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A rapid spectrophotometric method for quantitative determination of lactulose in heated milk and milk products

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(Received 20 August 1990; accepted 20 February 1991)

Summary — The reddish-brown colour produced on heating protein-free filtrate of heated milks containing less than 1.0% lactose and free of ions in 6.4 N sulphuric acid was used for quantitative determination of lactulose. The absorptivity of the coloured solution of lactulose at 330 nm was 14-fold more than that obtained from 62.5-fold more concentration of lactose and other aldoses. By following the optimum conditions established in this method, a minimum of 10 mg/100 ml concentration of lactulose can be determined in high heated unsweetened milk and milk products. The method was found to be very useful and sufficiently sensitive under dairy industry conditions where the ion removed step can be omitted if raw milk filtrate is taken for blank preparation.

Résumé — Méthode rapide de dosage par spectrophotométrie du lactulose dans le lait et les produits laitiers chauffés. La méthode de dosage du lactulose proposée est basée sur sa propriété à produire une coloration rouge brune au cours du chauffage (100 °C – 20 min) en milieu sulfurique (6,4 N). L’absorption de la solution de lactulose colorée à 330 nm est 14 fois supérieure à celle d’une solution de lactose et d’autres aldoses dont la concentration est 62,5 fois supérieure. Cette méthode permet, en suivant les conditions optimales établies, de déterminer une concentration minimale de 10 mg/100 ml de lactulose dans des laits et produits laitiers non sucrés ayant subi un fort traitement thermique. Elle est en outre très utile et suffisamment sensible à l’échelle industrielle où l’étape de désionisation peut être supprimée si le filtrat de lait cru est pris comme blanc.

lactulose / carbohydrate / heat treatment / dried milk / UHT milk

lactulose /hydrate de carbone / traitement thermique / poudre de lait / lait UHT
INTRODUCTION

Lactulose (4-0-β-D-galactopyranosyl-β-fructofuranose) is formed in heated milks by the alkaline isomerization of lactose catalysed by the free amino group of casein (Richards and Chandrasekhar, 1960; Adachi and Patton, 1961). Lactulose occurs in 2 forms: as lactulose in free solution, and as N-deoxy lactulosyl-L-lysine which is covalently bound to the amino group of the milk proteins (Henry et al, 1948).

The formation of lactulose in heated milks is dependent on time and temperature of heating and is also significantly affected by the pH of the environment (Adachi, 1959; Martinez-Castro and Olano, 1980). The lactose concentration in raw milk plays an important role in the formation of lactulose (Andrews, 1989).

Several methods have been used for the determination and quantification of lactulose in milk and milk products. The major problem with quantification of lactulose arises from the fact that the concentration of lactose is far in excess of that in milk and milk products. Most of the methods therefore include preliminary separation of carbohydrate mixtures or application of a specific aldose reaction to remove the lactose and thereafter quantification of lactulose either by a general carbohydrate method or by TLC, GLC or HPLC methods. Such methods are not only cumbersome and expensive, but also time-consuming when a large number of samples have to be analysed.

The spectrophotometric method of Adachi (1965) utilizes a lengthy procedure of oxidation of aldoses to aldonic acid by hypoiodate, whereas the anthrone method of Zagrodzki et al (1968) is reported to give slightly higher values as anthrone reacts with 5-hydroxymethylfurfural. The methylamine method of Nikerson et al (1976) is not very useful in milk where the lactose content is much higher than that of lactulose. The spectrophotometric method of Parrish et al (1980) is time-consuming and requires separate determination of all carbohydrates.

This paper describes a simple and rapid spectrophotometric method which does not require separation of lactulose from lactose or other aldoses and can be conveniently applied for quantification of lactulose in unsweetened milk and milk products.

MATERIALS AND METHODS

The principle of this method is based on the fact that ketoses are decomposed far more easily than aldoses by the action of mineral acids which produces an intense red or violet colouration, while the colour appears relatively slowly and is less intense when mineral acids act upon aldoses (Staněk et al, 1963). As heated milks contain no ketose other than lactulose (Gordon et al, 1956), the principle applied in this method is specific and consistent.

Materials

AR grade lactulose (98.0%) was obtained from Sigma Chemical Co, USA. 6.4 N H₂SO₄ (AR grade) and 40.0% TCA (AR grade) solution were freshly prepared before use. Amberlite IR-120 and Amberlite IR-458 (Sigma Chemical) were used as cation and anion exchanger respectively. HCl 3 N and NaOH 2 N (AR grade) were used for washing the column.

Standard methods used for comparative study

Lactulose was separated by TLC method of Martinez-Castro and Olano (1981) followed by quantification by the method of Adachi (1965) and cysteine-hydrochloride-carbazole colorimet-
The volume of the effluent was adjusted to 100 ml by removing excess water under vacuum at 65–70 °C.

**Development of colour**

Five ml of the above effluent was taken into a Pyrex 15 x 1.5 cm-test tube fitted with a glass stopper treated with 5.0 ml of 6.4 N H₂SO₄ and placed into a constantly boiling water bath (100 ± 0.5 °C) for 20 min. The tube was cooled immediately to 37 ± 2 °C and absorbance was measured within 30 min at 330 nm by a spectrophotometer (Spectronic 20D) against a blank prepared in a similar manner but with 5 ml of 40% TCA solution in place of the heated milk filtrate. For dried unsweetened milk powders, 1.0 g of the sample was dissolved in 10 ml of warm distilled water, diluted to 100 ml with 40% TCA solution (dilution coefficient 10⁻²) and the above procedure was followed for quantification.

**Calibration curve**

The calibration curve for lactulose was prepared by taking standard lactulose solution (conc 0.2–1.2 mg/5 ml) and developing the colour in a similar manner as for the milk filtrate.

**RESULTS AND DISCUSSION**

The action of mineral acids on sugars led to a number of decomposition products, some of which were coloured (Staněk et al., 1963). Besides a certain amount of humus-type decomposition, both aldohexoses and ketohexoses yield HMF as the main intermediate, which on further decomposition with acid produces furfural, a colouring substance.

**Absorbance spectra (λ_max)**

Absorbance spectra of standard lactulose solution (conc 2.92 x 10⁻³ mol/l) was ex-
examined in the range of 310–345 nm wavelength by a Spectronic 20 D spectrophotometer (fig 1). The absorbance maxima ($\lambda_{max}$) was slightly broad and noted at 330 nm of $0.616 \times 10^2$ mol/l absorptivity.

**Effect of concentration of acid on colour development**

The effect of the strength of sulphuric acid on lactulose and other aldoses after heating 20 min in a constantly boiling water bath ($100 \pm 0.5 ^\circ C$) on colour development is depicted in figure 2. It was observed that up to an acid concentration of 6.4 N, colour development for 1.0% lactose, glucose and galactose solution was significantly less (OD ≤ 0.01), whereas 0.016% solution of lactulose gave nearly 14 times more colouration (OD = 0.14). This reveals that the lactulose concentration of 62.5 times less than lactose and other aldoses produces 14 times more colouration with sufficient specificity.

**Effect of the length of heating time on colour development**

Rapid colour development was observed in the initial 20 min of heating of lactulose solution in 6.4 N H$_2$SO$_4$, whereas the colour development for other aldoses was less (≤ 0.012 OD) than that for lactulose for a similar extent of heat treatment (fig 3). Lactulose concentration of 0.02% produces 14 times more colouration (OD = 0.178)

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**Fig 1.** Absorption spectra of lactulose ($2.92 \times 10^{-3}$ mol/l) in 6.4 N H$_2$SO$_4$ at different wavelengths.

**Fig 2.** Effect of strength of H$_2$SO$_4$ on the colour development of lactulose and other aldoses. **a:** lactulose (0.016%), **b:** lactose (1.0%), **c:** glucose (1.0%), **d:** galactose (1.0%).
Fig 3. Effect of length of heating time on the colour development of lactulose and other aldoses. a: lactulose (0.02%), b: lactose (1.0%), c: glucose (1.0%), d: galactose (1.0%).

Effet de la durée du temps de chauffage sur l'intensité de la coloration produite par le lactulose et d'autres aldoses. a: lactulose (0,02%), b: lactose (1,0%), c: glucose (1,0%), d: galactose (1,0%).

Calibration curve for lactulose

The plot of absorbance versus lactulose concentration (0.2–1.2 mg/5 ml) was linear at 330 nm and obeyed the Beer-Lambert's law in this range. The concentration of the acid used and the time of heating were important determinant factors in the linearity of the curve. The higher the acid concentration and the time of heating, the greater the irregularities in the colour development observed. The regression equation developed for lactulose concentration ($Y = mg$ lactulose/5 ml) and absorbance ($X = observed$ optical density) was calculated as:

$$Y = 28.50X - 0.10$$

Colour contribution of different aldoses in relation to lactulose equivalent

Table I shows the relative colour development of lactose, glucose and galactose at

Table I. Colour contribution of different aldoses with respect to lactulose equivalent. Absorption à 330 nm du lactose du glucose et du galactose en fonction de leur concentration dans le filtrat.

<table>
<thead>
<tr>
<th>Concentration (%) in the filtrate</th>
<th>Lactose</th>
<th>Interfering Aldoses</th>
<th>Glucose</th>
<th>Galactose</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.50</td>
<td>0.000</td>
<td>0.000 (0.00)</td>
<td>0.000</td>
<td>0.000 (0.00)</td>
</tr>
<tr>
<td>0.75</td>
<td>0.001</td>
<td>0.000 (0.00)</td>
<td>0.000</td>
<td>0.001 (0.00)</td>
</tr>
<tr>
<td>1.00</td>
<td>0.004</td>
<td>0.002 (0.00)</td>
<td>0.003</td>
<td>0.000 (0.00)</td>
</tr>
<tr>
<td>1.25</td>
<td>0.006</td>
<td>0.004 (0.18)</td>
<td>0.007</td>
<td>0.007 (1.89)</td>
</tr>
<tr>
<td>1.50</td>
<td>0.018</td>
<td>0.009 (3.03)</td>
<td>0.016</td>
<td>0.016 (7.02)</td>
</tr>
<tr>
<td>2.00</td>
<td>0.039</td>
<td>0.021 (9.89)</td>
<td>0.029</td>
<td>0.029 (14.43)</td>
</tr>
</tbody>
</table>

* Figures in parentheses are mg equivalent lactulose per 100 ml of filtrate.
Les chiffres entre parenthèses représentent des mg d'équivalent lactulose pour 100 ml de filtrat.
Table II. Effect of interfering aldoses on colour contribution at the same absorbance value. 
Poids relatif des différents aldoses pour une même valeur d'absorption.

<table>
<thead>
<tr>
<th>Lactulose (mg/100 ml filtrate)</th>
<th>Absorbance at 330 nm</th>
<th>Relative weight equivalent of aldoses (mg/100 ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Lactose</td>
</tr>
<tr>
<td>10</td>
<td>0.02</td>
<td>-</td>
</tr>
<tr>
<td>20</td>
<td>0.04</td>
<td>2000</td>
</tr>
</tbody>
</table>

Effect of anion exchange resin on reducing sugars and raw milk filtrate

The effect of anion exchange resin on reducing sugars is presented in Table V. The result shows that on passing 1% lactose solution through the anion exchange resin there was a negligible increase in optical density at 330 nm. The initial value of 0.010 increased to 0.013 on passing 1% lactose solution, while the initial value of 0.181 dropped slightly to 0.174 after passing the lactulose solution (conc 1 mg/ml) through the anion exchange resin. At the same time when the raw milk filtrate was various concentration in relation to mg equivalent lactulose/100 ml of milk filtrate. It is evident from the table that 1.0% of lactose, galactose and glucose in milk filtrate contribute to the similar colour intensity of $= 0.18, 0.00$ and $0.00$ mg of lactulose per 100 ml in the respective milk filtrates. This clearly reveals that the intensity of colour contribution by aldoses is much less than that of the colour developed by lactulose even up to the 1.0% level. Table II represents the colour contribution of mg equivalent weight of lactulose in comparison to the mg equivalent of aldoses. It is evident from the results that $= 100, 180$ and $200$ times weight equivalent of lactose, galactose and glucose contribute a similar intensity of colouration to that of lactulose on following the optimum conditions prescribed in this method.

Recovery studies

Recovery studies were carried out by adding standard lactulose solution in heated milk and milk filtrate (in case of milk products). The results obtained are presented in table III. Approximately 97–101% recovery was observed, indicating high accuracy and reproducibility of the method developed.

<table>
<thead>
<tr>
<th>Lactulose added (mg/5 ml)</th>
<th>Lactulose recovered (mg/5 ml)</th>
<th>% Recovered</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.0</td>
<td>0.98</td>
<td>97.79</td>
</tr>
<tr>
<td>2.0</td>
<td>2.03</td>
<td>101.61</td>
</tr>
<tr>
<td>3.0</td>
<td>3.00</td>
<td>100.04</td>
</tr>
<tr>
<td>4.0</td>
<td>3.91</td>
<td>97.83</td>
</tr>
</tbody>
</table>
Table IV. Lactulose content of heated milk and milk products – comparison of 3 different methods. 
*Teneur en lactulose de laits et produits laitiers chauffés. Comparaison de 3 méthodes différentes.*

<table>
<thead>
<tr>
<th>Sample</th>
<th>TLC method</th>
<th>Cysteine - HCl - carbazole method</th>
<th>H₂SO₄ method</th>
</tr>
</thead>
<tbody>
<tr>
<td>Boiled milk (10 min)</td>
<td>84.24 (2.50)*</td>
<td>83.02 (2.75)</td>
<td>85.40 (2.55)</td>
</tr>
<tr>
<td>Sterilized milk (15 min)</td>
<td>166.77 (2.22)</td>
<td>162.60 (1.80)</td>
<td>165.24 (2.08)</td>
</tr>
<tr>
<td>UHT milk (indirect type)</td>
<td>58.60 (1.80)</td>
<td>57.24 (1.35)</td>
<td>56.04 (1.56)</td>
</tr>
<tr>
<td>SMP (roller-dried)</td>
<td>478.28 (2.90)</td>
<td>474.30 (2.75)</td>
<td>479.80 (2.90)</td>
</tr>
<tr>
<td>SMP (spray-dried)</td>
<td>266.17 (2.05)</td>
<td>261.40 (2.25)</td>
<td>268.34 (2.60)</td>
</tr>
</tbody>
</table>

* Figures in parentheses are standard deviations.

Table V. Effect of anion exchange resin on reducing sugars (lactose and lactulose) and raw milk filtrate. 
*Effet de la résine échangeuse d’anions sur l’absorption des sucres réducteurs (lactose et lactulose) et du filtrat de lait cru.*

<table>
<thead>
<tr>
<th>Standard lactose solution (1%)</th>
<th>Raw milk filtrate</th>
<th>Standard lactose solution (conc 1 mg/ml)</th>
<th>Added lactulose (mg/5 ml)</th>
<th>Practical Determined</th>
<th>% loss</th>
</tr>
</thead>
<tbody>
<tr>
<td>Untreated</td>
<td>Treated</td>
<td>Untreated Treated</td>
<td>Untreated Treated</td>
<td>Practical Determined</td>
<td>% loss</td>
</tr>
<tr>
<td>0.010</td>
<td>0.010</td>
<td>0.155</td>
<td>0.030</td>
<td>0.185</td>
<td>0.178</td>
</tr>
<tr>
<td>0.005</td>
<td>0.010</td>
<td>0.160</td>
<td>0.035</td>
<td>0.178</td>
<td>0.170</td>
</tr>
<tr>
<td>0.015</td>
<td>0.020</td>
<td>0.145</td>
<td>0.020</td>
<td>0.160</td>
<td>0.175</td>
</tr>
<tr>
<td>Mean</td>
<td>0.010</td>
<td>0.153</td>
<td>0.028</td>
<td>0.181</td>
<td>0.174</td>
</tr>
</tbody>
</table>

Passed through the anion exchanger, the initial value of 0.153 dropped sharply to 0.028 due to removal of interfering ions and the subsequent colour development was very insignificant in contributing to lactulose value. This reveals that neither were the reducing sugars adsorbed nor was the lactose isomerized to lactulose while passing through the anion exchanger. Turton and Pacsu (1955) observed a negligible change in mutarotation and in the nature of the sugar solution (2%) when passed through an anion exchange resin IRA-400-OH. However, they observed a rapid adsorption of D-glucose, D-fructose and D-mannose in the hydroxide form of resin; D-fructose was transformed to D-psicose, but no evidence was found of changes in reducing sugars like lactose or lactulose. Adachi (1965) reported no adsorption or
isomerization of lactulose while passing it through strong cation and anion exchange resins for a long time to remove interfering ions and thereafter successfully determined lactulose content spectrophotometrically. Our findings also showed a negligible adsorption of lactulose and no isomerisation of lactose to lactulose while passing through the anion exchange resin. This might be due to the strong acidic environment present in resin provided by the milk filtrate containing 40% TCA solution.

Omission of the ion exchange step: effect of changes in absorbance with raw milk filtrate and addition of lactulose solution

To study the efficacy of the method by omitting the ion exchange step, raw milk filtrate replaced heated milk filtrate and the lactulose solution was added at an increasing concentration, (fig 4) with the same quantity of raw milk filtrate to make up the total volume to 5 ml. Both the systems were treated in a similar manner for colour development as depicted in figures 5 and 6. The linearity of both the slopes curves reveals that they obeyed the Beer-Lambert's law accurately in the given concentration range of lactulose and raw milk filtrate. This signifies the sensitivity and accuracy of the method, provided the raw milk filtrate is used in place of 40% TCA solution needed for the preparation of the blank.

Efficacy ln the determination of lactulose content by using raw milk filtrate as blank

Table VI shows the values of lactulose concentration added and determined in raw milk filtrate when the same raw milk filtrate concentrations were used for blank preparation. The results reveal an accura-

![Fig 4. Calibration curve for lactulose (conc 1.16 x 10^{-4} to 7.01 x 10^{-4} mol/l).](attachment:fig4.png)

*Fig 4. Calibration curve for lactulose (conc 1.16 x 10^{-4} to 7.01 x 10^{-4} mol/l).*  
*Courbe de calibration pour le lactulose (conc 1,16 x 10^{-4} - 7,01 x 10^{-4} mol/l).*

![Fig 5. Absorbance of raw milk filtrate.](attachment:fig5.png)

*Fig 5. Absorbance of raw milk filtrate.*  
*Absorption de filtrat de lait cru.*
Spectrophotometric determination of lactulose

Fig 6. Absorbance of lactulose solution heated with raw milk filtrate. Absorption de la solution de lactulose chauffée avec un filtrat de lait cru.

Precision of 101.85% (mean value) in determining the lactulose concentration when raw milk filtrate was taken for blank preparation instead of 40% TCA solution. Slightly higher values may be attributed towards the effect of interfering ions, lactose and aldoses, which might have some influence on colour development in this context.

**Comparison with standard methods**

The method developed was compared with the procedure described by Martinez-Castro and Olano (1981) and the cysteine-hydrochloride-carbazole method of Fujita and Iwatake (1931). The results obtained (table IV) by the method developed were comparable, consistent and reproducible with sufficient accuracy (standard deviation of 2.9) as compared to the other 2 methods. The slightly higher values obtained for milk powders may be attributed to the higher pre-heating temperatures followed under Indian conditions.

**CONCLUSIONS**

The rapid and simple colorimetric method developed can successfully be performed within 2.5 to 3 h for routine quantitative determination of lactulose in unsweetened raw milk.

**Table VI.** Lactulose content in raw milk filtrate, when raw milk filtrate was taken for blank preparation. Détermination des teneurs en lactulose ajouté dans un filtrat de lait cru (filtrat de lait cru pris comme blanc).

<table>
<thead>
<tr>
<th>Vol raw milk filtrate taken (ml)</th>
<th>Vol lactulose solution added (ml)</th>
<th>Lactulose content (mg/5 ml)</th>
<th>% Accuracy (b/a x 100)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Added (a)</td>
<td>Determined (b)</td>
</tr>
<tr>
<td>1</td>
<td>4</td>
<td>16</td>
<td>16.28</td>
</tr>
<tr>
<td>2</td>
<td>3</td>
<td>12</td>
<td>12.44</td>
</tr>
<tr>
<td>3</td>
<td>2</td>
<td>8</td>
<td>8.28</td>
</tr>
<tr>
<td>4</td>
<td>1</td>
<td>4</td>
<td>3.94</td>
</tr>
<tr>
<td>5</td>
<td>0</td>
<td>0</td>
<td>0.24</td>
</tr>
</tbody>
</table>

Mean 101.85
heated milk and milk products, and is also highly suitable under dairy industry conditions where raw milk is readily available. The ion exchange step can be omitted provided that the raw milk filtrate is taken for blank preparation and the colour developed in a similar manner to the sample. Hence a large number of samples can conveniently be analysed for their lactulose content to determine the severity of heat treatment with sufficient accuracy under dairy industry conditions.

REFERENCES