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Dioxin-like, non-dioxin like PCB and PCDD/F contamination in European eel (*Anguilla anguilla*) from the Loire estuarine continuum: spatial and biological variabilities

I. Blanchet Letrouvé, A. Zalouk Vergnoux, A. Vénisseau, M. Couderc, B. Le Bizec, Pierre Elie, C. Herrenknecht, Catherine Mouneyrac, L. Poirier

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26 15 Abstract

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29 16 To characterize the eel contamination by dioxin-like (dl) and non dioxin-like (ndl) PCBs and
30 17 PCDD/Fs, 62 eels from the Loire estuary (France) were analyzed. PCB contamination
31 18 significantly increased from glass eel stage (3.7 ± 1.9 and 15.2 ± 4.2 ng.g⁻¹ dw) to other life
32 19 stages (for yellow eels: 62.8 ± 34.4 and 381.8 ± 181.8 ng.g⁻¹ dw; for silver eels: 93.7 ± 56.3 and
33 20 463.2 ± 244.6 ng.g⁻¹ dw respectively for dl and ndl PCB). An inter-site variability based on
34 21 PCB levels and fingerprints was observed between the three studied sites. The glass eel
35 22 pattern was mainly characterized by the less chlorinated PCBs contrarily to the other eels,
36 23 underlying a different bioaccumulation pathway. Overall, eels from this estuary showed an
37 24 intermediate contamination level compared to other international/national areas. However,
38 25 more than 60% of studied silver eels displayed WHO₂₀₀₅ PCDD/F and dl-PCB TEQ values
39 26 higher than the recommended level of 10 pg.g⁻¹ ww. This statement indicates a potential
40 27 exposure to PCBs through eel consumption, especially with silver individuals, and could
41 28 potentially lead to damages for the eel population.
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30 1. Introduction

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2 31 Since the 1980s, monitoring studies in European countries have shown the decline of glass
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4 32 eels arriving in the coastal waters and estuaries (ICES, 2006). The disappearance of the
5
6 33 prepubertal European eel (*Anguilla anguilla*) occurred as well a few decades earlier and
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8 34 stocks were estimated to be divided by ten (Dekker, 2003; Moriarty and Dekker, 1997).
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10 35 Several factors were brought forward to explain this decrease such as overfishing, obstacles to
11
12 36 migration (Robinet and Feunteun, 2002), pathogens (Palstra et al., 2007b), climate change
13
14 37 (Castonguay et al., 1994) and contaminants (Geeraerts et al., 2011; Palstra et al., 2007a;
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16 38 Roosens et al., 2010; van Ginneken et al., 2009)

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18 39 Among these different causes, polychlorinated biphenyls (PCBs), polychlorinated dibenzo-p-
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20 40 dioxins and furans (PCDD/Fs) seem to be particularly suspected because of their potential as
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22 41 estrogenic and anti-estrogenic disruptors (Canapa et al., 2002) and their neuroendocrine
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24 42 effects (Kodavanti and Curras-Collazo, 2010), endangering several fish species and notably
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26 43 the eel population (van Ginneken et al., 2009). PCBs represent a particularly persistent
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28 44 chlorinated chemical group of 209 congeners, ubiquitous in the environment and from
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30 45 anthropological origin exclusively. Two classes of PCBs were distinguished according to their
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32 46 toxicological properties: the dioxin-like PCBs (dl-PCBs) which present analogous toxicity as
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34 47 dioxin compounds and the non dioxin-like PCBs (ndl-PCBs) (European Union, 2011). These
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36 48 classes were related to chemical structures such as the number and chlorine positions. Due to
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38 49 their chemical stability, insulating and fire retardant properties, PCBs were used in the
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40 50 manufacturing of electrical equipment, heat exchangers, hydraulic systems, and several other
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42 51 specialized applications. In spite of the ban on their production during the eighties, the
43
44 52 accumulated production all over the world was estimated at 1,200,000 tons and approximately
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46 53 30 % of this production is scattered in the environment, essentially in the oceanic environment
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48 54 (Voltura and French, 2000). The contamination of aquatic organisms depends on the chemical
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50 55 properties of each congener. The exposure level in the environment and various biotic factors
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52 56 such as the metabolic capacity influence the bioaccumulation processes (Hubaux and
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54 57 Perceval, 2011).

53
54 58 Considered as a bottom dwelling fish, showing a high body lipid content, an important
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56 59 longevity and a carnivorous status, the European eel is extremely exposed to lipophilic
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58 60 persistent contaminants, such as PCBs, and represents a species sensitive to bioaccumulation
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60 61 (Roche et al., 2000). Moreover, eels constitute an important economic value nearby estuaries

62 and rivers and a significant food resource (Perraudeau and Després, 2009). Significant levels
63 of PCBs were detected in European eels from the Gironde and Adour estuary (France) (Tapie
64 et al., 2011), in the Mondego estuary (Portugal) (Nunes et al., 2011), in the rivers of Italy
65 (Mezzetta et al., 2011) and could be responsible for migration or reproduction impairments
66 (van Ginneken et al., 2009). Assessing PCB contamination of the European eel is therefore of
67 great interest since their level is threatening public health, beyond a maximal value (European
68 Union, 2011) and is also a potential risk for its own health (for review, Geeraerts and
69 Belpaire, 2010). The present study aims to assess PCB contamination of the European eel
70 from the Loire estuary (France) which the basin (117,800 km²) drains a lot of tributaries.
71 Moreover, the Loire estuary runs through important urban sites (Nantes, Saint-Nazaire) with
72 shipping, industrial and agricultural activities. It displays a diffusive pollution including a
73 mixture of contaminants such as heavy metals (Grobois et al., 2012), pesticides (Marchand et
74 al., 2004), PAHs and PCBs (Hubaux and Perceval, 2011). For European eels, this estuary
75 constitutes one of the most important continental migration path of glass eels. The
76 preservation of its chemical quality is therefore essential for eel health. However, a real lack
77 of data on the POPs contamination levels of European eels exists in this ecosystem. Only few
78 individuals, sampling on the whole Loire river, have been analyzed in the French PCB
79 framework (ONEMA, 2012). These results cannot be sufficiently representative of eels living
80 in the estuary. In the present study, dl-PCB, ndl-PCB and PCDD/F levels were investigated in
81 European eels fished in the Loire estuary. This work was set out to reach three objectives :
82 i) to get a representative trend of PCB contaminants over life stages, from glass eels to silver
83 eels; ii) to assess spatial PCB contamination variations on yellow individuals (similar size
84 class distributions), along three different Loire estuary sites (Fig. 1), iii) to evaluate health
85 risks for local consumers with PCDD/F and dl-PCB TEQs quantification according to WHO
86 recommendations (van den Berg et al., 2006).

88 2. Material and methods

89 2.1. Sampling sites

90 As shown in Fig.1, three sampling sites were selected in this study. Varades is a small city
91 (about 3550 locals), located upstream in the estuary at the limit of the salinity (100 km from
92 the Loire mouth); it also presents few industrial activities and is particularly under agricultural
93 pressure. The intermediate site is close to an important city, Nantes (about 600,000 locals)

94 located at 50 km from the mouth, characterized by an industrial harbor and an urban zone
95 including two incineration factories. The third site, Cordemais, is downstream of Nantes with
96 a strong influence of the North Atlantic Ocean and is well-known for its industrial activities,
97 particularly the presence of a coal-fired power plant and closed to an industrial complex
98 including oil refineries. These three sampling sites were chosen in order to represent the
99 estuary displaying different kinds of human activities.

100 2.2. Sampled animals

101 During one year and a half, *i.e.* from May 2009 to January 2011, European eels were
102 captured by local fishermen according to the fishing authorizations, in the three sampling sites
103 described above. Using specific methods, 62 yellow and silver eels were collected with fyke
104 and stow nets respectively. The aim of the sampling procedure was to evaluate the potential
105 spatial variability of contaminant levels in eels on these 3 different sites, to upstream from
106 downstream. Consequently, 16 yellow eels were captured in Varades, 16 in Nantes and 17 in
107 Cordemais. The captured eels were preferentially selected in order to obtain a similar size
108 class distribution, *i.e.* about 4 to 5 eels per size class and par site. To evaluate the trend in
109 contaminant level over life stage, glass eels and 13 silver eels were also captured. Individuals
110 were transported to the laboratory in aerated 200 L tanks filled with water from the sampling
111 site. They were maintained in the laboratory few hours until dissection under a natural
112 photoperiod (L15/D9) and at a temperature around 12 ± 2 °C, equivalent to the fishing site
113 conditions. Glass eels were collected with a specific fishing net (authorized mesh size) in
114 January 2011 in the estuary entry, near Cordemais. These glass eels had no pigment and
115 corresponded to a stage before the onset of the feeding (Elie et al., 1982). They were directly
116 frozen at -20°C in aluminum foil after fishing and later divided into two different pools.

117 118 2.3. Biometric parameters and life stages of the biological samples

119 Eels were anesthetized in a water bath of 10 L added with 1.5 to 2 mL of clove oil solution
120 dissolved in ethanol (70%), according to the weight of eels (Palstra et al., 2007a). Once
121 anesthetized, the body length (BL in mm) and the body weight (BW in g) of each European
122 eel were measured. The animals were then sacrificed, skinned and dissected in order to collect
123 filets and otoliths. Biometric parameters were recorded to evaluate the Fulton's condition

124 factor ($K = (BW \times 10^5) / BL^3$ with BW and BL respectively expressed as g and mm) (Fulton,
1 125 1904).

126 The otoliths were utilized to determine the age of the organisms. The pair of otoliths named
2 127 sagitta were removed from the eel's head. After extraction, otoliths were cleaned of all organic
3
4 128 membranes in distilled water, dried with ethanol, and then stored in Eppendorf tubes. The
5
6 129 otoliths were later embedded in synthetic resin (Synolithe), and then polished to the nucleus
7
8 130 with a polishing wheel (Streuers Rotopol-35) using two different grits of sandpaper (1200 and
9
10 131 2400). Fine polishing was done by hand with alumina (1 μ m grain) on a polishing cloth.
11
12 132 Etching was done using 10% EDTA. A drop of this solution was applied on the mold during
13
14 133 fifteen minutes. Then, the otoliths were rinsed with distilled water and stored in dry condition.
15
16 134 Yearly increments were revealed by staining with a drop of 5% Toluidine blue on the otolith
17
18 135 and letting it dry. Growth rings were then counted under binocular magnifier. The age of each
19
20 136 eel was determined by the number of increments starting from the nucleus, which was
21
22 137 considered as year 1 of the eel's life. The otolithometry was realized in partnership with the
23
24 138 IRSTEA (Cestas, France). Silver stage was determined by macroscopic characteristics such as
25
26 139 the differentiated lateral line (presence of black corpuscles), a contrasting skin color (dark
27
28 140 dorsal surface and a white ventral surface), the ocular diameter and the pectoral fin length.
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33 142 2.4. PCB and PCDD/F analysis

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35 143 Eel filets and pools of glass eels were analyzed for 18 PCBs (n= 62 and 2 pools of glass eels).
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37 144 Among them, 12 are dl-PCBs (#77; 81; 105; 114; 118; 123; 126; 156; 157; 167; 169; 189) and
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39 145 6 are ndl-PCBs (#28; 52; 101; 138; 153; 180). Ndl-PCB and dl-PCB levels in eel filets and
40
41 146 pools of glass eels were expressed as a sum of all congeners. In order to assess a potential
42
43 147 health risk, PCDD/F analyses were achieved on 11 out of 62 eels (5 yellow and 6 silver
44
45 148 individuals) and on the 2 pools of glass eels. The PCDD/Fs analyzed were the 17 congeners
46
47 149 regulated by the European Union (EC/1259/2011). PCB and PCDD/F levels were expressed
48
49 150 by congeners or as a sum of all congeners in ng.g⁻¹ dry, lipid or wet weight (dw, lw or ww).
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52 151 2.4.1 Reagents and Chemicals

53
54 152 All organic solvents (Promochem) were Picograde[®] quality. Silica (Fluka), sodium sulfate
55
56 153 (Merck), and sulfuric acid (SDS) were of superior analytical quality. Native and ¹³C-labeled
57
58 154 standards were purchased from Cambridge Isotope Laboratories (CIL) and Wellington
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60

155 Laboratory. Standard solutions were prepared in toluene. All reference solutions were stored
156 in darkness at a temperature < 6°C.

157 2.4.2 Sample preparation procedure

158 Eel filets and pools of glass eels were homogenized, weighed and freeze-dried. Five grams of
159 filets and pools of glass eels were cut, dehydrated, and milled using a turbo-mixer with glass
160 bowl. Each experiment was realized with disposable material. Then, samples were powdered
161 and transferred into cells in order to be extracted by Accelerated Solvent Extraction (ASE)
162 using a Dionex ASE 300. Before extraction, eighteen ¹³C-labelled PCB congeners were added
163 to the samples for internal standard calibration and quantification by the isotope dilution
164 method. Pressure and temperature were set to 100 bars and 120°C respectively. The extraction
165 solvent was a mixture of toluene/acetone 70:30 (v/v), and three successive extraction cycles
166 (5 min each) were performed. The extract was evaporated to dryness by rotary evaporation
167 (40°C), allowing the gravimetric determination of the fat content, in order to assess the filet
168 lipid weight (LW in % of wet weight). The extracts were dissolved in 25 mL of hexane for
169 sample clean-up.

170 Three purification steps were then performed, using successively acid silica, Florisil[®] and
171 celite/carbon columns. After removal of fat on the first silica gel column activated with
172 sulfuric acid, PCBs were separated from PCDDs/PCDFs on the second Florisil[®] column. The
173 separation of coplanar (non-ortho) PCBs from non coplanar PCBs was achieved on an
174 activated mixture of Florisil[®]/ Carbopack C/Celite 545 (overnight at 130°C). After the
175 addition of external standards for the recovery calculation (¹³C₁₂-PCB #111 for PCBs), final
176 sample extracts were evaporated under a nitrogen stream to dryness and reconstituted in 20
177 μL, 50 μL and 10 μL of toluene for coplanar PCBs, non coplanar PCBs and PCDD/Fs
178 respectively.

179 2.4.3 GC-HRMS measurement

180 PCB and PCDD/F measurements were performed by gas chromatography coupled to high
181 resolution mass spectrometry (GC-HRMS) using an 7890A gas chromatograph (Agilent)
182 coupled to a JMS 700D or a JMS 800D double electromagnetic sector high resolution mass
183 spectrometer (Jeol, Tokyo, Japan). A DB5MS (30 m x 0.25 mm x 0.25 μm) capillary column
184 (J&W) was used in the splitless mode. The GC program for PCBs was 120°C (3 min),
185 20°C/min to 170°C (0 min), 3°C/min to 245 °C (0 min) and finally 20°C/min to 275°C

186 (7 min). Ionization was achieved in the electron ionization mode (42 eV electron energy). The
187 spectrometric resolution was set at 10,000 (10% valley), and the signal acquisition was
188 performed in the Single Ion Monitoring (SIM) mode focusing on the two most abundant
189 signals from each target molecular ion (^{35}Cl and ^{37}Cl isotopic contributions). Signals were
190 integrated by JEOL Diok software (v.4). The detection and quantification limits (LOD and
191 LOQ respectively) are calculated by JEOL Diok software according to the regulation for
192 dioxin compounds analysis (LOD=LOQ at Signal/Noise=3). A LOD is calculated for each
193 congener and each sample (according to the sample mass).

194 2.4.4 Toxic equivalency calculation

195 Toxic Equivalent Quotient values (TEQ) were calculated according to the 2005 World Health
196 Organization Toxic Equivalency Factors (van den Berg et al., 2006) and basically expressed
197 on a fresh weight basis.

198 2.4.5. Quality assurance/quality control

199 All these procedures integrated quality control parameters to fulfill the requirements of the
200 Commission Directive 2002/69/EC and 2002/70/EC of July 2002, laying down the sampling
201 methods and the methods of analysis for the official control of dioxins and the determination
202 of dl-PCBs in foodstuffs and feedingstuffs respectively. Moreover, all analyses were
203 performed upon a double quality management system associated with an accreditation system
204 according to the ISO 17025:2005 standard for analytical measurements.

205 2.5. Statistical analysis

206 The Shapiro-Wilk and the Kolmogorov-Smirnov tests were employed to determine the
207 normality of the results. Consequently to these tests, non-parametric tests (Kruskal-Wallis and
208 pair-wise comparison tests) were used in order to highlight significant differences of PCB
209 levels and fingerprints in filets of eels with different life stages and from different sites. The
210 significant level of each test was determined according to Bonferroni correction (corrected
211 significant level of 0.005). To compare PCB levels in eel filets from different sites and
212 facilitate their discrimination, Principal Component Analysis (PCA) were performed.
213 WHO₂₀₀₅ PCDD/F and dl-PCB TEQ values were compared according to life stages using
214 Mann-Whitney test at a significant level of 5%. All statistical treatments were realized with
215 XLstat software.

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1
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3 217 3. Results and discussion

4
5 218 3.1 Biometric parameters

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7
8 219 Table 1 shows biometric parameters of the European eels collected in the Loire estuary
9
10 220 according to life stage, sampling site and size class. The increase of BW is positively
11
12 221 correlated with the increase of BL whatever the life stage (yellow or silver), and the sampling
13
14 222 site. The linear regression equations for yellow eels are: Varades $BL=126.18 \ln BW-154.9$
15 223 $R^2=0.97$ (n=16); Nantes $BL=152.8 \ln BW-292.5$ $R^2=0.97$ (n=16); Cordemais $BL=113.14$
16
17 224 $\ln BW-105.2$ $R^2=0.98$ (n=17) and those of silver eels was $BL=220.94 \ln BW-685.9$ $R^2=0.98$
18
19 225 (n=13). The age of eels is associated to BL and BW, only for yellow individuals from
20
21 226 Cordemais ($BL=60.9 \text{ Age}+109.2$ $R^2=0.77$ (n=17)). No significant correlation was observed
22
23 227 for yellow individuals from others sites and for silver eels. Fulton's condition factor values
24
25 228 (K) are roughly similar in the range of the different size classes studied as well as according to
26
27 229 the sampling site and the life stage, with values ranging from 0.13 to 0.17. According to
28
29 230 (Feunteun, 2002), these values are representative of eel good health of in the Loire estuary.
30
31 231 Such values are similar to the Fulton's condition factor found in other studies about European
32
33 232 areas (Gravato et al., 2010; Palstra et al., 2007b; Tapie et al., 2011). Nevertheless, better eel
34
35 233 conditions were calculated in some other studied sites like the River Rhine watershed and
36
37 234 Lake Ijsselmeer (Haenen et al., 2010).

38 235 3.2 Influence of life stage, sampling site and size class on dl and ndl-PCB levels

39
40
41 236 Table 1 shows the PCB levels (dl and ndl-PCBs) according to the life stage, the sampling site
42
43 237 and the size class. As it was already reported in a previous work (Tapie et al., 2011), PCB
44
45 238 levels determined for glass eels were higher than the limit of quantification of the analytical
46
47 239 methods. The sums of dl and ndl-PCBs are $3.7 \pm 1.9 \text{ ng.g}^{-1} \text{ dw}$ and $15.2 \pm 4.2 \text{ ng.g}^{-1} \text{ dw}$
48
49 240 respectively. These levels could be the result of a contamination via the food web during the
50
51 241 leptocephali stage (plankton) and to a direct exposure from the aquatic compartment. Another
52
53 242 hypothesis could be an intergenerational transfer of contaminants (Palstra and van den
54
55 243 Thillart, 2011).

56 244 Regarding yellow eels, the PCB contamination increases and becomes significantly higher
57
58 245 compared to glass eels whatever the sampling site and the size class considered. Regarding
59
60 246 each site, the trends of ndl and dl-PCB levels expressed as $\text{ng.g}^{-1} \text{ dw}$ or lw are similar and

247 showed no significant difference according to the size classes. This observation could be
1 248 attributed to the low sample number per size class. Nevertheless, it is possible to conclude that
2 249 PCB levels were not correlated to BL and BW, except for eels from Nantes (Table 1).
3
4
5 250 Considering the results depicted for eels from Nantes and the eel ecology (bottom dwelling
6
7 251 fish), the increasing contamination with BL and BW could be attributed to the continental
8
9 252 phase longevity and consequently to the time spent in the estuary environment, in close
10
11 253 contact with potentially contaminated sediments. It could be also related to the trophic chain
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13 254 based on a more or less contaminated food.

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15 255 Regarding the silver eels, dl-PCB levels, expressed as ng.g^{-1} dw, were significantly higher
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17 256 than results for yellow eels from Varades and Cordemais. Considering the same unit, ndl-PCB
18
19 257 levels for silver eels were significantly higher than levels for yellow eels from Varades only.
20
21 258 Considering dl- and ndl-PCB levels expressed as ng.g^{-1} lw, the results tend to decrease but are
22
23 259 only significantly different to those for yellow eels from Nantes, and whatever all the size
24
25 260 classes. This results can be explained by the highly lipid content in silver eels leading to a
26
27 261 dilution of the contaminants.

28
29 262 In a previous work (Tapie et al., 2011), a review about marker PCB levels in *Anguilla*
30
31 263 *anguilla* filets was achieved from the literature. Marker PCB congeners are #28, 52, 101, 118,
32
33 264 138, 153 and 180. To compare with this synthetic review, the values obtained in this study
34
35 265 for the last congeners were summed, and expressed as ng.g^{-1} ww and lw (Table 1). The PCB
36
37 266 congener #118 is usually used as a marker PCB until now (ANSES, 2011). The percentage of
38
39 267 this congener was relatively constant and represented an average of 9.39 ± 2.7 % of total PCB
40
41 268 marker level.

42
43 269 At the international scale, eels from the Loire estuary appear to be more contaminated than
44
45 270 those from some other sites in Poland, Ireland, Spain, Italy and the UK (Bordajandi et al.,
46
47 271 2003; Corsi et al., 2005; McHugh et al., 2010; Santillo et al., 2005). However, other sites are
48
49 272 more contaminated than the Loire estuary (twice to 10 times higher), i.e. the River Elbe in
50
51 273 Czech Republic and Germany, the Tevere and Gagliarino rivers in Italy, Flanders in Belgium
52
53 274 and different lakes in Finland (Belpaire et al., 2011; Maes et al., 2008; Tulonen and Vuorinen,
54
55 275 1996; van der Oost et al., 1996). Throughout France, eels from the Loire estuary are slightly
56
57 276 more contaminated than those from the Vacares lagoon and about three times more than those
58
59 277 from the Thau pound (Oliveira Ribeiro et al., 2008; Santillo et al., 2005), whereas they are

278 less contaminated than eels from the Rhone River (about ten times less) and the Gironde
279 estuary (about two times less, whatever the life stage and the size class) (Tapie et al., 2011).

280 In the study of Tapie et al. (2011), a significant decrease of marker-PCB levels expressed as
281 ng.g^{-1} dw was observed for eels exceeding 600 mm. These authors hypothesized that this
282 decrease could be induced by two parameters regarding the sexual maturity of the individuals
283 of this size class. On the one hand, eels could be at the onset of the silvering and coming from
284 upstream areas, less contaminated. On the other hand, silver eels could be already in
285 starvation and start to mobilize lipid stores as fuel energy to ensure the sexual maturation and
286 swimming towards spawning areas. This mobilization of lipids was already proposed to
287 explain a decrease in lipid contents observed in filets of eels larger than 800 mm (Durif et al.,
288 2005). In this present work, no decrease is observed for eels with length superior than 600
289 mm whatever the unit expression. .

290
291 In order to evaluate the correlations between biometric parameters and PCB levels as well as
292 the sampling site effect, a principal component analysis (PCA) was performed by using
293 biometric parameters (age, BW, BL and LW) and dl and ndl-PCB levels expressed as ng.g^{-1}
294 dw. Since silver eels are not strictly territorial, due to their downstream migration, they could
295 be originated from other sites than sampling ones. For that reason, the PCA was performed
296 with yellow eels only. As it was shown in the Table 1, the size class distributions between the
297 3 studied sites are comparable. Consequently, it is possible to study and discuss the presence
298 of an eventual sampling site effect on yellow eel impregnation.

299 The correlation loading and sample representation are shown on figure 2 (respectively Fig.2 A
300 and Fig.2 B). The first two principal components (respectively PC1 and PC2) describe
301 82.97% of the total variability among eels. PC1 and PC2 represent respectively 62.65 and
302 20.32%.

303 The correlation loading (Fig.2A) highlights that biometric parameters (BW, BL and age) are
304 correlated to each other as it was depicted in Table 1. Concerning LW, it appears to be quite
305 correlated to both levels of dl- and ndl-PCBs. This observation was expected and already
306 well-known according to the lipophilic properties of PCBs (van der Oost et al., 1996).
307 Regarding the sample presentation in Fig.2B, the eels are relatively clustered according to the
308 three different sampling sites. The comparison of Fig.2A and Fig.2B underlines that eels from

309 Varades are the lowest contaminated by dl and ndl-PCBs closely related with lower LW. The
1 310 eels from Nantes and some of those from Cordemais are more contaminated, showing a
2 311 higher LW. However, eels from Nantes present a higher heterogeneity. The inter-site
3 312 differences observed could be also related to differences of biometric parameters such as the
4 313 BL and the age, characterizing a different exposition time (9.8 ± 1.9 years for eels from
5 314 Nantes compared to 4.4 ± 1.4 years and 5.9 ± 1.9 years for eels from Cordemais and Varades,
6 315 respectively).

13 316 Moreover, Varades is a small city (about 3550 locals), relatively preserved, located upstream
14 317 in the estuary, and with few industrial activities. It is probably for these reasons that the eels
15 318 from this sampling site are less contaminated than the others. Nantes is indeed an important
16 319 city (about 600000 locals) and Cordemais is downstream of Nantes and well-known for its
17 320 industrial activities. In this study, the living area of eels seems to affect their contamination
18 321 level as it was already shown in the Gironde estuary (Tapie et al., 2011). These inter-site
19 322 differences would be highlighted in the next section dealing with eels PCB fingerprints.
20 323

27 324 3.3 PCB fingerprints in eels according to the sampling site and life stage

28 325 A sampling site effect has been previously demonstrated (Fig.2). A second PCA was then
29 326 performed using individual PCB levels expressed as ng.g^{-1} dw. For the same reason that 3.2
30 327 paragraph, this second PCA was realized with yellow eels only. Consequently, this PCA was
31 328 useful in order to evaluate the influence of the sampling site on PCB fingerprints in yellow
32 329 eels.
33 330

34 331 The result of the PCA correlation loading is shown in Fig.3A. The first two principal
35 332 components of the PCA (respectively PC1 and PC2) describe 85.74% of the total variability
36 333 among eels. PC1 and PC2 respectively represent 68.85 and 16.89%. This figure highlights
37 334 that the first principal component is positively correlated to all the individual PCB levels. The
38 335 second one is negatively correlated to low chlorinated PCBs and positively correlated to
39 336 highly chlorinated ones.
40 337

41 338 Each PCB congener is represented around the right part of the correlation circle. Nevertheless,
42 339 the repartition of the different PCBs seems to be due to their chemical structure, i.e. the
43 340 number and the position of Cl atoms. Low chlorinated PCBs with few Cl atoms in meta and
44 341 para-positions are in the right bottom of the circle. The more the number of total Cl atoms and
45 342

340 of Cl atoms in meta- and para- positions are important, the upper their localization is. Ndl-
1 341 PCBs are considered as particularly persistent and present in the environment, representing
2 342 about 50% of all of the PCB congeners found in food from animal origin (AFSSA, 2006).
3 343 According to Fig.3A, they were well distributed around the right part of the correlation circle
4 344 and among all the other PCBs, emphasizing their qualitative representativeness of all the PCB
5 345 congeners (Cariou et al., 2010).

11 346 Fig.3B enhances this result showing the accumulation patterns of marker-PCBs in yellow eels
12 347 from the three sampling sites comparatively to those in the glass and silver eels.

16 348 Concerning the main congener #153, it contributes to an average of 42% of the contamination
17 349 of the yellow eels whatever the sampling site. This PCB is interesting because it is non
18 350 metabolizable and a tracer of bioaccumulation process. The percentages of this congener are
19 351 not significantly different for glass and silver eels compared to yellow eels from Varades but
20 352 they are lower than those found for yellow eels from Cordemais and Nantes.

26 353 Considering the sampling site influence, an inter-site variability is observed, particularly for
27 354 eels from Cordemais which display patterns with significantly lower relative proportion of
28 355 PCBs #28 and 118 whereas the relative proportion of #180 is significantly higher than those
29 356 of the other sites. About eels from Nantes, the relative proportion of the PCB #101 is
30 357 significantly higher to the detriment of the #180 with a relative proportion significantly lower.
31 358 Finally, the PCB #180 is the only one able to discriminate the sampling sites. Its proportion is
32 359 higher in Cordemais eels, lower in Nantes eels and intermediate in Varades eels. For the other
33 360 marker-PCBs (#153, 138 and 52), no significant specific trend is observed according to the
34 361 sampling site.

43 362 The PCB contamination variability between the three sampled sites could be partially
44 363 explained by the anthropogenic activities existing in the area. Indeed, Varades is a relatively
45 364 small city, marked by several agricultural activities where PCB sources are probably less
46 365 important than in Nantes or Cordemais. On the contrary, Nantes is an important urban and
47 366 industrial city, with an important economic and demographic development, where two
48 367 incinerator factories and various maritime and industrial activities exist. A lot of domestic,
49 368 industrial and agricultural effluents are discharged into the Loire estuary more or less
50 369 previously depolluted in sewage treatment plants. Cordemais presents another profile because
51 370 it is a very small rural city but dominated by its economic and industrial activities, directly

371 based on the presence of the Loire estuary such as a coal-fired power plant and close to an
1 372 industrial complex which includes oil refineries.
2

3
4 373 The PCB sources are therefore multiple along the estuary and the PCB contamination of this
5
6 374 ecosystem could be done following atmospheric or aquatic routes. This complexity prevents
7
8 375 from establishing easy correlations between the congener profiles and the sources. Motelay-
9
10 376 Massei et al. (2004) showed, in the Seine river basin, that less chlorinated PCB congeners are
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12 377 transported over longer distances from the source sites because of their longer residence time
13
14 378 in the atmosphere, whereas the heaviest PCBs tend to be adsorbed on particles and to settle
15
16 379 near production sources. Therefore, our results could suggest that eels from Cordemais were
17 380 living closer to a PCB source than eels from the other sites.
18

19
20 381 Moreover, the Loire estuary presents strong hydrodynamic, sedimentary and abiotic
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22 382 parameters (Dauvin, 2008). Today, the effects of the tide can be observed within 97 km from
23
24 383 the estuary mouth and the salinity moves upstream. This modifies also the temperature from
25
26 384 the mouth of the estuary to the upstream front of the salinity which varies of 5°C from
27
28 385 downstream to upstream. The Loire estuary is also characterized by a fluid mud which
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30 386 extends over 20 km; it is an important factor in the rapid sedimentation of the estuary, so the
31
32 387 turbidity varies from 2 g.L⁻¹ at the surface to 20 g.L⁻¹ near the bottom. All these parameters
33
34 388 (salinity, turbidity, temperature and tidal amplitude) could affect the exposition level of PCBs
35
36 389 potentially present in the estuary, and their chemical bioavailability for eels. Moreover, the
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38 390 biological variability of eel PCB levels could also be explained by differences of diets,
39
40 391 ecology, physiology or metabolism capacities in relation to polyhaline, mesohaline and
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42 392 oligohaline ecozones.
43

44 393 Regarding the difference of the relative proportions found for silver eels compared to yellow
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46 394 eels, the only significant one is for PCB #28 which is higher in silver eels compared to yellow
47
48 395 eels whatever the sampling site considered. The patterns of silver eels are closer to those of
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50 396 eels from Nantes, then Varades, whereas they display many significant differences
51
52 397 comparatively to those of eels from Cordemais: all the relative proportions are significantly
53
54 398 different except the one of PCB #138.
55

56 399 Finally, concerning the glass eels, a contrasting pattern is noticed, underlying a different
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58 400 bioaccumulation phenomenon, characterized mainly by an important proportion of less
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60 401 chlorinated PCBs to the detriment to the heaviest PCBs. This was already shown in a previous
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62 402 study (Tapie et al., 2011) in which congeners 28, 50, 52, 101, 118 represented 51% of
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403 accumulation pattern for glass eels sampled in the Gironde estuary. Comparatively, the
1 404 relative proportion of these congeners (without the PCB #50 not analyzed in this study)
2 405 represent 53% for glass eels in our study. The less chlorinated PCBs are transported over
3 406 longer distances from the PCB sources sites because of their longer residence time in the
4 407 atmosphere (Motelay-Massei et al., 2004). Moreover, in aquatic environments, these PCBs
5 408 that are more polar, are found to be dominant in dissolved phase and particulate organic
6 409 matter (Cailleaud et al., 2007). Glass eels come from the oceanic platform after the
7 410 metamorphosis of larvae leptocephali stage. These transparent larvae move with currents
8 411 (pelagic compartment) for months, or years, in seawaters far from important pollution
9 412 sources. During their travel, the larvae were mainly contaminated by feeding uptake. When
10 413 they approach the continental shelf claim, the metamorphosis in glass eels occurs. They stop
11 414 then to feed and the contamination is then by direct exposure which leads to a pattern similar
12 415 to that of water column dominated by less chlorinated compounds. It is likely that the specific
13 416 PCB pattern of glass eels could be the result of these two phenomena. Another hypothesis is
14 417 the transfer of PCBs from adult eels to eggs and, consequently, to glass eels, low chlorinated
15 418 PCBs being more efficiently transferred than the heavy chlorinated ones (Bargar et al., 2001;
16 419 Verreault, 2006).

31 32 420 3.5 PCDD/F, PCB levels and public health

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34 421 Mean PCDD/F and dl-PCB levels expressed according to the 2005 WHO recommendations
35 422 were 5.21 ± 1.78 and 9.88 ± 4.14 $\text{pg}\cdot\text{g}^{-1}$ WHO₂₀₀₅ PCDD/F and dl-PCB TEQ (toxic
36 423 equivalents) ww for yellow (n=5) and silver individuals (n=6) respectively. Glass eels
37 424 depicted a mean level significantly lower (0.27 ± 0.03 $\text{pg}\cdot\text{g}^{-1}$ WHO₂₀₀₅ PCDD/F and dl-PCB
38 425 TEQ ww). The maximum level established for the level of PCDD/Fs in eel filets is currently
39 426 3.5 $\text{pg}\cdot\text{g}^{-1}$ WHO₂₀₀₅ PCDD/F TEQ ww and 10 $\text{pg}\cdot\text{g}^{-1}$ WHO₂₀₀₅ PCDD/F and dl-PCB TEQ wet
40 427 weight (European Union, 2011). These values were not reached regarding the yellow eels.
41 428 Nevertheless, in the case of silver ones, biological variability was high and 4 out of 6 studied
42 429 eels displayed WHO₂₀₀₅ PCDD/F and dl-PCB TEQ values higher than 10 pg WHO₂₀₀₅
43 430 PCDD/F and dl-PCB TEQ per gram of wet filet.

44 431 Regarding the congeners # 28, 52, 101, 138, 153 and 180 (ndl-PCB), sampled eels did not
45 432 present levels superior than the 2005 WHO recommendation of 300 $\text{ng}\cdot\text{g}^{-1}$ ww. Silver eels and
46 433 yellow individuals from Nantes depicted the highest levels (mean of 204.6 ± 113.3 and 175.7
47 434 ± 90.7 $\text{ng}\cdot\text{g}^{-1}$ ww, respectively), but few individuals (3/29) presented concentrations higher

435 than the recommended level (Table 1). Yellow individuals from Cordemais presented
436 intermediate levels (mean of $117.9 \pm 47.7 \text{ ng.g}^{-1} \text{ ww}$) and those from Varades the lowest ones
437 (mean of $75.5 \pm 25.2 \text{ ng.g}^{-1} \text{ ww}$).

438 Our results indicate a potential exposure to PCBs through eel consumption in this estuary, and
439 especially with silver ones. The French Food Safety Agency proposed a tolerable daily intake
440 (TDI) of 10 ng/ kg body weight/day (for the 6 ndl-PCB congeners), which represents 700
441 ng/day for a 70 kg person or 150 ng/day for a child of 15 kg (under 3 years) (French Food
442 Safety Agency, 2010). We could thus recommend to limit the consumption of eel from the
443 Loire estuary to one portion (150 g) per month for the general population, which represent an
444 average dietary daily intake of 694 ng/day. This is more restricted than the French Food
445 Safety Agency recommendations which limit the consumption of PCB bioaccumulating fish
446 to two portions per month for the general population. Specific recommendations (a portion of
447 60 g every two months) exist for the most sensitive populations (pregnant and breastfeeding
448 women, young and adolescent girls, women of childbearing age, and children under 3) and are
449 in agreement with our results, representing an average dietary daily intake of 139 ng/day.

450 A national study assessing the PCB impregnation of freshwater fish consumers performed on
451 six investigation sites including the Loire (French Food Safety Agency, 2011), revealed that
452 only 13% of participants (on a total of 606 amateur anglers and members of their households
453 and 16 professional anglers) are strong PCB-bioaccumulator freshwater fish consumers, with
454 a moderate consumption frequency of 1 time per month. Among the strong PCB-
455 bioaccumulator freshwater fish species, eel is consumed with a mean annual frequency of 2.6
456 times per year. Considering these local practices and our results, a dietary daily intake of ndl-
457 PCBs varying from 22 to 504 ng/day with a mean of 150 ng/day could be estimated. The TDI
458 value is not exceeded and the risk seems then to be moderate for an adult consumer but really
459 present for the most sensitive populations.

461 Conclusion

462 This study gives a first assessment of the PCB contamination of a European eel population
463 fraction from the Loire estuary, along a hundred-km long portion of ecosystem. The
464 quantitative and qualitative contents of PCBs in eel filets are different depending on their life
465 stage and the sampling sites. The eels sampled in the site next to Varades (small city under

466 agricultural pressure) appeared less contaminated than the two other sites, *i.e.* Nantes (an
1 467 important city) and Cordemais (a town hosting a coal-fired power station). Regarding the
2
3 468 PCB patterns, the sampled sites of Varades and Nantes could be associated to urban
4
5 469 influences whereas the one of Cordemais, more impacted by heavy chlorinated PCBs, would
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7 470 be nearer from a PCB industrial source. Compared to other international or national areas, the
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9 471 ndl-PCB levels in eels from Loire estuary show an intermediate contamination. Our results
10
11 472 indicate a potential exposure to PCBs through eel consumption, and especially with silver
12
13 473 ones. According to TDI value, the consumption must be limited to once per month for the
14
15 474 general population and to once every two month for the most sensitive ones.

16
17 475 Apart from an eventual sanitary problem, the contamination of eels could lead to damages for
18
19 476 the eel population by affecting their reproduction and by a transfer of pollutants to eggs.
20
21 477 Indeed, since these compounds are lipophilic, the results showed that the PCB levels are
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23 478 correlated to the lipid content in the filets. Lipids are essential compounds for the migration
24
25 479 reproduction (van Ginneken et al., 2009) and for both fat deposition in the oocytes and later
26
27 480 incorporation of vitellogenic stores in eggs. Moreover, acclimation of eels to seawater,
28
29 481 silvering process and reproduction migration are under different endocrine controls and fuel
30
31 482 consuming. This energetic cost is described to increase significantly when lipid filets of
32
33 483 swimming eels are charged in PCB mixture (after intraperitoneally injection of 5000 ng g⁻¹
34
35 484 PCB # 153, 7 ng g⁻¹ PCB # 126 and 50 ng g⁻¹ PCB # 77) (Thillart et al., 2009). PCB levels
36
37 485 determined in eels from the Loire estuary could thus potentially have an impact on the
38
39 486 reproduction success of European eels.

40 487 The comparison of eel biomonitoring studies highlighted heterogeneity in sampled
41
42 488 individuals. To better correlate studies at the international level, it appears necessary to
43
44 489 standardize parameters such as age, length, sex and sexual maturation stage. To preserve this
45
46 490 endangered species and such as recommended by scientists (van Ginneken et al., 2009), the
47
48 491 environmental quality of its habitats should be restored and protected. Considering our results,
49
50 492 the European eels from the Loire estuary appeared moderately contaminated compared to eels
51
52 493 from other major international estuaries, suggesting a moderate PCB contamination of the
53
54 494 Loire estuarine system. These conditions could contribute therefore to preserve genitors.

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4
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11 502 References

12
13 503 AFSSA. Avis de l'Agence française de sécurité sanitaire des aliments relatif à l'établissement
14
15 504 de teneurs maximales pertinentes en polychlorobiphényles qui ne sont pas de type
16
17 505 dioxine (PCB « non dioxin-like », PCB-NDL) dans divers aliments In: N°2006-SA-
18
19 506 0305 S, editor. AFSSA, 2006, pp. 28.

20
21 507 ANSES. <http://www.anses.fr> 2011.

22 508 Bargar TA, Scott GI, Cobb GP. Maternal transfer of contaminants: case study of the
23
24 509 excretion of three polychlorinated biphenyl congeners and technical-grade
25
26 510 endosulfan into eggs by white leghorn chickens (*Gallus domesticus*). Environ.
27
28 511 Toxicol. Chem. 2001; 20: 61-67.

29
30 512 Belpaire C, Geeraerts C, Roosens L, Neels H, Covaci A. What can we learn from monitoring
31
32 513 PCBs in the European eel? A Belgian experience. Environ. Int. 2011; 37: 354-364.

33
34 514 Bordajandi LR, Gomez G, Fernandez MA, Abad E, Rivera J, Gonzalez MJ. Study on PCBs,
35
36 515 PCDD/Fs, organochlorine pesticides, heavy metals and arsenic content in freshwater
37
38 516 fish species from the River Turia (Spain) Chemosphere 2003; 53: 163-171.

39 517 Cailleaud K, Forget-Leray J, Souissi S, Lardy S, Augagneur S, Budzinski H. Seasonal
40
41 518 variation of hydrophobic organic contaminant concentrations in the water-column of
42
43 519 the Seine Estuary and their transfer to a planktonic species *Eurytemora affinis*
44
45 520 (Calanoid, copepod). Part 2: Alkylphenol-polyethoxylates. Chemosphere 2007; 70:
46
47 521 281-287.

48 522 Canapa A, Barucca M, Celeste A, Olmo E, Regoli F. Preliminary investigations on
49
50 523 vitellogenin m-RNA induction in some bioindicator Mediterranean fish species. Mar.
51
52 524 Environ. Res. 2002; 54: 673-677.

53
54 525 Cariou R, Marchand P, Vénisseau A, Brosseau A, Bertrand D, Qannari EM, et al. Prediction
55
56 526 of the PCDD/F and dl-PCB 2005-WHO-TEQ content based on the contribution of six
57
58 527 congeners: Toward a new screening approach for fish samples? Environ. Pollut. 2010;
59
60 528 158: 941-947.

- 529 Castonguay M, Hodson P, Moriarty C, Drinkwater K, Jessop B. Is there a role of ocean
1 530 environment in American and European eel decline? *Fish Oceanogr.* 1994; 3: 197-203.
2
3 531 Corsi I, Mariottini M, Badesso A, Caruso T, Borghesi N, Bonacci S, et al. Contamination and
4
5 532 sub-lethal toxicological effects of persistent organic pollutants in the European eel
6
7 533 (*Anguilla anguilla*) in the Orbetello lagoon (Tuscany, Italy). *Hydrobiologia* 2005; 550:
8
9 534 237-249.
- 10 535 Dauvin J-C. The main characteristics, problems, and prospects for Western European coastal
11
12 536 seas. *Mar. Pollut. Bull.* 2008; 57: 22-40.
13
- 14 537 Dekker W. Did lack of spawners cause the collapse of the European eel, *Anguilla anguilla*?
15
16 538 *Fisheries Manag. Ecol.* 2003; 10: 365-376.
17
- 18 539 Durif C, Dufour S, Elie P. The silvering process of *Anguilla anguilla*: a new classification
19
20 540 from the yellow resident to the silver migrating stage. *J. Fish Biol.* 2005; 66: 1025-
21
22 541 1043.
- 23 542 Elie P, Lecomte-Finiger R, Cantrelle I, Charlon N. Définition des limites des différents stades
24
25 543 pigmentaires durant la phase civelle d'anguilles *Anguilla* sp.(poisson téléostéen
26
27 544 anguilliforme). *Vie et Milieu* 1982; 32: 149-157.
28
- 29 545 European Union CR. No 1259/2011 of 2 December 2011 amending Regulation (EC) N°
30
31 546 1881/2006 as regards maximum levels for dioxins, dioxin-like PCBs and non dioxin-
32
33 547 like PCBs in foodstuffs, 2011, pp. 18-23.
- 34 548 Feunteun E. Management and restoration of European eel population (*Anguilla anguilla*): An
35
36 549 impossible bargain. *Ecol. Eng.* 2002; 18: 575-591.
37
- 38 550 French Food Safety Agency. Opinion of the French Food Safety Agency on interpreting the
39
40 551 health impact of PCB concentration levels in the French population 2010, pp. 20.
41
- 42 552 French Food Safety Agency. Étude nationale d'imprégnation aux polychlorobiphényles des
43
44 553 consommateurs de poissons d'eau douce, 2011, pp. 176.
- 45 554 Fulton TW. The rate of growth of fishes. 22nd Annual Report of the Fishery Board of
46
47 555 Scotland. 1904, 3: 141-241.
48
- 49 556 Geeraerts C, Belpaire C. The effects of contaminants in European eel: a review.
50
51 557 *Ecotoxicology* 2010; 19: 239-266.
52
- 53 558 Geeraerts C, Focant JF, Eppe G, De Pauw E, Belpaire C. Reproduction of European eel
54
55 559 jeopardised by high levels of dioxins and dioxin-like PCBs? *Sci. Tot. Environ.* 2011;
56
57 560 409: 4039-4047.
- 58 561 Gravato C, Guimarães L, Santos J, Faria M, Alves A, Guilhermino L. Comparative study
59
60 562 about the effects of pollution on glass and yellow eels (*Anguilla anguilla*) from the
61

- 563 estuaries of Minho, Lima and Douro Rivers (NW Portugal). *Ecotoxicol. Environ. Saf.*
1 2010; 73: 524-533.
2
- 3 565 Grosbois C, Meybeck M, Lestel L, Lefèvre I, Moatar F. Severe and contrasted polymetallic
4 contamination patterns (1900-2009) in the Loire River sediments (France). *Sci. Tot.*
5 6 *Environ.* 2012; 435-436: 290-305.
7
- 8 568 Haenen OLM, Lehmann J, Engelsma MY, Stürenberg FJ, Roozenburg I, Kerkhoff S, et al.
9 The health status of European silver eels, *Anguilla anguilla*, in the Dutch River Rhine
10 Watershed and Lake IJsselmeer. *Aquaculture* 2010; 309: 15-24.
11
- 12 570 Hubaux N, Perceval O. Pollution des milieux aquatiques par les PCBs en France."PCB dans
13 les milieux aquatiques: enjeux de gestion et lacunes dans les connaissances".
14 ONEMA, 2011, pp. 52.
15
- 16 571 ICES ICftEotSac. annual congres. <http://www.ices.dk> 2006.
17
- 18 574 Kodavanti PRS, Curras-Collazo MC. Neuroendocrine actions of organohalogens: Thyroid
19 hormones, arginine vasopressin, and neuroplasticity. *Front. Neuroendo.* 2010; 31: 479-
20 575 496.
21
- 22 576 Maes J, Belpaire C, Goemans G. Spatial variations and temporal trends between 1994 and
23 2005 in polychlorinated biphenyls, organochlorine pesticides and heavy metals in
24 European eel (*Anguilla anguilla* L.) in Flanders, Belgium. *Environ. Pollut.* 2008; 153:
25 577 223-237.
26
- 27 578 Marchand J, Quiniou L, Riso R, Thebaut M-T, Laroche J. Physiological cost of tolerance to
28 toxicants in the European flounder *Platichthys flesus*, along the French Atlantic Coast.
29 *Aquat. Toxicol.* 2004; 70: 327-343.
30
- 31 585 McHugh B, Poole R, Corcoran J, Anninou P, Boyle B, Joyce E, et al. The occurrence of
32 persistent chlorinated and brominated organic contaminants in the European eel
33 (*Anguilla anguilla*) in Irish waters. *Chemosphere* 2010; 79: 305-313.
34
- 35 588 Mezzetta S, Cirlini M, Ceron P, Tecleanu A, Caligiani A, Palla G, et al. Concentration of DL-
36 PCBs in fish from market of Parma city (north Italy): Estimated human intake.
37 *Chemosphere* 2011; 82: 1293-1300.
38
- 39 590 Moriarty C, Dekker W. Management of the European Eel. *Fisheries Bulletin* 1997; 15: 110.
40
- 41 592 Motelay-Massei A, Ollivon D, Garban B, Teil MJ, Blanchard M, Chevreuil M. Distribution
42 and spatial trends of PAHs and PCBs in soils in the Seine River basin, France.
43 *Chemosphere* 2004; 55: 555-565.
44
- 45 593
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

- 595 Nunes M, Marchand P, Vénisseau A, Bizec BL, Ramos F, Pardal MA. PCDD/Fs and dioxin-
1 596 like PCBs in sediment and biota from the Mondego estuary (Portugal). *Chemosphere*
2 597 2011; 83: 1345-1352.
3
4
5 598 Oliveira Ribeiro CA, Vollaïre Y, Coulet E, Roche H. Bioaccumulation of polychlorinated
6 599 biphenyls in the eel (*Anguilla anguilla*) at the Camargue Nature Reserve - France.
7 600 *Environ. Pollut.* 2008; 153: 424-431.
8
9 601 ONEMA. Les données du plan national d'action PCB. 2012.
10 602 <http://www.pollutions.eaufrance.fr/pcb/resultats-xls.html>.
11
12
13 603 Palstra AP, Curiel D, Fekkes M, de Bakker M, Székely C, van Ginneken V, et al. Swimming
14 604 stimulates oocyte development in European eel. *Aquaculture* 2007a; 270: 321-332.
15
16 605 Palstra AP, Heppener DFM, van Ginneken VJT, Svakely C, van den Thillart GEEJM.
17 606 Swimming performance of silver eels is severely impaired by the swim-bladder
18 607 parasite *Anguillicola crassus*. *J. Exp. Mar. Biol. Ecol.* 2007b; 352: 244-256.
19
20 608 Palstra AP, van den Thillart GEEJM. Swimming physiology of European silver eels (*Anguilla*
21 609 *anguilla* L.): energetic costs and effects on sexual maturation and reproduction. *Fish*
22 610 *Physiol. Biochem.* 2011; 36: 297-322.
23
24
25 611 Perraudau Y, Després L. L'estuaire de la Loire: un territoire en développement durable?
26 612 Rennes: Presses Universitaires de Rennes; 2009.
27
28 613 Robinet TT, Feunteun EE. Sublethal Effects of Exposure to Chemical Compounds: A Cause
29 614 for the Decline in Atlantic Eels? *Ecotoxicology* 2002; 11: 265-277.
30
31 615 Roche H, Buet A, Jonot O, Ramade F. Organochlorine residues in european eel (*Anguilla*
32 616 *anguilla*), crucian carp (*Carassius carassius*) and catfish (*Ictalurus nebulosus*) from
33 617 Vaccarès lagoon (French National Nature Reserve of Camargue) - effects on some
34 618 physiological parameters. *Aquat. Toxicol.* 2000; 48: 443-459.
35
36 619 Roosens L, Geeraerts C, Belpaire C, Van Pelt I, Neels H, Covaci A. Spatial variations in the
37 620 levels and isomeric patterns of PBDEs and HBCDs in the European eel in Flanders.
38 621 *Environ. Int.* 2010; 36: 415-423.
39
40 622 Santillo D, Johnston P, Labunska I, Brigden K. Swimming in Chemicals Widespread.
41 623 Presence of Brominated Flame Retardants and PCBs in eels (*Anguilla anguilla*)
42 624 from Rivers and Lakes in 10 European Countries. In: Note T, editor. 12. Greenpeace
43 625 International; 2005, pp. 56.
44
45 626 Tapie N, Le Menach K, Pasquaud S, Elie P, Devier MH, Budzinski H. PBDE and PCB
46 627 contamination of eels from the Gironde estuary: From glass eels to silver eels.
47 628 *Chemosphere* 2011; 83: 175-185.
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

- 629 Thillart G, Dufour S, Rankin JC, Palstra A. Artificial Maturation and Reproduction of the
1 630 European Eel. Spawning Migration of the European Eel. Springer Netherlands; 2009.
2
3 631 Tulonen J, Vuorinen PJ. Concentrations of PCBs and other organochlorine compounds in eels
4
5 632 (*Anguilla anguilla*, L.) of the Vanajavesi watercourse in southern Finland, 1990-1993.
6
7 633 *Sci. Tot. Environ.* 1996; 187: 11-18.
8
9 634 van den Berg M, Birnbaum LS, Denison M, De Vito M, Farland W, Feeley M, et al. The 2005
10
11 635 World Health Organization Re-evaluation of Human and Mammalian Toxic
12
13 636 Equivalency Factors for Dioxins and Dioxin-like Compounds *Toxicol. Sci.* 2006; 93:
14
15 637 223-241.
16
17 638 van der Oost R, Opperhuizen A, Satumalay K, Heida H, Vermeulen NPE. Biomonitoring
18
19 639 aquatic pollution with feral eel (*Anguilla anguilla*) I. Bioaccumulation: biota-sediment
20
21 640 ratios of PCBs, OCPs, PCDDs and PCDFs. *Aquat. Toxicol.* 1996; 35: 21-46.
22
23 641 van Ginneken V, Palstra A, Leonards P, Nieveen M, van den Berg H, Flik G, et al. PCBs and
24
25 642 the energy cost of migration in the European eel (*Anguilla anguilla* L.). *Aquat.*
26
27 643 *Toxicol.* 2009; 92: 213-220.
28
29 644 Verreault J, Villa, R.A., Gabrielsen, G.W., Skaare, J.U., Letcher, R.J. Maternal transfer of
30
31 645 organohalogen contaminants and metabolites to eggs of arctic-breeding glaucous
32
33 646 gulls. *Environ. Pollut.* 2006; 144: 1053-1060.
34
35 647 Voltura MB, French JB. Effects of dietary polychlorinated biphenyl exposure on energetics of
36
37 648 white-footed mouse, *Peromyscus leucopus*. *Environ. Toxicol. Chem.* 2000; 19: 2757-
38
39 649 2761.

40 650
41
42 651
43 652
44
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Table 1: Means and standard deviations of PCB levels (dl, ndl and marker) and biometric parameters (Body Length BL, Body Weight BW, Lipid Weight LW), Fulton's condition factor (K) and age of sampled European eels (n = 62) from the three studied sites in the Loire estuary according to life stage and size classes.

Life stage	Sampling site	Size class (mm)	n	BL (mm)	BW (g)	Age (year)	LW (%)	K	dl-PCB levels (ng.g ⁻¹ dw)	ndl-PCB levels (ng.g ⁻¹ dw)	dl-PCB levels (ng.g ⁻¹ lw)	ndl-PCB levels (ng.g ⁻¹ lw)	ndl-PCB levels (ng.g ⁻¹ ww)	Marker-PCB levels (ng.g ⁻¹ ww)	Marker-PCB levels (ng.g ⁻¹ lw)
Glass eels		< 200	2 pools	≤ 90	62±12	< 1	4.0 ± 0.8	n.d.	3.7±1.9	15.2±4.2	18.6±8.3	78±26	3.0±0.4	3.5±0.2	89±21
	Varades	200-300	5	279±14	30±3	5.2±0.8	4.9±2.3	0.14±0.02	41.4±12.4	253.7±78.1	286.9±69.2	1764±458	79.7±28.6	86.8±31.3	1918±491
		300-400	5	349±31	59±17	5.7±1.8	6.4±4.9	0.14±0.01	37.6±9.3	228.0±46.9	349.3±372.9	1183±510	72.7±22.2	79.3±24.7	1284±546
		400-500	4	433±26	111±21	5.6±1.8	6.0±4.4	0.14±0.01	48.2±6.5	261.0±12.0	334.5±198.8	1770±1006	76.7±14.2	84.5±15.2	1953±1111
500-600		2	533±31	207±2	9.0±2.1	10.1±11.7	0.14±0.02	29.0±13.8	195.1±118.1	169.7±121.4	1041±619	69.1±58.8	74.6±63.0	1132±682	
Yellow eels	Nantes	300-400	5	366±37	81±28	8.6±0.7	10.4±5.7	0.16±0.02	71.8±15.7	448.5±78.5	266.8±131.2	1657±722	144.8±49.1	158.3±54.7	1810±799
		400-500	4	452±33	129±30	9.5±0.5	10.2±3.2	0.14±0.01	82.2±10.3	482.2±74.4	278.5±103.6	1669±797	151.9±16.4	166.6±17.3	1827±852
		500-600	4	546±24	259±49	10.5±1.2	11.2±6.7	0.16±0.01	97.8±31.0	542.8±171.3	344.4±135.5	1909±767	180.8±77.2	199.0±84.5	2100±828
		>600	3	678±63	551±183	11.0±3.9	11.6±8.9	0.17±0.03	134.9±82.9	734.6±401.9	488.0±244.5	2706±1365	252.0±187.9	278.6±210.0	2986±1508
Cordemais	200-300	5	272±10	26±2	3.1±0.7	6.6±2.8	0.13±0.01	45.6±11.4	326.7±78.8	238.7±126.2	1291±490	95.8±23.5	102.3±25.1	1910±1258	
	300-400	5	342±36	61±19	3.8±0.6	12.0±3.6	0.15±0.02	61.8±22.7	403.8±166.4	172.5±45.5	1130±378	135.7±63.1	146.2±67.7	1217±401	
	400-500	4	455±17	147±12	5.5±0.4	7.7±3.3	0.16±0.01	46.1±6.7	307.3±42.6	191.1±68.8	1275±476	88.1±21.0	94.9±22.4	1372±508	
	500-600	3	522±11	227±22	6.3±1.0	15.0±6.8	0.16±0.00	76.1±17.2	479.3±68.9	185.1±54.4	1170±285	164.8±37.0	177.9±40.9	1263±312	
Silver eels		>500	13	659±124	517±344	12.4±3.8	25.6±3.5	0.16±0.01	93.7±56.3	463.2±244.6	161.6±96.1	800±425	204.6±113.3	229.0±130.3	895±485

n.d.: non determined

Figure 1: Studied area: the Loire estuary (France). Three sampling locations (Cordemais; Nantes and Varades)

Figure 2: Principal Component Analysis of biometric parameters and dl- and ndl-PCB levels expressed in ng.g^{-1} dw in muscles of yellow eels from 3 sampling sites ($n = 49$): Varades, Nantes and Cordemais.

A: correlation loadings (BW: body weight; BL: body length; LW: lipid weight);

B: sample representation (circles = eels from Varades; triangles = eels from Nantes; squares = eels from Cordemais).

Figure 3: Representation of PCB patterns.

A: correlation loadings of Principal Component Analysis of dl and ndl-PCB muscle levels expressed in ng.g^{-1} dw in yellow eels from 3 sampling sites ($n = 49$): Varades, Nantes and Cordemais (dl-PCBs: black circles; ndl-PCBs: white circles);

B: Relative proportion (in %) of marker-PCBs in eel filets according to the life stage and the sampling site.

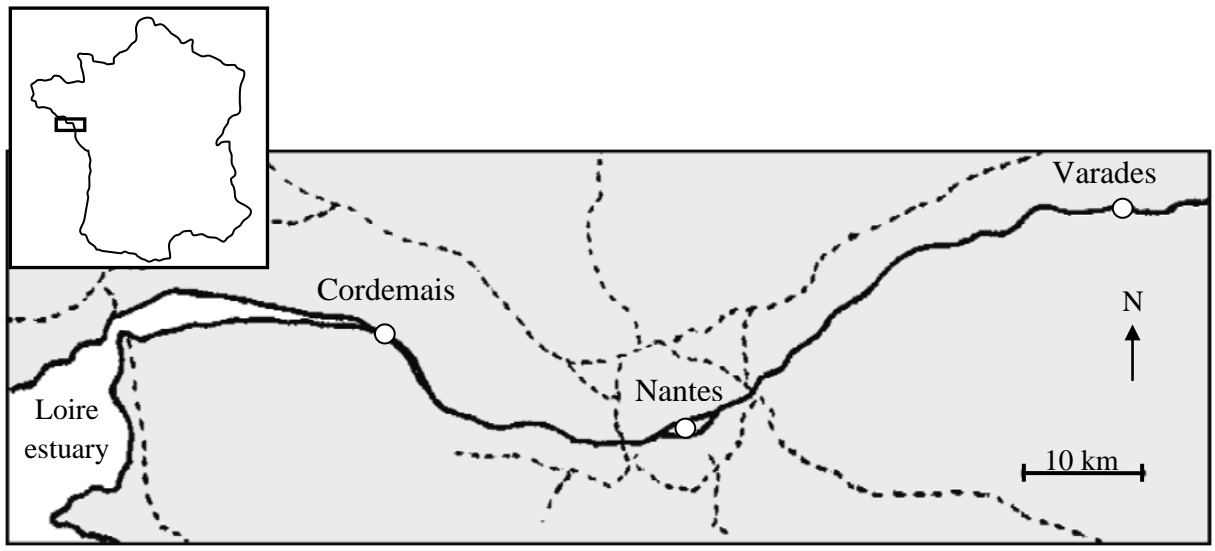


Figure 1

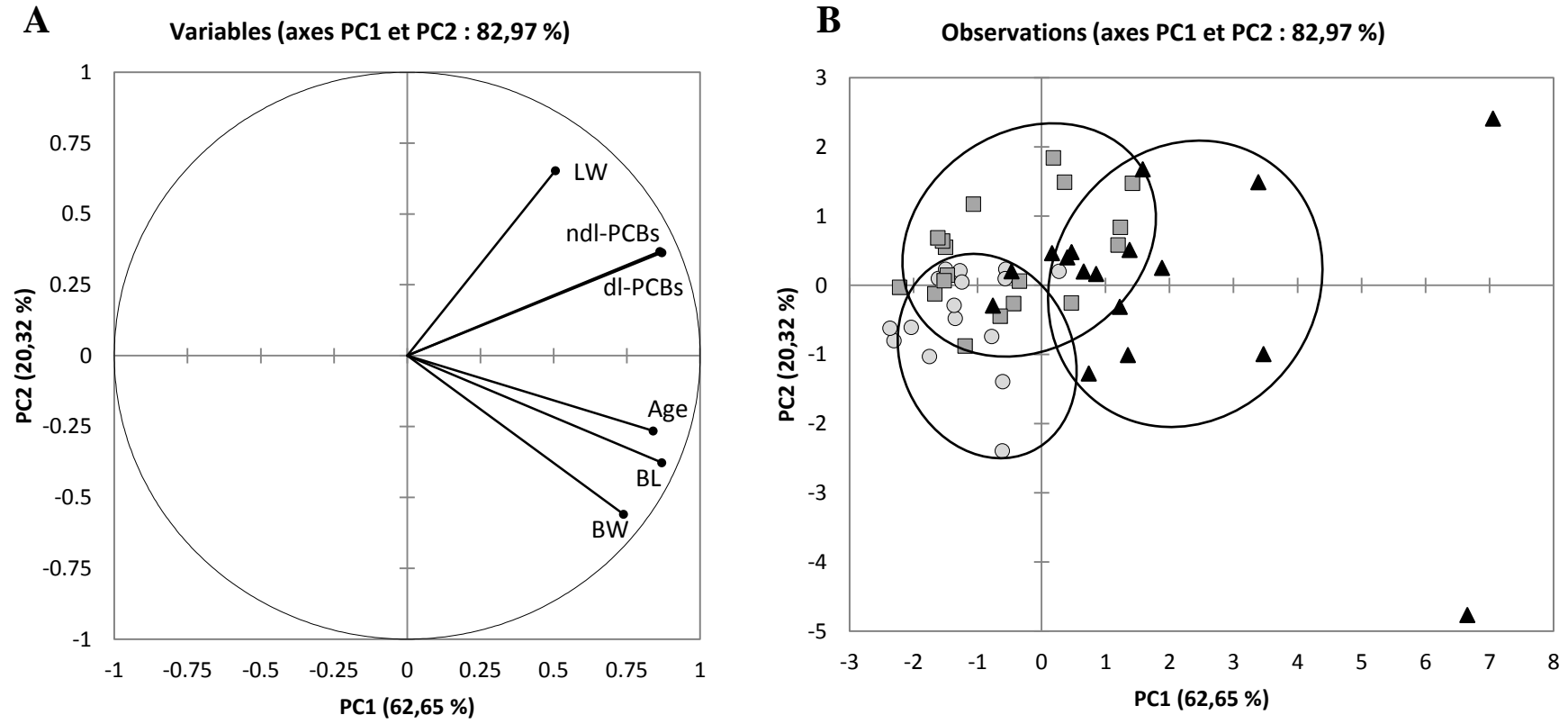


Figure 2

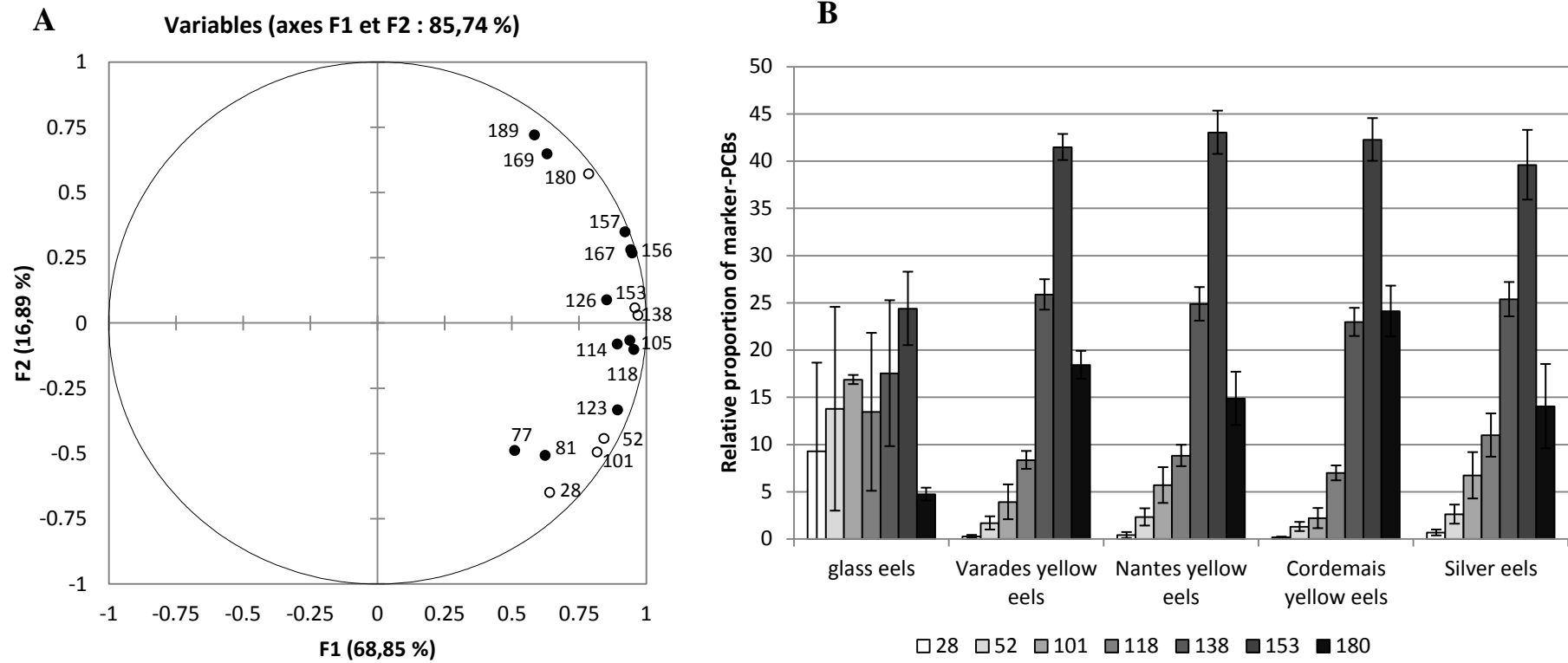


Figure 3

AUTHOR DECLARATION

We wish to confirm that there are no known conflicts of interest associated with this publication entitled " Dioxin-like, non-dioxin like PCB and PCDD/F contamination in European eel (*Anguilla anguilla*) from the Loire estuarine continuum: spatial and biological variabilities" and there has been no significant financial support for this work that could have influenced its outcome.

We confirm that the manuscript has been read and approved by all named authors and that there are no other persons who satisfied the criteria for authorship but are not listed. We further confirm that the order of authors listed in the manuscript has been approved by all of us.



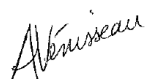

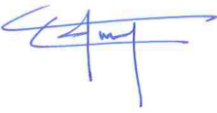



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We further confirm that any aspect of the work covered in this manuscript that has involved experimental animals has been conducted with the ethical approval of all relevant bodies and that such approvals are acknowledged within the manuscript.

We understand that the Corresponding Author is the sole contact for the Editorial process (including Editorial Manager and direct communications with the office). She is responsible for communicating with the other authors about progress, submissions of revisions and final approval of proofs.

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