20 expression on CD19+ cells (P=0.0007 and P<0.0001, respectively). In silico analyses of genes putatively targeted by both micro-ribonucleic acids revealed a central role of the interleukin-10 pathway and CD20 (MS4A1) family members. Interestingly, both micro-ribonucleic acids were negatively correlated with MS4A1 expression, while they were positively correlated with MS4A3 and MS4A7. Our results identify novel circulating predictive biomarkers for rituximab-mediated lymphodepletion efficacy in chronic lymphocytic leukemia, and suggest a novel molecular mechanism responsible for the rituximab mode of action that bridges miR-125b and miR-532-3p and CD20 family members. (clinicaltrials.gov Identifier: NCT01370772).
Introduction

Micro-ribonucleic acids (miRNAs) are a class of small noncoding RNAs that regulate gene expression at the post-transcriptional level and play an important regulatory role in many cellular processes.1 Deregulated expression of miRNAs could play a critical oncogenic or tumor-suppressor role and has therefore been associated with cancer, including hematological malignancies and, especially, B-cell lymphomas.2,3 MiRNAs correlate with the clinical characteristics or outcome of chronic lymphocytic leukemia (CLL) patients, allowing the identification of CLL subgroups with worse outcomes.2,3 Some of these miRNAs contribute to the deregulation of pathways involved in CLL oncogenic processes, such as PI3K/Akt (miR-22), NF-κB (miR-9 family), or toll-like receptor 9 (miR-17–92 family).4,5 The B-cell receptor (BCR) signaling pathway was recently shown to be directly regulated by miR-34, miR-150, and miR-155 in CLL,6,7 in addition to BCL2 (miR-15a/16), TCL1 (miR-29 and miR-181), P53 (miR-15a/miR-16-1 cluster, miR-17-5p, miR-29c and miR-34a), or PTEN (miR-125a and miR-21).8,9 Response to a CLL treatment could also be regulated by miRNAs. Thus, patients refractory to fludarabine exhibit significantly higher expression levels of miR-21, miR-148a and miR-222 than fludarabine sensitive patients.10 The activation of P53-responsive genes was found only in fludarabine resistant patients, suggesting a possible link between abnormal miRNA expression and P53 pathway dysfunction in non-responder patients. Links between miRNAs, fludarabine-refractory CLL and genomic abnormalities were further demonstrated, underlining the crucial role of MYC and P53 regulatory networks in determining cell response to fludarabine in CLL.11,12 Finally, in a prospective clinical trial aiming at evaluating the contribution of 17p deletion or TP53 mutation in fludarabine-refractory CLL, miR-34a expression at baseline was lower than in a control cohort of CLL non-refractory patients.13 The detection of miRNAs in serum and other body fluids under physiological and pathological conditions raises the possibility of using them as diagnostic or prognostic biomarkers.14,15 Visone et al. found that blood expression levels of miR-181b decreased in progressive CLL patients but not in patients with stable disease.16 Recently, we have shown that high miR-125b blood concentration can predict clinical benefit of rituximab (MabThera®, Rituxan®) treatment in CLL patients. The study herein aimed at identifying a blood-based miRNA signature at diagnosis, before rituximab monotherapy, that predicts rituximab efficacy in CLL patients. It also aimed at shedding light on the miRNA-mediated molecular mechanisms involved in rituximab’s mode of action in vivo.

Methods

CLL2010FMP protocol

A prospective, randomized, open-label, phase II study (CLL2010FMP, clinicaltrials.gov Identifier: 01370772) included 140 treatment-naive patients (aged 18-65 years) diagnosed with confirmed chronic lymphocytic leukemia, according to the International Workshop on Chronic Lymphocytic Leukemia (IWCLL) 2008 criteria, and Binet stage C, or with active Binet stage A or B.23 An additional inclusion criteria was the absence of 17p deletion, assessed by fluorescence in situ hybridization (FISH) (<10% positive nuclei). Each patient provided written informed consent before enrolment. Participating centers are listed in the Online Supplementary Information. Patients were stratified according to IGHV mutational status, FISH analysis (11q deletion or not) and were randomly assigned to receive either 6 cycles of chemioimmunochemotherapy combining fludarabine, cyclophosphamide, and rituximab (FCR) (rituximab 375mg/m² for the first course, Day (D) 1 and 500mg/m² for the others, fludarabine 40mg/m²/D1-D4, cyclophosphamide 250mg/m²/D1-D4) every 28 days, or Dense-FCR with an intensified rituximab pre-phase (500mg on D0, and 2000mg on D1, D8 and D15) before initiating the standard FCR treatment. The main objective was to increase the complete response rate with undetectable minimal residual disease three months after treatment, as published previously.16 In the study herein we have explored miRNA signature in a cohort of 123 patients, 61 receiving rituximab pre-phase before immunochemotherapy (Dense-FCR arm) and 62 receiving the FCR chemotherapy (standard arm). The study was approved by the institutional ethics committee of each participating center according to the principles of the Declaration of Helsinki.

Gene expression analysis

Details are described in the Online Supplementary Information.

FCGR3A genotyping

Single-step multiplex allele-specific PCR assays were performed as initially described by Dell’Ozzo et al. introducing minor modifications (for details see the Online Supplementary Information).

IL-10 competent CLL cells identification

Interleukin (IL)-10-competent CLL cell counts were determined by flow cytometry analysis of IL-10 production. Peripheral blood
mononuclear cells (PBMCs) were purified from peripheral blood samples of the Dense-FCR arm of the protocol using Ficoll-Hypaque density gradients (Eurobio, Courtaboeuf, France). Clonal CLL cells were identified as CD19+ CD5+ CD20 int lymphocytes with a previously described protocol. Analyses were performed on a CyAnTM ADP flow cytometer (Beckman Coulter, Brea, CA, USA).

### Cell surface CD20 expression analysis

CD20 expression was quantified using CD20-PE QuantiBRITE™ reagents (Ratio 1:1) according to manufacturer's recommendations (BD Biosciences, Le Pont-de-Claix, France). Calibration and quantification were performed using a FACSCANTO II cytometer (BD, Biosciences, Le Pont-de-Claix, France), and details are in the Online Supplementary Information. Circulating CD20 antigen was evaluated by considering the lymphocyte count and blood volume for each patient.

### Statistical analysis

The distribution of data was tested with the Shapiro-Wilk test. A X² or Fisher’s test were used to compare categorical data. For numerical data, medians were compared using a Student’s *t*-test or Mann-Whitney test. All variables with a *P* value <0.10 in univariate analysis were included in an intermediate model. The final model variables were determined by backward selection using a Student’s *t*-test (*P* <0.05 as significant model). The Spearman’s correlation test was used to assess the association between two numerical data. All statistical analyses were performed at the conventional two-tailed α level of 0.05 using R software version 3.0.2.10.

### Results

#### Patients’ characteristics

Sixty-one CLL patients were allocated to receive rituximab pre-phase. Their clinical and biological characteristics are presented in Table 1. They did not differ from the entire cohort. Median age was 58 years (interquartile range (IQR): 55-61), 28% were females and 77% were

### Table 1. Patients’ characteristics for the protocol CLL2010FMP

<table>
<thead>
<tr>
<th>Cohort (n=123)</th>
<th>FCR (n=62)</th>
<th>Dense FCR (n=61)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Age (years)</strong></td>
<td>-</td>
<td>58.45 (52.82-61.83)</td>
</tr>
<tr>
<td><strong>Women</strong></td>
<td>33 (26.83)</td>
<td>16 (25.81)</td>
</tr>
<tr>
<td><strong>Binet stage AB</strong></td>
<td>91 (73.98)</td>
<td>44 (70.97)</td>
</tr>
<tr>
<td><strong>ECOG 0</strong></td>
<td>86 (69.92)</td>
<td>41 (66.13)</td>
</tr>
<tr>
<td><strong>IGHV unmutated</strong></td>
<td>75/119 (63.03)</td>
<td>39/59 (66.10)</td>
</tr>
<tr>
<td><strong>β2 microglobulin (mg/L)</strong></td>
<td>114 (92.68)</td>
<td>3.05 (2.35-4.00)</td>
</tr>
<tr>
<td><strong>IL-10-competent B cells</strong></td>
<td>44 (35.77)</td>
<td>2.97 (0.98-9.58)</td>
</tr>
<tr>
<td><strong>FCGR3A</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>V/V</strong></td>
<td>14 (12.17)</td>
<td>8 (14.04)</td>
</tr>
<tr>
<td><strong>V/F</strong></td>
<td>55 (47.83)</td>
<td>26 (45.61)</td>
</tr>
<tr>
<td><strong>F/F</strong></td>
<td>46 (40.00)</td>
<td>23 (40.35)</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th><strong>miRNA ID</strong></th>
<th>Ct value in high lymphocyte count (mean)</th>
<th>Fold Change low/high lymphocyte count (mean)</th>
<th><em>P</em></th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Cluster 1</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>has-miR-323-3p</td>
<td>31.8</td>
<td>9.6</td>
<td>0.022</td>
</tr>
<tr>
<td>has-miR-99b</td>
<td>26.5</td>
<td>8.7</td>
<td>0.003</td>
</tr>
<tr>
<td>has-miR-326</td>
<td>31.3</td>
<td>7.5</td>
<td>0.001</td>
</tr>
<tr>
<td>has-miR-365</td>
<td>31.6</td>
<td>7.5</td>
<td>0.004</td>
</tr>
<tr>
<td>has-miR-125b</td>
<td>28.6</td>
<td>5.2</td>
<td>0.033</td>
</tr>
<tr>
<td>has-miR-193a</td>
<td>28.4</td>
<td>4.3</td>
<td>0.022</td>
</tr>
<tr>
<td>has-miR-193b</td>
<td>25.6</td>
<td>3.3</td>
<td>0.009</td>
</tr>
<tr>
<td>has-miR-211</td>
<td>28.0</td>
<td>2.8</td>
<td>0.012</td>
</tr>
<tr>
<td>has-miR-193a-5p</td>
<td>26.5</td>
<td>2.1</td>
<td>0.020</td>
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<tr>
<td>has-miR-212</td>
<td>26.4</td>
<td>1.8</td>
<td>0.012</td>
</tr>
<tr>
<td>has-miR-184</td>
<td>29.1</td>
<td>0.2</td>
<td>0.041</td>
</tr>
<tr>
<td><strong>Cluster 2</strong></td>
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<td></td>
<td></td>
</tr>
<tr>
<td>has-miR-328</td>
<td>22.9</td>
<td>5.1</td>
<td>0.015</td>
</tr>
<tr>
<td>has-miR-486</td>
<td>18.9</td>
<td>4.7</td>
<td>0.013</td>
</tr>
<tr>
<td>has-miR-332-3p</td>
<td>22.1</td>
<td>4.4</td>
<td>0.010</td>
</tr>
<tr>
<td>has-miR-484</td>
<td>17.4</td>
<td>3.7</td>
<td>0.028</td>
</tr>
<tr>
<td>has-miR-334-3p</td>
<td>22.2</td>
<td>2.9</td>
<td>0.004</td>
</tr>
<tr>
<td>has-miR-92a</td>
<td>17.2</td>
<td>2.9</td>
<td>0.001</td>
</tr>
<tr>
<td>has-miR-339-5p</td>
<td>23.3</td>
<td>2.8</td>
<td>0.010</td>
</tr>
<tr>
<td>has-miR-223</td>
<td>14.5</td>
<td>2.5</td>
<td>0.034</td>
</tr>
<tr>
<td>has-miR-423-5p</td>
<td>22.8</td>
<td>2.3</td>
<td>0.013</td>
</tr>
<tr>
<td>has-miR-652</td>
<td>21.2</td>
<td>2.2</td>
<td>0.038</td>
</tr>
<tr>
<td>has-miR-30c</td>
<td>16.6</td>
<td>2.0</td>
<td>0.029</td>
</tr>
<tr>
<td>has-miR-16</td>
<td>16.6</td>
<td>0.4</td>
<td>0.044</td>
</tr>
<tr>
<td>has-miR-26a</td>
<td>17.6</td>
<td>0.3</td>
<td>0.031</td>
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<tr>
<td>has-miR-29c</td>
<td>19.6</td>
<td>0.1</td>
<td>0.002</td>
</tr>
<tr>
<td>has-miR-29a</td>
<td>18.9</td>
<td>0.1</td>
<td>0.002</td>
</tr>
</tbody>
</table>

The cycle threshold (Ct) value is defined as the number of cycles required for the fluorescent signal to cross the background level. Fold Change is the fold ratio of the geometric means of miRNA expression from low and high lymphocyte count patients. The *P* value is determined by *t*-test and considered significant for *P* <0.05.
Binet stage A or B. Cytogenetic analysis demonstrated del11q in 20% of patients and median lymphocyte count was 89 G/L (IQR: 43-123).

**Blood miRNA expression profile discriminates CLL patients with high and low lymphocytosis**

Because miR-125b expression is reduced in CLL patients compared with healthy donors and is a key regulator of lymphoproliferation, we performed real-time PCR-based high-throughput miRNA arrays comparing CLL patients with high lymphocyte counts (>93.93 G/L, Q3) versus low lymphocyte counts (<11.66 G/L, Q1) at baseline. Q1 and Q3 were interquartile values of lymphocyte counts at D0 for patients of the Dense-FCR arm analyzed by the TaqMan low-density array (TLDA). After filtering (fold change ≥1.5 and cycle threshold (Ct) values <32) on the differentially expressed miRNAs, we found 5 miRNAs downregulated and 21 miRNAs upregulated in CLL patients with low lymphocyte counts compared with those with high lymphocyte counts (P<0.05) (Table 2). The heat map showed results of the unsupervised hierarchical clustering based on the significantly differentially expressed miRNAs (Figure 1A). Two patterns of miRNA expression profile named cluster 1 and cluster 2 were clearly identified according to lymphocyte counts. To confirm this finding, we selected four miRNAs (two from each cluster) based on technical criteria (21<Ct<29 and 2-fold difference between high and low lymphocyte count at D0 between two miRNAs in each cluster), and on the literature. We therefore assessed their expression in a cohort of 123 CLL patients (Figure 1B). Expression patterns of miR-193b, miR-125b, miR-652 and miR-532-3p were consistent with the array data. Scatter plots confirmed that increased lymphocyte counts were inversely correlated with the expression levels of miR-193b and miR-125b (P=0.03 and P=0.001, respectively) for cluster 1, and miR-652 and miR-532-3p (P=0.0017 and P=0.0001, respectively) for cluster 2 (Figure 1B). No significant correlation was found between individual miRNAs (miR-125b, miR-193b, miR-532-3p, miR-652) and clinical (age, Binet stage, Eastern Cooperative Oncology Group (ECOG)) or biological (IGHV mutation, cytogenetic abnormalities (del11q, del13q, trisomy 12), B10 frequency, or FcγRIIIa-158V/F polymorphism) parameters (data not shown). Finally, our results demonstrated that all 4 miRNAs were markedly downregulated in the blood of CLL patients displaying high lymphocyte counts.

miR-125b and miR-532-3p expression levels correlate with lymphodepletion observed after rituximab treatment

We hypothesized that lymphocyte depletion observed after rituximab infusions was related to *in vivo* rituximab activity, and then monitored the lymphocyte depletion in our CLL cohort following four rituximab infusions at D22. We thus determined whether a correlation existed between the miRNA expression profile that discriminates CLL patients with high lymphocytosis before treatment and the efficacy of lymphocyte depletion with rituximab. The median lymphocyte count was 89 G/L (range: 4-351) before rituximab treatment and 3 G/L (range: 0.1-189) after four rituximab infusions, bringing the median lymphocyte depletion after rituximab treatment up to 95.9% (range: 5.0-99.6).
Only 2 out of the 4 validated miRNAs, namely miR-125b and miR-532-3p, were significantly correlated with lymphodepletion, whereas miR-652 and miR-193b did not correlate with lymphodepletion. Thus, the lymphodepletion rate was inversely correlated with the expression levels of miR-125b and miR-532-3p ($P=0.020$ and $P=0.001$, respectively), as shown in the scatter plots in Figure 2. However, no correlation was found between miRNAs and the clinical response assessed 3 months after immunochemotherapy by FCR.

Our group recently showed that the frequency of IL-10-competent B cells adversely impacts lymphodepletion following rituximab treatment of CLL patients and also correlates with clinical response assessed 3 months after immunochemotherapy by FCR. Because we reported that the IL-10-competent regulatory B cells frequency predicts efficacy of rituximab-mediated lymphodepletion and clinical CLL outcome, we integrated this third variable in our analysis. Logistic regression analyses showed that only IL-10-competent B cells frequency and miR-532-3p were associated with 90% lymphodepletion after rituximab monotherapy (odds ratio (OR)=0.87; 95% confidence interval (CI)=0.76-0.97; $P=0.014$, and OR=0.0002; 95% CI=<10-4-0.34; $P=0.029$, respectively). Receiver operating characteristic curve (ROC) using IL-10-competent B cells frequency and miR-532-3p expression levels showed a highly discriminative power (area under the curve (AUC)=0.795; 95% CI=0.652-0.939), allowing one to predict patients who will have more than 90% of lymphodepletion after rituximab pre-phase.

**Putative and validated target genes of miR-125b and miR-532-3p**

Using the miRWalk database, a tool that compares miRNAs binding sites resulting from 5 main existing miRNA-target prediction programs (DIANA, RNA22, PicTar, miRanda and TargetScan), we investigated putative target genes for the two miRNAs associated with rituximab-induced lymphodepletion. Two lists of putative target genes were obtained: 5053 genes for miR-125b and 6652 for miR-532-3p. The Venny program, an interactive tool for comparing lists, identified 3151 common genes targeted by both miR-125b and miR-532-3p. We then compared them with transcriptomic datasets available for IL-10-competent B cells. Among the 104 genes differentially expressed in the study that compared IL-10- and IL-10 human regulatory B cells, 33 and 46 genes overlapped with miR-125b and miR-532-3p putative targeted genes, respectively. Importantly, 26 genes were common targets of both miRNAs (Figure 3A). Consequently, in the context of rituximab, which is known to target the pan-B-cell marker CD20/MS4A1, we wondered whether this gene could also be targeted by miR-125b and miR-532-3p. We found that both miRNAs were predicted to target MS4A1.

Pathway enrichment analysis was performed using the web-based bioinformatics application Ingenuity Pathway Analysis (IPA Ingenuity Systems) based on the in silico 26 predicted target genes common to miR-125b, miR-532-3p and differentially regulated in human IL-10/IL-10 regulatory B cells, as well as on MS4A1. A hierarchical layout was built with only miRNA/mRNA interactions displaying high predicted scores and for which the correlation was experimentally observed in humans (Figure 3B). All the 9 genes presented in this figure were associated with the IL-10 pathway (EGR3, IL1A, IL10, IL10RA, IRF4, IRF5, MS4A1, TLR7 and TSC22D3).

**Expression of CD20 family members, miR-125b and miR-532-3p on CLL cells**

We analyzed the association between miR-125b and miR-532-3p expression levels and the CD20 surface expression on CD19+/CD5+ CLL cells. In both cases, a significant inverse correlation was observed between CD20 protein and miRNA expression levels (Figure 4). A high expression level of CD20 at the surface of CLL cells correlated with a low expression level of miR-125b and miR-532-3p ($P=0.0007$ and $P<0.0001$, respectively). Since miRNAs are negative regulators of gene expression, we monitored the mRNA level of CD20 in the blood of CLL patients. CD20 (MS4A1) mRNA tended to be inversely correlated with both miR-125b and miR-532-3p (Figure 5A). Interestingly, the expression of two other members of the CD20 family, namely MS4A3 and MS4A7, might also be controlled by miR-125b and miR-532-3p. Indeed, both MS4A3 and MS4A7 mRNAs present putative binding sites for miR-125b and miR-532-3p, not only at the 3’UTR.

---

**Figure 2. Efficacy of lymphodepletion after rituximab treatment inversely correlates with miR-125b and miR-532-3p expression levels.** miR-125b ($r=-0.42$) (A) and miR-532-3p ($r=-0.49$) (B) expression levels were quantified by RT-qPCR. The lymphodepletion was measured 22 days after 4 doses of rituximab infusion and correlated with miRNAs (Spearman’s correlation test) (n=61).
region, but also in the promoter, 5’UTR and coding regions as shown in Figure 5B. MS4A7 mRNA levels were positively correlated with both miR-125b and miR-532-3p expression levels, whereas MS4A3 mRNA levels were positively correlated with miR-125b expression only. Overall, these results suggest that miR-125b and miR-532-3p might differentially control the expression of the CD20 family members.

Discussion

In the study herein, we investigated whether a blood-based miRNA signature can predict the efficacy of rituximab-mediated lymphodepletion in CLL patients, and provide some clue as to the underlying molecular mechanisms. We showed that miR-125b, miR-193b, miR-652 and miR-532-3p expression levels were inversely correlated with lymphocyte counts in untreated patients, and that miR-125b and miR-532-3p negatively correlated with lymphocyte depletion after rituximab monotherapy. Finally, our data suggest that both miR-125b and miR-532-3p might differentially control the expression of the CD20 family members.

Both miR-125b and miR-532-3p have been previously described in leukemia disorders. Recently, it has been shown that miR-125b was involved in specific subtypes of leukemia, either through IRF4 silencing, genetic deletion or chromosomal translocation as evidenced in B-cell leukemia or myeloid leukemia, respectively.40-42 One of the two genes encoding for the mature form of miR-125b, namely miR-125b-1, maps at 11q24, a chromosomal region that is close to the epicenter of 11q23 deletions found in CLL, and might explain why miR-125b expression is reduced in CLL patients compared to healthy donors.28 In the study herein, although the number of patients with a del11q was small, no correlation was
found with miR-125b expression levels. In a recent study investigating miRNA changes upon B-cell receptor stimulation in distinct subclasses of CLL patients, the expression of miR-532-3p was increased at 48 hours exclusively in CLL patients with stable disease. Like miR-125b, the role and implication of miR-532-3p in CLL are established, especially as it is strongly associated with progression-free survival in CLL.

Our data thus identify a novel prognostic relevance for miRNAs, and specifically for miR-125b and miR-532-3p, which are able to predict the efficacy of rituximab-mediated lymphodepletion, independently of the clinical outcome. In the study herein, logistic regression analysis showed that IL-10-competent B cells frequency and miR-532-3p were associated with lymphodepletion after rituximab pre-phase, whereas no correlation was found between miRNAs and the clinical response assessed 3 months after immunochemotherapy by FCR.

Taking advantage of the signatures of these 2 miRNAs for predicting the effect of rituximab, we searched for putative target genes according to these variables. Using available software and databases, we identified 3151 common putative target genes for miR-125b and miR-532-3p, which are able to predict the efficacy of rituximab-mediated lymphodepletion, independently of the clinical outcome. In the study herein, logistic regression analysis showed that IL-10-competent B cells frequency and miR-532-3p were associated with lymphodepletion after rituximab pre-phase, whereas no correlation was found between miRNAs and the clinical response assessed 3 months after immunochemotherapy by FCR.

An indirect implication of miR-532-3p on TLR7 gene expression mediated by an upregulation of IL-4 was reported in peripheral blood samples from CLL patients. The dysregulation of miR-532-3p was also evidenced in Binet A stage CLL patients as compared with a normal B-cell subset population. Among the miRNAs tested in relation with clinical data, miR-532-3p is part of a miRNA-based signature strongly associated with progression-free survival.

In humans, the MS4A4 gene includes CD20 and 18 other genes. Rituximab is a chimeric type I monoclonal antibody.
that specifically binds to inter-tetramers of CD20. CD20 could form homo- or hetero-oligomeric complexes with CD20 family members. Herein, we show that miR-125b and miR-532-3p putatively target other members of the MS4A family, including MS4A1 (coding CD20), MS4A3 (alias Htm4) and MS4A7. MS4A3 is a cell surface signaling molecule involved in the cell cycle of hematopoietic and tumor cells, whereas MS4A7 is expressed in lymphoid tissues. However, their role in the mode of action of rituximab is unexplored. Herein, we showed a significant correlation between miR-125b and miR-532-3p expression levels and three MS4A family members in CLL patients; CD20 being negatively correlated and MS4A3 and MS4A7 being positively correlated. Negative regulation of gene expression by miRNAs through translational repression and deadenylation-dependent decay of messengers is widely described. Emerging evidence reveal that miRNAs and their associated multiprotein complexes can directly or indirectly stimulate gene expression. GW182 is an essential component of the repressive miRNA complex that interacts with Ago1/2, leading to mRNA degradation. In conditions of quiescence, the absence of GW182 favors the interaction of FXR1 with Ago1/2 and induces the translation of the target-ed mRNA. In the context of CLL, which is a unique malig-nancy where quiescent B cells accumulate in the peripheral blood, we may hypothesize that miR-125b and miR-532-3p might differently affect the expression of the 3 MS4A family members and thus modulate the lymphodepletion outcome upon rituximab treatment. In Figure 6, we propose a schematic explanation on how miR-125b and miR-532-3p can act in CLL patient cells to modulate rituximab efficacy on lymphodepletion.

Overall, our results suggest that miR-125b and miR-532-3p are potential non-invasive biomarkers, detectable in the peripheral blood of CLL patients before treatment, which predict rituximab-mediated lymphodepletion efficacy. These miRNAs might also play a role in the molecular mechanisms involved in the rituximab-mediated mechanism of action, through their implication in the IL-10 pathway, including IL-10-competent B cells, and through the modulation of the MS4A family members’ expression. Further investigations in an independent cohort are needed to further explore these hypotheses.

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**References**
Increased chronic lymphocytic leukemia proliferation upon IgL stimulation is sus-


