Systematic molecular and cytogenetic screening of 100 patients with marfanoid syndromes and intellectual disability


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The association of marfanoid habitus (MH) and intellectual disability (ID) has been reported in the literature, with overlapping presentations and genetic heterogeneity. A hundred patients (71 males and 29 females) with a MH and ID were recruited. Custom-designed 244K array-CGH (Agilent®; Agilent Technologies Inc., Santa Clara, CA) and MED12, ZDHHC9, UPF3B, FBN1, TGFBR1 and TGFBR2 sequencing analyses were performed. Eighty patients could be classified as isolated MH and ID: 12 chromosomal imbalances, 1 FBN1 mutation and 1 possibly pathogenic MED12 mutation were found (17%). Twenty patients could be classified as ID with other extra-skeletal features of the Marfan syndrome (MFS) spectrum: 4 pathogenic FBN1 mutations and 4 chromosomal imbalances were found (2 patients with both FBN1 mutation and chromosomal rearrangement) (29%). These results suggest either that there are more loci with genes yet to be discovered or that MH can also be a relatively non-specific feature of patients with ID. The search for aortic complications is mandatory even if MH is associated with ID since FBN1 mutations or rearrangements were found in some patients. The excess of males is in favour of the involvement of other X-linked genes. Although it was impossible to make a diagnosis in 80% of patients, these results will improve genetic counselling in families.

Conflict of interest

The authors declare no conflict of interest.
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Marfan syndrome (MFS) is a multisystem genetic disease that can affect the cardiovascular, skeletal, ophthalmic, and integumentary systems and the dura mater. The diagnosis is based on an international classification (1–3). The severity of the condition lies in the risk of dilation and subsequent dissection of the ascending aorta. The *FBN1* gene is the major gene in patients with MFS, and variable expression of the disease has been largely identified within and between families (4, 5). Patients with atypical presentations not fulfilling the criteria involving only one organ system exist but are rare (6). Patients with MFS do not usually fulfill the international criteria (ID) (7). Patients with MFS can also display mutations in the *TGFB1* and *TGFB2* genes (8).

The term marfanoid habitus (MH) is used to describe patients with skeletal signs suggestive of MFS but who do not meet the international criteria (5, 7). The association of a MH and ID has been reported in the literature in syndromes with overlapping features. ID is defined, according to the American Association of Intellectual and Developmental Disabilities as significant limitations both in intellectual functioning (referring to general mental capacity such as learning, reasoning, and problem solving) and in adaptive behaviour (comprising conceptual, social and practical skills), and originates before the age of 18. Its incidence is 1.26/100 (9). The Lujan-Fryns syndrome (LJS) was first described in male patients with a family history suggesting an X-linked mode of inheritance (10, 11). The term LJS is now recognized to describe patients with MH (long, hyperlax fingers and toes, tall stature, dolicostenomelia), facial dysmorphism (prominent forehead contrasting with a long, narrow face, maxillary hypoplasia, small mandible, long nose with a high narrow nasal root); mild to moderate ID with behavioural abnormalities, that could include emotional instability, shyness, or even psychotic disturbances, generalized hypotonia, nasal speech, normal development of sexual features (12, 13). This syndrome has also been reported occasionally in females (14, 15). Thoracic aortic aneurysm (TAA) is usually absent, but has been reported in two families, including one patient who required aortic surgery (16, 17). In 2007, the systematic screening of 737 genes annotated in the Vertebrate Genome Annotation database on the human chromosome X in 250 families with syndromic or non-syndromic X-linked ID led to the identification of hemizygous mutations in the *MED12*, *ZDHHC9*, and *UPF3B* genes in a very small number of familial cases (18–20). The *MED12* gene (MIM 300188, also called *HOPA* or *TRAP230*), located in Xq13, encodes for a subunit of the macromolecular complex known as Mediator, which is required for thyroid hormone-dependent activation and repression of transcription by RNA polymerase II (21). Med12-deficient zebra fish embryos show defects in the brain and neural crest and do not survive beyond 1 week after fertilization (22). *MED12* mutation have been found in only one large family with LJS, which was the index family reported by Lujan, and carrier females displayed no bias in chromosome X inactivation (11, 19). Mutations in the *ZDHHC9* gene (MIM 300646, zinc finger, and DHHC-type containing 9), located in Xq26.1, have been found in three families with LJS (18). It encodes a palmitoyl transferase, highly expressed in the brain, which catalyzes the post-translational modification of rat sarcoma viral oncogene homolog (RAS). The mechanism by which loss of function mutations of *ZDHHC9* lead to ID is unclear, but it may be through alteration of the relative proportion of the RAS proteins within the different compartments of nerve cells (23). Finally, mutations of the *UPF3B* gene (MIM 200298), located on Xq24, have been found in two families with LJS (20). The UPF3B protein is an important component of the nonsense-mediated mRNA decay surveillance machinery (24) and the mechanism by which loss of function mutations of *UPF3B* lead to ID is also unknown. Mutations of these three genes have also been found in patients with Opitz–Kaveggia (FG) syndrome (18, 20, 25) and in non-syndromic ID for *UPF3B* (20). These results have not been replicated to date in LJS.

ID and MH can also be found in Shprintzen–Goldberg syndrome (SGS) associated with...
craniosynostosis (26, 27) and rare de novo mutations have been described in the FBN1 or TGFBR2 genes. These results are still a subject of debate in the literature (28–30). ID is only reported very occasionally in Loeys-Dietz syndrome (LDS), secondary to mutations in the TGFBR1 and TGFBR2 genes (31). Finally, chromosomal imbalances have also been reported in sporadic observations, including one 15q21.1q21.3 deletion involving the FBN1 gene (32) and two cases of terminal deletion of chromosome 5p (33, 34).

Therefore, the genetic aetiology of the association of a MH and ID represents a true challenge for individual diagnosis, follow-up and genetic counselling, particularly for sporadic presentations. Besides the clinical overlap between these different entities, the implication of each gene and its clinical spectrum in such a phenotype is unknown and little is known about the risk of aortic involvement. The aim of this research project was to conduct a clinical, cytogenetic and molecular study of a large cohort of 100 probands presenting with a skeletal marfanoid phenotype and ID, in order to provide answers to these questions, and help in the management of these patients.

**Materials and methods**

**Patients**

This study was prospectively designed. Patients were included by a clinical geneticist over an 18 months period. Inclusion criteria were the presence of ID and certain skeletal features of the Marfan spectrum, including at least two of the following clinical signs: height greater than the 97th centile in the absence of tall stature in parents, long, thin habitus, long, thin fingers, pectus abnormalities, and dolichostenomelia. A cardiac ultrasound and anterior eye chamber examination were required for all patients. The presence of heart and eye manifestations of the Marfan spectrum was not an exclusion criterion. Neuropsychological examination in order to evaluate non-subjectively the degree of ID could be performed in 15% of the patients only. In the other cases, ID was graduated according to the geneticist experience. Patients with a family history of MFS without ID were excluded from the study because the segregation was in favour of a different cause for MH and ID, and homocystinuria also needed to be ruled out. Inclusion was validated also after standard karyotype and **FMR1** gene study (Fragile-X) did not show any anomaly, although patients with apparently balanced chromosomal rearrangements were not excluded. Patients with the MH and ID were mainly recruited through the network of reference centres for rare diseases in France. Informed consent was obtained for all patients and the study was approved by the local ethics committee.

A detailed clinical evaluation was performed for every patient during a specialized consultation, and a detailed clinical file specifically designed for this project was completed by the referring geneticist, describing the family history, clinical features of MFS and information regarding developmental data. Patients were divided into two groups, depending on the absence (group 1) or presence (group 2) of associated features characteristic of MFS (TAA, mitral valve prolapse or ectopia lentis). The median systemic score was calculated in the total cohort and among groups, according to the 2010 new Ghent criteria for the diagnosis of MFS and related disorders. A systemic score above 7 indicated systemic involvement (3).

**Molecular and cytogenetic analyses**

For etiological purposes, all patients were screened for mutations of the ZDHHC9, **UPF3B**, **MED12**, **TGFBR1**, **TGFBR2**, and **FBN1** genes and for chromosomal micro-rearrangements using 244 K custom microarray. Female patients were also screened for X-inactivation bias.

**Direct sequencing of ZDHHC9, UPF3B, MED12, FBN1, TGFBR1 and TGFBR2 genes**

Genomic DNA was extracted from blood samples of the patients and parents when available. Exons and exon–intron boundaries sequencing analyses of the **MED12**, **ZDHHC9**, **UPF3B**, **FBN1**, **TGFBR1** and **TGFBR2** genes were performed according to previously reported methods (8, 18, 20, 35, 36). Corresponding reference of the genomic DNA sequences were downloaded using Ensembl Genome Browser, www.ensembl.org. Sequence analyses were searched for between consensus and reference sequences using **SeqScape®** software v2.5 package (Applied Biosystems, Foster City, CA). Mutation nomenclature numbering was based on the current Ensembl transcript (http://www.hgvs.org). The pathogenic nature of a mutation was determined according to a database search, bioinformatic predictions and segregation of the mutation in the family. The Universal Mutation Database (UMD) was used for **FBN1** (37) and **TGFBR2** (38) requests. When a mutation was suspected to lead to the creation of a splice site, additional mRNA studies were performed. All patients were screened for **MED12**, **ZDHHC9**, **UPF3B**, **FBN1** and **TGFBR2** gene mutations. Only patients with TAA were screened for **TGFBR1** mutations.

**X-inactivation assay in female patients**

This assay was performed as previously reported (39) with some slight modifications, notably the use of fluorescent primers and the detection mode. A control was used in the analyses. X-inactivation assay in female patients was considered as biased when it was superior or equal to 85%, or even 95%.

**Array-CGH experiments**

A custom-designed array was designed with approximately 240,000 oligonucleotides manufactured by Diagnogene™ (Division Imaxio, Biopôle Saint-Beauzire, France). This array contains oligonucleotides
selected from the Agilent online library (earray; https://earray.chem.agilent.com/earray) and has been further empirically optimized. In addition, we increased probe density within seven selected genes (12,307, Table S1). The entire genome was covered with an average resolution of 20 kb (NCBI, hg 18). The procedures of array-CGH were performed according to the Agilent instructions with minor modifications (40). Slides were scanned using the Agilent G2565 Microarray scanner and images quantified using Agilent feature extraction (v9.0) and a graphical overview was obtained using Genomic Workbench software (v5.0). Mapping data were analysed on the human genome sequence using Ensembl (www.ensembl.org; hg18). Copy Number Variations were assessed in the Database of Genomic Variants (http://projects.tcag.ca/variation/). When a chromosomal imbalance was detected, quantitative polymerase chain reaction (qPCR) or fluorescence in situ hybridization (FISH) studies with probes derived from bacterial artificial chromosomes were performed in order to confirm the chromosomal imbalances, as well as the family segregation. When a chromosomal imbalance was suspected to be pathogenic, the gene content was determined in order to find candidate genes for the phenotype, with a particular interest for genes expressed in the central nervous system and connective tissues (http://www.ensembl.org/biomart/). We also checked if a known ‘microdeletion/microduplication syndrome’ was described through the decipher database (http://sanger.ac.uk) and the available literature, and if additional patients with a similar phenotype had already been reported/gathered.

Results

Patients

Among the 100 patients with marfanoid phenotype and ID, 71 were males and 29 females (sex ratio: 2.45), originating from 98 families. They were aged from 2 to 45 years with a mean of 19.1 ± 8.6 years. Fifty percent of patients were adults. A family history of ID was found in 23/98 families, including 48% in a first-degree relative (91% in a sibling and 9% in a parent), 17% in a second-degree relative (including 50% compatible with an X-linked inheritance) and 35% in other degrees (including 12% compatible with an X-linked inheritance). MH was associated with ID in only four familial cases. Two patients had apparently balanced de novo reciprocal translocations: t(12;19)(q13.3;p12), and t(2;22)(q33;q11.2).

Detailed description of the overall cohort is given in Table 1. A minority was screened for dural ectasia and protrusio acetabulae. Other miscellaneous features included nasal speech (n = 18), abnormal genitalia (n = 15), ptosis (n = 5), vertebral instability (n = 4), deafness (n = 3), hypertrophic cardiomyopathy (n = 2), pneumothorax (n = 1), atrial septal defect (n = 1), aneurysm of the interventricular septum (n = 1), cleft lip (n = 1), absent uvula (n = 1), optic nerve atrophy (n = 1), horseshoe kidney (n = 1) and supernumerary mammary (n = 1).

General clinical, molecular and cytogenetic characteristics of patients belonging to group 1 and 2 are presented in Table 1. Clinical details of patients with a molecular or cytogenetic abnormality are presented in Tables 2–4. The systemic score for patients with a positive cytogenetic or molecular result was not significantly different from patients with normal results (7.6 ± 3.2 vs 6.6 ± 2.9).

Molecular and cytogenetics results

FBN1 sequencing analysis

Five pathogenic FBN1 mutations were found, one splicing and four missense mutations (Table 2). Their pathogenicity was verified according to the UMD-predictor tool (41) and except the c.2534G>A mutation (p.Cys845Tyr), they had already been reported at least once in the literature. Interestingly, the c.4270C>G (p.Pro1424Ala) mutation was found in two unrelated patients with no aortic manifestation at 18 and 36 years. The segregation of the mutation could not be verified in either family. The mutation had already been reported in six other instances in the UMD-FBN1 database (8, 42–46), and no ID was noticed when clinical data were available. The pathogenic nature of the FBN1 c.8176C>T (p.Arg2726Trp) mutation in exon 64 was demonstrated by in vitro studies (47) and reported in a few instances in patients with isolated skeletal features and even incomplete penetrance (48). Within the UMD-FBN1 database, we found 11 families with this mutation, and aortic dilatation was reported in two patients (37, 49). The mutation was also found in the father and sister of the proband. When re-examined, they displayed some very mild skeletal features of the Marfan spectrum, including dolichostenomelia, arachnodactyly, high arched palate, dental crowding and myopia in the father, as well as dolichostenomelia, mild scoliosis, significant striae and high arched palate in the sister. The presence of ID in the proband was finally explained by the co-occurrence of a FBN1 mutation and a 17q21.31 microdeletion.

Four variants were considered as polymorphic or probably non-pathogenic, according to UMD-FBN1 predictor and segregation analyses (Table S2). Of note, although considered a polymorphism in this study, the c.6700G>A (p.Val2234Met) variant was considered pathogenic in various publications (45, 50). Additional investigations were necessary to draw conclusions regarding the pathogenicity of the synonymous variant c.5097 C>T (p.Tyr1699Tyr), based on the prediction of a potential exonic splicing enhancers (ESE) disruption by the Human Splicing Finder tool (51). This hypothesis was ruled out as mRNA studies on fibroblasts were not in favour of abnormal splicing. The median systemic score of patients with FBN1 mutations or deletions was above 7, whereas the median systemic score of patients without FBN1 mutations or deletions was below 7, but the results were not significantly different (8.1 ± 4.1 vs 6.7 ± 2.9).
Table 1. Description of the population and results obtained in the total cohort, and depending on the classification of patients

<table>
<thead>
<tr>
<th></th>
<th>Group 1 Isolated MH and ID n = 80</th>
<th>Group 2 MH, ID and another MFS feature(a) n = 20</th>
<th>Overall series n = 100</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Clinical characteristics</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sex ratio ((n = 100))</td>
<td>2.8</td>
<td>1.5</td>
<td>2.4</td>
</tr>
<tr>
<td>Mean age ± SD ((n = 100))</td>
<td>18.4 ± 7.8</td>
<td>21.8 ± 11.1</td>
<td>19.1 ± 8.6</td>
</tr>
<tr>
<td><strong>Cardiovascular</strong></td>
<td></td>
<td></td>
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</tr>
<tr>
<td>TAA ((n = 94))</td>
<td></td>
<td></td>
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</tr>
<tr>
<td>Mitral valve prolapse ((n = 94))</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Other valvular abnormalities ((n = 94))</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Eye</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ectopia lentis ((n = 92))</td>
<td>0/73 (0%)</td>
<td>2/19 (10%)</td>
<td>2/92 (2%)</td>
</tr>
<tr>
<td>Myopia ((n = 92))</td>
<td>20/73 (27%)</td>
<td>3/19 (16%)</td>
<td>23/92 (25%)</td>
</tr>
<tr>
<td>Other eye abnormality (strabismus, astigmatism, hypermetropia, and nystagmus) ((n = 79))</td>
<td>46/62 (74%)</td>
<td>8/17 (47%)</td>
<td>54/79 (68%)</td>
</tr>
<tr>
<td><strong>Skeleton</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Long and thin habitus ((n = 99))</td>
<td>76/80 (95%)</td>
<td>16/19 (84%)</td>
<td>92/99 (93%)</td>
</tr>
<tr>
<td>Arachnodactyly ((n = 98))</td>
<td>61/79 (77%)</td>
<td>13/19 (68%)</td>
<td>74/98 (75%)</td>
</tr>
<tr>
<td>Dolichostenomelia ((n = 96))</td>
<td>51/78 (65%)</td>
<td>14/18 (78%)</td>
<td>65/96 (68%)</td>
</tr>
<tr>
<td>Scoliosis ((n = 99))</td>
<td>44/80 (55%)</td>
<td>12/19 (63%)</td>
<td>56/99 (56%)</td>
</tr>
<tr>
<td>Pectus abnormalities ((n = 100))</td>
<td>38/80 (48%)</td>
<td>15/20 (75%)</td>
<td>53/100 (53%)</td>
</tr>
<tr>
<td>Joint laxity ((n = 99))</td>
<td>39/80 (49%)</td>
<td>8/19 (42%)</td>
<td>47/99 (47%)</td>
</tr>
<tr>
<td>Flat foot ((n = 98))</td>
<td>35/79 (44%)</td>
<td>8/19 (42%)</td>
<td>43/98 (44%)</td>
</tr>
<tr>
<td>Limitation of extension of the elbow ((n = 97))</td>
<td>16/78 (20%)</td>
<td>1/19 (5%)</td>
<td>17/97 (17%)</td>
</tr>
<tr>
<td>Camptodactyly ((n = 91))</td>
<td>10/74 (13%)</td>
<td>3/17 (18%)</td>
<td>13/91 (14%)</td>
</tr>
<tr>
<td><strong>Others</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Striae ((n = 98))</td>
<td>13/79 (16%)</td>
<td>5/19 (26%)</td>
<td>18/98 (18%)</td>
</tr>
<tr>
<td>Translucent skin ((n = 88))</td>
<td>2/70 (3%)</td>
<td>2/18 (11%)</td>
<td>4/88 (4%)</td>
</tr>
<tr>
<td><strong>Median systemic score</strong></td>
<td>6.6 ± 2.9</td>
<td>7.3 ± 3.1</td>
<td>6.8 ± 3.0</td>
</tr>
<tr>
<td><strong>Cerebral and cognitive</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Degree of ID ((n = 95))(b)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>• Mild ID</td>
<td>13/76 (17%)</td>
<td>4/19 (21%)</td>
<td>17/95 (18%)</td>
</tr>
<tr>
<td>• Moderate ID</td>
<td>43/76 (56%)</td>
<td>13/19 (68%)</td>
<td>56/95 (59%)</td>
</tr>
<tr>
<td>• Severe ID</td>
<td>20/76 (26%)</td>
<td>2/19 (10%)</td>
<td>22/95 (23%)</td>
</tr>
<tr>
<td>Behavioural abnormalities (including anxiety, hyperactivity, psychotic troubles, and ASD) ((n = 98))</td>
<td>56/79 (71%)</td>
<td>13/19 (68%)</td>
<td>69/98 (70%)</td>
</tr>
<tr>
<td>**Abnormal MRI ((n = 45))(c)</td>
<td>28/77 (36%)</td>
<td>5/18 (28%)</td>
<td>33/95 (35%)</td>
</tr>
<tr>
<td><strong>Molecular and cytogenetic results ((n = 100)</strong></td>
<td>10/40 (25%)</td>
<td>3/6 (50%)</td>
<td>13/45 (29%)</td>
</tr>
<tr>
<td>Submicroscopic chromosome rearrangements</td>
<td>11/80 (14%)(d)</td>
<td>5/20 (25%)(d)</td>
<td>16/100 (16%)</td>
</tr>
<tr>
<td>FBN1</td>
<td>2/80 (2%)(d)</td>
<td>3/20 (15%)(d)</td>
<td>5/100 (5%)</td>
</tr>
<tr>
<td>TGFB1R1, TGFB2</td>
<td>0/80 (0%)</td>
<td>0/20 (0%)</td>
<td>0/100 (0%)</td>
</tr>
<tr>
<td>MED12</td>
<td>1/80 (1%)</td>
<td>0/20 (0%)</td>
<td>1/100 (1%)</td>
</tr>
<tr>
<td>UPF3B, ZDHHC9</td>
<td>0/80 (0%)</td>
<td>0/20 (0%)</td>
<td>0/100 (0%)</td>
</tr>
<tr>
<td>X-inactivation bias in females</td>
<td>• &gt;85% skewing</td>
<td>0/20 (0%)</td>
<td>28/100 (28%)</td>
</tr>
<tr>
<td>• &gt;95% skewing</td>
<td>1/80 (1.2%)(e)</td>
<td>0/20 (0%)</td>
<td>1/100 (1%)</td>
</tr>
<tr>
<td>Total</td>
<td>14/80 (18%)</td>
<td>6/20 (30%)(f)</td>
<td>20/100 (20%)</td>
</tr>
</tbody>
</table>

ASD, autism spectrum disorder; ID, intellectual deficiency; MFS, Marfan syndrome; MH, marfanoid habitus; MRI, magnetic resonance imaging; TAA, thoracic aortic aneurysm.

\(a\)Including TAA, ectopia lentis, sinuous aorta and/or mitral valve prolapse.

\(b\)Twenty-three percent of patients had neuropsychological evaluation.

\(c\)Including absent corpus callosum, thin corpus callosum, corpus callosum dysgenesis, vermis hypoplasia, Arnold Chiari malformation, pituitary stalk interruption syndrome, asymmetric ventricles, cortical atrophy, hydrocephaly, arachnoid cyst, enlarged Virchow Robin spaces, and enlarged cisterna major.

\(d\)One patient had both a FBN1 mutation and a chromosomal rearrangement.

\(e\)Patient P20 not carrying any cytogenetic nor molecular abnormality.
<table>
<thead>
<tr>
<th>Identifier</th>
<th>Nomenclature</th>
<th>Segregation</th>
<th>Number of reports in UMD-FBN1 database</th>
<th>Clinical features</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Group 1</strong></td>
<td></td>
<td></td>
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</tr>
</tbody>
</table>
| 86 | c.8176C>T 64 | Inherited from pauci symptomatic father | 11 | • Male, 21 years, second- and third-degree relatives with ID on both sides  
  • Skeletal features: 168 cm, dolichostenomelia, scoliosis, joint laxity, arachnodactyly, hyperlaxity with recurrent sprains, crowded teeth with irregular shape  
  • Other MFS features: myopia  
  • Moderate ID, epilepsy, abnormal pronunciation, attention deficit disorder, abnormal co-ordination, abnormal behaviour (automutilation and shyness), corpus callosum dysgenesis  
  • Other features: VSD, recurrent ENT infections, anal fistula, right pyelocaliceal dilatation, dysmorphism, cryptorchidism, hyperelastic skin, pescavus, fractures, gingival enlargement |
| 97 | c.4270C>G 34 | Parents not available | 6 | • Male, 18 years, no FH  
  • Skeletal features: 188 cm, asymmetric pectus excavatum, dolichostenomelia, arachnodactyly, joint laxity, severe thoraco-lumbar kyphosis, protrusio acetabulae  
  • Other MFS features: none  
  • Moderate ID  
  • Other features: nasal speech, perceptive deafness |
| **Group 2** | | | | |
| 56 | c.247+1G>A 2_3 | De novo | 13 | • Female, 21 years, no FH  
  • Skeletal features: 188 cm, long and thin habitus, dolichostenomelia, arachnodactyly, pectus excavatum, scoliosis, hyperlaxity, flat feet, typical dysmorphism, high arched palate. Surgery for metatarsus varus at age 2  
  • Other MFS features: TAA diagnosed at age 5 years, mitral valve prolapse with mitral insufficiency that required surgery at age 4 years, ectopia lentis, striae, dural ectasia  
  • Moderate ID with Chiari malformation  
  • Other features: nasal speech, perceptive deafness |
| 83a | c.2534A>G 20 | Father not available | 0 | • Male, 54 years, father unknown  
  • Skeletal features: 180 cm, scoliosis, pectus excavatum, arachnodactyly, high-arched palate with dental malposition  
  • Other MFS features: lens dislocation that required surgery, mitral valve prolapse  
  • Mild ID  
  • Other features: hypertrophic cardiomyopathy  
  • ASD, ventricular septal defect; ID, intellectual disability; ENT, ear, nose and throat; FH, family history; MFS, Marfan syndrome; TAA, thoracic aortic aneurysm; VSD, ventricular septal defect. |
| 96 | c.4270C>G 34 | Parents not available | 6 | • Male, 36 years, no FH  
  • Skeletal features: long and thin habitus, pectus excavatum, dolichostenomelia, arachnodactyly  
  • Other MFS features: severe mitral valve prolapse with massive mitral insufficiency requiring surgery at age 36 years, myopia  
  • Moderate ID, epilepsy  
  • Other features: ASD, vascular embolization of two intracranial sylvian aneurysms, dolicohcephaly, plagiocephaly, down slanting palpebral fissures, small and round ears |

ASD, ventricular septal defect; ID, intellectual disability; ENT, ear, nose and throat; FH, family history; MFS, Marfan syndrome; TAA, thoracic aortic aneurysm; VSD, ventricular septal defect.
aThe patients also carries a submicroscopic chromosomal rearrangement that was already reported in the literature for patient 86 (61) EVS, exome variant server: http://evs.gs.washington.edu/EVS/.
Table 3. Possibly pathogenic MED12 mutation obtained from direct sequencing in the cohort of interest

<table>
<thead>
<tr>
<th>Identifier</th>
<th>Nomenclature</th>
<th>Segregation</th>
<th>Clinical features</th>
</tr>
</thead>
<tbody>
<tr>
<td>68</td>
<td>c.3884G&gt;A, (p.Arg1295His)* exon 28</td>
<td>Inherited from his mother with mild ID, and not found in the two asymptomatic brothers</td>
<td>• Male, 18 years, mild phenotype in her mother (skeletal features, mitral valve prolapse) • Skeletal features: 187 cm, long and thin habitus, pectus excavatum, joint laxity, malar hypoplasia, down slanting palpebral fissures, teeth malposition • Other MFS features: none • Mild ID • Other features: small and low-set ears, nasal speech, bilateral ptosis</td>
</tr>
</tbody>
</table>

TGFBRI and TGFBRII sequencing analysis

Neither pathogenic mutations nor known variants were found in the TGFBRI gene. One missense and three synonymous variants were found in the coding sequence of the TGFBRII gene. For three of them, their presence in an asymptomatic parent combined with database predictions were in favour of a polymorphism (Table S2). The c.1039C>T (p.Leu347Leu) variant was found to be de novo. On the basis of the prediction of a potential ESE disruption by the Human Splicing Finder tool (51), mRNA studies on lymphocytes were performed but the results were not in favour of abnormal splicing.

MED12, UPF3B, and ZDHHC9 sequencing analysis

One possibly pathogenic MED12 mutation was found (c.3884G>A, p.Arg1295His) in patient 68 (Table 3). The use of different databases gave opposite results, databases against the pathogenicity, taking into account the fact that a histidine is present at this position in another gene of the MED family. However, this amino acid is highly conserved across species (Fig. S1), and was not found in 200 male controls tested in the laboratory, or in the 1000 Genomes Project Consortium or in the EVS database (http://evs.gs.washington.edu/EVS/). Segregation in the family showed that the variant was inherited from the mother, who had a milder phenotype, which could be compatible with an X-linked inheritance, and was absent in the two healthy brothers. Additional functional studies should be performed in order to arrive at a definitive conclusion.

Table S2 summarizes the likely non-pathogenic variants found in this cohort. The c.3692-22A>C MED12 variant was predicted to break the potential branch point (tccctAt, -22 position in the intron) according to the in silico splice site analysis program Human Splicing Finder (HSF) (51), but this hypothesis could not be confirmed by normal mRNA studies.

X-inactivation studies

An X-inactivation bias >85% was found in seven females of group 1 (7/20, 35%), but absent in both other groups. When considering X-inactivation bias >95%, only one female was concerned.

Targeted array-CGH

Among the 100 patients, 84 displayed normal results, with no copy number variants (CNVs) or only the presence of benign CNVs. For 16 patients, genomic imbalances were detected ranging from 500 kb to 11.6 Mb, (i) ten deletions, including one deletion 15q21.1q21.3 encompassing the FBN1 gene; (ii) five duplications; and (iii) one patient with an unbalanced translocation (Table 4). The overviews of copy number changes of all patients along the whole genome are shown in Fig. 1. Two patients had both chromosomal rearrangements and a pathogenic FBN1 mutation. Five patients had a known microdeletional syndrome (3q29, 15q13.2q13.3 and 17q12 microdeletions, 15q11.2 and 16p11.2 microduplications).

Discussion

The systematic screening of genes that may be involved in MH and ID in this study provides useful information regarding the work-up necessary in such patients. From a study of 100 patients, we showed that: (i) submicroscopic rearrangements are the most prevalent abnormalities, (ii) the presence of ID should not rule out the possibility of an FBN1 mutation; (iii) the association of two pathogenic abnormalities was possible, which means that extreme caution must be exercised when considering relating the overall phenotype to a given anomaly, unless additional data become available in the literature; (iv) the X-linked MED12, ZDHHC9 and UPF3B genes are not major genes for MH and ID, although a skewed sex ratio in favour of males was found.

Submicroscopic rearrangements were identified in 16/100 patients. This percentage is in the range of the overall detection rate of genomic imbalances by array-CGH in patients with ID and/or developmental delay (52). Retrospectively, the use of targeted custom oligonucleotide array-CGH did not permit the identification of intragenic rearrangement in the genes of interest. The chromosomal imbalances identified were all different, strengthening the hypothesis that high genetic heterogeneity exists in this phenotype. For each abnormality, we tried to determine if there was evidence that the rearrangement might explain the MH, and
Table 4. Submicroscopic chromosomal rearrangements detected by array-CGH in the cohort of patients with marfanoid habitus (MH) and mental retardation according to the group concerned (in grey, rearrangement that has already been described in the literature in association with a MH)

<table>
<thead>
<tr>
<th>Identifier</th>
<th>Copy number change</th>
<th>Chromosomal location</th>
<th>Size of rearrangement (Mb)</th>
<th>Sequences co-ordinates (hg18)</th>
<th>Inheritance</th>
<th>Clinical features</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>31</td>
<td>Loss</td>
<td>2q24.1q24.2</td>
<td>6.44</td>
<td>155397362–161841920</td>
<td>De novo</td>
<td>Male, 15 years, FH of ID in maternal uncle • Skeletal features: 167 cm, long and thin habitus, arachnodactyly, dolichostenomelia, hyperlaxity, high arched palate, coxovalga • Other features of MFS: none • Severe ID, hyperactivity • Other features: facial dysmorphism, strabismus</td>
<td>(62, 63)</td>
</tr>
<tr>
<td>103</td>
<td>Loss</td>
<td>3q23.3</td>
<td>1.5</td>
<td>137357633–138147814</td>
<td>De novo</td>
<td>Male, 15 years, no FH • Skeletal features: size − 1 SD, long and thin habitus, pectus excavatum, scoliosis, flat feet • Other features of MFS: none • Mild ID (IQ 66), autistic features • Other features: mild myopia</td>
<td></td>
</tr>
<tr>
<td>52</td>
<td>Loss</td>
<td>3q29.1a</td>
<td>1.42</td>
<td>197216153–198643540</td>
<td>De novo</td>
<td>Male, 16 years, no FH • Skeletal features: 172 cm, long and thin habitus, dolichostenomelia, arachnodactyly, pectus excavatum, scoliosis, flat feet, high arched palate • Other features of MFS: pneumothorax • Mild ID (IQ 54), anxiety, psychiatric manifestations with auditory hallucinations, dysarthria • Other features: mild camptodactyly</td>
<td>(53, 54)</td>
</tr>
<tr>
<td>101</td>
<td>Loss</td>
<td>5p15.31p15.2a</td>
<td>1.5</td>
<td>8824218–10286748</td>
<td>De novo</td>
<td>Female, 18 years, no FH • Skeletal features: 180 cm, long and thin habitus, dolichostenomelia, arachnodactyly, incomplete extension of the elbows, facial dysmorphism • Other features of MFS: striae • Mild ID, severe epilepsy • Other features: none</td>
<td>(33, 34)</td>
</tr>
<tr>
<td>13</td>
<td>Loss</td>
<td>6q27</td>
<td>2.42</td>
<td>168323802–170748862</td>
<td>Not known (father dead)</td>
<td>Male, 14 years, no FH • Skeletal features: 167.5 cm, long and thin habitus, dolichostenomelia, arachnodactyly, pectus excavatum, scoliosis, hyperlaxity, facial dysmorphism • Other features of MFS: none • Moderate ID (IQ 48), hyperactivity, anxiety, autistic traits, mild ventricular dilatation • Other features: microcephaly, apparent blood vessels</td>
<td>(64)</td>
</tr>
<tr>
<td>Identifier</td>
<td>Copy number change</td>
<td>Chromosomal location</td>
<td>Size of rearrangement (Mb)</td>
<td>Sequences co-ordinates (hg18)</td>
<td>Inheritance</td>
<td>Clinical features</td>
<td>References</td>
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<tr>
<td>102&lt;sup&gt;D&lt;/sup&gt;</td>
<td>Gain 9q33.3q34.3&lt;sup&gt;a&lt;/sup&gt;</td>
<td>11.66</td>
<td>128531258–140193874</td>
<td>Not verified</td>
<td>Male, 20 years, younger brother with the same appearance at birth who died of a cardiopathy at 2 years of age • Skeletal features: 169 cm, long and thin habitus, dolichostenomelia, limited extension of the elbows, scoliosis, high arched palate • Other features of MFS: none • Severe ID (IQ 35), sociable, jovial, very affectionate, nervous • Other features: mitral regurgitation, hypernasal voice, malrotation of the right kidney, epilepsy, delayed puberty, dysmorphism, hypodontia, hammer-shaped of the second toe, previously by Mégarbané et Chammas, (1997)</td>
<td></td>
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</tr>
<tr>
<td>47</td>
<td>Loss 11q23.1q23.2</td>
<td>5.18</td>
<td>110122508–115306206</td>
<td>De novo</td>
<td>Male, 18 years, no FH • Skeletal features: 190.5 cm, long and thin habitus, arachnodactyly, abnormal pectus, scoliosis, hyperlaxity, flat feet, high arched palate • Other features of MFS: mild myopia • Mild ID, hyperactivity, epilepsy • Other features: mild camptodactyly, mild dysmorphism, nasal speech, stammering and dysarthria</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>Gain 12q13.3q14.4 (at the breakpoint of a de novo translocation 46,XY,t(12;19)(q13.3;p12)</td>
<td>0.701</td>
<td>56177083–56878427</td>
<td>De novo</td>
<td>Male, 14 years, no FH • Skeletal features: &gt;97th centile, long and thin habitus, arachnodactyly, asymmetric pectus excavatum, scoliosis, high-arched palate with crowded teeth, facial dysmorphism • Other features of MFS: surgery for inguinal herniae, ascending aorta upper the normal range at the level of the sinus of Valsalva, redundant mitral valve with mitral insufficiency • Mild ID, schizophrenia • Other features: nasal speech</td>
<td>(66)</td>
<td></td>
</tr>
<tr>
<td>99</td>
<td>Gain 15q11.2&lt;sup&gt;a&lt;/sup&gt;</td>
<td>5.2</td>
<td>20811696–26138064</td>
<td>De novo</td>
<td>Male, 16 years, no FH • Skeletal features: 181 cm, long and thin habitus, dolichostenomelia, arachnodactyly, scoliosis • Other features of MFS: none • Mild ID, schizophrenia • Other features: hypodontia</td>
<td>(58)</td>
<td></td>
</tr>
</tbody>
</table>
### Table 4. Continued

<table>
<thead>
<tr>
<th>Identifier</th>
<th>Copy number change</th>
<th>Chromosomal location</th>
<th>Size of rearrangement (Mb)</th>
<th>Sequences co-ordinates (hg18)</th>
<th>Inheritance</th>
<th>Clinical features</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>6</td>
<td>Gain</td>
<td>16p11.2a</td>
<td>1.70</td>
<td>28399426–30107008</td>
<td>De novo</td>
<td>Male, 23 years, FH of 2 maternal cousins with autistic behaviour</td>
<td>(56)</td>
</tr>
<tr>
<td></td>
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<td></td>
<td></td>
<td></td>
<td></td>
<td>• Skeletal features: 193 cm, long and thin habitus, arachnodactyly, scoliosis, asymmetric pectus excavatum, flat feet, and high arched palate</td>
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<td></td>
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<td></td>
<td></td>
<td>• Other features of MFS: none</td>
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<td></td>
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<td></td>
<td></td>
<td></td>
<td>• Mild ID, obsessive compulsive behaviour, anxiety, autistic features and psychosis episodes</td>
<td></td>
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<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>• Other features: mild dysmorphic features</td>
<td></td>
</tr>
<tr>
<td>100</td>
<td>Loss</td>
<td>17q12a</td>
<td>1.3</td>
<td>31891335–33242358</td>
<td>Not verified</td>
<td>Female, 25 years, FH of ID in her mother and maternal uncle</td>
<td>(60)</td>
</tr>
<tr>
<td></td>
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<td></td>
<td></td>
<td></td>
<td>• Skeletal features: 193 cm, long and thin habitus, arachnodactyly, multiple surgery for severe scoliosis</td>
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<td></td>
<td>• Other features of MFS: none</td>
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<td></td>
<td></td>
<td></td>
<td>• Mild ID, psychotic behaviour</td>
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<td></td>
<td></td>
<td></td>
<td>• Other features: interventricular septal aneurysm with abnormal cardiac rhythm, recurrent fractures of the fingers with osteoporosis, facial dysmorphism (round face with hypertelorism), strabismus, pescavum, thenar and hypotenar hypoplasia</td>
<td></td>
</tr>
<tr>
<td>86c</td>
<td>Loss</td>
<td>17q21.31a</td>
<td>0.504</td>
<td>41062469–41566740</td>
<td>De novo</td>
<td>Male, 20 years, no FH</td>
<td>(61)</td>
</tr>
<tr>
<td>69</td>
<td>Gain</td>
<td>19p13.3</td>
<td>1.66</td>
<td>4899227–6563324</td>
<td>De novo</td>
<td>Male, 23 years, brother with mild school difficulties</td>
<td>(67)</td>
</tr>
<tr>
<td></td>
<td></td>
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<td></td>
<td></td>
<td></td>
<td>• Skeletal features: 187 cm, dolichostenomelia, hyperlaxity, limited elbow extension, kyphosis, high arched palate and facial dysmorphism</td>
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<td></td>
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<td></td>
<td></td>
<td>• Other features of MFS: none</td>
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<td></td>
<td></td>
<td></td>
<td>• Mild ID, abnormal behaviour including anxiety, hypersomnia and obsessive compulsive behaviour necessitating treatment</td>
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<td></td>
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<td></td>
<td></td>
<td></td>
<td></td>
<td>• Other features: chiari I malformation and supra tentorial white matter hypersignals</td>
<td></td>
</tr>
<tr>
<td>Group 2</td>
<td></td>
<td></td>
<td></td>
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</tr>
<tr>
<td>83h</td>
<td>Gain</td>
<td>2q37.3a</td>
<td>4.51</td>
<td>238189799–242701179</td>
<td>Unknown</td>
<td>Male, 23 years, brother with mild school difficulties</td>
<td>(40, 59)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>238189799–242701179</td>
<td></td>
<td>• Skeletal features: 194 cm, long and thin habitus, scoliosis, hyperlaxity, facial dysmorphism</td>
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<td></td>
<td></td>
<td></td>
<td>183993225–191028016</td>
<td></td>
<td>• Other features of MFS: TAA and bicuspid aortic valve</td>
<td></td>
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<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>183993225–191028016</td>
<td></td>
<td>• Mild ID, sleep disturbances with anxiety</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>28718936–30298296</td>
<td>De novo</td>
<td>• Other features: hypermetropia</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Loss</td>
<td>4q35.1q35.2</td>
<td>7.03</td>
<td>183993225–191028016</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>39</td>
<td>Loss</td>
<td>15q13.2q13.3a</td>
<td>1.57</td>
<td>28718936–30298296</td>
<td>De novo</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
if the results could provide some information regarding the genes responsible for MH. In our series, the presence of a marfanoid phenotype was recurrent in 4/16 rearrangements, including the 3q29 microdeletion (53, 54), the 5p15.31p15.2 microdeletion (33, 34), the 15q21.1q21.3 microdeletion (55) and the 16p11.2 microduplication (56) (Table 4). Except for the FBN1 gene located in the 15q21.1q21.3 microdeletion, none of the other rearrangements comprised a good candidate gene for the MH. The recurrent de novo 3q29 microdeletion had a commonly deleted region of 1.6 Mb, most likely secondary to non-allelic homologous recombination. Besides mild/moderate ID and a long, narrow face, chest-wall deformity has been described in 3/15 patients, long, tapering fingers in 3/15 patients, and ligamentous hyperlaxity in 1/15 patient (53, 54).

Of note, our patient had a more generalized MH and pneumothorax, which can be part of the MFS spectrum. Some patients with the 5pter microdeletion and with various marfanoid features in adolescence have already been described (33, 34). Unfortunately, previous patients were not studied using high resolution techniques. It was therefore not possible to determine the smallest region of overlap. Finally, it is highly probable that the phenotype found in the patient with chromosome 16p11.2 duplication could be fully explained by the cytogenetic abnormality as it has been recently shown that it was the countertype of patients presenting with chromosome 16p11.2 microdeletion with obesity and a wide range of behavioural abnormalities (57).

Conversely, we have data to conclude that the rearrangements in five patients were certainly responsible for ID but cannot explain the MH. Four of them are well-known frequent genomic rearrangements, comprising the 15q11.2 microduplication including UBE3A/SNRPN (58), the 15q13.3 microdeletion including CHRNA7 (59), the 17q12 microdeletion (60) and finally the 17q21.31 microdeletion including MAPT (61), and manifestations of MFS have never been described in large series. The best argument against the causality of the 17q21.31 microdeletion in MH is that we identified in the same patient a concomitant pathogenic FBN1 mutation. Similar conclusions can be drawn for the patient with an unbalanced translocation responsible for a 2q37.3 duplication and a 4q35.1q35.2 deletion, since the patient also carried a pathogenic FBN1 mutation. These observations especially emphasize the importance of a systematic screening approach for accurate genetic counselling and clinical follow-up. Unfortunately, no conclusion can be drawn for the other rearrangements.

The second take home message from this study is that the presence of ID should not rule out the diagnosis of MFS with its aortic risk, in a patient with MH. Indeed, 6% of our patients had a FBN1 mutation or a rearrangement involving FBN1, i.e. 35% of the abnormalities found in the study. This conclusion is of importance as approximately a third of the patients were not screened by echocardiography prior to the study. When an FBN1 mutation was found, we have data showing that ID could be attributable to a different
cause, since two of the patients also had a chromosomal micro-rearrangement that explained the ID. Also, a large study of patients with MFS did not reveal an increased risk of ID (7). Given the very high allelic heterogeneity of the \textit{FBN1} gene, it was surprising to find the same mutation (c.4270C>G, p.Pro1424Ala) in two unrelated patients of the study. However, six additional patients with the same mutation and with no ID were reported in the UMD-\textit{FBN1} database. The majority of \textit{FBN1} mutations/rearrangements was found in patients with TAA, ectopia lentis and/or mitral valve prolapse (four out of five were from group 2), but only skeletal manifestations were diagnosed at 22 years of age (patient 97, Table 2), confirming the high clinical variability of patients with \textit{FBN1} mutations, who can present with isolated skeletal features (47).

By combining all these data it was possible to create a flow chart (Fig. 2) with the following recommendation:

\begin{itemize}
\item 1.
\item 2.
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\item 10.
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\item 35.
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\item 37.
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\item 40.
\item 41.
\item 42.
\item 43.
\item 44.
\item 45.
\item 46.
\item 47.
\end{itemize}

}\begin{figure}
\centering
\includegraphics[width=\textwidth]{chart.png}
\caption{Flow chart for the diagnosis of patients with MH and ID. TAA, thoracic aortic aneurysm. *The list of genes to be tested will lengthen as new syndromic TAA genes are identified.}
\end{figure}
echocardiography and eye examination should be systematically performed in patients with MH and ID. If TAA or ectopia lentis is found, FBN1 screening is warranted for genetic counselling and follow-up. The systemic score in patients with an FBN1 mutation did not appear to be significantly different from that in other patients of the cohort. If the echocardiography and eye examination are normal, they should be performed at least every 5 years, to take into account the features that evolve with time.

The third answer from this study is that the three X-linked genes found in association with LFS are not major genes in this clinical population, at least in sporadic patients. Indeed, only one probably pathogenic MED12 mutation (c.3884G>A, p.Arg1295His) was identified in our cohort of 100 patients. However, very few familial cases were included, and we therefore cannot draw any conclusions about the frequency of MED12 involvement in cases with X-linked inheritance. Nevertheless, the role of other X-linked genes appears probable because there was an obvious skewed sex ratio in favour of males in our cohort of patients.

Finally, no molecular or cytogenetic abnormalities were found in 80% of the patients (82% in group 1 and 70% in group 2). Chromosomal micro-rearrangements found in this study could point to new candidate regions to be studied to identify new genes responsible for the phenotype. Further studies with next-generation sequencing technology such as the exome approach using a trio strategy will hopefully help in the diagnosis of such patients. In particular, a distinct entity which associates TAA and intellectual deficiency may exist because no FBN1 mutation was found in the majority of the patients. Besides to the hypothesis that more loci with genes are yet to be discovered, an alternative explanation would be that MH is a relatively non-specific feature of patient with ID. Future studies on the subject would be useful to know if a diagnostician should give importance or not on the marfanoid findings in association to ID, given that it can be a common genetic problem in the population.

In conclusion, this collaborative work suggests a practical diagnostic pathway resulting in a better clinical differentiation, and providing a basis for more effective management and appropriate genetic counselling for families with MH and ID.

**Supporting Information**

The following Supporting information is available for this article:

Fig S1. Interspecies conservation of the amino acid concerned by the MED12 missense mutation in patient 68.

Table S1. Candidate genes with numbers of clones added to the custom array-CGH design.

Table S2. Non-pathogenic variant obtained from direct sequencing of the FBN1, TGFBR1, TGFBR2, MED12, UPF3B and ZDHHC9 in the cohort of interest.

Additional Supporting information may be found in the online version of this article.

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**References**


