



HAL
open science

Up to 92 % increase of cancer-preventing lunasin in organic spring barley

Linda Legzdina, Ilva Nakurte, Inga Kirhnere, Jana Namniece, Liga Krigere, Kristine Saleniece, Indra Beinarovica, Ruta Muceniece

► To cite this version:

Linda Legzdina, Ilva Nakurte, Inga Kirhnere, Jana Namniece, Liga Krigere, et al.. Up to 92 % increase of cancer-preventing lunasin in organic spring barley. *Agronomy for Sustainable Development*, 2014, 34 (4), pp.783-791. 10.1007/s13593-013-0203-4 . hal-01234823

HAL Id: hal-01234823

<https://hal.science/hal-01234823>

Submitted on 27 Nov 2015

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Up to 92 % increase of cancer-preventing lunasin in organic spring barley

Linda Legzdina · Ilva Nakurte · Inga Kirhnere ·
Jana Namniece · Liga Krigere · Kristine Saleniece ·
Indra Beinarovica · Ruta Muceniece

Accepted: 26 November 2013 / Published online: 8 January 2014
© INRA and Springer-Verlag France 2013

Abstract Lunasin is a plant peptide that has health benefits such as cancer-preventing, antioxidant, anti-inflammatory, and cholesterol-lowering effects. However, there is actually no information on the influence of cropping on the lunasin content of cereals. Therefore, we studied lunasin in 22 spring barley genotypes grown both organically and conventionally during two seasons. We found that lunasin content of barley grain averaged 44.8 µg/g, ranging from 5.0 to 189.0 µg/g. Organic farming increased average lunasin content by 47–92 %. Ten out of 22 genotypes produced significantly more lunasin under organic farming in both years. Our findings evidence positive effects of organic farming on lunasin content in barley.

Keywords Lunasin · Organic agriculture · Conventional agriculture · Spring barley · Genotype × environment interaction · Heritability

1 Introduction

Lunasin is a novel peptide originally identified in soybean (Galvez and de Lumen 1999) and meanwhile also found in cereals, barley (Jeong et al. 2002, 2010a), wheat (Jeong et al. 2007a), rye (Jeong et al. 2009), triticale (Nakurte et al. 2012),

and oat (Nakurte et al. 2013), as well as non-cereals, e.g., amaranth (Silva-Sánchez et al. 2008) and *Solanum nigrum* (Jeong et al. 2007b). The presence of the lunasin peptide has been reported in many soybean varieties, with concentrations ranging from 4.4 to 70.5 mg/g of protein or 0.5 to 8.1 mg/g of seed (de Mejia et al. 2004; de Lumen 2005). In cereals, the lunasin content ranged from 13.6 to 21.5 µg/g in barley (Jeong et al. 2002), 211–249 µg/g in wheat (Jeong et al. 2007a), 50–150 µg/g (Jeong et al. 2009) and 732–1510 µg/g in rye, 429–6,458 µg/g in triticale (Nakurte et al. 2012), and 64–197 µg/g in oat (Nakurte et al. 2013).

Recent scientific evidence indicates that lunasin has antioxidant, anti-inflammatory, and cholesterol-lowering effects (Hernández-Ledesma et al. 2009, 2013) and has influence on central nervous system (CNS) (Dzirkale et al. 2013). Studies in animals have shown that lunasin can be administered orally and can enter target tissues (de Lumen 2005; Jeong et al. 2009). Findings obtained on the bioavailability, bioactivity, and thermostability of lunasin support the inclusion of lunasin-containing products in the human diet (Park et al. 2005; Jeong et al. 2010a). Synthetic lunasin and that isolated from soybean were first investigated as a factor that might prevent cancer cells from dividing and multiplying. Anticarcinogenic activity of the lunasin has been demonstrated both in in vitro and in vivo assays (Hernández-Ledesma et al. 2013). Lunasin is also a valuable peptide for the cardiovascular system. It has been reported to lower serum low density lipoprotein (LDL) cholesterol levels by selectively disrupting a necessary step in the production of a key enzyme, 3-hydroxy-3-methyl-glutaryl-CoA reductase, and by upregulating the expression of the LDL-receptor gene (Hernández-Ledesma et al. 2009). Recent data show that internalization of lunasin into macrophages is amplified in inflammatory conditions and is primarily mediated by endocytic mechanisms that involve integrin signaling, clathrin-coated structures, and macropinosomes. Lunasin may be responsible for

L. Legzdina (✉) · I. Kirhnere · I. Beinarovica
State Priekuli Plant Breeding Institute, Zinatnes Str. 1a,
Priekuli 4126, Latvia
e-mail: Linda.Legzdina@priekuliselekcija.lv

I. Nakurte
Faculty of Chemistry of University of Latvia, Kr.Valdemara Str. 48,
Riga 1013, Latvia

J. Namniece · L. Krigere · K. Saleniece · R. Muceniece
Faculty of Medicine, University of Latvia, Sarlotes Str. 1a,
Riga 1001, Latvia

attenuation of cardiovascular diseases risk factors by interacting with pathways involved in endocytosis and inflammation (Cam et al. 2013). We (Nakurte et al. 2013) and other authors (Jeong et al. 2010b; Hernández-Ledesma et al. 2013) have shown that lunasin isolated from plants exerts antioxidant effects similarly to the synthetic lunasin. Effects of the synthetic lunasin on CNS were described as markedly expressed neuroleptic/cataleptic action in male C57Bl/6 mice (Dzirkale et al. 2013). The authors suggest that the action of lunasin at least partially is provided via dopaminergic D1 receptor pathways.

Although several publications indicate that the lunasin content in crops depends on the genotype (Wang et al. 2008; Nakurte et al. 2012, 2013), very limited information is available with regard to the influence of the growing environment on the content of lunasin in crops. The effect of temperature and soil moisture on lunasin in soybean was reported (Wang et al. 2008), but no data on cereals are available to date.

Since the end of the last century, the market for organic products has grown. Organic farming is continuously becoming a widespread form of agriculture, and organic products are increasingly being requested by consumers. Additionally, people have shown a growing interest in food containing phytochemicals, commonly known as functional food. Conventional, organic, and integrated agricultural practices may induce differences in the plant phytochemical content; higher amounts of phytochemicals are usually produced in stressful growing conditions and farmers are challenged to achieve both crop yield and benefit for health (García-Mier et al. 2013). Many studies consistently report a lack of significant differences between organically and conventionally grown food in terms of safety and nutritional value, suggesting that crops and livestock products produced in both farming systems are comparable with regard to their nutrient content (Smith-Spangler et al. 2012). Nevertheless, the results obtained are contradictory: some authors report no significant differences, yet many studies have shown different amounts of nutritionally important and health-promoting compounds in crops grown under organic and conventional management (Woese et al. 1997; Worthington 2001; Bour and Prescott 2002; Magkos et al. 2003).

Interest in research on the content of biologically active substances in organic farming has increased dramatically (Huber et al. 2011). For instance, Benbrook (2005) estimated that the content of biologically active compounds is, on average, 30 % higher in organically grown plants in comparison to those grown conventionally. In most studies, the content of vitamin C was found to be significantly higher in an organic farming system (Worthington 2001). However, the carotenoid content in potato was found not to be influenced by organic and conventional farming systems (Murniece et al. 2012), whereas the level of carotenoids in other vegetables and fruits was higher when they were grown organically (Huber et al.

2011). The majority of studies have reported a higher content of phenolic compounds in organically grown fruits, vegetables (Huber et al. 2011), and potatoes (Lombardo et al. 2012); organically generated products contain higher amounts of flavonoids, which are important antioxidants (Koh et al. 2008; Carbonaro and Mattera 2001).

To our best knowledge, no data about differences in the lunasin content of organic and conventional crops were published so far.

Therefore, the aim of this study was to compare the content of lunasin peptide in the grain of organically and conventionally grown spring barley genotypes and to estimate the effect of the genotype and environment.

2 Material and methods

2.1 Barley genotypes and experimental conditions

A total of 22 spring barley varieties and breeding lines differing in their plant morphological characteristics, origin, and year of registration (Table 1) were grown in field trials under organic and conventional management in Priekuli (lat. 57°19' N, long. 25°20'E). Eight of the genotypes were selected for organic farming. The experiment was conducted during 2010 and 2011 in 6.5 m² plots with three replications and a randomized complete block design.

The trials were arranged on sod-podzolic loamy sand; the soil properties are summarized in Table 2. In the conventional field, the precrop was potato. Mineral fertilizer before sowing at 80–83 kg/ha N, 45–48 kg/ha P₂O₅, and 75–84 kg/ha K₂O; the herbicide Secator (100 g/L amidosulfuron and 25 g/L iodosulfuron) at 0.15 L/ha; and insecticide Karate Zeon 5 m.s. (50 g/L lambda-cyhalothrin) at 0.15 L/ha were applied. In the organic field, the precrop was pea for green manure grown as main season crop. Weed control was performed by harrowing at the beginning of the tillering stage.

The grain yield was assessed by combine harvest of the whole plots after cleaning with 1.5 mm sieve and recalculating to a 14 % moisture content; the test weight, content of crude protein, and (1→3) and (1→4) β-D-glucans (beta-glucans) in the dry matter were determined using a Infratec 1241 Near-Infrared Transmittance grain analyzer (Foss, Högenäs, Sweden).

The meteorological conditions were, in general, favorable for barley development in both years. The mean air temperature during the barley vegetation period surpassed the long-term average (14.1 °C) by 2.6 and 2.4 °C in 2010 and 2011, respectively, resulting in a comparatively early maturity. In 2010, the amount of rainfall was 143 % of the long-term average (221.7 mm), which promoted lodging in the conventional field, whereas the precipitation was close to the average (93 %) in 2011.

Table 1 Barley genotypes included in the study

Variety, breeding line	Country of origin	Pedigree	Year of first release	Specific information
Abava	LV ^a	Mari/Elsa//Domen	1978	
Annabell	DE	90014-DH/Krona	1999	
Anni	EE	Lola/Lisa	1991	
BZ12-83	LV	Primus/Idumeja		Selected under OF ^b
BZ14-12	LV	Anni/Dziugiai		Selected under OF
BZ14-99	LV	Anni/Dziugiai		Selected under OF
Dziugiai	LT		1947	
Idumeja	LV	Imula/Ida	2000	Early maturity
Inari	FI	JO-1263/Triumph	1994	
Irbe	LV	Filippa/CDC McGwire//Kristaps	2011	Hulless barley
PR-3605	LV	Rūja/Prestige/3/L-2233//Linus/ Annabell		
PR-4121	LV	Tunika/L-3118		
PR-4181	LV	Hydrogen/H-155		
PR-4407	LV	Roxane/Danuta//Idumeja		
PR-4812	LV	Rubiola/L-3118		Selected under OF
PR-4814	LV	Danuta/L-3008//Rubiola		Selected under OF
PR-4825	LV	Abava/Annabell		
PR-5145	LV	Peggy/L-3118//Rubiola		Selected under OF
Primus	SE		1901	Tall plants, late maturity
Rasa	LV	Frankengold/KM-R-54/72	1996	
Rubiola	LV	Rūja/Run8/458	2011	Recommended for OF
Vienna	AT		2007	Registered for OF

^a ISO 3166-1-alpha-2 code^b Organic farming

In 2010, in the conventional field, an unusually high infection level of Barley Yellow Dwarf Virus was observed (average score of 5.7, range of 2.3–8.3; scale from 0=0 % to 9=100 % infection level).

2.2 Materials and reagents

Acetonitrile, methanol and hexane, both gradient grade, formic acid ($\geq 99\%$), trifluoroacetic acid ($\geq 99\%$), and protease cocktail were supplied by Sigma-Aldrich. The water used in this work was purified using a Milli-Q water purification system from Millipore. The standard for synthetic lunasin was purchased from CASLO Laboratory ApS (c/o Scion Denmark Technical University, Lyngby, Denmark), and working

Table 2 Soil agrochemical properties at the study site

Soil characteristics	2010		2011	
	Organic	Conventional	Organic	Conventional
pH KCL	5.7	5.5	5.4	5.4
Humus content, %	2.8	2.6	2.1	3.0
K ₂ O, mg/kg	144	132	98	165
P ₂ O ₅ , mg/kg	111	100	116	187

solutions were prepared before the sample analyses. Standard addition method was used to solve the matrix effect problem by diluting the stock solution with sample solution of known concentration of the lunasin. The stock solution of the standard at a concentration of 40 $\mu\text{g/mL}$ was prepared by dissolving the peptide in water and storing at 4 °C.

2.3 Instrumentation

The chromatographic analysis was performed using a modular high-performance liquid chromatography (HPLC) system, Waters 2690 Alliance, consisting of quaternary pump, autosampler, and column thermostat, coupled to an electrospray ionization tandem mass spectrometer (Waters Micromass Quattro MicroTM API (Atmospheric Pressure Ionization)). The HPLC separations were achieved using a reverse-phase Phenomenex Synergi Hydro-RP 4 μm , (150 \times 2.0 mm inner diameter) analytical column (30 °C), with a mobile phase composed of 0.1 % formic acid in water (A) and 0.1 % formic acid in acetonitrile (B) in gradient mode at a flow rate 300 $\mu\text{L/min}$. The separation of the peptide was accomplished using a linear gradient of 20 to 60 % B over 8.0 min, 60 % B was maintained for 3.0 min, followed by 60–20 % for another 1.0 min, and 20 % B was maintained for 8.0 min to reach the initial conditions. The injection volume was 50 μL .

The quadrupole protonated molecular ion, with m/z 1258 for lunasin, was detected by single-ion recording. The mass spectra were measured using a Micromass Quatro Micro triple-quadrupole spectrometer equipped with an electrospray ionization source. The analyses were performed in the positive ion mode. The source temperature was 120 °C, and the desolvation temperature was 250 °C. Nitrogen was used as the nebulizing gas (600 L/h). The electrospray capillary was set at 3.0 kV. The mass spectrometry analyses were performed at a cone voltage of 60 V. The data analyses were performed using MasLynx version 4.1 software (Waters Corporation).

2.4 Measurement of lunasin content

Reverse-phase chromatography coupled to an electrospray ionization source was used to separate and ionize lunasin. The grain samples were pooled over the three field replications in equal amounts and ground using a Falling number Laboratory mill 3100 (Perten Instruments) with a 0.6-mm sieve. To isolate lunasin, we used an assay similar to that of Jeong et al. (2007a) and Nakurte et al. (2012).

In brief, 5 g of flour was extracted with 50 mL of 0.1 M phosphate-buffered saline buffer, pH 7.4, supplemented with fresh protease inhibitor cocktail (Sigma, St. Louis, USA) at a concentration of 1 % v/v by stirring on a magnetic stirrer for 48 h at 4 °C. The extract was centrifuged at 15,000 rpm for 30 min at 4 °C, and the supernatants were collected. The mixture was transferred to a new tube and twice extracted with 20 mL of hexane and cleared by centrifugation (15,000 rpm for 5 min). The upper layer was discarded, and the lower layer was purified by solid-phase extraction (SPE) using Strata X 3 mL, 60 mg (Phenomenex) cartridges. The SPE columns were pretreated with 3.0 mL of methanol, followed by 3.0 mL of water and sample. The column was washed with 3.0 mL of 0.1 % formic acid in water, and 0.1 % trifluoroacetic acid in 50 % acetonitrile (3.0 mL) was used to elute the analytes from the extraction column. The eluate was dissolved 1:1 (v/v) with mobile phase A and injected in the liquid chromatography-mass spectrometry/mass spectrometry system. Measurement for each sample was performed in three technical replications.

2.5 Data analysis

Analysis of variance (ANOVA) was carried out using GENSTAT 14.0 (2011); data of the three technical replications were submitted, and model GENOTYPE (G)+MANAGEMENT SYSTEM (M)+YEAR (Y)+G×Y+G×M+Y×M+G×Y×M was applied. Partitioning of sum of squares was calculated from the ANOVA for each factor and their interaction as percentage from the total.

To estimate stability of lunasin concentration, ecovalence (W_i) was computed using Microsoft Excel software as

described by Becker and Léon (1988) and expressed in percentage of the total interaction sum of squares.

Pearson phenotypic correlation coefficients were calculated using Microsoft Excel between the values of lunasin in both management systems and between lunasin and the plant traits assumed to influence lunasin, i.e., grain yield, thousand grain weight, grain test weight, beta-glucans, and crude protein content for each environment; correlation between lunasin and Barley Yellow Dwarf Virus infection score under conventional management in 2010 was calculated.

The broad-sense heritability was estimated from the variance components using the following formula:

$$h^2 = 100V_g / (V_g + V_{gm}/m + V_{gy}/y + V_{gmy}/my + V_e/rmy)$$

where V_g is the genotypic variance, V_{gm} is the variance of genotype × management system interaction, V_{gy} is the variance of genotype × year interaction, V_{gmy} is the variance of genotype × management system × year interaction, V_e is the error variance, r is the number of replications, m is the number of management systems, and y is the number of years;

$$\begin{aligned} V_g &= (MS_g - MS_{gm} - MS_{gy} - MS_{gmy}) / rmy \\ V_{gm} &= (MS_{gm} - MS_{gmy}) / ry \\ V_{gy} &= (MS_{gy} - MS_{gmy}) / rm \\ V_{gmy} &= (MS_{gmy} - MS_e) / r \\ V_e &= MS_e \end{aligned}$$

where MS is the respective mean square from the ANOVA.

3 Results and discussion

3.1 Range of lunasin content and effect of genotype

The content of lunasin in the individual barley grain samples ranged from 5.0 to 189.0 $\mu\text{g/g}$ (Table 3).

The effect of the genotype on the lunasin content was significant (Table 4). The mean lunasin content in both management systems and in both years significantly surpassed the average value of the respective environment for four genotypes, ‘Dziugiai,’ ‘Idumeja,’ ‘Rubiola,’ and PR-4121, whereas it was below the average for nine genotypes. A low lunasin content of 10 $\mu\text{g/g}$ and less in all environments was registered for varieties ‘Rasa’ and ‘Inari.’ Jeong et al. (2009) showed that lunasin was present in 15 out of 21 cultivars of rye analyzed. Such varying amounts of lunasin in different rye genotypes grown in identical conditions confirm that genotype is a primary determinant of the composition of plant secondary metabolites.

Table 3 Lunasin content (micrograms per gram) in barley grain during 2010–2011 in organic (O) and conventional (C) management systems and ecovalence ($W_i\%$) over all environments and over organic and conventional management systems

Genotype	2010		2011		$W_i\%$	$W_i\%$ O	$W_i\%$ C
	O	C	O	C			
Abava	34.2 a –	27.1 b	31.4 a –	20.5 b –	0.80	1.22	1.58
Annabell	84.1 a +	18.7 b –	79.8 a +	22.9 b –	3.25	1.60	1.00
Anni	17.3 –	19.1 –	19.5 b –	92.3 a +	10.57	0.29	3.52
BZ12-83	9.9 –	10.0 –	11.3 –	14.6 –	1.21	0.39	0.40
BZ14-12	7.4 b –	17.3 a –	13.5 b –	54.6 a +	5.05	0.01	1.75
BZ14-99	13.6 –	9.1 –	23.7 a –	12.4 b –	0.50	0.12	0.34
Dziugiai	108.6 a +	82.3 b +	95.4 b +	168.0 a +	9.66	5.11	22.12
Idumeja	61.3 b +	80.7 a +	72.6 +	73.1 +	2.41	0.24	12.58
Inari	5.0 –	8.0 –	5.1 –	5.0 –	1.39	0.60	0.24
Irbe	24.7 –	19.1 –	19.1 b –	30.3 a –	1.37	2.00	1.19
PR-3605	12.0 –	10.0 –	13.3 –	9.2 –	0.93	0.40	0.34
PR-4121	151.3 a +	131.6 b +	156.0 b +	189.0 a +	3.58	0.06	42.05
PR-4181	68.9 a +	11.7 b –	80.6 a +	16.2 b –	3.07	0.28	0.50
PR-4407	99.8 a +	18.2 b –	139.0 a +	15.6 b –	14.12	13.06	0.84
PR-4812	23.9 a –	3.5 b –	16.7 a –	6.6 b –	0.38	2.49	0.12
PR-4814	30.9 a –	14.4 b –	36.0 a –	10.7 b –	0.28	0.05	0.56
PR-4825	47.0 a	12.8 b –	54.5 a	20.8 b –	0.31	0.00	0.62
PR-5145	86.3 a +	13.4 b –	166.0 a +	19.4 b –	20.56	66.55	0.63
Primus	10.1 –	13.0 –	18.0 –	13.9 –	1.06	0.01	0.53
Rasa	5.0 –	7.0 –	8.0 –	10.0 –	1.25	0.20	0.24
Rubiola	169.3 a +	61.8 b +	178.1 a +	69.2 b +	14.77	0.04	8.42
Vienna	78.9 a +	11.3 b –	65.5 a +	12.3 b –	3.48	5.26	0.43
Average	52.3	27.3	59.2	40.3			

The mean values of the respective genotype marked with a different letter (a, b) are significantly different between the management systems ($p < 0.05$) within the respective year; the mean values significantly above or below the average value of all genotypes ($p < 0.05$) in the respective column are marked with “+” and “–,” respectively; $LSD_{0.05} = 6.22$ when comparing means with the same level of year and management system; $LSD_{0.05} = 6.14$ for other comparisons; minimum and maximum values of the column are shown in italics

The range of lunasin content in the barley genotypes analyzed in this study was larger than that reported by Jeong et al. (2002), showing that the genetic diversity of barley is considerably wider than previously supposed. The mean lunasin content value in our experiment was 44.8 $\mu\text{g/g}$, but the maximal value was 189.0 $\mu\text{g/g}$. In comparison with other cereals, this value was slightly less than that reported for wheat (Jeong et al. 2007a; Nakurte et al. 2012) and was similar to that for rye reported by Jeong et al. (2009) and oat reported by us (Nakurte et al. 2013) but considerably less than found in rye

and triticale grown under similar conditions in Latvia (Nakurte et al. 2012).

The broad-sense heritability for the lunasin content was 70.4 %. Correlations between the lunasin content in both systems and in both harvest years were significant in all cases, $p < 0.01$, except $p < 0.05$ for the correlation between the organic and conventional systems in 2011. The correlation coefficients were the highest between both years in each management system (0.94 for organic and 0.90 for conventional) and ranged between 0.44 and 0.69 in the other cases. The possibility of selecting and breeding soybean with a high lunasin content due to the noticeable variation of lunasin concentrations was suggested by Hernández-Ledesma et al. (2009). The comparatively large effect of the genotype, significant correlations between the lunasin content in different environments, and the high broad-sense heritability estimate also support the possibility of selecting and breeding lunasin-rich barley genotypes.

Although barley is not as rich in lunasin as soybean and triticale, we suggest that lunasin improvement in barley for functional food purposes may provide an additional value to this health-promoting cereal. Barley contains well-balanced protein, minerals, vitamins, especially vitamin E, and

Table 4 Partitioning of the sum of squares, mean square, and significance of the genotype (G), management system (M), year (Y), and their interaction affecting the lunasin content in barley grain

G	M	Y	G × M	G × Y	G × M × Y
Partitioning of sum of squares, %					
68.41	5.21	1.08	19.36	2.21	3.22
Mean square					
19917	31826	6615	5636	642	938
p value					
<0.001	<0.001	<0.001	<0.001	<0.001	<0.001

insoluble and soluble fiber including beta-glucans, as well as much greater amounts of phenolic compounds (0.2–0.4 %) than other cereal grains. Renewed interest in barley for food uses largely focuses on the effects of beta-glucans which are found almost exclusively in barley and oats on lowering blood cholesterol levels and glycemic index (Baik and Ullrich 2008). Noteworthy, lunasin peptide also possesses antioxidant, anti-inflammatory, and cholesterol-lowering properties. Hulless barley could be of special interest because of generally higher content of beta-glucans and some other nutritionally valuable substances than covered barley and no need for dehulling and pearling for use in wholegrain foods. Unfortunately, the lunasin content in hulless variety 'Irbe' was significantly below the average.

Although barley was presumably first among the cereals used as human food, only 2–3 % of barley crop is used for food nowadays (Ullrich 2011) and can be called the most underestimated among small grain cereals. In addition to various food uses in porridges, soups, stews, flatbreads, muffins, pasta, etc., Kinner et al. (2011) have shown that the baking quality of hulless barley flour can be sufficient to bake pure hulless barley bread.

The maximal lunasin content found by us in triticale (Nakurte et al. 2012) surpassed that of barley by approximately 34-fold; therefore, the breeding of this cereal species as a functional food with an improved lunasin content is a good prospect. However, the conclusions from the present study on barley may be useful for future investigations in cereals; it is possible that the interconnections in triticale are similar to those in barley. Triticale is a good source of protein and energy and is used mainly for animal feed and very little for human consumption (Peña 2004); a little breeding effort was made for developing varieties suitable for food hitherto, but there is a potential and discovery of high lunasin content is one of the reasons to work in this direction. Barley is described as one of the most genetically diverse cereal crops (Baik and Ullrich 2008) with wide differences in physical and compositional characteristics and accordingly has different processing properties and end-use quality. Therefore, it is possible that testing a larger number of genotypes may result in the identification of barley with an even higher lunasin content.

No significant correlations between the content of lunasin and grain quality parameters such as content of crude protein, beta-glucans, thousand grain weight, and test weight or the grain yield were observed. We also did not find significant correlations between lunasin and beta-glucan content in oat (Nakurte et al. 2013) and between protein content and lunasin content in triticale, wheat, and rye (Nakurte et al. 2012). As there was no negative interconnection found, it allows to breed varieties with improved health benefit combining both high content of lunasin and beta-glucans.

3.2 Effect of management system

The mean lunasin content was higher by 25.0 and 18.9 $\mu\text{g/g}$ (92 and 47 %) in the years 2010 and 2011, respectively, in the organic farming system when compared to the conventional system (Fig. 1), and the management system had a significant effect on the lunasin content (Table 4). Our results are in agreement with Wang et al. (2008), reporting that the lunasin concentration in soybean depends mainly on the genotype and, to some extent, on environmental factors. The effect of the farming system was significant in our experiment, even though only 5.2 % of the difference in the lunasin content was explained by the farming system in contrast to the 68.4 % for the genotype and 19.4 % for the interaction between both factors. The mean lunasin content difference between both management systems in both years exceeded 100 $\mu\text{g/g}$ for variety 'Rubiola.' 'Rubiola' was the most lunasin-rich genotype under organic management in both years, whereas line PR-4121 was prominent under conventional conditions and was the second best in the organic system, showing a maximum lunasin content throughout the experiment in 2011 (189.0 $\mu\text{g/g}$).

The obtained results support the value of organically grown food products for the improvement of human health, at least with regard to lunasin, as the mean lunasin content was significantly higher in the organic management system. This result is in agreement with Grønder-Pedersen et al. (2003), who noted that cereals grown in organic farming systems contain significantly more antioxidants and other biologically active substances. The possible reason for this finding can be the increased natural defense system of plants to the different stress factors that are encountered to a greater extent in organic farming, as no agrochemicals are applied and plants have to compete with weeds, diseases, and insects and to be able to uptake the nutrients from soil without supplement of easy accessible mineral fertilizers. An increase in phenolic compounds production by plants with the purpose of increasing the natural defenses is described by García-Mier et al. (2013). Additionally, Huber et al. (2011) reviewed several studies showing higher antioxidative and antimutagenic activities and better inhibition of cancer cell proliferation of organically produced foods when compared to conventionally produced foods. In our experiment, the largest differences between the systems were because of the nutrient supply and weed management. Our findings support the hypothesis that organic cereal foods may contain higher levels of phytonutrients including anticancer substances.

3.3 Genotype \times management system interaction and stability

The genotype and management system interaction was found to be significant showing that there are genotypes which synthesize more lunasin under organic farming and also

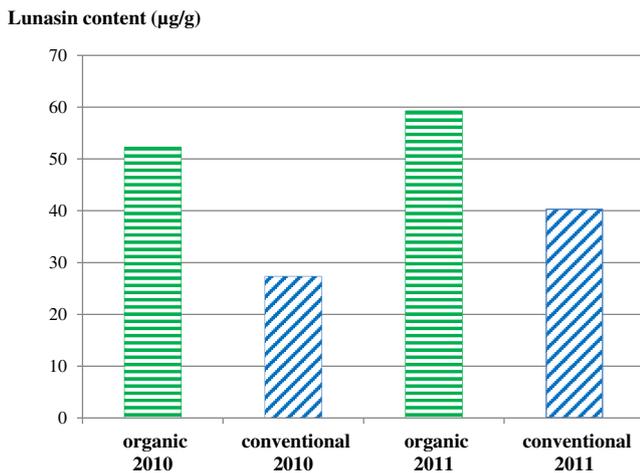


Fig. 1 Average lunasin content of 22 barley genotypes grown under organic and conventional management systems in 2010 and 2011; the difference between the management systems was significant, $LSD_{0.05}=1.08$

genotypes with higher lunasin under conventional farming. Ten genotypes had a significantly higher lunasin under the organic conditions in comparison to the conventional conditions in both years, whereas varieties ‘Idumeja,’ ‘Anni,’ and line BZ14-12 with ‘Anni’ in its pedigree exhibited an opposite tendency; the difference for ‘Idumeja’ and ‘Anni’ was significant in 1 year only (Table 3). Similarly to our results, contrasting responses to organic practices with respect to the phytochemical content was reported by Picchi et al. (2012) for two cauliflower genotypes.

The most promising barley genotypes with a high lunasin content exclusively under organic management were ‘Rubiola,’ PR-4407, and PR-5145; however, only ‘Rubiola’ was stable over both years according to the ecovalence, but PR-5145 was highly unstable. Ecovalence measures the contribution of genotype to genotype \times environment interactions; a genotype with W_i close to 0 is considered as stable (Becker and Léon 1988). ‘Rubiola’ is a recently registered variety with agronomic traits that are appropriate for growth in organic farming (Legzdina et al. 2008) (Fig. 2). The mean decrease of the mean lunasin content under conventional conditions was 62 % for ‘Rubiola’ and more than 80 % for breeding lines PR-4407 and PR-5145. Breeding line PR-4121 and an old Lithuanian variety, ‘Dziugiai,’ provided comparatively high lunasin content in both farming systems. PR-4121 was relatively stable over all environments and over organic environments, but the most unstable between the years in the conventional system, whereas ‘Dziugiai’ was fairly stable under organic system but unstable under conventional one. We presume that the lunasin content was not influenced by the year of variety release and was not lost during the intensive breeding work performed because the other old variety ‘Primus’ had a low lunasin content in both systems.



Fig. 2 Above: spring barley variety ‘Rubiola’ registered as appropriate for organic farming in Latvia. In the grain of ‘Rubiola,’ the highest lunasin content was found in trials under organic management system. Below: field experiment in organically managed field in 2011 provided the highest mean lunasin content. Photo by L. Legzdina

3.4 Effect of the year and genotype \times year interaction

A higher mean lunasin content was found in 2011 than in 2010 in both management systems; the effect of a particular harvest year conditions including meteorological, soil, and other peculiarities on the differences in the lunasin content was significant and was estimated at 1.08 %. The difference between the years was notably larger under conventional conditions than under organic ones; the reduction in 2010 if compared to 2011 was 32 and 11.8 %, respectively. One of the possible reasons for this effect could be the unusually high incidence of Barley Yellow Dwarf Virus infection in the conventional field in 2010, as there is a possibility that this virus could hinder lunasin synthesis in the grain. However, no correlation between the lunasin content and virus infection score was found in the particular environment. No other substantial differences in environmental factors such as average temperatures, amount of precipitation and soil characteristics between the years were recorded; the vegetation period of 2011 was fairly drier if compared to 2010. Effect of temperature, soil moisture, and the interactions between these factors and the genotype were found in soybean by Wang et al. (2008); significant effects of genotype \times environment interactions were reported in respect to other biologically active chemicals in cereals, e.g., 5-*n* alkylresorcinols and free phenolics in durum wheat (Bellato et al. 2013) and vitamin E isomers in barley (Ehrenbergerová et al. 2006). Genotype \times year and genotype \times management system \times year interactions were also significant and explained 2.21 and 3.22 % of the differences in lunasin content, respectively. Two genotypes

with high lunasin content, PR-4121 and ‘Dziugiai,’ showed opposite reaction to the management systems in both years.

To summarize, our findings provide new information on variation of lunasin in barley grain, proves higher average lunasin content in organically grown grain, and shows that the reaction of barley genotypes to organic and conventional management and to the environmental factors of the year in connection with the synthesis of lunasin in the grain can be different.

4 Conclusions

The lunasin content in barley grain samples ranged from 5.0 to 189.0 µg/g. The effects of the genotype, crop management system, year, and the interaction between the factors on the lunasin content in barley grain were significant with the largest proportion of genotype. The mean lunasin content was higher under organic management than that in the conventional system. The barley genotypes responded differently to the management system: 10 of the 22 genotypes synthesized significantly more lunasin under organic management in both years, whereas one genotype synthesized more lunasin under conventional management and two genotypes had conversely results between the years.

Acknowledgments This research was supported by European Social Found co-financed project Nr. 2009/0218/1DP/1.1.1.2.0/09/APIA/VIAA/099. The authors thank Aina Kokare for the help in data statistical analysis.

References

- Baik BK, Ullrich SE (2008) Barley for food: characteristics, improvement, and renewed interest. *J Cer Sci* 48:233–242. doi:10.1016/j.jcs.2008.02.002http
- Becker HC, Léon J (1988) Stability analysis in plant breeding. *Plant Breed* 101:1–23
- Bellato S, Ciccioritti R, Del Frate V, Sgrulletta D, Carbone K (2013) Influence of genotype and environment on the content of 5-n alkylresorcinols, total phenols and on the antiradical activity of whole durum wheat grains. *J Cer Sci* 57:162–169
- Benbrook CM (2005) Elevating antioxidant levels in food through organic farming and food processing. The Organic Center for Education and Promotion. http://www.organic-center.org/reportfiles/Antioxidant_SSR.pdf. Accessed 10 Oct 2012
- Bour D, Prescott J (2002) Comparison of the nutritional value, sensory qualities, and food safety of organically and conventionally produced foods. *Crit Rev Food Sci Nutr* 42:1–34. doi:10.1080/10408690290825439
- Cam A, Sivaguru M, Gonzalez de Mejia E (2013) Endocytic mechanism of internalization of dietary peptide lunasin into macrophages in inflammatory condition associated with cardiovascular disease. *PLoS One* 5:8. doi:10.1371/journal.pone.0072115
- Carbonaro M, Mattera M (2001) Polyphenoloxidase activity and polyphenol levels in organically and conventionally grown peach (*Prunus persica* L., cv. Regina bianca) and pear (*Pyrus communis* L., cv. Williams). *Food Chem* 72:419–424. doi:10.1016/S0308-8146(00)00248-X
- de Lumen BO (2005) Lunasin: a cancer-preventive soy peptide. *Nutr Rev* 63:16–21. doi:10.1301/nr.2004.janr.16-21
- de Mejia EG, Vasconez M, de Lumen BO, Nelson R (2004) Lunasin concentration in different soybean genotypes, commercial soy protein, and isoflavone products. *J Agric Food Chem* 52:5882–5887. doi:10.1021/jf0496752
- Dzirkale Z, Rumaks J, Svirskis S, Mazina O, Allikalt A, Rinken A, Jekabsons K, Muceniece R, Klusa V (2013) Lunasin-induced behavioural effects in mice: focus on the dopaminergic system. *Behav Brain Res* 256:5–9. doi:10.1016/j.bbr.2013.08.002
- Ehrenbergerová J, Belcrediová N, Prýma J, Vaculová K, Newman CW (2006) Effect of cultivar, year grown, and cropping system on the content of tocopherols and tocotrienols in grains of hulled and hullless barley. *Plant Foods Hum Nutr* 61:145–150
- Galvez AF, de Lumen BO (1999) A soybean cDNA encoding a chromatin-binding peptide inhibits mitosis of mammalian cells. *Nat Biotechnol* 17:495–500. doi:10.1038/8676
- García-Mier L, Guevara-González RG, Mondragón-Olguín VM, Verduzco-Cuellar BR, Torres-Pacheco I (2013) Agriculture and bioactives: achieving both crop yield and phytochemicals. *Int J Mol Sci* 14:4203–4222. doi:10.3390/ijms14024203
- Grinder-Pedersen L, Rasmussen SE, Bügel S, Jørgensen LV, Dragsted LO, Gundersen V, Sandström B (2003) Effect of diets based on foods from conventional versus organic production on intake and excretion of flavonoids and markers of antioxidative defense in humans. *J Agric Food Chem* 51:5671–5676. doi:10.1021/jf030217n
- Hernández-Ledesma B, Hsieh CC, de Lumen BO (2009) Lunasin, a novel seed peptide for cancer prevention. *Pept* 30:426–430. doi:10.1016/j.peptides.2008.11.002
- Hernández-Ledesma B, Hsieh CC, de Lumen BO (2013) Chemopreventive properties of peptide lunasin: a review. *Protein Pept Lett* 20:424–432. doi:10.2174/092986613805290327
- Huber M, Rembiałkowska E, Średnicka D, Bügel S, van de Vijver LPL (2011) Organic food and impact on human health: assessing the status quo and prospects of research. *NJAS—Wagen. J Life Sci* 58:103–109
- Jeong HJ, Lam Y, de Lumen BO (2002) Barley lunasin suppresses ras-induced colony formation and inhibits core histone acetylation in mammalian cells. *J Agric Food Chem* 50:5903–5908. doi:10.1021/jf0256945
- Jeong HJ, Jeong JB, Kim DS, Park JH, Lee JB, Kweon DH, Chung GY, Seo EW, de Lumen BO (2007a) The cancer preventive peptide lunasin from wheat inhibits core histone acetylation. *Cancer Lett* 255:42–48. doi:10.1016/j.canlet.2007.03.022
- Jeong JB, Jeong HJ, Park JH, Lee SH, Lee JR, Lee HK, Chung GY, Choi JD, de Lumen BO (2007b) Cancer-preventive peptide lunasin from *Solanum nigrum* L. inhibits acetylation of core histones H3 and H4 and phosphorylation of retinoblastoma protein (Rb). *J Agric Food Chem* 55:10707–10713. doi:10.1021/jf072363p
- Jeong HJ, Lee JR, Jeong JB, Park JH, Cheong Y, de Lumen BO (2009) The cancer preventive seed peptide lunasin from rye is bioavailable and bioactive. *Nutr and Cancer* 61:680–686. doi:10.1080/01635580902850082
- Jeong HJ, Jeong JB, Hsieh CC, Hernández-Ledesma B, de Lumen BO (2010a) Lunasin is prevalent in barley and is bioavailable and bioactive in vivo and in vitro studies. *Nutr Cancer* 62:1113–1119. doi:10.1080/01635581.2010.515529
- Jeong JB, De Lumen BO, Jeong HJ (2010b) Lunasin peptide purified from *Solanum nigrum* L. protects DNA from oxidative damage by suppressing the generation of hydroxyl radical via blocking fenton reaction. *Cancer Lett* 293:58–64. doi:10.1016/j
- Kinner M, Nitschko S, Sommeregger J, Petrasch A, Linsberger-Martin G, Grausgruber H, Berghofer E, Siebenhandl-Ehn S (2011) Naked barley-optimized recipe for pure barley bread with sufficient beta-glucan according to the EFSA health claims. *J Cereal Sci* 53:225–230. doi:10.1016/j.jcs.2011.01.001

- Koh E, Wimalasiri KMS, Renaud ENC, Mitchel AE (2008) A comparison of flavonoids, carotenoids and vitamin C in commercial organic and conventional marinara pasta sauce. *J Sci Food Agric* 88:344–354. doi:10.1002/jsfa.3097
- Legzdina L, Gaike M, Gaile Z, Bērziņa I (2008) Testing results of spring barley variety ‘Rubiola’. *Latv J Agron* 11:94–101
- Lombardo S, Pandino G, Mauromicale G (2012) Nutritional and sensory characteristics of “early” potato cultivars under organic and conventional cultivation systems. *Food Chem* 133:1249–1254. doi:10.1016/j.foodchem.2011.10.005
- Magkos F, Arvaniti F, Zampelas A (2003) Organic food: nutritious food or food for thought? A review of the evidence. *Int J Food Sci Nutr* 54:357–371. doi:10.1080/09637480120092071
- Murniece I, Kruma Z, Skrabule I (2012) Carotenoids and colour before and after storage of organically and conventionally cultivated potato genotypes in Latvia. *World Acad Sci Eng Technol* 67:1201–1205
- Nakurte I, Klavins K, Kirhnere I, Namniece J, Adlere L, Matvejevs J, Kronberga A, Kokare A, Strazdina V, Legzdina L, Muceniece R (2012) Discovery of lunasin peptide in triticale (X *Triticosecale* Wittmack). *J Cereal Sci* 56:510–514. doi:10.1016/j.jcs.2012.04.004
- Nakurte I, Kirhnere I, Namniece J, Saleniece K, Krigere L, Mekss P, Vicupe Z, Bleidere M, Legzdina L, Muceniece R (2013) Detection of the lunasin peptide in oats (*Avena sativa* L.). *J Cereal Sci* 57:319–324. doi:10.1016/j.jcs.2012.12.008
- Park JH, Jeong HJ, de Lumen BO (2005) Contents and bioactivities of lunasin, Bowman-Birk inhibitor, and isoflavones in soybean seed. *J Agric Food Chem* 53:7686–7690
- Peña RJ (2004) Food uses of triticale. In: Mergoum M, Gómez-Macpherson H (eds), *Triticale improvement and production*. FAO Plant Production and Protection Paper 179, pp 37–49. [ftp://ftp.fao.org/docrep/fao/009/y5553e/y5553e01.pdf](http://ftp.fao.org/docrep/fao/009/y5553e/y5553e01.pdf). Accessed 18 Nov 2013
- Picchi V, Migliori C, Scalzo RL, Campanelli G, Ferrari V, Di Cesare LF (2012) Phytochemical content in organic and conventionally grown Italian cauliflower. *Food Chem* 130:501–509. doi:10.1016/j.foodchem.2011.07.036
- Silva-Sánchez C, de la Rosa AP, León-Galván MF, de Lumen BO, de León-Rodríguez A, de Mejía EG (2008) Bioactive peptides in amaranth (*Amaranthus hypochondriacus*) seed. *J Agric Food Chem* 56:1233–1240. doi:10.1021/jf072911z
- Smith-Spangler C, Brandeau ML, Hunter GE, Bavinger JC, Pearson M, Eschbach PJ, Sundaram V, Liu H, Schirmer P, Stave C, Olkin I, Bravata DM (2012) Are organic foods safer or healthier than conventional alternatives?: a systematic review. *Ann Intern Med* 157:348–366. doi:10.7326/0003-4819-157-5-201209040-00007 DOI:10.7326%2F0003-4819-157-5-201209040-00007
- Ullrich SE (2011) Significance, adaptation, production, and trade of barley. In: Ullrich SE (ed) *Barley: production, improvement, and uses*. Wiley-Blackwell, Ames, pp 3–13
- Wang W, Dia VP, Vasconez M, de Mejia EG, Nelson RL (2008) Analysis of soybean protein-derived peptides and the effect of cultivar, environmental conditions, and processing on lunasin concentration in soybean and soy products. *J AOAC Int* 91:936–946
- Woese K, Lange D, Boess C, Bögl KW (1997) A comparison of organically and conventionally grown foods—results of a review of the relevant literature. *J Sci Food Agric* 74:281–293. doi:10.1002/(SICI)1097-0010(199707)74:3<281::AID-JSFA794>3.0.CO;2-Z
- Worthington V (2001) Nutritional quality of organic versus conventional fruits, vegetables, and grains. *J Altern Complement Med* 7:161–173