

# Cause or consequence? Exploring the role of phenotypic plasticity and genetic polymorphism in the emergence of phenotypic spatial patterns of the European eel

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M. Mateo, Patrick Lambert, S. Tétard, M. Castonguay, B. Ernande, et al.. Cause or consequence? Exploring the role of phenotypic plasticity and genetic polymorphism in the emergence of phenotypic spatial patterns of the European eel. Canadian Journal of Fisheries and Aquatic Sciences, 2017, 74 (7), pp.987-999. 10.1139/cjfas-2016-0214. hal-01548904

# HAL Id: hal-01548904 https://hal.science/hal-01548904

Submitted on 28 Jun2017

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## 34 1 Abstract

35 The European eel (Anguilla anguilla), and generally, temperate eels, are relevant species for 36 studying adaptive mechanisms to environmental variability because of their large distribution areas 37 and their limited capacity of local adaptation. In this context, GenEveel, an individual-based 38 optimization model, was developed to explore the role of adaptive phenotypic plasticity and genetic-39 dependent habitat selection, in the emergence of observed spatial life-history traits patterns for eels. 40 Results suggest that an interaction of genetically and environmentally controlled growth may be the 41 basis for genotype-dependent habitat selection, whereas plasticity plays a role in changes in life-42 history traits and demographic attributes. Therefore, this suggests that those mechanisms are 43 responses to address environmental heterogeneity. Moreover, this brings new elements to explain 44 the different life strategies of males and females. A sensitivity analysis showed that the parameters 45 associated with the optimization of fitness and growth genotype were crucial in reproducing the 46 spatial life-history patterns. Finally, it raises the question of the impact of anthropogenic pressures 47 that can cause direct mortalities but also modify demographic traits, and act as a selection pressure.

48

Keywords: phenotypic plasticity, *Anguilla anguilla*, genetic polymorphism, life history theory,
modeling

## 51 2 Introduction

52 Life-history theory posits that the schedule and duration of life-history traits are the result of 53 natural selection to optimize individual fitness (Clark 1993; Giske et al. 1998). Optimal solutions 54 greatly depend on environmental conditions, and consequently, living organisms have developed 55 different adaptive mechanisms to address environmental variability. Among them, local adaptation 56 theory posits that natural selection favors the most well adapted genotypes in each type of 57 environment. In a context of limited genetic exchange between environments, this may lead to 58 isolation and speciation (Williams 1996; Kawecki and Ebert 2004). Phenotypic plasticity might also be 59 an adaptive response to an heterogeneous environment (Levins 1963; Gotthard and Nylin 1995; 60 Pigliucci 2005). Phenotypic plasticity refers to the possibility of a genotype to produce different 61 phenotypes depending on environmental conditions. In some cases, increases in fitness occur 62 because of plastic phenotypes compared to non-plastic ones, and that consequently, phenotypic 63 plasticity may be selected by natural selection (Schlichting 1986; Sultan 1987; Travis 1994).

64 Adaptation to environment heterogeneity is a key issue for temperate anguilids, Anguilla 65 anguilla, A. japonica, A. rostrata, three catadromous species that display remarkable similarities in 66 their life-history traits (Daverat et al. 2006; Edeline 2007). The European eel (A. anguilla) is widely 67 distributed from Norway to Morocco, grows in contrasting environments, and displays considerable 68 phenotypic variation. The species displays a complex life cycle: reproduction takes place in the 69 Sargasso Sea, larvae (or leptocephali) are transported by ocean currents to European and North 70 African waters, where they experience their first metamorphosis to become glass eels. These 71 juveniles colonize continental waters and undergo progressive pigmentation changes to become 72 yellow eels. The growth phase lasts between two and 20 years depending upon the region and sex of 73 the eels (Vollestad 1992). At the end of this stage, yellow eels metamorphose again into silver eels, 74 which mature during the migration to their spawning area in the Sargasso Sea. The population is 75 panmictic, resulting in a homogeneous population, from a genetic viewpoint (Palm et al. 2009; Als et

al. 2011; Côté et al. 2013). This panmixia combined with a long and passive larval drift limit the
possibility of adaptation to local environments. However, spatial patterns of different life traits,
including growth rate (Daverat et al. 2012; Geffroy and Bardonnet 2012), sex (Helfman et al. 1987;
Tesch 2003; Davey and Jellyman 2005), length at maturity (Vollestad 1992; Oliveira 1999), and habitat
use (De Leo and Gatto 1995; Daverat et al. 2006; Edeline 2007) are observed and correlated with
environmental patterns.

82 Growth rates greatly vary depending on latitude, temperature, sex (Helfman et al. 1987) but 83 also on habitat characteristics (Cairns et al. 2009). Indeed, eel can settle in a wide range of habitats 84 (De Leo and Gatto 1995; Daverat et al. 2006; Geffroy and Bardonnet 2012) and faster growth is 85 observed in brackish waters than in freshwater (Daverat et al. 2012). Slower growth in freshwater 86 habitats is sometimes assumed to be compensated for by lower mortalities and Edeline (2007) 87 suggested that habitat choice could be the result of a conditional evolutionary stable strategy. 88 However, Cairns et al. (2009) questioned this assumption because they did not observe strong 89 variation in mortality rates between habitats. Spatial patterns were also observed with respect to sex 90 ratios, with female biased sex ratios in the upper part of river catchments (Tesch 2003) and in the 91 northern part of the distribution range (Helfman et al. 1987; Davey and Jellyman 2005). However, sex 92 is not determined at birth but is determined by environmental factors (Oliveira 2001; Davey and Jellyman 2005; Geffroy and Bardonnet 2012). Population density also plays a role in this mechanism: 93 94 males are favored at high densities, whereas low densities favor females (Tesch 2003). This is 95 important because males and females have different life-history strategies (Helfman et al. 1987). The 96 reproductive success of a male does not vary with body size, and consequently, males are assumed 97 to follow a time-minimizing strategy, leaving continental waters as soon as they have enough energy 98 to migrate to the spawning grounds (Vollestad 1992). However, a female's reproductive success is 99 constrained by a trade-off between fecundity, which increases with length, and survival, which 100 decreases with length. Consequently, females are assumed to adopt a size-maximizing strategy

101 (Helfman et al. 1987). Strong differences in female length at silvering were observed among habitats102 and latitudes (Oliveira 1999).

103 Because local adaptation is impossible, this raises two questions: (i) are those life-history 104 trait patterns resulting from an adaptive response to environmental heterogeneity, and (ii) which 105 adaptation mechanisms have been selected. Despite panmixia, previous researchers (Gagnaire et al. 106 2012; Ulrik et al. 2014; Pavey et al. 2015) have detected genetic differences correlated with 107 environmental gradients and assumed that those differences were reshuffled at each generation. 108 Common garden experiments have been used to test the respective contributions of genetic and 109 plastic mechanisms on phenotypic differences observed in glass eels found in distinct locations. The 110 results revealed genetic patterns related to geographic zones in American eels, whereas individual 111 growth rates had a genetic basis and could be sex-dependent (Côté et al. 2009, 2014, 2015). Building 112 on this, Boivin et al. (2015) studied the influence of salinity preferences and geographic origin on 113 habitat selection and growth in American eels, demonstrating genetic-based differences for growth 114 between glass eels from different origins. However, these experiments also confirmed the 115 contribution of phenotypic plasticity that allowed individuals to develop quick and effective 116 responses to environmental variability (Hutchings et al. 2007). Several traits have been proposed as 117 plastic: growth habitats (Daverat et al. 2006; Edeline 2007), growth rates (Geffroy and Bardonnet 118 2012), and length at silvering (Vollestad 1992). Understanding the adaptive mechanisms that explain 119 this diversity is crucial to environmental conservation and management (Brodersen and Seehausen 120 2014).

As a result of a decline observed since the 1980s, the European eel is now listed as critically endangered in the IUCN Red List (Jacoby and Gollock 2014) and the European Commission enforced a European Regulation, which requires a reduction in all sources of anthropogenic mortality (obstacles, loss of habitat, fisheries, pollution, and global change) (Council of the European Union 2007). However, those anthropogenic pressures are not uniformly distributed (Dekker 2003) and acts on specific fractions of the stock isolated in river catchments (Dekker 2000), with heterogeneous life-

history traits because of the spatial phenotypic variability. This strong spatial heterogeneity of anthropogenic pressures affecting the eel population in Europe combined with this spatial phenotypic variability at both the distribution area and river catchment scales causes specific challenges for management, because it impairs our ability to assess the effect of anthropogenic pressures on the whole stock and to coordinate management actions (Dekker 2003, 2009).

132 Recently, a model called EvEel (evolutionary ecology-based model for eel) was developed to 133 explore the contribution of adaptive phenotypic plasticity in the emergence of observed phenotypic 134 patterns: sex ratio, length at silvering, and habitat use (Drouineau et al. 2014). Assuming fitness 135 maximization, the model was able to mimic most observed patterns at both river catchment and 136 distribution area scales. The result confirmed the probable role of adaptive phenotypic plasticity in 137 response to environmental variability. However, recent findings demonstrated the existence of 138 genetic differences in growth traits in a wide range of different habitats (Côté et al. 2009, 2014, 2015; 139 Boivin et al. 2015). Building on these new results, we developed GenEveel, a new version of EvEel, 140 which introduces a bimodal growth distribution (fast and slow growers) for individuals, as observed 141 by Côté et al. (2015), and considers phenotypic plasticity in life-history traits and demographic 142 attributes as in EvEel. Because individuals have different intrinsic growth and mortality rates, they 143 can be favored differently among environments, opening the door to conditional habitat selection. In 144 this study, we used GenEveel to test whether simultaneously considering genetically distinct 145 individuals and phenotypic plasticity improves model performance. Pattern orienting modelling was 146 used to detect the reproduced spatial patterns of EvEel and other patterns based on the distribution 147 of the different types of individuals.

### 149 **3** Materials and methods

### 150 3.1 Model description

151 The model description follows the Overview, Design concepts, and Details (ODD) protocol 152 (Grimm et al. 2006, 2010):

#### 153 **3.1.10verview**

#### 154 **3.1.1.1 Purpose**

155 GenEveel is a model based on a former model called EvEel (Drouineau et al. 2014), but 156 includes a genetic component. It is an individual-based population model that predicts emergent life-157 history spatial patterns depending on adaptive mechanisms and environmental heterogeneity. 158 Emergent patterns can later be compared to observed spatial patterns in freshwater life stages of 159 European eels in order to (i) confirm that observed phenotypic patterns can plausibly result from 160 adaptive responses to environmental heterogeneity, (ii) validate that phenotypic plasticity for length 161 at silvering, sex determination, habitat choice, and genetic polymorphism (slow growers and fast 162 growers) with conditional habitat selection can explain those patterns.

#### 163 **3.1.1.2 State variables and scales**

164 *Temporal scales:* the model simulates a population generation. It has no *sensu stricto* time 165 steps, but rather successive events: sex-determination and habitat selection, survival, and growth 166 until maturation.

167 *Entities and spatial scales:* a Von Bertalanffy growth function is assumed for individual 168 growth. Each individual *i* is characterized by an intrinsic Brody growth coefficient *K<sub>i</sub>* and a natural 169 mortality rate *M<sub>i</sub>*. Based on Côté et al. (2015), who observed two clusters in growth rates, we build a 170 simple quantitative-genetic model assuming that growth is coded for by a single gene with two 171 variations. Therefore, we assumed that there are two types of individuals called (i) fast-growing

individuals for which *Ki* = *Kfast* and *Mi* = *Mfast* and (ii) slow-growing individuals for which *Ki* = *Kslow*and *Mi* = *Mslow*. At the end of the simulation, individuals are characterized by a sex, length at
silvering, corresponding fecundity (if female), position in the river catchment, and survival rate until
silvering.

The river catchment environment was represented by a sequence of cells of the same size. The first cell represents the river mouth, whereas the *n*th cell represents the source of the river. Because it was observed that an individual grows faster downstream than upstream (Acou et al. 2003; Melià et al. 2006), we assumed that realized growth rate in a cell depends on both intrinsic growth rate and position in the catchment (i.e., cell) (see submodel section). Realized natural mortalities depend both on individual intrinsic mortality rates, position of the cell in the catchment, and number of individuals in the catchment (to mimic density-dependent mortality).

#### 183 **3.1.1.3 Process overview and scheduling**

184 The model has two main steps. In a first step, individuals select their growth habitat (a cell in 185 the catchment) and determine a sex (male or female) one after another (random order). To do that, 186 fitness is calculated for each combination of sex and cell (a quasi-Newton algorithm is used to 187 estimate the lengths at silvering that optimize female fitness in each cell). Individuals are assumed to select the combination with highest fitness given the choices made by former individuals. Once this 188 189 step is finished (i.e., all individuals have a growth habitat and sex), using the quasi-Newton algorithm 190 we estimated the optimal length at silvering for all females (males have a constant length at silvering) 191 given the positions of fishes from step 1, and then compute corresponding survival rates until 192 silvering and fecundity determination (Fig. 1). The process mentioned above is defined by the 193 computer algorithm (Figure 2):

194 (1) For each individual i:

195 - For each cell *x*:

196 • Compute  $\pi_m(x)$  given positions of individuals  $\{1, ..., i-1\}$ 

- Compute  $\max_{L_{s_f}} \pi_f(x, L_{s_f})$  given positions of individuals {1,...,i-1} 197 Put individual and determine sex by selecting maximum values within  $\max_{L_{s_f}} (x, L_{s_f})$  and  $\pi_m(x)$ 198 \_ (2) For each individual i: 199 200 For each cell x: \_ 201 if sex(i) = male 202  $L_{s}(i) = L_{sm}$ 203 • else 204  $L_s(i) = \arg_L \max(\pi_f(x, L_{sf}))$  given positions of individuals  $\{1, ..., n\}$
- where fitness is defined in equations 9 and 10 for females and males respectively.

#### 206 3.1.2 Design concept

#### 207 **3.1.2.1 Basic principles**

208 Consistent with life-history theory and optimal foraging theory, the model uses an 209 optimization approach in which individuals "respond to choices" so as to select and fix the adaptive 210 traits, maximizing their expected fitness given their environment (Parker and Maynard Smith 1990; 211 McNamara and Houston 1992; Giske et al. 1998; Railsback and Harvey 2013).

#### 212 **3.1.2.2** Emergence

Using the pattern-oriented modelling approach (Grimm and Railsback 2012), GenEveel compares predicted spatial patterns with those observed in real river catchments. Five emergent

- 215 population spatial patterns were analyzed from the literature:
- 216 (i) higher density downstream than upstream
- 217 (ii) higher length at silvering upstream than downstream
- 218 (iii) male-biased sex ratio downstream and female-biased sex ratio upstream

- (iv) more individuals characterized with the fast-growing genotype downstream than upstream,
- 220 which was mainly characterized by the slow-growing genotype
- 221 (v) the phenotypic response led to faster growth rate downstream than upstream.

#### 222 **3.1.2.3 Adaptation**

223 Individuals have three adaptive traits: sex-determination, length at silvering for females, and

224 choice of growth habitat (cell in the grid). These traits are assumed to maximize the predicted

225 objective function (i.e., the individual fitness).

#### 226 **3.1.2.4 Predictions**

- 227 We assumed that individuals could perfectly predict expected fitness given previous choices
- and could make the most appropriate choices.

#### 229 **3.1.2.5 Sensing**

In the model, individuals are able to "sense" fitness, which was a function of a densitydependent mortality and growth rate. In the real world, temperature and density would probably be the proximal cues because natural mortality and growth rates are strongly influenced by temperature (Bevacqua et al. 2011; Daverat et al. 2012).

#### 234 **3.1.2.6** Interaction

Interactions occurred through growth habitat selection, sex determination, and density-dependent mortality.

#### 237 **3.1.2.7 Stochasticity**

Stochasticity occurred at two levels. First, individuals were randomly affected by a slowgrowing genotype (Pr = 0.5) or by a fast-growing genotype (Pr = 0.5). Then stochasticity occurred in the order of individuals for step 1.

#### 241 **3.1.2.8 Observations**

- 242 Five spatial patterns were computed at the end of the simulation:
- 243 (i) number of individuals per cell
- 244 (ii) mean length at silvering per cell
- 245 (iii) sex ratio (proportion of females) per cell
- 246 (iv) ratio of fast-growing genotype per cell
- 247 (v) phenotypic response of mean realized growth rate per cell.
- 248 These five patterns corresponded to five patterns available in the literature. Simulated patterns (i),
- (iv), (v) was said to be consistent with the literature when a negative trend from downstream to
- 250 upstream was observed, whereas patterns (ii), (iii) were said to be consistent with the literature
- when a positive trend was observed from downstream to upstream.

#### 252 **3.1.3 Details**

#### 253 **3.1.3.1 Initialization**

At the beginning of the simulation, the catchment was empty. *N* individuals were created and attributed to the slow-growing or fast-growing genotype with probability 0.5 and had a length 7.5 cm. They had not yet entered the river catchment.

#### 257 3.1.3.2 Input data

We tested the model using a reference simulation. Values of parameters were obtained from the literature (Table 1). The outputs of the model were identified based on spatial patterns as previously defined in *Observations*.

#### 261 **3.1.3.3 Submodels**

- 262 Most of the submodels were similar to submodels from EvEel. Consequently, we provide 263 here only the novelties and the equations that are required for a better understanding of the model. 264 Further details are provided in Drouineau et al. (2014).
- Growth and silvering

Growth rate was assumed the outcome of an intrinsic Brody growth coefficient (*Ki*), which is modulated by an environmental effect. This combination resulted in a phenotypic growth rate. Within the river, growth rates were significantly faster downstream than upstream (even for the same individual). Therefore, we assumed that individual *i* would have a growth rate K(i, x) in cell *x* given by:

271 (1) 
$$K(\mathbf{i}, \mathbf{x}) = r_K \cdot K(\mathbf{i}, \mathbf{n}) + (K(\mathbf{i}, 1) - r_K \cdot K(\mathbf{i}, \mathbf{n})) \cdot \operatorname{cauchi}\left(\frac{x}{n}, \gamma_K\right)$$

272 (2) cauchit(x,  $\gamma$ ) = 1 -  $\frac{2}{\pi} \cdot \operatorname{atan}\left(\frac{x^2}{\gamma}\right)$ 

where  $r_k$  defined the ratio between upstream and downstream growth rate, K(i, 1) is the growth rate in cell 1, n is the total cells in the river catchment and *cauchit* was a mathematical function similar to the sigmoid function, but which allowed asymmetrical patterns (by modifying the parameter  $\gamma$ ) to model, for example, a small brackish area in the downstream part of the catchment and a large freshwater zone upstream.

278 Individual's growth was simulated by a Von Bertalanffy function:

**279** (3) 
$$L(t, i, x) = L_{\infty} \left[ 1 - e^{-K(i, x)(t-t_0)} \right]$$

where L(t, i, x) was the length at time t and  $L_{\infty}$  and K(i, x), the Von Bertalanffy parameters in cell x for individual i.

282 From this equation, we could calculate the time required to reach the length at silvering.

283 (4) As(i, x) = 
$$\frac{1}{K(i, x)} \cdot \log\left(\frac{L_{\infty} - Lg}{L_{\infty} - Ls(i, x)}\right)$$

where Lg was the length at recruitment and Ls(i, x) was the length at silvering, which was constant
for males, and a fitness maximizing variable for females.

• Survival

287 Mortality rate was assumed the result of three factors: density-dependence, intrinsic growth 288 rate, and *Mi* modulated by an environmental effect. Because natural mortality was sometimes 289 assumed to be smaller upstream than downstream (Moriarty 2003; Daverat and Tomás 2006), we 290 assumed that the instantaneous natural mortality without density-dependence in cell *x* for individual 291 *i*, M(i, x) was:

292 (5) 
$$M(\mathbf{i}, \mathbf{x}) = r_M \cdot M(\mathbf{i}, \mathbf{n}) + (M(\mathbf{i}, 1) - r_M \cdot M(\mathbf{i}, \mathbf{n})) \cdot \operatorname{cauchi}\left(\frac{x}{n}, \gamma_M\right)$$

where  $r_m$  is the ratio between upstream and downstream instantaneous mortality rate and M(i, 1) was the natural mortality in cell 1.

295 To account for the additional density-dependent mortality, we assumed that natural mortality 296 increased linearly with an intensity of density  $\alpha$  as in EvEeI:

**297** (6) 
$$M_d(i, x) = M(i, x) + N(i, x) \cdot \alpha$$

where *N* was the number of competitors in cell *x*. An eel was assumed a competitor if it had an intrinsic growth rate greater or equal to *Ki*. This corresponded to an asymmetric growth rate with larger individuals harassing smaller individuals. The basis of this assumption was the intraspecific competition, which leads to compete for limited resources between individuals of different sizes (Francis 1983; Juanes et al. 2002).

Given equation (4) and this survival rate, we could calculate the probability of surviving until silveringas:

**305** (7) 
$$p(i, x) = e^{-M_d(i, x)} \operatorname{As}(i, x) = \left(\frac{L_{\infty} - Lg}{L_{\infty} - \operatorname{Ls}(i, x)}\right)^{-\frac{M_d(i, x)}{K(i, x)}}$$

306 • Fitness

In any optimization model, an important component is the computation of the fitness.
 Because sexes adopt different life strategies, and following Drouineau et al. (2014), we assumed sex-

309 specific fitness functions. Males were known to adopt a time minimizing strategy (Helfman et al. 310 1987), with constant length at silvering. Therefore, male fitness was proportional to survival rate 311 until length at silvering. However, females follow a size-maximizing strategy in which length at 312 silvering was constrained by a trade-off between survival and fecundity (Helfman et al. 1987). 313 Consequently, we assumed that female fitness was the product of fecundity at an optimal length at 314 silvering (based on an allometric relationship, fecundity is assumed to be a power function of length) 315 multiplied by the probability of survival until this length at silvering. In the model, individuals were 316 assumed to determine their sex according to the relative potential male and female fitness. To make 317 fitness values comparable, we rescaled male fitness (which was the probability of survival) into an 318 expectation of egg production (the scale of female fitness). To do that, we multiplied the male 319 survival by a constant that would be similar to fertility. Hence, we had to specify a value for fertility 320 with an order of magnitude similar to fecundity. The first solution might be to fix the fertility value 321 equal to the fecundity of silver females having a length equal to male length at silvering. However, 322 with this solution, female fitness will always be greater (because females can optimize their length at 323 silvering). Consequently, fertility has to be slightly greater such that male fitness can be sometimes 324 be greater than female fitness (but not too much, to avoid male fitness always being superior). These 325 resulted in the following equations:

326 (8) fecundity 
$$\left(L_{s_f}(i,x)\right) = \left(a_1 + a \cdot L_{s_f}(i,x)^b\right)$$

where a1, a and b are the parameters of the allometric relationship linking fecundity and female length at silvering  $L_{sf}(i, x)$  (Andrello et al. 2011; Melia et al. 2006).

329 (9) 
$$\pi_f(i, x) = fecundity\left(L_{s_f}(i, x)\right) \cdot \left[\frac{L_{\infty} - L_g}{L_{\infty} - L_{s_f}(i, x)}\right]^{-M_f(i, x)}$$

330 (10) 
$$\pi_m(i, x) = fertility \cdot \left[\frac{L_{\infty} - L_g}{L_{\infty} - L_{s_m}(i, x)}\right]^{-M_m(i, x)}$$

### 331 3.2 Model exploration

### 332 3.2.1 Reference simulation

The reference simulation consisted of a simulation using parameter values in Table 1, i.e. the best set of values found in the literature. After simulating this scenario, we analyzed the different patterns. Mann-Kendall tests were implemented on each pattern to detect a monotonic upward or downward trend of the variable of interest confirming the spatial patterns previously defined. The correlation coefficient of this non-parametric test was denoted by τ.

338 **3.2.2 Experimental design** 

339 Simulation design is a classical tool to explore complex models. Typically, the goal is to assess 340 the sensitivity of results to uncertain model parameters. We developed such an experimental design 341 to (i) assess the influence of uncertain parameters on the simulated patterns (Table 1) and (ii) derive 342 environmental and population dynamics for all the patterns that were correctly modelled.

343 Seventeen uncertain parameters were identified in the model (Table 1) and they were dispatched 344 into twelve groups: number of glass eel entering the catchment freshwater (N), parameters that 345 impact the male fitness (fertility and male length at silvering,  $L_{sm}$ ), fast growing genotype ( $K_{fast}(i, 1)$ ) 346 and  $M_{fast}(i, 1)$ , slow-growing genotype ( $K_{slow}(i, 1)$  and  $M_{slow}(i, 1)$ ), proportion of individuals that grow 347 slowly (*propK*), intensity of density-dependence ( $\alpha$ ), cells of river catchment (*n*), regression 348 coefficient from fecundity at length (b), asymptotic length ( $L_{\infty}$ ), length at recruitment ( $L_{q}$ ), ratio 349 between upstream and downstream instantaneous growth and mortality rates ( $r_k$  and  $r_m$ ), and the 350 shape parameter of growth and mortality ( $\gamma_k$  and  $\gamma_m$ ). Groups were composed of parameters that 351 have are assumed to influence the model in similar directions, a method called group-screening 352 (Kleijnen 1987). A low and high value was set for each parameter around the reference value, with 353 20% variation (Drouineau et al. 2006; Rougier et al. 2015), except for three sets of parameters: 354 fertility and L<sub>sm</sub>(as a minimum value, fertility corresponded to the fecundity of a female with a length

355 at silvering equals to male length at silvering; otherwise, female fitness would always be superior to 356 male fitness), and growth genotypes (to avoid overlap between them), where the range of variation was less. We then conducted a fractional factorial design of resolution V ( $2^{12-4} = 256$  combinations). 357 358 This kind of orthogonal designs allows to explore main effects and first order interactions without 359 confusion. To account for model stochasticity, we conducted 10 replicates for each of the 256 360 combinations leading to 2560 simulations. The five patterns were calculated for each simulation 361 producing an output table with 2560 lines (one per simulation) and five columns containing the tau 362 value of the Mann-Kendall trend tests for each pattern (a negative tau value indicates a negative 363 trend from downstream to upstream while a positive tau indicates a positive trend from downstream 364 to upstream).

365

366 **4 Results** 

### 367 4.1 Reference simulation

368 In the reference simulation, GenEveel mimicked the five spatial patterns at the catchment 369 scale (Fig. 3). Males were concentrated in the downstream section of the river where density was 370 higher (Helfman et al. 1987; Tesch 2003; Davey and Jellyman 2005). Fast growers preferentially 371 settled in downstream habitats, whereas slow growers tended to move upstream to avoid 372 competition (De Leo and Gatto 1995; Daverat et al. 2006, 2012; Drouineau et al. 2006; Edeline 2007; 373 Geffroy and Bardonnet 2012). Regarding mean length at silvering (for males and females), a smaller 374 size at maturity was simulated in the downstream section of the river, whereas larger lengths were 375 occurred gradually throughout the catchment (Vollestad 1992; Oliveira 1999).

The Mann-Kendall test confirmed that the five patterns were mimicked in the simulation.More specifically, negative tau values confirmed a decreasing trend for density, ratio of fast growers

and mean realized growth rate; while positive tau values pointed to an increasing trend for ratio offemales and mean length at silvering (Table 2).

#### 380 4.2 Model exploration

For each combination, the 10 replicates provided the same results, confirming that thepatterns were not sensitive to stochasticity.

Interestingly, 310 simulations produced only females while 640 simulations produced only males. Simulations with only females corresponded to simulation where density-dependence  $\alpha$ ,  $L_{\infty}$ and the fecundity exponent *b* were simultaneously strong. Conversely, simulations with only males corresponded to simulations with a low *b* and a low  $L_{\infty}$ . With only one sex, it was not possible to calculate a spatial trend in sex ratio and with only males, it was not possible to calculate a trend of length at silvering.

Two questions were addressed here. In a first time, we compared the five patterns to see which of those patterns were frequently mimicked and which were less frequently mimicked. Then, we compared the sensitivity of the model to each group of parameters. To quantify this sensitivity to a group of parameters values, we compared the number of simulations that reproduce a given pattern when the group had modality (-) with the number of simulations and when the group had modality (+). A strong discrepancy indicated a high sensitivity to the group of parameters.

The Mann-Kendall tests of spatial patterns confirmed that the simulated patterns of abundance, ratio of fast growers, and mean realized growth rate were consistent with the literature in each of the 2560 combinations (Table 3). This result indicates that these model outputs do not depend on parameters values in the parameter space considered. Consequently, the assumptions about asymmetrical density-dependence and growth genotypes were enough to simulate catchment colonization.

401 Regarding length at silvering pattern, patterns were consistent in 1300 simulations of the 402 1920 simulations for which it was possible to calculate a pattern (Table 3, Fig. 4 for several examples).

403 This meant that, in situations where some females were produced, the pattern was consistent in 404 about 2/3 of the simulations. Length at silvering pattern appeared to be sensitive to most of the 405 parameters. The two most important were  $L_{\infty}$  and b: consistent pattern were much more frequent 406 with a modality (+) (respectively 990 and 940 simulations) for these two parameters than with 407 modality (-) (respectively 310 and 360 simulations). This is not surprising since with modality (-) for 408 those parameters, the model produced only males in 640 simulations. Two other groups of 409 parameters had a strong influence: male fertility/ male length at silvering and density-dependence. 410 Consistent patterns were more frequent with low male fertility and length at silvering (800 with 411 modality (-) vs 500 with modality (+)), and with limited density dependence (810 with modality (-) vs 412 490 with modality (+)).

For the female ratio pattern, 130 simulations produced consistent patterns over the 1610 simulations for which it was possible to calculate a pattern (Table 3). This pattern was mostly sensitive to four groups of parameters which correspond to the four most influential groups for the pattern of length at silvering. Patterns were consistent only when male fertility/length at silvering had modality (-) whereas Kslow/Mslow had modality (+). Moreover, consistent patterns were more frequent when  $L_{\infty}$  had a modality (-) and *b* a modality (+).

On the whole, 130 of the 2560 combinations produced results which were consistent for all
the five patterns (Table 4, Fig. 4 and Table S1). These 130 simulations corresponded exactly to the
130 simulations that produced consistent sex ratio patterns, demonstrating that this last pattern was
the more constraining (Fig. 5). Consequently, the interpretation regarding sensitive parameters was
similar.

To make a summary of those results: in situations where females' fitness was favored because of a strong  $L_{\infty}$  or a strong *b*, i.e. a high fecundity, the model produced only females. Conversely, when females were too penalised, model produced only males. Therefore, an equilibrium was required between males and females fitnesses to mimic all patterns. The patterns in length at silvering and sex ratio were the two most constraining patterns and were mainly sensitive to four

groups of patterns. These groups of parameters set the equilibrium between males and females fitnesses (male fertility and length at silvering, *b* and  $L_{\infty}$ ) and the advantages between slow and fast growers. Density-dependence was also important regarding the pattern on length at silvering. We can observed that the five patterns were consistent mostly when slow growers and females were not too penalized with respect to males and fast-growers.

434 Some of the patterns were indeed very constrained by model assumptions so it is hardly 435 surprising that they were mimicked by the model. For example, our constraints on mortality and 436 growth really constrained the distribution of fishes and probably the pattern of realized growth rates 437 in the catchment. However, those constraints were based on various observations in the literature 438 that have rarely been considered together to see if they make sense in a context of adaptive 439 response. We do not specify any constraints on the sex ratio, length at maturity and relationships 440 between sex ratio and slow/fast growers. Those results are really emerging patterns that are 441 consistent with the literature.

442

### 443 **5 Discussion**

#### 444 5.1 Adaptation to environmental variability: phenotypic plasticity and

445

### genetic polymorphism of European eel

The European eels, and more generally, temperate eels, display fascinating characteristics: catadromy with a long larval drift, large distribution area with contrasted growth habitats, panmixia, and strong phenotypic and tactic variability at different spatial scales. Consequently, this species is relevant to explore adaptive mechanisms to environmental variability. Phenotypic plasticity has been proposed as one such mechanism because of random mating and larval dispersal that prevent local selection pressures to generate habitat-specific adaptations, or local adaptation, from one generation to the next. Drouineau et al. (2014) developed the first model to explore the major role of

453 phenotypic plasticity in both life-history traits and tactical choices as an adaptive response to 454 spatially structured environments and density dependence. However, recently Gagnaire et al. (2012), 455 Pujolar et al. (2014), Boivin et al. (2015), Côté et al. (2015) and Pavey et al. (2015) demonstrated the 456 existence of genetic differences correlated with the environment, suggesting that part of the 457 observed phenotypic variability had a genetic basis.

Based on the approach developed by Drouineau et al. (2014), the objective of this study was to propose a model based on life-history theory and optimal foraging theory to explore the role of both adaptive phenotypic plasticity and genetic polymorphism with genetic-dependent habitat selection, in the emergence of phenotypic patterns. To that end, we used a pattern-oriented modelling approach, as developed by Grimm et al. (1996). This kind of approach compared field observed patterns to simulated patterns and postulated that those patterns are similar, the model is likely to contain the mechanisms generating these patterns.

#### 465 **5.2** *In which conditions were the patterns mimicked?*

Similarly to Eveel, a main limitation of our approach was that it was based on a simulation model with a pattern-oriented approach. Consequently, our results demonstrated that our assumptions were plausible, but did not demonstrate that they were correct. Such a demonstration would require demonstrating underlying mechanisms, for example by conducting complementary controlled experiments.

We built a full experimental design to explore the model. This type of approach is classical in complex model exploration (de Castro et al. 2001; Faivre et al. 2013). For example, in the context of sensitivity analysis of complex simulation models (Drouineau et al. 2006). Our exploration goals were to generate simulations from the parameter space and analyze the qualitative differences in the model output to (i) study the impact of parameters on the model output, (ii) determine which parameters were the most important, and (iii) identify the combinations of parameters required to

477 mimic all observed spatial patterns. In this study, 17 parameters grouped in 12 set of parameters, 478 were chosen to define the region of the parameter space where all spatial patterns were reproduced. 479 To assess the influence of stochasticity, we made 10 replicates per combination. This can 480 appear limited, however, it was impossible to increase the number of simulations and we preferred 481 to have a better exploration of uncertainty due to uncertain parameters rather than on stochasticity 482 which is rather limited in our model. Stochasticity occurs during the initialization process when 483 randomly building slow or fast value with a given probability. This corresponds to a binomial 484 distribution which has, given the large number of individuals, a very small variance. Stochasticity also 485 occurs in the order of individuals for step 1, but this is closely linked to the previous process and 486 consequently also has a limited variability. This limited effect of stochasticity was confirmed by our 487 results since patterns per combination were always consistent among replicates (Fig. 4 and 5).

488 One hundred thirty simulations among the 2560 mimicked the five spatial patterns. The 489 fourth pattern stated that fast growers and slow growers had different spatial distributions. Fulfilling 490 this pattern demonstrated that genetically different individuals have different habitat selection 491 strategies to maximize their respective fitnesses. Consequently, fulfilling the five patterns suggested 492 that, at least in certain conditions, genotype-dependent habitat selection and phenotypic plasticity 493 could explain observed phenotypic patterns. The level of sensitivity was variable among groups of 494 parameters, but four main groups of parameters were crucial: males' fertility and length at silvering, 495 growth and mortality rates of slow growers, fecundity, and  $L_{\infty}$ . Density-dependence was also an 496 important parameter regarding length at silvering. In summary, the patterns were mimicked in 497 simulations with dominants and dominated but when dominated individuals, mainly females, were 498 not too penalized with respect to dominants, mainly males.

Regarding the spatial patterns, higher density, higher proportions of fast growers, and faster growth rates in downstream regions were mimicked for all combinations of parameters. This suggested that in the range of variation considered, none of the parameters had effects on model outputs. This probably means that the gradient in environmental conditions and the population

dynamics in the model were sufficient to reproduce these patterns, regardless of the competitive advantage of fast growers with respect to slow growers, confirming that phenotypic plasticity plays an important role in environmentally induced changes in life-history traits and demographic attributes. Concerning the two other patterns (sex ratio and length at silvering), additional hypotheses are needed regarding competition and genetic polymorphism. They were fulfilled in conditions of weak competition and when growth differences were not too strong between the two genotypes.

510

### 5.3 Consequences of intra-specific competition

511 In our model, we assumed the existence of asymmetrical density dependence between fast 512 and slow growers. We assumed that smaller individuals would avoid engaging in competition with 513 larger ones (regardless of sex) and would consequently be more affected by density dependence. 514 This assumption seems ecologically realistic. Asymmetrical density dependence has been observed in 515 plants (Weiner 1990), insects (Varley et al. 1973), and fish (DingsØr et al. 2007). Intraspecific 516 competition is a very common mechanism of density dependence, favoring large body size in fishes 517 (Francis 1983; Juanes et al. 2002). In anguillid eels, this may be manifested through agonistic 518 interactions (Knights 1987; Bardonnet et al. 2005), including cannibalism (Edeline and Elie 2004). 519 Such behaviors have been observed in yellow eels under artificial rearing conditions (Peters et al. 520 1980; Degani and Levanon 1983; Knights 1987).

We modelled this asymmetric competition by specifying different levels of densitydependent mortality for slow and fast growers. Interestingly, the spatial patterns were still reproduced when setting these parameters to a similar value (not presented here). Indeed, even with similar intensity of density dependence, slow growers needed more time to reach their length at silvering and consequently, suffered competition longer. Thus, even if competition has the same impact on instantaneous mortality rates of slow and fast-growers, density dependence produces asymmetric impacts on their respective fitness. In EvEel, Drouineau et al. (2014) assumed the

existence of asymmetric competition between males and females, with females being more affected by competition. Interestingly, we observed in our results that females had a higher proportion of slow growers than males. This means that the gender-based asymmetry proposed by Drouineau et al. (2014) may be an indirect result of an asymmetry between two genetically distinct types of individuals with respect to growth.

533 The asymmetric competition implies that fitness of individuals having a given growth 534 genotype depends on the number of individuals having the other growth genotype, which may lead 535 to frequency-dependent selection (Heino et al. 1998). This has several implications. In the model, we 536 assumed that individual fitness corresponded to the lifetime reproductive success called R0, and that 537 this fitness is maximized. However, in a frequency-dependent selection context (i) natural selection 538 does not necessarily lead in fitness maximization (Mylius and Diekmann 1995; Metz et al. 2008), and 539 (ii) fitness may need to be defined as an invasion criterion (Metz et al. 1992). Even when fitness 540 maximization applies, r, the population growth rate, may be a more appropriate measure of fitness 541 than R0 depending on how density-dependence acts (Mylius and Diekmann 1995). To ensure that 542 our assumptions about fitness definition and maximization were valid would require a multi-543 generational model at the scale of the population distribution area. This would allow computing 544 fitness for the whole life-cycle across all potential habitat types of the distribution area while 545 accounting for population structure in terms of genotypes or clusters. At this point, it would be 546 interesting to explore the heritability of the different traits and the intra-generational spatially 547 varying selection, a mechanism suggested by the SNP differences according to latitude (Pujolar et al. 548 2011; Gagnaire et al. 2012; Ulrik et al. 2014).

This was not possible because of difficulties to develop a whole life-cycle model. More specifically, the fractal dimension of the eel population makes it very difficult to develop a population dynamics model for the continental phase at the distribution area scale. Moreover, such a model would require the use of stock-recruitment relationships, which is very difficult for the European eels because of insufficient data, long larval drift, and different recruitment trends through the

distribution area. In this context, we had to use intra-generational model and a R0 fitness function,
restricted to a single catchment and a portion of the whole life-cycle, and to postulate that this R0
was maximized.

557

### 5.4 Reinterpreting the time-minimizing and size-maximizing strategies

558 To summarize the results for combinations of parameters that mimicked observed patterns, 559 we observed a high proportion of individuals, mainly fast growers, in the downstream environment, 560 which corresponded to marine or brackish water. These individuals were mainly males with a 561 constant length at silvering. In upstream areas, we found mainly slow growers, primarily females with 562 higher length at silvering. This can aid in the reinterpretation of gender difference in life tactics (i.e., 563 males with a time-maximizing strategy and females with a size-maximizing strategy). Our results 564 suggest that these tactics were possibly based on the existence of two genotypes for growth. Fast 565 growers grow fast but suffer higher mortality (because they inhabit downstream habitats with higher 566 mortality and density); a time-minimizing strategy is suitable for them. Slow growers grow slowly but 567 suffer lower mortality, consequently they can stay longer in continental habitats, and a size-568 maximizing strategy is suitable for them.

569 Another interesting question is whether cues are used by individuals to select their growth 570 habitat. In the model, individuals were omnipotent and omniscient: they were able to assess the 571 potential fitness in each cell and move in the most suitable cell. This would mean that they were able 572 to assess the natural mortality, growth rate, and density in each cell. Drouineau et al. (2014) 573 suggested that temperature might be one of the main proximal cue used by individuals to assess the 574 suitability. Regarding density-dependence, reaction to aggressiveness (Geffroy and Bardonnet, 2012) 575 or cons-specific odors (Schmucker et al. 2016) were observed on growth and propensity to migrate. 576 Vélez-Espino and Koops (2010) also revealed temperature as main factor explaining variation in life-577 history traits. Our model suggested that density in various habitats was also probably a main cue, 578 especially for slow growers, which tended to minimize competition.

#### 579 5.5 Perspectives

### 580 **5.5.1 Exploring conditions in which phenotypic plasticity is adaptive**

581 It has been demonstrated that phenotypic plasticity allows short-term adaptation to 582 environmental heterogeneity for many species (Schlichting 1986; Sultan 1987; Scheiner 1993; 583 Pigliucci 2005). However, the fitness gain arising from phenotypic plasticity should overcome its cost 584 to be selected. This last point has not been demonstrated for eels. One possibility would be to 585 simulate the evolution of a plastic reaction norm, for example length at silvering, close to the model 586 developed by Marty et al. (2011). Following Ernande et al. (2004) and based on adaptive dynamics 587 models (Mylius and Diekmann 1995), it would be interesting to explore in which environmental and 588 density-dependence conditions, phenotypic plasticity may be selected as an adaptive mechanism 589 despite its costs, and if plasticity is still adaptive in a context of low densities after a population 590 collapse.

#### 591 **5.5.2** Assessing the impact of anthropogenic pressures at the distribution

#### 592 area scale

Another perspective is to assess the impact of anthropogenic pressures on eel populations. Drouineau et al. (2014) mentioned that, because of phenotypic plasticity, anthropogenic pressures are not only a source of mortality, but may also affect sex ratio or mean length at silvering. The existence of two genotypes for growth suggests that anthropogenic activities may act as selective forces. Recently, Podgorniak et al. (2015) demonstrated that human-induced obstacles to migration could act as an evolutionary pressure. Concerning this, Boulenger et al. (2016) highlighted that human pressures impact survival, leading to different life-history strategies.

To conclude, our model provided new insights on eel adaptive mechanisms to heterogeneous environments. Phenotypic plasticity and genotype-dependent habitat selection are two types of mechanisms that can explain the patterns in life-history traits observed in natural environments at the river catchment scale. A better understanding of these mechanisms is crucial to interpret the observations made in the environment, the effects of anthropogenic pressures on the population, and to understand if eels are still adapted in the context of depleted population size and climate change.

608

609 6 Acknowledgements

This study was supported by the Hynes project between Irstea and EDF R&D. We would like to thank Christian Rigaud and Laurent Beaulaton and two anonymous referees for their contribution to the discussion.

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## 614 7 References

- Acou, A., Lefebvre, F., Contournet, P., Poizat, G., Panfili, J., and Crivelli, A.J. 2003. Silvering of female
  eels (*Anguilla anguilla*) in two sub-populations of the Rhone Delta. Bull. Fr. Pêche Piscic. 368:
  55–68.
- Als, T.D., Hansen, M.M., Maes, G.E., Castonguay, M., Riemann, L., Aarestrup, K., Munk, P., Sparholt,
  H., Hanel, R., and Bernatchez, L. 2011. All roads lead to home: panmixia of European eel in
  the Sargasso Sea. Mol. Ecol. 20(7): 1333–1346. doi:10.1111/j.1365-294X.2011.05011.x.
- Andrello, M., Bevacqua, D., Maes, G.E., and De Leo, G.A. 2011. An integrated genetic-demographic
  model to unravel the origin of genetic structure in European eel (*Anguilla anguilla* L.):
  Genetic-demographic model for the European eel. Evol. Appl. 4(4): 517–533.
  doi:10.1111/j.1752-4571.2010.00167.x.
- Bardonnet, A., Rigaud, C., and Labonne, J. 2005. Etude expérimentale des comportements de civelles
  d'Anguilla anguilla L. influence de la densité et de la disponibilité en abris. Bull. Fr. Pêche
  Piscic. 378–379: 47–65.

- Bevacqua, D., Melià, P., De Leo, G.A., and Gatto, M. 2011. Intra-specific scaling of natural mortality in
  fish: the paradigmatic case of the European eel. Oecologia 165(2): 333–339.
  doi:10.1007/s00442-010-1727-9.
- Boivin, B., Castonguay, M., Audet, C., Pavey, S.A., Dionne, M., and Bernatchez, L. 2015. How does
  salinity influence habitat selection and growth in juvenile American eels *Anguilla rostrata*?:
  salinity preference in *Anguilla rostrata* glass eels. J. Fish Biol. **86**(2): 765–784.
  doi:10.1111/jfb.12604.
- Boulenger, C., Acou, A., Gimenez, O., Charrier, F., Tremblay, J., and Feunteun, E. 2016. Factors
  determining survival of European eels in two unexploited sub-populations. Freshw. Biol.
  637 61(6): 947–962. doi:10.1111/fwb.12759.
- Brodersen, J., and Seehausen, O. 2014. Why evolutionary biologists should get seriously involved in
  ecological monitoring and applied biodiversity assessment programs. Evol. Appl. 7(9): 968–
  983. doi:10.1111/eva.12215.
- 641 Cairns, D.K., Secor, D.A., Morrison, W.E., and Hallett, J.A. 2009. Salinity-linked growth in anguillid eels
  642 and the paradox of temperate-zone catadromy. J. Fish Biol. 74(9): 2094–2114.
  643 doi:10.1111/j.1095-8649.2009.02290.x.
- de Castro, L.A.B., Petrere Jr., M., and Comune, A.E. 2001. Sensitivity of the BEAM4 fisheries
  bioeconomic model to the main biological input parameters. Ecol. Model. 141(1–3): 53–66.
  doi:10.1016/S0304-3800(01)00241-1.
- 647 Clark, C.W. 1993. Dynamic Models of Behavior: An extension of Life-history Theory. Tree 8(6): 205–
  648 209.
- 649 Côté, C.L., Castonguay, M., Kalujnaia, M., Cramb, G., and Bernatchez, L. 2014. In absence of local
  650 adaptation, plasticity and spatially varying selection rule: a view from genomic reaction
  651 norms in a panmictic species (*Anguilla rostrata*). BMC Genomics 15(1): 403.
  652 doi:10.1186/1471-2164-15-403.
- Côté, C.L., Castonguay, M., Verreault, G., and Bernatchez, L. 2009. Differential effects of origin and
  salinity rearing conditions on growth of glass eels of the American eel *Anguilla rostrata* :
  implications for stocking programmes. J. Fish Biol. **74**(9): 1934–1948. doi:10.1111/j.10958649.2009.02291.x.
- Côté, C.L., Gagnaire, P.-A., Bourret, V., Verreault, G., Castonguay, M., and Bernatchez, L. 2013.
  Population genetics of the American eel (*Anguilla rostrata*): F<sub>ST</sub> = 0 and North Atlantic
  Oscillation effects on demographic fluctuations of a panmictic species. Mol. Ecol. 22(7):
  1763–1776. doi:10.1111/mec.12142.

Author-produced version of the article published in

Canadian Journal of Fisheries and Aquatic Sciences, 2017, 74 (7), 987-999.

The original publication is available at http://www.nrcresearchpress.com/doi/10.1139/cjfas-2016-0214#.WVIIMevyhyw

doi : 10.1139/cjfas-2016-0214

- Côté, C.L., Pavey, S.A., Stacey, J.A., Pratt, T.C., Castonguay, M., Audet, C., and Bernatchez, L. 2015.
  Growth, Female Size, and Sex Ratio Variability in American Eel of Different Origins in Both
  Controlled Conditions and the Wild: Implications for Stocking Programs. Trans. Am. Fish. Soc.
  144(2): 246–257. doi:10.1080/00028487.2014.975841.
- 665 Council of the European Union. 2007. Council Regulation (EC) No 1100/2007 of 18 September 2007 666 establishing measures for the recovery of the stock of European eel. Brussels: 7pp.
- Daverat, F., Beaulaton, L., Poole, R., Lambert, P., Wickström, H., Andersson, J., Aprahamian, M.,
  Hizem, B., Elie, P., Yalçın-Özdilek, S., and Gumus, A. 2012. One century of eel growth: changes
  and implications. Ecol. Freshw. Fish 21(3): 325–336. doi:10.1111/j.1600-0633.2011.00541.x.
- Daverat, F., Limburg, K.E., Thibault, I., Shiao, J.-C., Dodson, J.J., Caron, F., Tzeng, W.-N., Iizuka, Y., and
  Wickstrom, H. 2006. Phenotypic plasticity of habitat use by three temperate eel species,
  Anguilla anguilla, A. japonica and A. rostrata. Mar. Ecol. Prog. Ser. 308: 231–241.
- Daverat, F., and Tomás, J. 2006. Tactics and demographic attributes in the European eel Anguilla
   *anguilla* in the Gironde watershed, SW France. Mar. Ecol.-Prog. Ser. **307**: p–247.
- Davey, A.J.H., and Jellyman, D.J. 2005. Sex Determination in Freshwater Eels and Management
  Options for Manipulation of Sex. Rev. Fish Biol. Fish. 15(1–2): 37–52. doi:10.1007/s11160005-7431-x.
- De Leo, G.A., and Gatto, M. 1995. A size and age-structured model of the European eel (*Anguilla* 679 *anguilla* L.). Can. J. Fish. Aquat. Sci. 52: 1351–1367.
- Degani, G., and Levanon, D. 1983. The influence of low density on food adaptation, cannibalism and
  growth of eels (*Anguilla anguilla* (L.)). Bamidgeh **35**: 53–60.
- Dekker, W. 1998. Long-term trends in the glass eels immigrating at Den Oever, The Netherlands. Bull.
  Fr. Pêche Piscic. (349): 199–214. doi:10.1051/kmae:1998045.
- 684 Dekker, W. 2000. The fractal geometry of the European eel stock. ICES J. Mar. Sci. 57(1): 109–121.
  685 doi:10.1006/jmsc.1999.0562.
- Dekker, W. 2003. Status of the European Eel Stock and Fisheries. *In* Eel Biology. *Edited by* K. Aida, K.
  Tsukamoto, and K. Yamauchi. Springer Japan. pp. 237–254.
- 688 Dekker, W. 2009. Worldwide decline of eel resources necessitates immediate action.
- Desaunay, Y., and Guerault, D. 1997. Seasonal and long-term changes in biometrics of eel larvae: a
   possible relationship between recruitment variation and North Atlantic ecosystem
   productivity. J. Fish Biol. 51(sA): 317–339.
- Desaunay, Y., Lecomte-Finiger, R., and Guérault, D. 2012. Mean age and migration patterns of
   *Anguilla anguilla* (L.) glass eels from three French estuaries (Somme, Vilaine and Adour
   Rivers). Arch. Pol. Fish. 20(3). doi:10.2478/v10086-012-0023-1.

- DingsØr, G.E., Ciannelli, L., Chan, K.-S., Ottersen, G., and Stenseth, N.C. 2007. Density dependence
  and density independence during the early life stages of four marine fish stocks. Ecology
  88(3): 625–634.
- Drouineau, H., Mahévas, S., Pelletier, D., and Beliaeff, B. 2006. Assessing the impact of different
   management options using ISIS-Fish: the French *Merluccius merluccius Nephrops norvegicus* mixed fishery of the Bay of Biscay. Aquat. Living Resour. 19(1): 15–29.
   doi:10.1051/alr:2006002.
- Drouineau, H., Rigaud, C., Daverat, F. and Lambert, P. 2014. EvEel (evolutionary ecology-based model
   for eel): a model to explore the role of phenotypic plasticity as an adaptive response of three
   temperate eels to spatially structured environments. Can. J. Fish. Aquat. Sci. 71(10): 1561–
   1571. doi:10.1139/cjfas-2014-0090.
- Edeline, E. 2007. Adaptive phenotypic plasticity of eel diadromy. Mar. Ecol. Prog. Ser. **341**: 229–232.
- 707 Edeline, E., and Elie, P. 2004. Is salinity choice related to growth in juvenile eel *Anguilla anguilla*?
  708 Cybium 28: 77–82.
- Frnande, B., Dieckmann, U., and Heino, M. 2004. Adaptive changes in harvested populations:
  plasticity and evolution of age and size at maturation. Proc. R. Soc. Lond. B Biol. Sci.
  271(1537): 415–423.
- Faivre, R., Iooss, B., Mahévas, S., Makowski, D., and Monod, H. 2013. Analyse de sensibilité et
  exploration de modèles. Editions Quae.
- Francis, R.C. 1983. Experiential effects on agonistic behavior in the paradise fish, *Macropodus opercularis*. Behaviour **85**(3): 292–313.
- Gagnaire, P.-A., Normandeau, E., Cote, C., Moller Hansen, M., and Bernatchez, L. 2012. The Genetic
   Consequences of Spatially Varying Selection in the Panmictic American Eel (*Anguilla rostrata*). Genetics 190(2): 725–736. doi:10.1534/genetics.111.134825.
- Geffroy, B., and Bardonnet, A. 2012. Differential effects of behaviour, propensity to migrate and
  recruitment season on glass eels and elvers' growing performance: Young eel behaviour and
  growth pattern. Ecol. Freshw. Fish 21(3): 469–482. doi:10.1111/j.1600-0633.2012.00566.x.
- Giske, J., Huse, G., and Fiksen, O. 1998. Modelling spatial dynamics of fish. Rev. Fish Biol. Fish. 8: 57–
  91. doi:10.1023/A:1008864517488.
- Gotthard, K., and Nylin, S. 1995. Adaptive plasticity and plasticity as an adaptation: a selective review
  of plasticity in animal morphology and life-history. Oikos 74(1): 3–17. doi:10.2307/3545669.
- 726 Grimm, V., Berger, U., Bastiansen, F., Eliassen, S., Ginot, V., Giske, J., Goss-Custard, J., Grand, T., Heinz,
- 727 S.K., Huse, G., Huth, A., Jepsen, J.U., Jørgensen, C., Mooij, W.M., Müller, B., Pe'er, G., Piou, C.,
- 728 Railsback, S.F., Robbins, A.M., Robbins, M.M., Rossmanith, E., Rüger, N., Strand, E., Souissi, S.,

Author-produced version of the article published in

Canadian Journal of Fisheries and Aquatic Sciences, 2017, 74 (7), 987-999.

The original publication is available at http://www.nrcresearchpress.com/doi/10.1139/cjfas-2016-0214#.WVIIMevyhyw

doi : 10.1139/cjfas-2016-0214

- Stillman, R.A., Vabø, R., Visser, U., and DeAngelis, D.L. 2006. A standard protocol for
  describing individual-based and agent-based models. Ecol. Model. 198(1–2): 115–126.
  doi:10.1016/j.ecolmodel.2006.04.023.
- Grimm, V., Berger, U., DeAngelis, D.L., Polhill, J.G., Giske, J., and Railsback, S.F. 2010. The ODD
  protocol: A review and first update. Ecol. Model. 221(23): 2760–2768.
  doi:10.1016/j.ecolmodel.2010.08.019.
- Grimm, V., Frank, K., Jeltsch, F., Brandl, R., Uchmanski, J., and Wissel, C. 1996. Pattern-oriented
  modelling in population ecology. Sci. Total Environ. 183: 155–166.
- Grimm, V., and Railsback, S.F. 2012. Pattern-oriented modelling: a "multi-scope" for predictive
  systems ecology. Philos. Trans. R. Soc. B Biol. Sci. 367(1586): 298–310.
  doi:10.1098/rstb.2011.0180.
- Heino, M., Metz, J.A.J., and Kaitala, V. (1998). The enigma of frequency-dependent selection. Trends
  Ecol. Evol. (13): 367–370.
- Helfman, G.S., Facey, D.E., Stanton Hales, L., and Bozeman, E.L. 1987. Reproductive Ecology of the
  American eel. Am. Fish Soc. Symp. 1: 42–56.
- Hutchings, J.A., Swain, D.P., Rowe, S., Eddington, J.D., Puvanendran, V., and Brown, J.A. 2007. Genetic
  variation in life-history reaction norms in a marine fish. Proc. R. Soc. B Biol. Sci. 274(1619):
  1693–1699. doi:10.1098/rspb.2007.0263.
- Jacoby, D., and Gollock, M. 2014. *Anguilla anguilla*. The IUCN Red List of Threatened Species.
  Available from http://www.iucnredlist.org/details/60344/0.
- Juanes, F., Buckel, J.A., and Scharf, F.S. 2002. Feeding ecology of piscivorous fishes. *In* Handbook of
  fish biology and fisheries, Blackwell. Paul J.B. Hart and John D. Reynolds, UK. pp. 267–283.
- 751 Kawecki, T.J., and Ebert, D. 2004. Conceptual issues in local adaptation. Ecol. Lett. 7(12): 1225–1241.
  752 doi:10.1111/j.1461-0248.2004.00684.x.
- 753 Kleijnen, J.P.C. 1987. Statistical Tools for Simulation Practitioners. Marcel Dekker, Inc., New York, NY,
  754 USA.
- Knights, B. 1987. Agonistic behaviour and growth in the European eel, *Anguilla anguilla* L., in relation
  to warm-water aquaculture. J. Fish Biol. **31**(2): 265–276.
- Levins, R. 1963. Theory of fitness in a heterogeneous environment. II. Developmental flexibility and
  niche selection. Am. Nat. 97(893): 75–90. doi:10.1086/ 282258.
- Marty, L., Dieckmann, U., Rochet, M.-J., and Ernande, B. 2011. Impact of Environmental Covariation
  in Growth and Mortality on Evolving Maturation Reaction Norms. Am. Nat. 177(4): E98–
  E118. doi:10.1086/658988.

- McNamara, J.M., and Houston, A.I. 1992. State-dependent life-history theory and its implications for
  optimal clutch size. Evol. Ecol. 6(2): 170–185. doi:10.1007/BF02270710.
- Melià, P., Bevacqua, D., Crivelli, A.J., De Leo, G.A., Panfili, J., and Gatto, M. 2006. Age and growth of *Anguilla anguilla* in the Camargue lagoons. J. Fish Biol. 68: 876–890. doi:10.1111/j.10958649.2006.00975.x.
- 767 Metz, J.A.J., Mylius, S.D., and Diekmann, O. 2008. When does evolution optimize? Evol. Ecol. Res.
  768 **10**(5): 629–654.
- Metz, J.A.J., Nisbet, R.M., and Geritz, S.A.H. 1992. How should we define "fitness" for general
  ecological scenarios? Trends Ecol. Evol. 7(6): 198–202.
- 771 Moriarty, C. 2003. The eel. *In* Eel Biology. Springer Japan, Tokyo. pp. 89–105.
- Mylius, S.D., and Diekmann, O. 1995. On Evolutionarily Stable Life Histories, Optimization and the
   Need to Be Specific about Density Dependence. Oikos 74(2): 218–224.
- Oliveira, K. 1999. Life-history characteristics and strategies of the American eel, *Anguilla rostrata*.
  Can. J. Fish. Aquat. Sci. 56(5): 795–802.
- Oliveira, K. 2001. Regional variation and the effect of lake: river area on sex distribution of American
  eels. J. Fish Biol. 58(4): 943–952. doi:10.1006/jfbi.2000.1503.
- Palm, S., Dannewitz, J., Prestegaard, T., and Wickström, H. 2009. Panmixia in European eel revisited:
  no genetic difference between maturing adults from southern and northern Europe.
  Heredity 103(1): 82–89.
- Parker, G.A., and Maynard Smith, J. 1990. Optimality theory in evolutionary biology. Nature 348: 27–
  33.
- Pavey, S.A., Gaudin, J., Normandeau, E., Dionne, M., Castonguay, M., Audet, C., and Bernatchez, L.
  2015. RAD Sequencing Highlights Polygenic Discrimination of Habitat Ecotypes in the
  Panmictic American Eel. Curr. Biol. 25(12): 1666–1671. doi:10.1016/j.cub.2015.04.062.
- Peters, G., Delventhal, H., and Klinger, H. 1980. Physiological and Morphological Effects of Social
  Stress on the Eel, *Anguilla anguilla* L. *In* Fish Diseases. *Edited by* D.W. Ahne. Springer Berlin
  Heidelberg. pp. 225–227.
- Pigliucci, M. 2005. Evolution of phenotypic plasticity: Where are we going now? Trends Ecol. Evol.
  20(9): 481–486. doi:10.1016/j.tree.2005.06.001.
- Podgorniak, T., Angelini, A., Blanchet, S., de Oliveira, E., Pierron, F., and Daverat, F. 2015. Climbing
  experience in glass eels: A cognitive task or a matter of physical capacities? Physiol. Behav.
  151: 448–455. doi:10.1016/j.physbeh.2015.08.001.
- Pujolar, J.M., Bevacqua, D., Andrello, M., Capoccioni, F., Ciccotti, E., De Leo, G.A., and Zane, L. 2011.
   Genetic patchiness in European eel adults evidenced by molecular genetics and population

- 796dynamicsmodelling.Mol.Phylogenet.Evol.58(2):198–206.797doi:10.1016/j.ympev.2010.11.019.
- Pujolar, J.M., Jacobsen, M.W., Als, T.D., Frydenberg, J., Munch, K., Jónsson, B., Jian, J.B., Cheng, L.,
  Maes, G.E., Bernatchez, L., and Hansen, M.M. 2014. Genome-wide single-generation
  signatures of local selection in the panmictic European eel. Mol. Ecol. 23(10): 2514–2528.
  doi:10.1111/mec.12753.
- Railsback, S.F., and Harvey, B.C. 2013. Trait-mediated trophic interactions: is foraging theory keeping
  up? Trends Ecol. Evol. 28(2): 119–125. doi:10.1016/j.tree.2012.08.023.
- Rougier, T., Lassalle, G., Drouineau, H., Dumoulin, N., Faure, T., Deffuant, G., Rochard, E., and
  Lambert, P. 2015. The Combined Use of Correlative and Mechanistic Species Distribution
  Models Benefits Low Conservation Status Species. PLOS ONE 10(10): e0139194.
  doi:10.1371/journal.pone.0139194.
- Scheiner, S.M. 1993. Genetics and Evolution of Phenotypic Plasticity. Annu. Rev. Ecol. Syst. 24: 35–68.
- Schlichting, C.D. 1986. The Evolution of Phenotypic Plasticity in Plants. Annu. Rev. Ecol. Syst. 17(1):
  667–693. doi:10.1146/annurev.es.17.110186.003315.
- Sultan, S.E. 1987. Evolutionary Implications of Phenotypic Plasticity in Plants. *In* Evolutionary Biology.
   *Edited by* M.K. Hecht, B. Wallace, and G.T. Prance. Springer US. pp. 127–178.
- 813 Tesch, F.-W. 2003. The eel. *In* 3rd ed. Blackwell Science, Oxford, UK.
- Travis, J. 1994. Evaluating the adaptive role of morphological plasticity. *In* Ecological Morphology.
   *Edited by* P. Wainwright and S. Reilly. University of Chicago Press. pp. 99–122.
- 816 Ulrik, M.G., Pujolar, J.M., Ferchaud, A.-L., Jacobsen, M.W., Als, T.D., Gagnaire, P.A., Frydenberg, J.,
  817 Bøcher, P.K., Jónsson, B., Bernatchez, L., and others. 2014. Do North Atlantic eels show
  818 parallel patterns of spatially varying selection? BMC Evol. Biol. 14(1): 138.
- Varley, C.G., Gradwell, G.R., and Hassell, M.P. 1973. Insect population ecology an analytical approach.
  University of California Press, California.
- Vélez-Espino, L.A., and Koops, M.A. 2010. A synthesis of the ecological processes influencing variation
   in life-history and movement patterns of American eel: towards a global assessment. Rev.
   Fish Biol. Fish. 20(2): 163–186. doi:10.1007/s11160-009-9127-0.
- Vollestad, L.A. 1992. Geographic Variation in Age and Length at Metamorphosis of Maturing
   European Eel: Ennvironmental Effects and Phenotypic Plasticity. J. Anim. Ecol. 61(1): 41–48.
- Weiner, J. 1990. Asymmetric Competition in Plant Population. Tree **5**(11): 360–364.
- Williams, G.C. 1996. Adaptation and natural selection: a critique of some current evolutionary
  thought. Princeton Univ. Press, Princeton, NJ.
- 829

### 830 Tables

831 Table 1. GenEveel parameter descriptions with reference values and modalities (- and +) for the 17

832 parameters involved in the experimental design.

Darameter	Description	Reference	(-)	(+)	Poforonco		
Parameter	Description	value	modality	modality	Reference		
n	Cells of river catchment	30	24	36	(Drouineau et al. 2014)		
Ν.	Number of glass eels that	20.000	24.000	26.000	(Drouinogu at al. 2014)		
N	colonize freshwater	30 000	24 000	36 000	(Drouineau et al. 2014)		
a 1	Regression coefficient from	0 016			(Androllo at al. 2011)		
01	fecundity at length	8 840	-	-			
-	Regression coefficient from	1 2077110		-	(Melià et al. 2006, Andrello		
ŭ	fecundity at length	1.38//119	-		et al. 2011)		
h	Regression exponent from	2 22	2 576	2.064	(Malià at al. 2006)		
D	fecundity at length	3.22	2.570	3.804	(IVIEIIA EL AL 2006)		
L∞(cm)	Asymptotic length	76.2	60.96	91.44	(De Leo and Gatto 1995)		
<i>L<sub>sm</sub></i> (cm)	Male length at silvering	40.5	38.15	42.85	(Vollestad 1992)		
			6	9	(Desaunay and Guerault		
<i>L<sub>g</sub></i> (cm)	Length at recruitment	7.5			1997, Dekker 1998,		
					Desaunay et al. 2012)		
fertility	Constant of male fertility	43	40.5	45.5	-		
K <sub>fast</sub> (i,1), year⁻¹	Fast intrinsic growth rate	0.315	0.295	0.335	(De Leo and Gatto 1995)		
K <sub>slow</sub> (i,1), year <sup>-1</sup>	Slow intrinsic growth rate	0.253	0.233	0.273	(De Leo and Gatto 1995)		
ana a K	Proportion of individuals	0.5	0.4	0.6			
ριορκ	that grow slowly	0.5	0.4	0.6	-		
<i>M<sub>fast</sub>(i,1)</i> , year <sup>-1</sup>	Fast intrinsic mortality rate	0.38	0.405	0.355	-		
<i>M<sub>slow</sub>(i,1)</i> , year <sup>-1</sup>	Slow intrinsic mortality rate	0.138	0.15	0.127	(Dekker 2000)		
c,	Intensity of density-	0.0001	0.00000	0.00012	(Drouincou et al. 2014)		
ά	dependence	0.0001	0.00008		(Drouneau et al. 2014)		
	Ratio between upstream						
$r_{\kappa}$	and downstream growth	0.5	0.4	0.6	(Drouineau et al. 2014)		
	rate						
r	Ratio between upstream	1	0 0	1.02	(Drouinogu at al 2014)		
I M	and downstream mortality	Ţ	0.8		(Diouilleau et al. 2014)		

	rate				
$Y_{K}$	Shape parameter of growth	0.05	0.049	0.051	(Drouineau et al. 2014)
Y <sub>M</sub>	Shape parameter of mortality	0.05	0.049	0.051	(Drouineau et al. 2014)

#### 834 **Table 2.** Results of Mann-Kendall test of reference simulation.

Spatial pattern	Tau
Abundance	-1
Mean length at silvering	0.98
Sex ratio (proportion of females)	0.57
Ratio of fast growers	-0.78
Mean realized growth rate	-1

- Table 3. Number of simulations with consistent patterns for each modality of the groups of parameters, over the number of simulations for which it was possible to calculate a pattern. The columns represent the spatial patterns and the numbers of simulations for which it was possible to
- estimate a pattern.

		Moon longth	Sex ratio	Patio of fact	Mean	The five
Parameters	Abundance	at silvering	(proportion		realized	spatial
group	2560	1020	of females)	arco	growth rate	patterns
		1920	1610	2500	2560	1610
N	(-) 1280/1280	(-) 650/960	(-) 70/800	(-) 1280/1280	(-) 1280/1280	(-) 70/800
/•	(+) 1280/1280	(+) 650/960	(+) 60/810	(+) 1280/1280	(+) 1280/1280	(+)60/810
<i>fertility</i> and	(-) 1280/1280	(-) 800/960	(-) 130/800	(-) 1280/1280	(-) 1280/1280	(-)130/800
L <sub>sm</sub>	(+) 1280/1280	(+) 500/960	(+) 0/810	(+) 1280/1280	(+) 1280/1280	(+) 0/810
K <sub>fast</sub> (i, 1) and	(-) 1280/1280	(-) 650/960	(-) 70/810	(-) 1280/1280	(-) 1280/1280	(-) 70/810
M <sub>fast</sub> (i, 1)	(+) 1280/1280	(+) 650/960	(+) 60/800	(+) 1280/1280	(+) 1280/1280	(+) 60/800
<i>K<sub>slow</sub>(i, 1)</i> and	(-) 1280/1280	(-) 710/960	(-) 0/810	(-) 1280/1280	(-) 1280/1280	(-) 0/810
M <sub>slow</sub> (i, 1)	(+) 1280/1280	(+) 590/960	(+) 130/800	(+) 1280/1280	(+) 1280/1280	(+) 130/800
nronK	(-) 1280/1280	(-) 650/960	(-) 90/810	(-) 1280/1280	(-) 1280/1280	(-) 90/810
ριορκ	(+) 1280/1280	(+) 650/960	(+) 40/800	(+) 1280/1280	(+) 1280/1280	(+) 40/800
0	(-) 1280/1280	(-) 810/960	(-) 60/960	(-) 1280/1280	(-) 1280/1280	(-) 60/960
u	(+) 1280/1280	(+) 490/960	(+) 70/650	(+) 1280/1280	(+) 1280/1280	(+) 70/650
n	(-) 1280/1280	(-) 660/960	(-) 60/810	(-) 1280/1280	(-) 1280/1280	(-) 60/810
п	(+) 1280/1280	(+) 640/960	(+) 70/800	(+) 1280/1280	(+) 1280/1280	(+) 70/800
h	(-) 1280/1280	(-) 360/640	(-) 10/640	(-) 1280/1280	(-) 1280/1280	(-) 10/640
D	(+) 1280/1280	(+) 940/1280	(+) 120/970	(+) 1280/1280	(+) 1280/1280	(+) 120/970
	(-) 1280/1280	(-) 310/640	(-) 120/640	(-) 1280/1280	(-) 1280/1280	(-) 120/640
L∞	(+) 1280/1280	(+) 990/1280	(+) 10/970	(+) 1280/1280	(+) 1280/1280	(+) 10/970
,	(-) 1280/1280	(-) 650/960	(-) 60/800	(-) 1280/1280	(-) 1280/1280	(-) 60/800
Lg	(+) 1280/1280	(+) 650/960	(+) 70/810	(+) 1280/1280	(+) 1280/1280	(+) 70/810
r, and r	(-) 1280/1280	(-) 630/960	(-) 40/810	(-) 1280/1280	(-) 1280/1280	(-) 40/810
<b>r</b> <sub>k</sub> and <b>r</b> <sub>m</sub>	(+) 1280/1280	(+) 670/960	(+) 90/800	(+) 1280/1280	(+) 1280/1280	(+) 90/800
v. and v	(-) 1280/1280	(-) 730/960	(-) 60/810	(-) 1280/1280	(-) 1280/1280	(-) 60/810
Yk and Ym	(+) 1280/1280	(+) 570/960	(+) 70/800	(+) 1280/1280	(+) 1280/1280	(+) 70/800

840 **Table 4.** Results of the 13 combinations that generated five consistent patterns. The signs +/- refer to

841 the modalities of the parameters groups. The two last columns represent the five spatial patterns. An

- ascendant arrows stands for positive Mann-Kendall tau value (increasing trend from downstream to
- upstream). Conversely, a descendant arrow stands for a negative Mann-Kendall tau value.

			rtility, K <sub>fast</sub> (i, 1), L <sub>sm</sub> M <sub>fast</sub> (i, 1)					b	L∞	Lg			Abundance		
		fertility, N L <sub>sm</sub>		K <sub>slow</sub> (i, 1),	nronK	a					r <sub>ĸ,</sub>	<b>ү</b> к,	Ratio of fast	Sex ratio	
	N			M <sub>slow</sub> (i, 1)	ριορι	u	n				r <sub>M</sub>	Υм	growers	LS	
													$\overline{K(\iota,x)}$		
1	-	-	-	+	+	+	+	+	-	-	+	+	И	7	
2	+	-	-	+	-	+	+	+	-	+	+	+	Ц	7	
3	+	-	-	+	+	-	+	+	-	+	+	-	Ы	7	
4	-	-	-	+	-	-	+	+	-	-	+	-	Ы	7	
5	+	-	+	+	+	-	-	+	-	-	+	-	И	7	
6	+	-	-	+	-	-	-	+	-	+	-	-	И	7	
7	-	-	-	+	-	+	+	-	+	+	+	+	Ы	7	
8	-	-	+	+	-	-	-	+	-	+	+	-	Ы	7	
9	-	-	+	+	+	+	-	+	-	+	+	+	И	7	
10	-	-	+	+	-	+	+	+	-	+	-	+	Ц	7	
11	+	-	+	+	-	+	-	+	-	-	+	+	Ы	7	
12	-	-	-	+	-	+	-	+	-	-	-	+	Ы	7	
13	+	-	+	+	-	-	+	+	-	-	-	-	И	7	
844															

845 Figures



846

847 **Fig. 1.** Flow chart representing the fish biological pathway.



849

850 **Fig. 2.** Algorithm of the model GenEveel.











855

**Fig. 4.** Simulated mean length at silvering patterns in the 13 combinations of parameters that consistently mimic the pattern described in the literature. These 13 combinations correspond to the 13 combinations that generate consistent patterns for all the five spatial patterns. Each plot stands for a combination (the number is an identifier of the combination that can be found in table 4) and each line stands for a replicate.





Fig. 5. Simulated sex ratio (proportions of females) patterns in the 13 combinations of parameters
that consistently mimic the pattern described in the literature. These 13 combinations correspond to
the 13 combinations that generate consistent patterns for all the five spatial patterns. Each plot
stands for a combination (the number is an identifier of the combination that can be found in table
and each line stands for a replicate.

#### 868 Supplemental Information

- **Table S1.** Results of the 256 combinations. The signs +/- refer to the modalities of the parameters
- 870 groups . The last five columns represent the spatial patterns. An ascendant arrows stands for positive
- 871 Mann-Kendall tau value (increasing trend from downstream to upstream). Conversely, a descendant
- 872 arrow stands for a negative Mann-Kendall tau value.