Growth enhancement of maize (Zea mays L) through
Azospirillum lipoferum inoculation: effect of plant
genotype and bacterial concentration
Jf Arsac, C Lamothe, D Mulard, Jacques Fages

To cite this version:
Jf Arsac, C Lamothe, D Mulard, Jacques Fages. Growth enhancement of maize (Zea mays L) through
Azospirillum lipoferum inoculation: effect of plant genotype and bacterial concentration. Agronomie,
EDP Sciences, 1990, 10 (8), pp.649-654. <hal-00885325>

HAL Id: hal-00885325
https://hal.archives-ouvertes.fr/hal-00885325
Submitted on 1 Jan 1990

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L’archive ouverte pluridisciplinaire HAL, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d’enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.
Plant pathology

Growth enhancement of maize (Zea mays L) through Azospirillum lipoferum inoculation: effect of plant genotype and bacterial concentration

JF Arsac, C Lamothe, D Mulard, J Fages*

Pioneer France-Maïs, chemin de l’Enseigure, 31840 Aussonne, France

(Received 9 March 1990; accepted 5 August 1990)

Summary — Germination tests and pot experiments were carried out to study the specificity between an Azospirillum lipoferum strain and Zea mays hybrids. In both types of trial differences between hybrids were exhibited, and the influence of the plant genotype was clearly demonstrated. The optimal level for bacterial concentration was found to be $10^7$ bacteria per plant and the importance of an early inoculation was shown. Pot experiments were less precise than germination tests. Such germination tests for screening plant varieties using the criterion of plant-growth promotion appeared to be a very useful tool.

Zea mays / Azospirillum lipoferum / germination tests / plant genotype / plant-growth promotion

INTRODUCTION

Rhizospheric bacteria of the genus Azospirillum are known for their ability to improve grain and dry matter yields of crop plants (Yahalom et al., 1984; Wani et al., 1985; Fayez and Daw, 1987; Tilak and Subba Rao, 1987; Sarig et al., 1988).

The mechanism by which Azospirillum contributes to plant growth has remained a matter of controversy for years. Nowadays it is known that associative N$_2$ fixation is often negligible in temperate regions and that additional processes should be considered in the beneficial effects observed (Michiels et al., 1989). An early effect of inoculation on root morphology and physiology has been shown: localization on and within wheat roots (Levanony et al., 1989), wheat root-hair branching (Jain and Patriquin, 1984), enhancement of mineral absorption (Baldani et al., 1983; Lin et al., 1983; Morgenstern and Okon, 1987; Sarig et al., 1988), root length, respiration rate and ATP-saving on tomato (Hadas and Okon, 1987).

The specificity of the plant–bacterium association is still controversial. According to Jain and Patriquin (1984) and Baldani and Döbereiner (1980), the results depend on the strain–host plant specificity, whereas for Okon and Hadar (1987) the effect is not strain-dependent within a species of Azospirillum or within different plant species.
In Zea mays, enhancement of root and shoot length were observed under controlled conditions (Fallik et al., 1988); in the field, Hegazi et al. (1983) obtained yield increases in the Nile valley.

This paper describes the comparative use of 2 methods for measuring the inoculation effects of an Azospirillum lipoferum strain on Zea mays seedlings. The influence of bacterial concentration and plant genotype are investigated.

**MATERIALS AND METHODS**

**Cultivars**

In the germination tests and pot experiments, hybrids INRA 258, Dea (Pioneer Hi-bred Int) were used, and hybrids Pioneer Hi-bred Int, Marina, Lioea, Dea, Eva, Fulvia, Volga and Sirena in the comparison between cultivars.

**Micro-organism**

The strain used was Azospirillum lipoferum CRT1 isolated from maize rhizosphere (Fages and Mulard, 1988).

**Medium**

AFM 10 medium was used (Fages, 1990); the composition was as follows (in g/l): MgSO₄·H₂O, 0.2; NaCl, 0.1; CaCl₂, 0.02; FeSO₄·7H₂O, 0.01; Na₂MoO₄·2H₂O, 0.002; KH₂PO₄, 0.4; K₂HPO₄, 0.6; yeast extract, 3.0; glucose, 5.0; pH adjusted to 7.2 with NaOH 10 N. Sterilization was by autoclaving for 20 min at 121°C. Glucose was separately sterilized under the same conditions.

**Inoculum preparation**

Inoculum was prepared in a 250-ml Erlenmeyer flask containing 150 ml medium. These flasks were incubated at 30°C on a rotative shaker for 24 h. These cultures in late log-phase were centrifuged for 15 min at 4,000 g. Cells were resuspended in sterile distilled water, homogenized with a turbine (Ultraturrax, Ika) and diluted until the desired cell concentration was reached for plantlet inoculation. Microbial population was determined by dilution plate counts on AFM 10 medium complemented with agar. Results were expressed as colony forming units (cfu).

**Germination tests**

One hundred Zea mays seeds were placed in a polystyrene box (18 x 15 x 3 cm) containing sterile Loire sand (from the Loire river, France) humidified with bacterial suspension (sand: suspension ratio 5:1 v/v, equivalent to 2.5 ml inoculum per seed). The previously autoclaved inoculum was used as control; its concentration varied from one experiment to another. Seeds were covered with a 1-cm thick sand layer then placed in a germination chamber at 18°C with a relative humidity of 70% for 5 days in darkness. Percentage of germination as well as coleoptile and primary root lengths were measured. Ten replicates of each treatment were made. Germination percentages were determined by deduction of dead or abnormal plantlets from total plant count. Lengths were measured on 10 plants sampled randomly.

**Pot experiments**

Twelve or 20 Zea mays seeds were placed in a pot over a water tank containing 2.5 l of sterile Loire sand. The sand had been previously humidified with demineralized water (sand: water ratio, 5:1 v/v). Inoculation was achieved by placing 1 ml bacterial suspension on each seed. The previously autoclaved inoculum was used as control. When several inoculum concentrations were tested, the intermediate level (10⁷ cfu/ml) was chosen for autoclaving. Seeds were then covered with a 1-cm thick layer of sand. Ten replicates of each treatment were made. Pots were placed in a germination chamber for 14 days with a photoperiod of 14 h light and 10 h dark and a synchronous thermoperiod of 25/20°C. Plant nutritive requirements were covered by the following presterilized solution (in mg/l): KNO₃, 384; K₂HPO₄, 54; KH₂PO₄, 109; NaNO₃, 17; NaCl, 12; NH₄NO₃, 16; Ca(NO₃)₂·4H₂O, 732; MgSO₄·7H₂O, 185; FeSO₄·7H₂O, 1; ZnSO₄·H₂O, 1; H₃BO₃, 1; CuSO₄·5H₂O, 0.03; MnSO₄·4H₂O, 0.1; (NH₄)₂MoO₄, 0.03. At emergence (after 4 days), thinning out of seedlings was carried out leaving 9 out of 12 or 16 out of 20 plantlets per pot. At harvest after 14 days, the following measurements were performed: root surface area, root and shoot dry matter.

**Titrimetric measurement of root surface area**

(Carley and Watson, 1966)

The root system was washed with demineralized water to eliminate all the sand particles remaining on the root system. After excision, roots were drained for 3 h. Each root system was then soaked for 15 s in HCl 3 N. Excess acid elimination was performed in a manual centrifugal dryer. Roots were then left in 300 ml of demineralized water with gentle shaking. 100 ml of this solution was then titrated with NaOH 0.25 N and phenolphthalein as dye indicator. Results were ex-
pressed in ml of poured NaOH. Roots and shoots were then placed in a ventilated drying oven for 48 h at 85°C for dry matter determination.

**Statistical analysis**

Variance analyses were followed by Newman–Keuls test when Fisher test probability was < 5%.

**RESULTS**

**Germination tests**

No modification in the percentages of germination was observed (data not shown).

**Effect of bacterial concentration**

A preliminary experiment carried out with $6 \times 10^9$ cfu/ml (table I) showed an inhibition of root growth. A non-significant decrease in shoot-length was observed with both cultivars.

Another experiment was set up with 3 different bacterial concentrations of inoculum: $0.9 \times 10^9$, $0.9 \times 10^7$ and $0.9 \times 10^5$ cfu/ml. At the highest concentration, no difference in length from control was observed (table II). A significant increase in primary root elongation was obtained with the 2 other bacterial concentrations. However, a decrease in shoot length with the lowest bacterial density was obtained.

Table III shows results from trials using the same protocol but carried out at different dates. Four inoculum levels were tested in these trials.

<table>
<thead>
<tr>
<th>Hybrid</th>
<th>INRA 258</th>
<th>DEA</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Control</td>
<td>Inoculated</td>
</tr>
<tr>
<td>Shoot length (mm)</td>
<td>41.8 a</td>
<td>40.6 a</td>
</tr>
<tr>
<td>Primary root length (mm)</td>
<td>93.3 b</td>
<td>80.3 c</td>
</tr>
</tbody>
</table>

Mean values having common letter within each cultivar are not significantly different at 0.05 level (Newman–Keuls test).

<table>
<thead>
<tr>
<th>Bacterial concentration (cfu/ml)</th>
<th>Shoot length (mm)</th>
<th>Primary root length (mm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>$0.9 \times 10^9$</td>
<td>37.0 a</td>
<td>94.3 ab</td>
</tr>
<tr>
<td>$0.9 \times 10^7$</td>
<td>37.6 a</td>
<td>95.5 a</td>
</tr>
<tr>
<td>$0.9 \times 10^5$</td>
<td>33.8 b</td>
<td>95.8 a</td>
</tr>
<tr>
<td>Autoclaved control</td>
<td>36.3 a</td>
<td>91.2 b</td>
</tr>
</tbody>
</table>

Mean values having common letter within a column are not significantly different at 0.05 level (Newman–Keuls test). Control is the intermediate concentration culture autoclaved.

<table>
<thead>
<tr>
<th>Trial No</th>
<th>Bacterial concentration (cfu/ml)</th>
<th>Shoot length (mm)</th>
<th>Primary root length (mm)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Control</td>
<td>Inoculated</td>
<td>Control</td>
</tr>
<tr>
<td>1</td>
<td>$1.0 \times 10^6$</td>
<td>42.9 a</td>
<td>45.3 b</td>
</tr>
<tr>
<td>2</td>
<td>$5.0 \times 10^6$</td>
<td>47.1 a</td>
<td>48.6 b</td>
</tr>
<tr>
<td>3</td>
<td>$1.0 \times 10^7$</td>
<td>34.7 a</td>
<td>37.6 b</td>
</tr>
<tr>
<td>4</td>
<td>$1.0 \times 10^8$</td>
<td>40.4 a</td>
<td>42.5 b</td>
</tr>
</tbody>
</table>

Mean values having common letter within a trial are not significantly different at 0.05 level (Newman–Keuls test). Length variations between trials are caused by test duration which may vary by a few hours after 5 days. For each trial, control is the bacterial culture autoclaved.
Apart from the lowest concentration which exhibited no difference, all the other inoculations led to a significant increase in root elongation. The highest increases were obtained with the 2 intermediate concentrations. The bacterial density effect on shoot length was less pronounced, since a significant increase was observed with all the inoculum concentrations.

**Effect of the genotypes of 7 maize hybrids**

To test host-plant genotype, the level of $1.0 \times 10^7$ cfu/ml was chosen. Besides the Dea hybrid already tested with this concentration, 6 new cultivars were tested (fig 1). Four hybrids, Eva, Fulvia, Dea and Marina, exhibited a positive response to both criteria. Shoot elongation was respectively increased by 13.1%, 10.2%, 8.4% and 7.8%. Root elongation was respectively increased by 4.3%, 3.0%, 5.0% and 3.1%. In Volga, a significant increase in root elongation of 3.8% was observed. In Licea, in spite of a 5% increase in shoot length, no significant difference was observed. Finally an inhibition of root growth was obtained with Sirena.

![Fig 1. Germination tests. Primary root and shoot lengths of Zea mays hybrids through inoculation with A. lipoforum CRT1 at 1.0 $\times 10^7$ cfu/ml. s: significantly different at 0.05 level (Newman–Keuls test); ns, not significantly different at 0.05 level (Newman–Keuls test).](image)

### Pot experiments

**Effect of bacterial concentration**

In the first experiment, 3 bacterial concentrations of $0.9 \times 10^9$, $0.9 \times 10^7$ and $0.9 \times 10^5$ cfu/ml were tested for 14 days with 16 plants per pot. As shown in table IV, a significant increase in root surface area (13.5%) was obtained for the intermediate level. No significant difference with the other parameters was observed, due to too wide a biological variation.

Nevertheless, in all cases measurements on inoculated plants gave higher values than those obtained with control plants. In a second trial carried out with 9 plants, 3 other concentrations, $1.0 \times 10^9$, $1.0 \times 10^7$ and $1.0 \times 10^6$ cfu/ml, were tested.

There was too great a dispersion in the measurements for the statistical threshold to be reached on root dry matter and root surface area. However, the intermediate inoculum concentration gave the highest values. On aerial parts, a significant increase in dry matter was obtained whatever the inoculum concentration.

**Effect of the genotypes of 5 maize hybrids**

Two hybrids, Volga and Licea, tested in the germination tests, were not assayed here. The lar-

### Table IV. Pot experiment. Effect of bacterial concentration of A. lipoforum CRT1 on root surface area and dry weight of Zea mays (cv Dea).

<table>
<thead>
<tr>
<th>Bacterial concentration (cfu/ml)</th>
<th>Root surface area (ml NaOH/pot)</th>
<th>Shoot dry weight (g/pot)</th>
<th>Root dry weight (g/pot)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Trial 1</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>$0.9 \times 10^9$</td>
<td>14.8 a</td>
<td>2.45 a</td>
<td>2.07 a</td>
</tr>
<tr>
<td>$0.9 \times 10^7$</td>
<td>15.1 b</td>
<td>2.40 a</td>
<td>2.18 a</td>
</tr>
<tr>
<td>$0.9 \times 10^5$</td>
<td>13.5 a</td>
<td>2.36 a</td>
<td>2.01 a</td>
</tr>
<tr>
<td>Control</td>
<td>13.3 a</td>
<td>2.33 a</td>
<td>1.99 a</td>
</tr>
<tr>
<td><strong>Trial 2</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>$1.0 \times 10^8$</td>
<td>10.3 a</td>
<td>1.99 a</td>
<td>0.56 a</td>
</tr>
<tr>
<td>$1.0 \times 10^7$</td>
<td>11.1 a</td>
<td>2.02 a</td>
<td>0.61 a</td>
</tr>
<tr>
<td>$1.0 \times 10^6$</td>
<td>9.6 a</td>
<td>2.08 a</td>
<td>0.54 a</td>
</tr>
<tr>
<td>Control</td>
<td>9.4 a</td>
<td>1.75 b</td>
<td>0.57 a</td>
</tr>
</tbody>
</table>

Mean value having common letter within trial and within a column are not significantly different at 0.05 level (Newman–Keuls test). Control is the intermediate concentration culture autoclaved.
gest significant effects were obtained on Dea root surface area (+21% over control) and on Eva shoot dry matter (+7%). No other differences were obtained with these 2 cultivars (table V). Responses of the 3 other plant varieties to inoculation were non-significant.

**Table V. Pot experiments. Effect of inoculation with A. lipoferum CRT1 at 1.0 x 10^7 cfu/ml on root surface area and dry weight of different Zea mays hybrids.**

<table>
<thead>
<tr>
<th>Hybrid</th>
<th>Root surface area (ml NaOH/pot)</th>
<th>Root dry weight (g/pot)</th>
<th>Shoot dry weight (g/pot)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>9.6a</td>
<td>0.64c</td>
<td>0.65c</td>
</tr>
<tr>
<td>Inoc</td>
<td>9.7a</td>
<td>0.65c</td>
<td>0.68c</td>
</tr>
<tr>
<td>Dea</td>
<td>13.4a</td>
<td>17.0b</td>
<td>0.65c</td>
</tr>
<tr>
<td>Eva</td>
<td>8.3a</td>
<td>8.0a</td>
<td>0.60c</td>
</tr>
<tr>
<td>Fulvia</td>
<td>17.8a</td>
<td>17.1a</td>
<td>0.88c</td>
</tr>
<tr>
<td>Sirena</td>
<td>10.3a</td>
<td>10.6a</td>
<td>0.59c</td>
</tr>
</tbody>
</table>

Mean values with common letter within a hybrid are not significantly different at 0.05 level (Newman–Keuls test).

**DISCUSSION**

The different parameters checked in germination tests did not show the same sensitivity to bacterial inoculation. The percentage of germination was not affected. This qualitative criterion may not be suitable for demonstrating growth promotion. Moreover, it is related to seed physiology rather than to external effects.

Root elongation was the best parameter for displaying differences between inoculum concentrations. This is evidence that it is the root development of plantlets which is directly affected by bacterial inoculation. Under our experimental conditions, the optimal inoculum level was 1.0 x 10^7 cfu/ml or 2.5 x 10^7 cfu/plant. Lower bacterial densities led to lower responses. These results are in agreement with the findings of Fallik et al (1988) on Zea mays, while Kapulnik et al (1985) found an optimal inoculum level for wheat of between 10^5 cfu/plant and 10^6 cfu/plant. Beyond 10^8 cfu/ml, a clear inhibition was observed.

Shoot elongation was also increased through inoculation. However, this parameter was less sensitive to the number of bacteria applied.

These results show a very early effect of *Azospirillum* inoculation on *Zea mays*. With another crop (*Panicum miliaceum*), similar effects were obtained by Harari et al (1989) on root elongation and root-hair development.

The effect of *Zea mays* inoculation by *Azospirillum lipoferum* CRT1 was host-plant genotype dependent since coleoptile elongation varied from −5.6% to +13.1% and primary root length varied from −2.8% to +5.8% over control. Figure 1 illustrates these results, showing that germination tests are a powerful tool for the screening of plant–bacteria associations on the plant-growth promotion criterion. This is a decisive step forward compared with classical screening methods performed with indirect criteria such as biological nitrogen fixation (Ela et al, 1982; Thomas-Bauzon et al, 1982) or phytohormone production (Zimmer and Böthe, 1989).

Pot experiment results showed that inoculation effects were still observable after 2 weeks' growth. The increases obtained had the same order of magnitude as those observed after 5 days. However, the biological variation was at this development stage more pronounced, and consequently, results were less often statistically significant.

For root surface area improvement, the optimal inoculum level appeared to be 1.0 x 10^7 cfu/plant. No correlation between root dry matter and root surface area was found. The better development of the root system which can be easily observed de visu was not converted into root dry matter variations measurable after 14 days of growth. Titrimetric measurement of root surface area is the best method for evidencing differences among inoculum concentrations. As shown by the trial with several plant varieties, this method was not suitable for hybrid screening. Compared with the germination tests method, it has 3 major disadvantages. It is longer, involves more work and is less discriminating.

*Azospirillum lipoferum* CRT1 growth-promotion effect on *Zea mays* began from the very early plant development stages and remained observable after 2 weeks. It could be measured on several parameters, and dependent on both the inoculum concentration and the host-plant genotype. These findings suggest that the yield enhancements in the field came, at least partly, from such early root-growth promotion effects.
The methods described here, particularly the germination tests, indicated which bacterial strain-cultivar pair should be preferably chosen and which inoculum level was optimal. Moreover, the precocity of the observed effects showed that the period immediately following planting is crucial. Hence, one wonders, if the fate of the inoculated strain in the rhizosphere is really important beyond the few days during which the growth-promotion effect acts. The inoculum formulation (Fages, 1990) and the methodology of field application is therefore very important and must allow a good root colonization at the beginning of plant growth. These methods are under current investigation in our laboratory.

ACKNOWLEDGMENT

We thank G Lafargue from Sica France-Maïs quality control laboratory for helping us with the protocol design.

REFERENCES


Fayez M, Daw ZY (1987) Effect of inoculation with different strains of Azospirillum brasilense on cotton (Gossypium barbadense). Biol Fertil Soils 4, 91-95


