

Characterization and evolution of a family of integrative and potentially conjugative or mobilizable elements from Streptococcus thermophilus

Guillaume Pavlovic, Vincent Burrus, Alexandre Toulmay, Frédéric Choulet, Bernard Decaris, Gérard Guédon

▶ To cite this version:

Guillaume Pavlovic, Vincent Burrus, Alexandre Toulmay, Frédéric Choulet, Bernard Decaris, et al.. Characterization and evolution of a family of integrative and potentially conjugative or mobilizable elements from Streptococcus thermophilus. Le Lait, 2004, 84 (1-2), pp.7-14. 10.1051/lait:2003042. hal-00895527

HAL Id: hal-00895527

https://hal.science/hal-00895527

Submitted on 11 May 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Review

Characterization and evolution of a family of integrative and potentially conjugative or mobilizable elements from *Streptococcus thermophilus*

Guillaume PAVLOVIC, Vincent BURRUS, Alexandre TOULMAY, Frédéric CHOULET, Bernard DECARIS, Gérard GUÉDON*

Laboratoire de Génétique et Microbiologie, UMR INRA 1128, IFR110, Université Henri Poincaré - Nancy 1, Bd des Aiguillettes, BP 239, 54506 Vandœuvre-lès-Nancy, France

Published online 12 December 2003

Abstract – The integrative and conjugative elements (ICEs) excise by site-specific recombination, self-transfer the resulting circular form by conjugation and integrate into the genome of the recipient bacterium. The 34.7-kb element from Streptococcus thermophilus CNRZ368, ICESt1, excises and integrates by site-specific recombination. This element also possesses a conjugation module distantly related to that of the conjugative transposon Tn916 from Enterococcus faecalis. Therefore, ICESt1 is probably an ICE. Four types of elements related to ICESt1 are integrated into the same location as ICESt1 in seven other strains of S. thermophilus. One of these elements, ICESt3, is probably an ICE whereas the three others (CIMEs) would have arisen from ICEs by deletion of the conjugation and recombination modules. These elements also encode functions that are not involved in element maintenance or transfer, such as restriction-modification systems. Each of these elements has a chimerical structure resulting from the acquisition of modules from different origins. Sequence analyses indicate that these elements are involved in horizontal transfers with various species of dairy or pathogenic lactic acid bacteria. Δ CIME308 has exchanged restrictionmodification and cadmium resistance modules with plasmids. CIME19258 has acquired a cadmium resistance module by the integration of an ICE related to Tn916 within the CIME. The site-specific recombination between an internal attL-related site and attR of ICESt1 leads to the excision of a circular molecule which could be another ICE, ICESt2, suggesting that ICESt1 has arisen by accretion of a CIME and ICESt2. CIME302, ICESt2 and ICESt3 would also have arisen by sitespecific accretion of CIMEs and ICEs and/or mobilization of CIMEs by ICEs. An ICE would integrate by site-specific recombination in the attR site of a CIME; then the CIME-ICE would excise by site-specific recombination and transfer by conjugation.

 $Site-specific \ recombination \ / \ conjugative \ transposon \ / \ modular \ evolution \ / \ horizontal \ transfer \ / \ genomic \ island$

Résumé – Structure et évolution d'une famille d'éléments intégratifs potentiellement conjugatifs ou mobilisables de *Streptococcus thermophilus*. Les éléments intégratifs conjugatifs (ICEs) s'excisent par recombinaison site-spécifique sous forme circulaire, se transfèrent par conjugaison puis s'intègrent dans la bactérie réceptrice. L'élément ICE*St1* de *Streptococcus thermophilus* CNRZ368 (34,7 kb) s'intègre et s'excise par recombinaison site-spécifique. Par ailleurs, il code un

^{*} Corresponding author: guedon@nancy.inra.fr

système de conjugaison putatif apparenté de facon lointaine à celui de Tn916 d'Enterococcus faecalis et constitue donc un ICE putatif. Quatre éléments apparentés à ICESt1 sont intégrés au même site que lui chez sept souches de S. thermophilus. L'un des éléments, ICESt3 est probablement un ICE. Les trois autres (CIMEs) dériveraient d'un ICE par délétion des modules de conjugaison et de recombinaison. Ces éléments codent des fonctions non impliquées dans le transfert comme des systèmes de restriction-modification. Chacun d'entre eux présente une structure chimérique résultant de l'acquisition de modules d'origines différentes. L'analyse de leur séquence indique qu'ils sont impliqués dans des transferts horizontaux entre S. thermophilus et diverses espèces de bactéries lactiques alimentaires ou pathogènes. \(\Delta \text{CIME} 308 \) a \(\text{échangé des modules de restriction-modification} \) et de résistance aux ions Cd⁺⁺ avec des plasmides. CIME19258 a acquis un module de résistance aux ions Cd++ par intégration d'un ICE apparenté à Tn916 dans un CIME. La recombinaison sitespécifique entre un site interne d'ICESt1 de type attL et le site attR provoque l'excision de la partie droite d'ICESt1 qui constituerait elle-même un ICE, ICESt2. ICESt1 serait ainsi apparu par accrétion d'un CIME et d'ICESt2. CIME302, ICESt2 et ICESt3 seraient également apparus par accrétion site-spécifique d'ICEs et de CIMEs et/ou mobilisation de CIMEs par des ICEs. Un ICE s'intégrerait par recombinaison site-spécifique dans le site attR d'un CIME formant ainsi un élément composite (CIME-ICE) par accrétion. Celui-ci s'exciserait par recombinaison site-spécifique puis se transférerait par conjugaison.

Recombinaison site-spécifique / transposon conjugatif / évolution modulaire / transfert horizontal / îlot génomique

1. INTRODUCTION

Horizontal gene transfer between strains and species constitutes a major driving force in the evolution of bacteria [6, 9]. In this way, pathogenic bacteria have acquired antibiotic resistance genes by horizontal transfer of various plasmids or transposons and the acquisition of pathogenicity islands, prophages and/or plasmids has turned non-pathogenic strains from various species into virulent strains. Sequence comparisons revealed that multiple horizontal transfers have recently occurred between S. thermophilus and other lactic acid bacteria such as Lactococcus lactis, probably in co-cultures from cheese manufacture [5].

2. CHARACTERIZATION OF AN INTEGRATIVE AND POTENTIALLY CONJUGATIVE ELEMENT, ICEST1

2.1. Characterization of a site-specific integrative element

Map comparison and probe hybridizations showed that the chromosome of S. thermophilus CNRZ368 contains a 34.7kb region, ICESt1, which is absent in its close relative A054 [1]. ICESt1 is flanked by a 27-bp direct repeat whereas only one copy of this sequence was found in the corresponding region of A054. The right 27-bp sequence of ICESt1 and the unique 27-bp sequence of A054 include the 3' end of fda, which encodes a putative fructose-1,6diphosphate aldolase. The ICESt1 right end (Fig. 1) codes for the protein Xis which shares 37-41% identity with the putative excisionase of the conjugative transposons Tn5252 from S. pneumoniae and Tn5276 from L. lactis [1]. It also encodes an integrase (Int) related to those of Tn5252, Tn5276 and numerous temperate phages of lactic acid bacteria [1].

The site-specific recombination between the two 27-bp sequences, included in the attachment sites *attL* and *attR*, generates a chromosomal integration site, *attB*, identical to that found in A054 and a recombination site, *attI*, carried by a circular molecule corresponding to an excised form of ICESt1 [1]. *attB* and *attI* were detected by PCR in CNRZ368. The PCR products carrying *attB* or *attI* were not obtained when the *int* gene of ICESt1 was disrupted by the integration

of a thermosensitive plasmid. The recombination module (i.e. xis-int-attI) and the integration module (i.e. int-attI) were cloned downstream from a promoter on the thermosensitive plasmid pG+host9. These plasmids integrate by site-specific recombination between their attI site and the chromosomal attB site. After integration, the plasmid containing the recombination module can excise by recombination between attL and attR, whereas the plasmid containing the integration module does not excise. Therefore, site-specific excision is catalyzed by both the integrase and excisionase, whereas the integrase but not the excisionase is needed for site-specific integration.

2.2. ICESt1 carries a putative conjugation module

Seven putative proteins encoded by the right part of ICESt1 are related to proteins encoded by conjugation modules of some elements from low G+C Gram-positive bacteria [4]. orfA codes for a protein related to a transfer protein encoded by the staphylococcal conjugative plasmid, pSK41. The putative products of six ORFs (orfC, orfD, orfE, orfG, orfJ and orfK) are related to transfer proteins encoded by the conjugative transposon Tn916 from Enterococcus faecalis [4]. Furthermore, the right part of ICESt1 also encodes four putative proteins (OrfF, OrfH, OrfL and OrfM) related to proteins encoded by the conjugation modules of putative conjugative elements which are integrated into the completely sequenced chromosome from various low G+C Gram-positive bacteria (data not shown). Therefore, the right region of ICESt1 is likely to carry a conjugation module. However, the translation products of two small ORFs belonging to this module, orfB and orfI, are not significantly related to known or putative proteins.

Various elements, which excise by sitespecific recombination, self-transfer the resulting circular form by conjugation and integrate by recombination between a specific site of this circular form and a site in the genome of their host, have been found in bacteria. All these elements were recently proposed to be grouped in Integrative and Conjugative Elements (ICEs) irrespective of the mechanisms of conjugative transfer or of integration [3, 4]. Therefore, ICESt1 is an ICE or derives from an ICE.

3. IDENTIFICATION AND STRUCTURE OF ELEMENTS RELATED TO ICEST1

Elements related to ICESt1 and/or integrated into the same location were searched for in twenty strains of *S. thermophilus* by hybridization with probes corresponding to the various regions of ICESt1 and by amplifications of the *fda* locus. Four types of elements were found in seven strains. All are integrated into exactly in the same site as ICESt1.

ICESt3 (28.1 kb) has a recombination module (including attL and attR) almost identical to that of ICESt1 (Fig. 1) and excises by site-specific recombination. Furthermore, it carries putative conjugative and regulation modules that are almost identical or closely related to those of ICESt1. Therefore, this element could be an ICE. The three other elements do not carry conjugation modules and their regulation and recombination modules are truncated (Fig. 1). Therefore, they are neither integrative nor conjugative. The sequence comparisons suggest that these three elements derive from ICEs closely related to ICESt1 and ICESt3 by deletion of these modules. Sequences related to attL and attR of ICESt1 were found to flank two of these elements, that were named CIMEs for CIs Mobilizable Elements, CIME19258 (15.0 kb) and CIME302 (13.1 kb). Another element, Δ CIME308 (16.8 kb), carries an *attR* site (Fig. 1) and a truncated integrase gene but does not harbor an attL site. Sequence comparisons suggest that it derives from a CIME by an IS1193-mediated deletion of its left end.

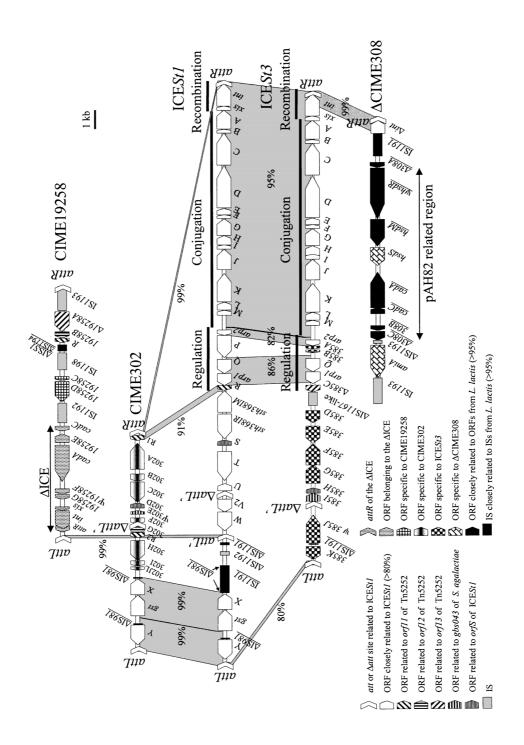


Figure 1. Comparison of ICESt1, ICESt3, CIME302, CIME19258 and ΔCIME308 from S. thermophilus. The location and orientation of ORFs, pseudogenes (ψ) or truncated ORFs (Δ) are indicated by arrowed boxes, complete or truncated ISs by rectangles, and complete or truncated att sites by arrowheads. Some very short truncated ORFs and the ORFs belonging to ISs are not indicated. The white boxes depict ORFs closely related to ORFs of ICESt1 (>80%). ORF names beginning with "orf" are abbreviated with the corresponding letter and/ or number. The gray areas join sequences that are closely related (>80% nucleotide identity); the identity percent is indicated. The close relationship between short non-coding internal regions or ISs is not displayed. The ORFs sharing 40–80% nucleotide identities are represented by boxes with identical symbols. The black boxes and rectangles correspond to ORFs and ISs almost identical to L. lactis sequences (>95% nucleotide identity); the names of these ORFs and ISs are underlined. The left limit of the ΔICE integrated into CIME19258 (corresponding to the right end of the DICE in the figure) cannot be defined exactly.

4. FUNCTIONS ENCODED BY ICES AND CIMES

ICEs and CIMEs carry modules that are not involved in the conjugation or the maintenance of the element. They encode phage resistance: ICESt1 encodes the restriction-modification (RM) system Sth368I (sth368IR and sth368IM) [2] and a putative abortive infection resistance mechanism (orfY), CIME302 a putative isoschizomer of the RM systems MvaI or BcnI (orf302A and orf302C), ICESt3 two methyltransferases which probably belong to an unknown putative RM system (orf385F and orf385G), and Δ CIME308 a type I RM system which is probably not functional (hsdS, hsdM and \psihsdR). Furthermore, ΔCIME308 encodes a putative binding subunit of an oligopeptide ABC carrier (amiA) and a putative cadmium resistance (cadA and cadC), CIME19258 a putative cadmium resistance (cadA and cadC), and ICESt1 and CIME302 a putative glutamate and/or aspartate carrier (gst). Some of these functions would be interesting for applied purposes. Therefore, conjugative transfers of ICEs or CIMEs could be useful for improving industrial strains.

5. ORIGIN OF MODULES AND MECHANISMS OF MODULE ACQUISITION

5.1. Exchanges with plasmids or unrelated ICEs and horizontal transfers

A large part of \triangle CIME308 (i.e. $\psi hsdR$, hsdM, cadA, cadC, orf308B and $\triangle orf308C$)

(Fig. 1) is almost identical to a region of the plasmid pAH82 from L. lactis [10]. Furthermore, some ORFs belonging to this region and the adjacent ORF $\Delta orf308A$ are almost identical to ORFs or genes found in other plasmids from L. lactis, S. thermophilus and Listeria innocua (data not shown). This suggests that genes were exchanged between CIMEs (or ICEs) and plasmids. The sequence analyses also reveal complete or truncated copies of insertion sequences (ISs) which are almost identical to complete or truncated ISs from other bacterial species. These species include lactic acid bacteria used in food fermentation such as *L. lactis* (IS1191, IS981, ISS1 α, ISS1 β and IS1194) or Leuconostoc mesenteroides (ISS α), pathogenic lactic acid bacteria such as Enterococcus faecalis (ISS1 a), Enterococcus hirae (IS1194) or Streptococcus pneumoniae (IS1194), and Listeria innocua (ISS1 α). Therefore, various sequences found in ICEs and CIMEs of S. thermophilus, and perhaps complete CIMEs or ICEs, have undergone horizontal transfers with other species of bacteria.

The region located at the right of the CIME19258 attL site is related (61% identity) to the recombination module of Tn916, including its attR site, its integrase gene, int, and excisionase gene, xis. The region located at the right of this module (orf19258G, \psi orf19258F, cadA, orf19258E and cadC) is related (83% identity) to the regulation module (lmo1098-lmo1099) and the cadmium resistance module (cadA-lmo1102) of a putative ICE found in the completely

sequenced genome of *Listeria monocytogenes* EGD-e (*lmo1097-lmo1115*), ICE*Lm1*. However, CIME19258 does not possess any sequences related to the conjugation modules of Tn916 or ICE*Lm1* and to their *attL* site. Therefore, CIME19258 bears a truncated ICE related to Tn916 and ICE*Lm1*. This suggests that CIME19258 has acquired a cadmium resistance module by the integration of an ICE within the CIME.

5.2. Evolution of ICEs and CIMEs by site-specific accretion

The *attR* sequences of ICE*St1* and CIME19258 (102 bp) share 67% identity, and their *attL* sequences (202 bp) 60%. A plasmid bearing the *xis* and *int* genes from ICE*St1* was introduced into the strain harboring CIME19258 (ATCC19258^T). In this recombinant strain, the *attB* and *attI* sites resulting from the excision of CIME19258 by site-specific recombination were detected by PCR, indicating that the *att* sites of CIME19258 are functional.

Furthermore, the CIME19258 attL sequence is almost identical (99% identity) to an internal sequence of ICESt1 (attL'). The site-specific recombination between attL' and attR of ICESt1 leads to the formation of a CIME flanked by two recombination sites (attL and attB') and to the excision of a 27 205-bp circular molecule carrying an attI' site. The sites resulting from the excision (i.e. attB' and attI') were detected by PCR in CNRZ368. The PCR products carrying attB' or attI' were not obtained when the int gene of ICESt1 was disrupted by the integration of a thermosensitive plasmid. The excised molecule includes the conjugation and recombination modules of ICESt1. Therefore, the sequences flanked by attL' and attR (i.e. the central and right regions of ICESt1) correspond to a novel putative ICE, ICESt2. This suggests that ICESt1 results from the accretion of an ICE and a CIME.

Furthermore, a truncated *attL*' internal site (71 bp, 54% identity with *attL* of

ICESt1) was also found in CIME302 (Fig. 1). This putative $\Delta attL$ ' site of CIME302 is associated with two ORFs, orfR2 and orf302G, which are related to the ORFs located at the left end of the complete or truncated regulation modules of ICESt1 (orfR), CIME302 (orfR1) and CIME19258 (orfR and orf19258B). This suggests that the region $orfR2-\Delta attL'$ results from the deletion of a region which initially included complete regulation, conjugation and recombination modules. Moreover, putative internal $\Delta attL$ ' sites are also found in ICESt2, i.e. in the putative ICE corresponding to the central and right regions of ICESt1, (71 bp, 69% identity) and in ICESt3 (44 bp, 93% identity). Therefore, ICESt1 and the related elements have evolved by accretions of ICEs and CIMEs.

In the proposed model (Fig. 2), some ICEs have lost their conjugation and recombination modules by deletion but have retained attL and attR, leading to CIMEs. Strains that harbor a CIME or an ICE (ICE1) have acquired a related ICE by conjugation (ICE2 in Fig. 2). Then, ICE2 integrated by recombination between its attI site and the attR site flanking the CIME (or of ICE1) leading to structures such as attL-CIME-attI'-ICE2-attR' (or attL-ICE1-attI'-ICE2-attR'). The resulting internal attI' site of the CIME-ICE2 structure or the regulation, conjugation and recombination modules (including the attI' site) of ICE1 found in the ICE1-ICE2 structure were partially deleted, leading to an attL' or a truncated $attL'(\Delta attL')$. The whole structure (ICE3 in Fig. 2) could excise by recombination between attL and attR' (generating attI'') and transfer to a new host bacterium, leading to composite ICEs such as ICESt1 or ICESt3. Alternatively, the regulation, conjugation and recombination modules could be deleted, leading to a composite CIME such as CIME302. In this model, the ICEs could acquire novel modules by the site-specific accretion of CIMEs and the ICEs could mobilize CIMEs.

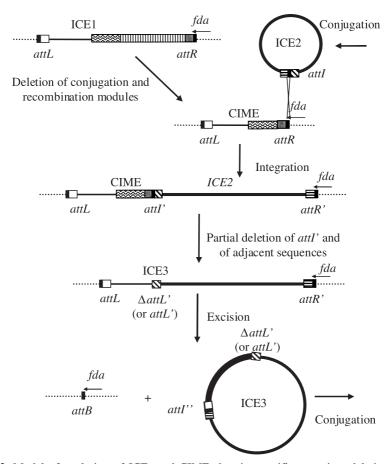


Figure 2. Model of evolution of ICEs and CIMEs by site-specific accretion, deletion and *cis*-mobilization. The black rectangles correspond to the 27-bp sequences (or the homologous sequences) found in all attachment sites of ICE*St1*-related elements. The deletion of the internal *att1* attachment site created by the site-specific accretion of the CIME and of the ICE can lead, not only to a functional *attL'*, but also to a truncated *attL'* as indicated in the figure or to the loss of the site (not shown). The *attL*-CIME-*attL'*-ICE-*attR'* could not only arise by partial deletion of the *att1'* from the structure *attL*-CIME-*att1'*-ICE-*attR'* but also by the tandem integration of two related ICEs followed by deletion of the regulation, conjugation and recombination modules (including a part of *att1'*) of the left ICE.

6. KNOWN ICES AND CIMES: THE TIP OF THE ICEBERG

Few ICEs have been described. However, a recent genomic search revealed 17 putative ICEs in completely or incompletely sequenced genomes from 23 various low G+C bacteria [4]. This suggests that

ICEs are widespread in bacteria. Therefore, like the conjugative plasmids and the prophages, ICEs could be one of the main types of elements responsible for horizontal gene transfer.

Elements which, like CIMEs, are related to ICEs, and lack conjugation and recombination modules but are flanked by

attL and attR, have not previously been reported. Various pathogenicity islands (PAIs) encode their own excision and integration by site-specific recombination but the mechanism of their transfer remains unknown. Other PAIs have lost the integrative functions by inactivation of the integrase by point mutation or deletion [6]. A PAI from Salmonella enterica is a putative ICE [11] and the putative ICE EfaD2 from Enterococcus faecalis is a PAI [3, 4, 12]. Some PAIs are flanked by direct repeats or are associated with tRNA genes but do not possess functional recombination modules. Like the CIMEs of S. thermophilus, some of them could derive from ICEs.

Accretion of CIMEs and ICEs has not previously been described. However, the ICE Tn5481 from *L. lactis* bears an internal 187-bp sequence sharing 88% identity with its *attL* site [8]. Furthermore, the ICE SXT from *Vibrio cholerae* was found to site-specifically integrate into the *attL* or *attR* sites of the related element R391 [7]. Therefore, site-specific accretion could be a major evolution mechanism of ICEs.

REFERENCES

- [1] Burrus V., Roussel Y., Decaris B., Guédon G., Characterization of a novel integrative element, ICESt1, in the lactic acid bacterium *Streptococcus thermophilus*, Appl. Environ. Microbiol. 66 (2000) 1749–1753.
- [2] Burrus V., Bontemps C., Decaris B., Guédon G., Characterization of a novel type II restriction-modification system, Sth368I, encoded by the integrative element ICESt1 of Streptococcus thermophilus CNRZ368,

- Appl. Environ. Microbiol. 67 (2001) 1522–1528.
- [3] Burrus V., Pavlovic G., Decaris B., Guédon G., Conjugative transposons: the tip of the iceberg, Mol. Microbiol. 46 (2002) 601–610.
- [4] Burrus V., Pavlovic G., Decaris B., Guédon G., The ICESt1 element of Streptococcus thermophilus belongs to a large family of integrative and conjugative elements that exchange modules and change their specificity of integration, Plasmid 48 (2002) 77.
- [5] Guédon G., Bourgoin F., Decaris B., Does gene horizontal transfer occur in lactic acid bacteria co-cultures?, Lait 78 (1998) 53–58.
- [6] Hacker J., Kaper J.B., Pathogenicity islands and the evolution of microbes, Annu. Rev. Microbiol. 54 (2000) 641–679.
- [7] Hochhut B., Beaber J.W., Woodgate R., Waldor M.K., Formation of chromosomal tandem arrays of the SXT element and R391, two conjugative chromosomally integrating elements that share an attachment site, J. Bacteriol. 183 (2001) 1124–1132.
- [8] Immonen T., Wahlstrom G., Takala T., Saris P.E., Evidence for a mosaic structure of the Tn5481 in Lactococcus lactis N8, DNA Seq. 9 (1998) 245–261.
- [9] Ochman H., Lawrence J.G., Groisman E.A., Lateral gene transfer and the nature of bacterial innovation, Nature 405 (2000) 299–304.
- [10] O'Sullivan D., Ross R.P., Twomey D.P., Fitzgerald G.F., Hill C., Coffey A., Naturally occurring lactococcal plasmid pAH90 links bacteriophage resistance and mobility functions to a food-grade selectable marker, Appl. Environ. Microbiol. 67 (2001) 929– 937
- [11] Pembroke J.T., MacMahon C., McGrath B., The role of conjugative transposons in the *Enterobacteriaceae*, Cell Mol. Life Sci. 59 (2002) 2055–2064.
- [12] Shankar N., Baghdayan A.S., Gilmore M.S., Modulation of virulence within a pathogenicity island in vancomycin-resistant *Entero*coccus faecalis, Nature 417 (2002) 746–750.