

Stereodifferentiation in the Intramolecular Singlet Excited State Quenching of Hydroxybiphenyl/Tryptophan Dyads

P. Bonancía, I. Vayá, D. Markovitsi, T. Gustavsson, M. C. Jiménez, M. A. Miranda

▶ To cite this version:

P. Bonancía, I. Vayá, D. Markovitsi, T. Gustavsson, M. C. Jiménez, et al.. Stereodifferentiation in the Intramolecular Singlet Excited State Quenching of Hydroxybiphenyl/Tryptophan Dyads. Organic & Biomolecular Chemistry, 2013, 11 (12), pp.1958-1963. 10.1039/C3OB27278H . hal-00807300

HAL Id: hal-00807300

https://hal.science/hal-00807300

Submitted on 3 Apr 2013

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Cite this: Org. Biomol. Chem. XXXX, XX, XX

www.rsc.org/obc PAPER

Stereodifferentiation in the Intramolecular Singlet Excited State Quenching of Hydroxybiphenyl/Tryptophan Dyads

Paula Bonancía, ^a Ignacio Vayá, ^a Dimitra Markovitsi, ^b Thomas Gustavsson, ^b M. Consuelo Jiménez ^{*a} and Miguel A. Miranda ^{*a}

5 Received (in XXX, XXX) Xth XXXXXXXX 20XX, Accepted Xth XXXXXXXX 20XX DOI: 10.1039/b000000x

The photochemical processes occurring in diastereomeric dyads (*S*,*S*)-1 and (*S*,*R*)-1, prepared by conjugation of (*S*)-2-(2-hydroxy-1,1'-biphenyl-4-yl)propanoic acid ((*S*)-**BPOH**) with (*S*)- and (*R*)-**Trp**, have been investigated. In acetonitrile, the fluorescence spectra of (*S*,*S*)-1 and (*S*,*R*)-1 were coincident in shape and position with that of (*S*)-**BPOH**, although they revealed a markedly stereoselective quenching. Since singlet energy transfer from **BPOH** to **Trp** is forbidden (5 kcal mol⁻¹ uphill), the quenching was attributed to thermodynamically favoured (according to Rehm-Weller) electron transfer or exciplex formation. Upon addition of 20% water, the fluorescence quantum yield of (*S*)-**BPOH** decreased, while only minor changes were observed for the dyads. This can be explained by an enhancement of the excited state acidity of (*S*)-**BPOH**, associated with bridging of the carboxy and hydroxy groups by water, in agreement with the presence of water molecules in the X-ray structure of (*S*)-**BPOH**. When the carboxy group was not available for coordination with water, as in the methyl ester (*S*)-**BPOHMe** or in the dyads, this effect was prevented; accordingly, the fluorescence quantum yields did not depend on the presence or absence of water. The fluorescence lifetimes in dry acetonitrile were 1.67, 0.95 and 0.46 ns for (*S*)-**BPOH**, (*S*,*S*)-1 and (*S*,*R*)-1, respectively, indicating that the observed quenching is indeed dynamic. In

BPOH, (*S*,*S*)-1 and (*S*,*R*)-1, respectively, indicating that the observed quenching is indeed dynamic. In line with the steady-state and time-resolved observations, molecular modelling pointed to a more favourable geometric arrangement of the two interacting chromophores in (*S*,*R*)-1. Interestingly, this dyad exhibited a folded conformation in the solid state.

Introduction

25 Bichromophoric compounds incorporating two photoactive moieties are useful models to gain insight into key mechanistic aspects of light-induced processes. In this context, intramolecular energy, proton or electron transfer, as well as exciplex formation and photochemical reactivity, have received considerable 30 attention. These processes have been investigated in a variety of systems, including dyads containing a non-steroidal antiinflammatory drug (NSAID) covalently linked to a biomimetic substructure (for instance, an amino acid or a nucleoside, as models for proteins or DNA).² Specifically, previous efforts have 35 been devoted to the photophysical study of dyads containing NSAIDs and the amino acid tryptophan (Trp), as valuable tools to provide relevant information on drug-protein interactions within the binding sites.³ In this context, steady-state and timeresolved fluorescence spectroscopy has been widely used to 40 address a variety of biological problems, including the excited state interactions between proteins and complexed ligands.⁵

The photophysical behaviour of diastereomeric dyads containing the biphenyl-like NSAID flurbiprofen ((S)-FBP, Chart

1) and **Trp** has recently been described. Upon irradiation at 266 mm, both chromophores are excited, but only **Trp** emission (E_S = 96 kcal mol⁻¹) is detected. Moreover, stereoselective intramolecular ¹**Trp*** charge transfer quenching is observed, accompanied by formation of exciplexes. Indeed, the absorption and emission properties of **Trp** are very sensitive to the local environment and have been employed to investigate structural changes of protein functions and to address the protein binding of endogenous and exogenous agents (such as drugs, fatty acids, metabolites), metabolites),

Attachment of a hydroxy group at the *ortho* position of a biphenyl results in a significant decrease of its singlet energy (*ca*. 10 kcal mol⁻¹) and provides an additional deactivation pathway, namely excited state intramolecular proton transfer (ESIPT). As ESIPT is in general a reversible process, ¹⁰ it has found application for the design of photostabilizers, solar filters, solar oenergy collectors, etc. ¹¹ In hydroxybiphenyls, ESIPT occurs from the acidic phenolic proton to the sp² hybridized 2'-carbon of the aromatic ring, giving rise to a reactive quinone methide. In addition, intermolecular excited state deprotonation is observed in

the presence of bulk water, to give the corresponding phenolate. 12 With this background, we have now investigated the photophysical processes occurring in diastereomeric dyads (S,S)-1 and (S,R)-1 (Chart 1), which contain a chiral 2-5 hydroxybiphenyl derivative covalently linked to **Trp**.

Results and Discussion

The required (S)-2-(2-hydroxy-1,1'-biphenyl-4-yl)propanoic acid ((S)-BPOH) was readily obtained by irradiation of (S)-FBP in aqueous media, through a photonucleophilic substitution.¹³ The 10 dyads (S,S)-1 and (S,R)-1 were prepared by conjugation of ((S)-**BPOH**) with the (S)- or (R)- methyl ester of tryptophan (**TrpMe**), using a carbodiimide as activating agent. The chemical structures are shown in Chart 1.

Chart 1

The UV-Vis absorption spectrum of (S,S)-1 in MeCN is shown 25 in Figure 1, together with those of (S)-FBP and (S)-BPOH for comparison. It can be observed that replacement of fluorine by hydroxyl induces a bathochromic shift of the longer wavelength band. The spectra of (S,S)-1 and (S,R)-1 were coincident and matched with that obtained upon addition of the absorption 30 spectra of the isolated chromophores ((S)-BPOH and (S)-**TrpMe**), indicating the absence of significant interactions between the two moieties in the ground state.

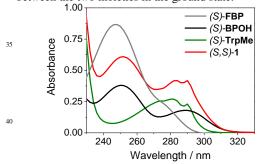
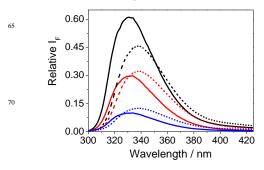


Figure 1. UV-Vis absorption spectra of (S)-FBP (grey), (S)-BPOH ₄₅ (black), (S)-**TrpMe** (green) and (S,S)-**1** (red) in acetonitrile at 5×10^{-5} M concentration.

The fluorescence spectra of (S,S)-1 and (S,R)-1, obtained after excitation at $\lambda = 267$ nm in MeCN/air, are shown in Figure 2 (solid lines) together with that of (S)-**BPOH**. All the emissions 50 consisted in a structureless band centered at ca. 330 nm (Table 1). In the dyads, the incident photons are absorbed by both the hydroxybiphenyl chromophore and the **Trp** moiety (Figure 1); however, no Trp contribution was observed in the fluorescence spectra, which were coincident in shape and position with that of 55 (S)-**BPOH**. From the intersection between the excitation ($\lambda_{em} =$ 330 nm) and emission ($\lambda_{exc} = 267$ nm) normalised bands (Figure

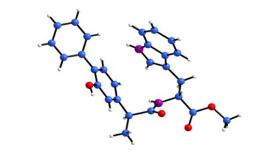
S4 in Supporting Information), a singlet energy value of ca. 91 kcal mol⁻¹ was obtained for (S)-**BPOH**, (S,S)-1 and (S,R)-1.

In the dyads, the most remarkable observation was a dramatic 60 quenching, which turned out to be stereoselective (see Table 1). Since SSET from (S)-**BPOH** to **Trp** is thermodynamically forbidden, the observed quenching was in principle attributed to electron transfer or exciplex formation.



75 Figure 2. Fluorescence spectra of (S)-BPOH (black), (S,S)-1 (red) and (S,R)-1 (blue) at $\lambda_{exc} = 267$ nm, under air. Solid lines: MeCN; dashed lines: MeCN/H2O 4:1.

The feasibility of formation of radical ion pairs or exciplexes 80 from the singlet excited state can be estimated by application of the Rehm-Weller equations. 14 Taking into account the corresponding oxidation (E_{OX}) and reduction (E_{RED}) potentials, ¹⁵ the appropriate dielectric constants $(\varepsilon)^{16}$ and the (S)-BPOH singlet energy ($E_S = 91 \text{ kcal mol}^{-1}$), a favourable thermodynamics 85 was anticipated for charge transfer quenching via both electron transfer and exciplex formation. These processes are more likely to occur in folded conformations, since a close contact is required between the donor and acceptor moieties. As a matter of fact, the (S,R)-diastereomer exhibited a folded conformation in the solid 90 state, as shown by X-ray diffraction (Figure 3). Interestingly, quenching was more efficient in this dyad than in (S,S)-1.



100

Figure 3. X-ray structure of compound (S,R)-1.

To check whether ESIPT from the acidic phenolic proton to the sp² hybridised 2'-carbon of the neighbouring aromatic ring could play a role in these systems, in competition with charge transfer quenching, a 20% of water was added (dashed lines in Figure 2). This water content should be low enough as to prevent intermolecular excited state deprotonation to bulk water, giving the corresponding phenolate.

Table 1. Relevant photophysical parameters for (S)-**BPOH**, (S)-**BPOHMe**, (S,S)-1 and (S,R)-1.

	MeCN			60	MeCN/H ₂ O			
	(S)-BPOH	(S)-BPOHMe	(S,S)-1	(S,R)-1	(S)-BPOH	(S)-BPOHMe	(S,S)-1	(S,R)-1
λ_{\max}^{em} (nm)	331	334	330	332	338	340	339	339
$\phi_F^{\ b}$	0.241	0.290	0.116	0.042	0.200	0.280	0.139	0.060
$\tau_F (ns)^c$	1.67	2.10	0.95	0.46	1.27	1.80	1.07	0.57
$k_{F} \times 10^{8} (s^{-1})^{d}$	1.43	1.38	1.22	0.945	1.57	1.55	1.29	1.05
$k_Q \cdot \times 10^8 (s^{-1})^e$	-	-	4.5	15.8	-	-	1.5	9.7
$\phi_{Q(ov)}^{f}$	-	-	0.52	0.83	-	-	0.31	0.70
$\varphi_{Q(dyn)}{}^g$	-	-	0.43	0.68	-	-	0.16	0.55

a The absorbance of the samples was 0.2 at the excitation wavelength. $^{\text{b}}\lambda_{\text{exc}} = 267$ nm, air, (S)-Trp in water as standard ($\phi_{\text{std}} = 0.13$); $^{\text{c}}\lambda_{\text{em}} = 330$ nm; $^{\text{d}}k_F = \phi_F/\tau_F$; $^{\text{e}}k_Q = 0.13$ $(1/\tau_{F(dyad)} - 1/\tau_{F(BPOH)}); \ ^f \varphi_{Q(ov)} = [1 - (\varphi_{F(dyad)}/\varphi_{F(BPOH)})]; \ ^g \varphi_{Q(dyn)} = \varphi_{F(dyad)} k_{Q(dyad)}/k_{F(dyad)}.$

In this solvent mixture, the emission maxima were red-shifted, due to the enhanced medium polarity. Remarkably, the 15 fluorescence quantum yield of (S)-**BPOH** was clearly lower than in dry MeCN, while only minor changes were observed for the dyads (Table 1). A similar trend has been reported for the model 2-hydroxybiphenyl under related compound conditions (acetonitrile containing 10 M water), although fluorescence 20 quenching occurs to a lesser extent. 12 The interesting effect observed with (S)-BPOH in the presence of water can be explained by an enhancement of the excited phenol acidity, associated with bridging of the carboxy and hydroxy groups by water. As a matter of fact, this situation is clearly observed in the 25 X-ray structure of (S)-BPOH, which actually crystallizes with a water molecule (Figure 4A). This effect should be, in principle, avoided by preventing water-mediated bridging between carboxy and hydroxy group.

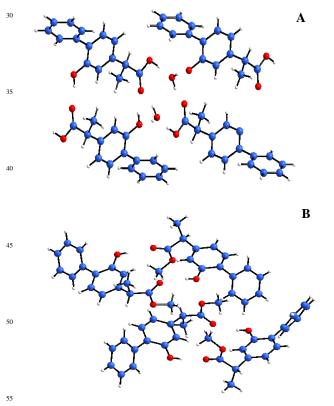
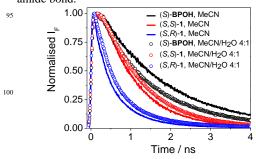


Figure 4. Fragment of the unit cell for A: (S)-BPOH and B: (S)-BPOHMe.

With this purpose, we synthesized the methyl esther (S)-**BPOHMe**. As anticipated, the fluorescence quantum yields of this model remained practically constant in the presence and absence of water (Table 1). Accordingly, water molecules were 75 absent in the X-ray structure of (S)-**BPOHMe** (Figure 4B). In the case of the dyads, the acidity of the phenolic group is not enhanced as in (S)-BPOH, because the carboxy group is employed in the construction of the amide linker and is not available for coordination with water. In fact, no water molecules 80 were found in the crystalline samples.

The fluorescence lifetimes were determined by the timecorrelated single photon counting (TCSPC) technique (Figure 5). In dry acetonitrile, τ_F ((S)-**BPOH**) was found to be 1.67 ns, while τ_F values of 0.95 and 0.46 ns were measured for (S,S)-1 and 85 (S,R)-1, respectively (Table 1). This indicates that most of the overall quenching observed in the steady-state experiments is indeed dynamic, as expected for a charge transfer process (compare the two sets of values in Table 1). In the presence of water, a decrease of the (S)-BPOH fluorescence lifetime was 90 noticed, confirming again a dynamic quenching, in agreement with an enhanced participation of the ESIPT route. As discussed above, this deactivation channel does not operate in the dyads, where the carboxylate function is involved in formation of the amide bond.



105 **Figure 5.** Kinetic decay traces ($\lambda_{\text{exc}} = 267 \text{ nm}$, $\lambda_{\text{em}} = 330 \text{ nm}$, $A_{267} = 0.2$) for (S)-BPOH (black), (S,S)-1 (red) and (S,R)-1 (blue) under air, in MeCN (solid line) and MeCN/H₂O 4:1 (open circles).

The quenching rate constants (k_O), determined from the fluorescence lifetimes $\tau_{\rm E}$, were in the range 10^8 - 10^9 s⁻¹; as regards 110 the radiative rate constants k_F, they were obtained from the values of τ_F and ϕ_F and found to be in the order of 10^8 s⁻¹ (Table 1).

In order to get insight into the geometrical arrangement of the two chromophores in the dyads, which is closely related with the fluorescence quenching by charge transfer, simple molecular modelling (PM3) was performed with both (S,S)-1 and (S,R)-1(see Figure 6).

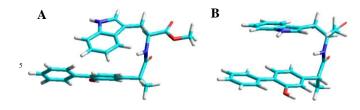


Figure 6. Geometry optimized (HyperChem Release 8.0.3 for Windows 10 Molecular Model System, PM3) structure for (S,S)-1 (A) and (S,R)-1 (B).

Thus, (S,R)-1 (Figure 6B) presented a more favourable geometric arrangement of the two interacting chromophores compared to the (S,S)-dyad (Figure 6A), whose geometry is more distorted, with the indole chromophore nearly orthogonal to the 15 biphenyl moiety, which makes more difficult interchromophoric interaction. This is in good agreement with the steady-state and time-resolved observations of lower ϕ_F and shorter τ_F values for the (S,R)-diastereomer.

Conclusions

20 The (S)-BPOH chromophore dominates the fluorescence spectra of diastereomeric dyads (S,S)-1 and (S,R)-1 in acetonitrile, which reveal a substantial intramolecular charge transfer quenching with remarkable stereodifferentiation. In the presence of water (20%), the fluorescence quantum yield of (S)-BPOH decreases; by 25 contrast, only minor changes are observed for (S,S)-1 and (S,R)-1. Being (S)-BPOH a phenol, this is attributed to a water-assisted enhancement of its excited state acidity, a hypothesis that is supported by the presence of water molecules linking the carboxy and hydroxy groups in the crystalline structure. This effect is 30 prevented in the dyads, whose fluorescence quantum yields do not depend on the presence or absence of water. The dynamic nature of the observed quenching is evidenced by a configuration-dependent shortening of the fluorescence lifetimes in (S,S)-1 and (S,R)-1 compared to (S)-BPOH. In agreement with 35 the steady-state and time-resolved experimental results, molecular modelling points to a more favourable geometric arrangement of the two interacting chromophores in (S,R)-1 than in (S,S)-1; this is validated by the X-ray analysis, indicating that the former dyad exhibits a folded conformation in the solid state.

40 Materials and Methods

Commercial (S)-**FBP**, (S)and (R)-TrpMe, dimethylaminopropyl)-N-ethylcarbodiimide hydrochloride (EDC) and 1-hydroxybenzotriazole (BtOH) were purchased from Sigma-Aldrich. Their purity was checked by ¹H NMR and HPLC 45 analysis. Spectrophotometric, HPLC or reagent grade solvents were obtained from Scharlab and used without further purification. Solutions of phosphate-buffered saline (PBS) (0.01 M, pH 7.4) were prepared by dissolving phosphate-buffered saline tablets (from Sigma) in Milli-Q water. The ¹H-NMR and ₅₀ ¹³C-NMR spectra were recorded in CDCl₃ at 300 and 75 MHz, respectively, using a Varian Gemini instrument; chemical shifts are reported in ppm. Exact mass values were obtained in Servicio de Espectroscopía de Masas de la Universidad de Valencia. The X-ray structures were determined at Unidade de Raios X, at the 55 Universidade de Santiago de Compostela. Crystallographic data

(excluding structure factors) for the structures of (S)-**BPOH**, (S)-**BPOHMe** and (S,R)-1 have been deposited at the Cambridge Crystallographic Data Centre as supplementary publication numbers CCDC 662956, CCDC 910283 and CCDC 910284, 60 respectively. Isolation and purification were done by conventional column chromatography on silica gel Merck 60 (0.063-0.200 mm), or by preparative layer chromatography on silica gel Merck 60 PF254, using hexane/ethyl acetate as eluent.

Synthesis of the new compounds

65 (S)-BPOH. A solution of (S)-FBP (100 mL, 5 mM) in PBS was irradiated for 9 h through quartz, inside a Luzchem multilamp photoreactor, with the light from ten 8W lamps emitting mainly at 254 nm. The photomixture was acidified with HCl, extracted with methylene chloride and dried over MgSO₄. The organic 70 phase was evaporated in vacuo and purified by preparative layer chromatography, using hexane/ethyl acetate 40/60 (v/v) as eluent, affording (S)-BPOH as white powder (60 % yield).

(S)-BPOHMe. A solution of (S)-BPOH (59 mg, 0.23 mmol) and SOCl₂ (2.4 mL) in MeOH (10 mL) was maintained under reflux 75 for 2 h. The crude was rotavaporated, dissolved in methylene chloride and washed consecutively with saturated NaHCO₃, 1 M HCl and brine, then dried over MgSO₄. The solvent was evaporated in vacuo. Purification was done by preparative layer chromatography (hexane/ethyl acetate 70/30, v/v), affording (S)-80 **BPOHMe** as a white powder in nearly quantitative yield.

Dyads (S,S)-1 and (S,R)-1. To a solution of (S)-BPOH (56 mg, 0.23 mmol) in acetonitrile (20 mL), 0.23 mmol of EDC and 0.23 mmol of BtOH in acetonitrile were added. The mixture was maintained under stirring, and then 0.23 mmol of (S)- or (R)-85 **TrpMe** in 2 mL of acetonitrile were added. After 3 h, the solvent was evaporated; the crude was dissolved in methylene chloride and washed consecutively with diluted NaHCO3, 1 M HCl, and brine, then dried over MgSO₄. Final purification was performed by preparative layer chromatography (methylene chloride/ethyl 90 acetate, 60/40, v/v), followed by recrystallization. The yields were 50 % and 47 % for (S,S)-1 and (S,R)-1, respectively.

Characterisation of the new compounds

All new compounds were characterised by ¹H and ¹³C NMR spectroscopy, as well as by high resolution mass spectrometry 95 (HRMS). Their purity was confirmed by gas chromatography (GC) and high performance liquid chromatograpy (HPLC). A summary of the most relevant data follows.

Methyl (S)-2-(2-hydroxy-1,1'-biphenyl-4-yl)propanoate

¹H-NMR (CDCl₃) (δ , ppm) 1.53 (d, 3H, J = 8.0 Hz), 3.70 (s, 3H), $_{100}$ 3.73 (q, 1H, J = 8.0 Hz), 5.19 (s, 1H), 6.92-6.94 (m, 1H), 7.18-7.21 (d, 1H), 7.37-7.51 (m, 6H). ¹³C-NMR (CDCl₃) (δ, ppm) 18.5, 45.2, 52.1, 114.9, 119.4, 127.0, 127.9, 129.0, 129.3, 130.8, 136.8, 141.8, 152.5, 174.7. Exact Mass (ESI) [MH+] Calcd. for C₁₆H₁₇O₃: 257.1172; Found: 257.1181.

N-[2-(S)-(2-Hydroxy-1,1'-biphenyl-4-yl)propanoyl]-(R)tryptophan methyl ester.

¹H-NMR (CDCl₃) (δ, ppm) 1.46 (d, 3H, J = 7.1 Hz), 3.13-3.26 (m, 2H), 3.45 (q, 1H, J = 7.1 Hz), 3.64 (s, 3H), 4.90 (m, 1H), 6.05 (d,

1H, J = 8.0Hz), 6.28 (s, 1H), 6.50 (d, 1H, J = 2.4 Hz), 6.80 (m, 2H), 7.01-7.23 (m, 4H), 7.36-7.53 (m, 10H), 8.11 (bs, 1H). ¹³C-NMR (CDCl₃) (δ, ppm) 18.3, 27.5, 46.8, 52.6, 52.8, 109.5, 111.4, 115.1, 118.6, 119.7, 120.2, 122.2, 123.2, 127.5, 127.9, 129.0, 5 129.2, 130.9, 136.1, 137.5, 142.2, 153.3, 172.5, 173.9. Exact Mass (EI) [M⁺.] Calcd. for C₂₇H₂₆N₂O₄: 442.1893; Found: 442.1897.

N-[2-(S)-(2-Hydroxy-1,1'-biphenyl-4-yl)propanoyl]-(S)tryptophan methyl ester.

¹⁰ H-NMR (CDCl₃) (δ , ppm) 1.47 (d, 3H, J = 7.2 Hz), 3.19-3.32 (m, 2H), 3.47 (q, 1H, J = 7.2 Hz), 3.61 (s, 3H), 4.85 (m, 1H), 6.05 (d, 1H, J = 7.8 Hz), 6.12 (s, 1H), 6.73-6.79 (m, 3H), 7.05-7.47 (m, 9H), 8.13 (bs, 1H). ¹³C-NMR (CDCl₃) (δ, ppm) 18.1, 27.4, 46.7, 52.4, 52.9, 109.9, 111.2, 115.1, 118.6, 119.7, 120.0, 15 122.2, 122.3, 127.2, 127.6, 127.7, 129.1, 130.7, 136.0, 137.1, 141.7, 152.9, 172.3, 173.3. Exact Mass (EI) [M⁺] Calcd. for C₂₇H₂₆N₂O₄: 442.1893; Found: 442.1891

Fluorescence measurements

Emission spectra were recorded on a spectrofluorometer system 20 which was provided with a monochromator in the wavelength range of 200-900 nm. The solutions were placed into 10 mm \times 10 mm quartz cells. The absorbance of the samples at the excitation wavelength was kept below 0.2. Steady state experiments were performed at 22 °C. For the fluorescence quantum yield 25 determination, Trp in water/air was used as a reference, with ϕ_F = $0.13.^{17}$

Time-resolved fluorescence measurements were performed using the time-correlated single photon counting (TCSPC) technique. The excitation source was the third harmonic (267 nm) 30 of a mode-locked Ti-Sapphire laser, delivering 100-fs pulses with a repetition rate of 4.75 MHz. A Schott WG 295 filter was placed in front of a SPEX monochromator. The detector was a microchannel plate (R1564 U Hamamatsu) providing an instrumental response function of 60 ps (fwhm). The average 35 laser power (0.1 mW) was measured with a Melles Griot broadband powermeter. Solutions were contained in a 10 mm × 10 mm quartz cell and continuously stirred. Successive recordings with the same sample gave identical decays which were eventually merged to improve the signal-to-noise ratio. 40 Such a procedure allowed us to ensure that the measured signals were not altered during the measurements due to a possible

Acknowledgements

accumulation of photoproducts.

Financial support from the Spanish Government (CTQ2010-45 14882, BES-2008-003314, JCI-2010-06204, PR2011-0581), from the Generalitat Valenciana (Prometeo 2008/090) and from the Universitat Politècnica de València (PAID 05-11, 2766) is gratefully acknowledged.

Notes and references

50 a Departamento de Química/Instituto de Tecnología Química UPV-CSIC, Universitat Politécnica de València, Camino de Vera s/n, 46022, Valencia, Spain. E-mail: mcjimene@qim.upv.es; mmiranda@qim.upv.es

- ^b CNRS, IRAMIS, SPAM, Francis Perrin Laboratory, URA 2453, 91191 Gif-sur-Yvette, France.
- a) M. C. Jiménez, U. Pischel and M. A. Miranda, J. Photochem. Photobiol. C, 2007, 8, 128-142; b) S. Abad, U. Pischel and M. A. Miranda, J. Phys. Chem. A, 2005, 109, 2711-2717; c) S. Abad, I. Vayá, M. C. Jiménez, U. Pischel and M. A. Miranda, ChemPhysChem, 2006, 7, 2175-2183.
- a) P. Bonancía, I. Vayá, M. J. Climent, T. Gustavsson, D. Markovitsi, M. C. Jiménez and M. A. Miranda, J. Phys. Chem. A, 2012, 116, 8807-8814; b) C. Paris, S. Encinas, N. Belmadoui, M. J. Climent and M. A. Miranda, Org. Lett., 2008, 10, 4409-4412; c) N. Belmadoui, S. Encinas, M. J. Climent, S. Gil and M. A. Miranda, Chem. Eur. J., 2006, 12, 553-561; d) V. Lhiaubet-Vallet, P. Boscá and M. A. Miranda, J. Phys. Chem. B, 2007, 111, 423-431.
- I. Vayá, R. Pérez-Ruiz, V. Lhiaubet-Vallet, M. C. Jiménez and M. A. Miranda, Chem. Phys. Lett., 2010, 486, 147-153.
- J. R. Lakowicz, *Principles of Fluorescence Spectroscopy*, 3rd Edition; Plenum Press: New York, 2006.
- a) N. Seedher and S. Bhatia, J. Pharm. Biomed. Anal., 2005, 39, 257-262; b) N. Seedher and S. Bhatia, Pharmacol. Res., 2006, 54, 77-84; c) R. K. Nanda, N. Sarkar and R. Banerjee, J. Photochem. Photobiol. A, 2007, 192, 152-158; d) B. Zhou, R. Li, Y. Zhang and Y. Liu, Photochem. Photobiol. Sci., 2008, 7, 453-459; e) H. Vahedian-Movahed, M. R. Saberi and J. Chamani, J. Biomol. Struct. Dyn., 2011, 28, 483-502; f) U. Katrahalli, V. K. A. Kalalbandi and S. Jaldappagari, J. Pharm. Biomed. Anal., 2012, 59, 102-108; g) M. El-Kemary, M. Gil and A. Douhal, J. Med. Chem., 2007, 50, 2896-2902; h) L. Tormo, J. A. Organero, B. Cohen, C. Martin, L. Santos and A. Douhal, J. Phys. Chem. B, 2008, 112, 13641-13647; i) S. Tardioli, I. Lammers, J. H. Hooijschuur, F. Ariese, G. van der Zwan and C. Gooijer, J. Phys. Chem. B, 2012, 116, 7033-7039.
- I. Vayá, M. C. Jiménez and M. A. Miranda, J. Phys. Chem. B, 2007, 111, 9363-9371.
- a) P. R. Callis and B. K. Burgess, J. Phys. Chem. B, 1997, 101, 9429-9432; b) J. R. Lakowicz, *Photochem. Photobiol.*, 2000, **72**, 421-437; c) B. Schuler and W. A. Eaton, Curr. Opin. Struct. Biol., 2008, 18, 16-26; d) X. H. Shen and J. R. Knutson, J. Phys. Chem. B, 2001, 105, 6260-6265.
- a) J. M. Beechem and L. Brand, Annu. Rev. Biochem., 1985, 54, 43-71; b) P. R. Callis, Methods Enzymol., 1997, 278, 113-150.
- a) T. Peters, All About Albumins: Biochemistry Genetics and Medical Applications, Academic Press, San Diego, 1995; b) D. C. Carter, J. X. Ho, in Advances in Protein Chemistry, Vol. 45, Academic Press, New York, 1994, pp. 152-203.
- 10 a) N. Basaric and P. Wan, J. Org. Chem., 2006, 71, 2677-2686; b) M. Lukeman and P. Wan, J. Am. Chem. Soc., 2003, 125, 1164-1165.
- a) J. Keck, H. E. A. Kramer, H. Port, T. Hirsch, P. Fischer and G. Rytz, J. Phys. Chem., 1996, 100, 14468-14475; b) F. Vollmer and W. Rettig, J. Photochem. Photobiol. A, 1996, 95, 143-155.
- M. Lukeman and P. Wan, J. Am. Chem. Soc., 2002, 124, 9458-9464.
- 13 M. C. Jiménez, M. A. Miranda, R. Tormos and I. Vayá, Photochem. Photobiol. Sci., 2004, 3, 1038-1041.
- 14 A. Weller, Z. Phys. Chem., 1982, 133, 93-98.
- 15 a) E_{OX} (2-phenylphenol) = 0.831 V, taken from: P. Winget, C. J. Cramer and D. G. Truhlar, *Theor. Chem. Acc.*, 2004, 112, 217; b) E_{RED} (Trp) = -1.55 V, taken from: S. Cakir and E. Bicer, J. Chil. $\textit{Chem. Soc.}, \ 2010, \ \textbf{55}, \ 236; \ c) \ E_{OX} \ (indole) \ = \ 1.015 \ \ V, \ E_{RED}$ (biphenyl) = -2.55 V, taken from S. L. Murov, I. Carmichael, and G. L. Hug, Handbook of Photochemistry, 2nd Edition; Marcel Dekker: New York, 1993. 8.
- 16 Handbook of Chemistry and Physics, 85th Edition; CRC Press: Boca Raton, FL, 2004.
- M. Montalti, A. Credi, L. Prodi and M. T. Gandolfi, Handbook of Photochemistry, CRC Press, Taylor and Francis Group, Boca Raton Fl., 2006.