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The effect of Echinacea preparations in three laboratory tests of anxiety. Comparison with chlordiazepoxide

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view

COVER SHEET

Title: The effect of Echinacea preparations in three laboratory tests of anxiety. Comparison with chlordiazepoxide

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Running head: Echinacea and anxiety

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Abstract

Echinacea preparations are traditionally used to treat upper respiratory infections and inflammations. No psychotropic effects of Echinacea were reported so far, although some recently reported active constituents are behaviorally active. Prompted by these findings, we evaluated the anxiolytic potential of 5 different Echinacea preparations. Three of these decreased anxiety but two of them had a very narrow effective dose range. Only one extract decreased anxiety within a wide dose-range (3-8 mg/kg). Anxiolytic effects were consistently seen in three different tests of anxiety, the elevated plus-maze, social interaction and shock-induced social avoidance tests. No locomotor suppressant effects were seen at any dose. Noteworthy, the doses that showed anxiolytic effects in the present study were much lower than those used in the laboratory models of the traditional indications. Chlordiazepoxide robustly decreased anxiety-like behavior in all tests but suppressed locomotion at higher doses. Perceived and real risks of conventional medications increase the demand for alternative therapies, provided that these are safe and efficient. Earlier evidence shows that Echinacea preparations have an excellent safety profile, while our findings suggest for the first time that certain preparations have a considerable anxiolytic potential. Further research is required to identify factors that differentiate efficient and inefficient preparations.

Keywords: Echinacea, anxiety, rat, elevated plus-maze, social interaction, stress-induced social avoidance

Introduction

Echinacea preparations have been used for centuries by Native Americans as anti-inflammatory agents and for the treatment of upper respiratory infections; such treatments were incorporated into Western medicine about a century ago (Barnes et al., 2005; Birt et al., 2008; Blumenthal and Busse, 1998; Borchers et al., 2000). No psychotropic effects of Echinacea preparations have yet been elucidated, but recent studies identified active constituents by which these may affect behavior, especially anxiety-like behavior.

Echinacea preparations contain a large number of different alkamides, the structures of which are very similar to that of the endocannabinoid anandamide (Bauer and Remiger, 1989; Wu et al., 2009). Certain alkamides behave as cannabinomimetics at both the cannabinoid CB1 and CB2 receptor, and inhibit the anandamide-degrading enzyme fatty acid amid hydrolase (FAAH) (Woelkart et al., 2005). Although the interaction between cannabinoids and anxiety is complex (Witkin et al., 2005), CB1 signaling has been repeatedly implicated in the control of anxiety (Freund, 2003; Haller et al., 2002). Additionally, FAAH inhibition is considered a promising new anxiolytic drug target (Piomelli et al., 2006). Unidentified components of Echinacea extracts have been shown to be agonists at the transient receptor potential vanilloid-1 (TRPV1) receptors (Birt et al., 2008), a mechanism that has also been implicated in the control of anxiety (Starowicz et al., 2008). Certain Echinacea preparations contain rosmarinic acid, which has decreased anxiety in laboratory models at low doses (Biber et al., 2009; Pereira et al., 2005). In addition, caffeic acid is one of the major constituents of Echinacea; this compound decreases anxiety by indirectly modulating α_{1A} adrenoceptors (Takeda et al., 2003). Also noteworthy is the fact that the above-listed constituents of Echinacea preparations readily cross the blood-brain barrier (Birt et al., 2008; Woelkart et al., 2009; Konishi et al., 2005).

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Some of the above-listed effects of Echinacea components are weak, and/or the brain concentrations after oral administration are too low to effectively modulate the mechanisms involved in anxiety control. For example, the K_i values of CB1 binding and the brain concentrations of alkamides exclude robust effects of orally administered alkamides on CB1 receptors (Woelkart et al., 2005, 2009). Yet, anandamide –the structure of which is very similar to that of alkamides– affects receptors other than the CB1, e.g. de PPAR α receptor (Mazzola et al., 2009). It is also worth noting that the TRPV1 agonist activity of Echinacea preparations is rather strong as it is produced at doses similar to those employed here (Birt et al., 2008). These considerations suggest that particular components of Echinacea preparations, alone or in combination with synergistic/additive effects, show great promise for anxiolytic drug development.

Prompted by the above findings, we started a series of studies to evaluate the anxiolytic potential of Echinacea preparations. It is noteworthy that, despite the extensive research on and wide use of this plant, no psychotropic effects have thus far been identified. We believe that this paucity of data is mainly due to two factors. First, the anxiolytic effect was detected at doses that were about one order of magnitude lower than those used for the traditional indications (see below). Secondly, only a fraction of Echinacea preparations showed consistent anxiolytic effects in our studies. In addition, the improvement of infectious diseases inherently improves mood, which may subjectively mask the intrinsic anxiolytic effects of the extracts that do show anxiolytic effects.

51 **Materials and Methods**

52 *Animals*

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Subjects were 3-month-old male Wistar rats provided by Charles River Laboratories (Budapest, Hungary), and weighing 250-300 g. Rats were fed on standard laboratory food (Charles River Laboratories). Water was available ad libitum. Temperature and relative

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3 humidity were kept at $22\pm 2^{\circ}\text{C}$ and $60\pm 10\%$, respectively. A light/dark cycle of 12 h was
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5 ensured with lights on at 0700 h. All rats were housed in 1354G Eurostandard Type 4 cages
6
7 (59.5×38×22 cm) in groups of four. Acclimatization to housing conditions lasted at least 1
8
9 week. All subjects were experimentally naive and used in only one experiment each (with no
10
11 drug history prior to the experiment).
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15 Experiments were carried out in accordance with the European Communities Council
16
17 Directive of 24 November 1986 (86/609/EEC) and were reviewed and approved by the
18
19 Animal Welfare Committee of the Institute of Experimental Medicine.
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21

22 *Experimental design*

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24
25 *Experiment 1* evaluated the anxiolytic potential of Extract No. 1 in the elevated plus-
26
27 maze test. This experiment was also used to establish effective dose-ranges; therefore, 7
28
29 doses were tested (vehicle, 0.5, 1, 1.5, 2, 3, and 6 mg/kg). Rats were submitted to the test 30
30
31 min after treatment. Sample size was 10 per treatment group.
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35 *Experiment 2-5* tested the efficacy of four additional extracts under similar conditions.
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37 Based on the results of Experiment 1, we tested these extracts in the 1-5 mg/kg dose-range,
38
39 except for Extract No. V, which proved to have a very narrow effective dose range in
40
41 preliminary studies. Therefore, the increment of doses was 0.5 mg/kg with this extract. The
42
43 four extracts were tested in separate experiments. Sample size was 8-10 per group, except for
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45 Extract No V, where sample size was 16 per group.
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49 Based on the results of Experiments 1-5, Extract No IV was selected for further
50
51 analysis. In *Experiment 6 and 7*, we studied a wider dose range (1-8 mg/kg) in the elevated
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53 plus-maze, 30 min and 1h after treatments, respectively. Sample size was 12 per group. The
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55 doses that proved most effective in Experiments 6 and 7 were also studied in the social
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57 interaction and shock-induced social avoidance tests (*Experiments 8-9*). Sample size was 12
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3 and 8 per group, respectively. Finally, locomotor effects were studied in the open-field test
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6 (*Experiment 10*). The doses tested were 2-16 mg/kg; sample size was 15-16 per group.
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8 The behavioral effects of the benzodiazepine chlordiazepoxide were also studied for
9
10 comparison. In *Experiment 11*, we studied the effects of 1, 2, 3, 4, 6, 8, 10, 12, 14 mg/kg
11
12 chlordiazepoxide in the elevated plus-maze, 30 min after treatment. This detailed dose-
13
14 response curve was taken as chlordiazepoxide was shown to have a biphasic effect on
15
16 locomotion. Sample size was 10-14 per treatment group. *Experiment 12* established the
17
18 effects of chlordiazepoxide in the elevated plus-maze 1h after treatments. The dose-range
19
20 studied was 2-8 mg/kg, and included doses that increased and decreased locomotion in
21
22 *Experiment 11*. Sample size was 10 per group. The effects of chlordiazepoxide in the social
23
24 interaction and shock-induced social avoidance tests were investigated in *Experiments 13 and*
25
26 *14*. In the social interaction test, rats were given 1, 2, 5, and 10 mg/kg chlordiazepoxide.
27
28 Sample size was 12 per group. A specific dose-range was investigated in the shock-induced
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30 social avoidance test. Previous experience showed that this test is especially sensitive to the
31
32 sedative effects of chlordiazepoxide. Therefore, rats were treated with 0.5, 1, 2.5, and 5
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34 mg/kg in this test. Because both the anxiolytic and sedative effects of chlordiazepoxide were
35
36 robust in earlier experiments, sample size was 7 in this experiment.
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43 *Pharmacological treatment*

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46 The following extracts were studied in the experiments: Echinacea purpurea root
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48 extract (EPO Istituto Farmacochimico Fitoterapico, Milano, Italy; batch No 0700326;
49
50 **extraction procedure: ethanol 4% V/V; ratio herbal drug : drug preparation: 5-12:1;**
51
52 **excipient: maize dextrin 30%; marker: Echinacoside 4%**), Echinacea purpurea herb
53
54 extract (Finzelberg GmbH and Co. KG, Andernach, Germany; Batch No. 07022307;
55
56 **extraction: ethanol 60% M/M; ratio herbal drug : native extract: 4-10 : 1; excipients:**
57
58 **maltodextrin 13% and colloidal silica anhydrate 2%; marker: total phenols 4%**), and
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3 Echinacea angustifolia root extract (Euromed SA, Millet del Valles, Spain; Batch No.
4 419061; **extraction: ethanol 85% V/V; ratio herbal drug : native extract: 6.5-8 : 1;**
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8 **excipient: maltodextrin 30%; marker: Echinacoside 4%**). All three extracts were dissolved
9
10 in 0.4% methylcellulose in saline. We also studied a hydroalcoholic extract of Echinacea
11
12 purpurea roots (FitoChem Ltd. Monor, Hungary; Batch No. FECH-010011205; **extraction:**
13
14 **ethanol 70% V/V; ratio herbal drug : native extract: 4-8 : 1; excipient: none; marker: not**
15
16 **standardized**). Before experimental use, the ethanol was evaporated at 4 °C, and the dry
17
18 residue was dissolved in 0.4% methylcellulose in saline. Finally, we investigated an
19
20 Echinacea angustifolia root extract prepared by the Department of Pharmacognosy, Faculty
21
22 of Pharmacy, University of Szeged (Hungary). Briefly, 80 g air-dried and powdered root of
23
24 Echinacea purpurea was extracted with 1x300 and 2x200 ml 70% EtOH using an ultrasonic
25
26 extractor for 10-10 min. The filtered extracts were combined, concentrated and dissolved in
27
28 15 ml water. The aqueous solution was lyophilized, yielding 8.67 g dark brown and semi-
29
30 fluid extract. Before experimental use, it was physically dispersed, dissolved in small
31
32 amounts of dimethyl sulphoxide (DMSO) and diluted to the final volume by 0.4%
33
34 methylcellulose. **The final concentration of DMSO was 3.3% at each test concentration.**
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36 Controls received a similar mixture of DMSO and methylcellulose. Chlordiazepoxide came
37
38 from Sigma (Budapest, Hungary) and was dissolved in 0.4% methylcellulose in saline.
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46 All treatments were administered *per os* in a volume of 2 ml/kg. Controls received
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48 0.4% methylcellulose except for the last mentioned extract (see above).
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50 51 *Behavioral tests*

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53 Behavioral studies were performed in the early hours of the dark (active) period
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55 between 1100 and 1300 h. Each experiment was performed in several series balanced over
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57 groups. Group assignment was random. Behavior was video recorded and later analyzed by
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59 an experimenter blind to treatment conditions.
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3 The tests were performed as described earlier (Haller et al., 2000; Haller and Bakos,
4 2002; Haller et al., 2003; Leveleki et al., 2006). Briefly, *the elevated plus-maze* (arm length
5 50 cm, arm width 15 cm, wall height 30 cm and platform height 80 cm) was illuminated by
6 dim red light. Rats were placed into the center of the apparatus and were allowed to explore it
7 for 5 min. Closed-arm entries were considered indicators of locomotor activity, whereas open
8 arm exploration (duration and % entries) was used as a measure of anxiety (Pellow et al.
9 1985). The *social interaction test* arena was a plastic box of 40x60x60 cm that was lit by
10 white light. Pairs of similarly treated rats were placed in the arena, and their behavior was
11 recorded for 10 min. Subjects were unfamiliar to each other. In this test, anxiety is shown by
12 the duration of social interactions defined as sniffing movements directed towards the partner
13 rat. Exploration/walking and resting are indicative of sedative or muscle-relaxant effects (File
14 and Hyde, 1978; File and Johnston, 1989; Guy and Gardner, 1985). *The social avoidance test*
15 was performed on two consecutive days. On the first day, rats were exposed to 10 electric
16 shocks (3 mA) over 5 min in a plastic box of 30x30x30 cm. Controls were placed into the
17 box, but received no shocks. On the next day, rats were studied in a three-compartment
18 plastic cage. The subject was placed in the habituation compartment (15 x 40 cm) that was
19 separated from the rest of the cage by an opaque sliding door. After 3 min, the sliding door
20 was removed and the rat was allowed to explore the test arena (40 cm x 40 cm) for 5 min.
21 The third compartment contained a large unfamiliar male confined behind a transparent,
22 perforated Plexiglas wall. In this test, anxiety is indicated by the number of entries into, and
23 the time spent in the test arena i.e. in the vicinity of the unfamiliar opponent. Shock exposure
24 strongly inhibits opponent visits, a response that is abolished by anxiolytics (Leveleki et al.,
25 2006). *The open-field* was a circular area with a diameter of 90 cm. Rats were placed next to
26 the wall of the open-field and were allowed to explore the arena for 10 min. The arena was
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3 divided into equal sub-areas by concentric and radial lines. Locomotor activity was shown by
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5 the number of line crossings.
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8 *HPLC analysis*

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10 **HPLC analysis was carried out on a Waters instrument (Milford, MA, USA;**
11 **solvent delivery system 6000A, pump 600E, UV detector type 2487, Rheodyne 7725i**
12 **injector, Empower software). The alkamide profile was investigated at 25 °C on**
13 **LiChrospher RP-18 column (5 µm, 125-4 mm, Merck) using linear gradient of**
14 **acetonitrile-H₂O 2:3→4:1 (0-30 min) at flow of 1 ml/min detected at 254 nm. Peak**
15 **assignment was made according to Bauer and Remiger (1989).**
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24 *Statistical analysis*

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27 **Statistical analysis was made by the STATISTICA software (Tulsa, USA). Data**
28 **were presented as means ± the standard error of the mean (SEM). Significance level**
29 **was set at p < 0.05. Behavioral data were analyzed by Kruskal-Wallis ANOVA. Mann-**
30 **Whitney post-hoc comparisons were also run where appropriate.**
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37 **Results**

38 *Screening experiments*

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42 Extract No I significantly increased open arm exploration with marginal effects on
43 locomotion as shown by closed arm entries (Table 1). However, only the 1.5 and 2 mg/kg
44 doses were effective; neither 1 nor 3 mg/kg decreased anxiety in the elevated plus-maze. In
45 addition, the time spent on the open arms was significantly increased, but open arm choice
46 (% open arm entries) showed non-significant variation only.
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54 Extracts No II and III failed to affect behavior (Table 2). Extract No IV appeared to
55 dose-dependently increase open arm exploration (Table 2). The effect was significant at the
56 highest dose tested. Closed arm entries were not changed by the treatments. Extract No V
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3 increased the duration of open arm exploration at 1 mg/kg only; neither 0.5 nor 1.5 mg/kg
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5 were effective (Table 2). In addition, % open arm entries showed no significant changes.
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8 *Behavioral effects of Extract No IV*

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10 The effects of extract No IV were investigated in the elevated plus-maze over a wider
11 dose-range, both 30 min and 1h after treatments (Fig. 1, upper panels). Locomotion as shown
12 by closed arm entries were not affected at either time-point ($H(7,120)= 5.18$; $p > 0.5$, and
13 $H(7,120)= 7.01$; $p > 0.4$, respectively). The duration of open arm entries increased at both
14 time points (30 min: $H(7,120)= 22.96$; $p < 0.01$); 1h: $H(7,120)= 19.65$; $p < 0.01$). Open arm
15 choice was affected significantly after 30 min ($H(7,120)= 23.70$; $p < 0.01$) but not after 1h
16 ($H(7,120)= 10.79$; $p = 0.14$). Post-hoc comparisons showed that the effective dose-range was
17 3-6 mg/kg after 30 min and 4-8 mg/kg after 1h.
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29 In the social interaction test, resting, exploration and agonistic interactions showed non-
30 significant changes ($H(3,80)$ values were 1.59, 3.79, and 0.42; p values were larger than 0.3
31 at least) (Fig. 1. lower left-hand panel). Social interactions were increased by both the 3 and 4
32 mg/kg dose ($H(3,80)= 14.72$, $p < 0.001$).
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39 In the shock-induced social avoidance model, the extract abolished social avoidance at
40 3 and 4 mg/kg ($H(6, 80)= 20.11$; $p < 0.005$) (Fig. 1. lower middle panel). Locomotion was not
41 affected by the extract within the 2-16 mg/kg dose range ($H(4,79)= 1.4$; $p > 0.8$) (Fig. 1. lower
42 left-hand panel).
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49 *The effects of chlordiazepoxide*

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51 As expected, chlordiazepoxide significantly decreased anxiety-like behavior in the
52 elevated plus-maze both 30 min and 1h after treatments (% open arm entries 30 min:
53 $H(9,132) = 29.27$; $p < 0.005$; % open arm entries 1h: $H(4,60) = 15.48$; $p < 0.005$; % time in
54 open arm 30 min: $H(9,132) = H(9,132) = 34.38$; $p < 0.0001$; % time in open arm 1h: $H(4,60)$
55 = 19.63; $p < 0.001$) (Fig. 2, upper panels). However, the anxiolytic effects of chlordiazepoxide
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3 were not devoid of locomotor effects (30 min: $H(9,132)= 33.13$; $p < 0.0001$; 1h: $H(4,60)=$
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5
6 9.83; $p < 0.04$). Post-hoc comparisons showed that 30 min after treatments, chlordiazepoxide
7
8 increased locomotion at 2 and 3 mg/kg, and decreased locomotion above 8 mg/kg. The dose
9
10 range within which chlordiazepoxide decreased anxiety without affecting locomotion was 4-6
11
12 mg/kg at 30 min, and 4 mg/kg at 1h.

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14
15 Chlordiazepoxide significantly increased social interactions in the social interaction test
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17 ($H(4,72)= 13.39$; $p < 0.01$) (Fig. 2, lower left hand panel). No locomotion-suppressing effects
18
19 were noticed in this test, as neither resting, nor exploration were affected significantly
20
21 ($H(4,72)= 8.89$; $p < 0.07$ and $H(4,72)= 3.24$; $p > 0.5$). We note that the marginally significant
22
23 difference in resting shown by the Kruskal-Wallis test was due to differences between
24
25 chlordiazepoxide-treated groups (the lowest resting value was seen at chlordiazepoxide 2
26
27 mg/kg). In contrast to extract No. IV, chlordiazepoxide dramatically reduced agonistic
28
29 interactions ($H(4,72)= 29.60$; $p < 0.0001$). In the shock-induced social avoidance test, 0.5 and
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31 1 mg/kg did, whereas 2.5 and 5 mg/kg did not abolish social avoidance ($H(5, 42)= 31.89$; $p <$
32
33 0.0001) (Fig. 2. lower right-hand panel). The failure of higher doses to abolish social
34
35 avoidance was likely due to sedation, as opponent entries were reduced from 5.14 ± 0.70 at 1
36
37 mg/kg chlordiazepoxide to 2.29 ± 0.57 and 1.14 ± 0.55 at 2.5 mg/kg and 5 mg/kg
38
39 chlordiazepoxide, respectively. Conversely, unshocked controls showed 5.57 ± 0.61 entries.

44 45 *HPLC measurements*

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48 The results of HPLC measurements were summarized in Fig. 3. Two of the
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50 extracts (No. 2 and No. 3), contained very low amounts of alkamides (Fig. 3 upper right
51
52 hand panel). Alkamides were not lacking in these extracts (see the insert of Fig. 3 upper
53
54 right hand panel), but their absorption curves became almost flat when the scale of the
55
56 absorption curve was set such to accommodate the high levels seen in other extracts.
57
58 Alkamide contents were considerably higher in Extracts No 1, 4, and 5. The level of
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3 **these compounds and, more importantly their fingerprint showed considerable**
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5 **differences in these extracts.**
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8 **Discussion**

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11 Out of the five Echinacea extracts studied, three affected anxiety-like behavior. Two of
12 these, however, had a very narrow dose range, encompassing no more than 0.5 mg/kg.
13
14 Neither lower nor higher doses proved effective. There was only one extract that robustly
15
16 decreased anxiety in three different tests, with an effective dose-range that was comparable
17
18 with that of chlordiazepoxide. Locomotion was not affected by this extract at doses that were
19
20 5 times larger than the minimally effective anxiolytic dose. Chlordiazepoxide robustly
21
22 decreased anxiety-like behavior but also suppressed locomotion.
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27 **The anxiolytic effects of chlordiazepoxide were consistent with those reported**
28 **earlier in the same tests (File and Hyde, 1978; Leveleki et al., 2006; Patel and Malick,**
29 **1982; Pellow et al., 1982; Treit et al., 1981). Locomotor effects were also similar to those**
30 **reported earlier; low doses (e.g. 2-3 mg/kg) increased, whereas larger doses (e.g. 5-10**
31 **mg/kg) suppressed locomotion (Davies and Steinberg, 1984; File and Pellow, 1985;**
32 **Fernandes and File, 1999; Martin et al., 1982; McElroy et al., 1982). Also consistent**
33 **with earlier findings, locomotor stimulation that was seen at 30 min disappeared when**
34 **effects were tested 1h after injections (Davies and Steinberg, 1984).**
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46 To our knowledge, this is the first study showing that Echinacea preparations possess an
47 anxiolytic potential. Unfortunately, however, this potential is not common, as only three out
48 of five preparations showed any anxiolytic effect. In addition, the effective dose range was
49 very narrow (0.5 mg/kg) in the case of two effective extracts. Noteworthy, very narrow dose
50 ranges are irrelevant from a therapeutic perspective as such data are difficult to extrapolate to
51 effective doses in humans. In addition, a very narrow dose-range involves the establishment
52 of precise weight-dependent dosage regimens, which appears unrealistic in a clinical setting.
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3 Extract No. 4, however, decreased anxiety at a low dose and over a wide, bell-shaped dose-
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5 response curve that was comparable with that of chlordiazepoxide. **Such bell-shaped dose-**
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7 **response curves are commonly seen with herbal extracts (Butterweck et al., 1997).**

8
9
10 **Moreover, this phenomenon □ also called hormesis □ appears to be a general feature of**
11
12 **drug effects (Calabrese and Baldwin, 2001, Calabrese, 2008).**

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14
15 Discrepant findings with different extracts are likely explained by the fact that while the
16
17 main constituents of various Echinacea species are essentially similar, the absolute amounts
18
19 and the quantitative relationships between different constituents are subject to major variation
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21 (Kapteyn et al., 2002; Pellati et al., 2005; Percival, 2000; Perry et al., 2001; Wu et al., 2009).
22
23 Noteworthy, the immunostimulant effect of Echinacea preparations also show extreme
24
25 variations (Tamta et al., 2008). **We hypothesized that Echinacea preparations affect**
26
27 **anxiety-related behaviors mainly due to their alkamide content (see Introduction). This**
28
29 **hypothesis was partly supported by HPLC findings, as the extracts that contained the**
30
31 **lowest amounts of alkamides were behaviorally inactive. Efficacy, however, was not**
32
33 **directly related to alkamide content. Extract No 5 contained high amounts of alkamides**
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35 **but had modest behavioral effects, while Extract No 4 strongly decreased anxiety**
36
37 **despite its relatively lower alkamide content. One can hypothesize that behavioral**
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39 **effects depend partly on the total amount, partly on the fingerprint of the different**
40
41 **alkamides. As such, behavioral effects may develop in conjunction with the**
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43 **additive/synergistic and possibly antagonistic effects of various alkamides. The**
44
45 **involvement of other components cannot be ruled out either.** Further research is required
46
47 to elucidate the compositions that differentiate efficient and inefficient preparations.
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56 **The perceived or real risks of conventional drug treatments –as well as personal**
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58 **beliefs about healthy lifestyles– lead to a strong demand for alternative therapies among**
59
60 **both patients and medical practitioners (Astin, 1998; Barrett et al., 2003; Ben-Arye et**

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2
3 al., 2008; Dilhuydy, 2003). Such alternative therapies would be welcome provided that
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5 the treatment is safe and its efficacy is proven. Among herbs, anxiolytic effects are not
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7 unique to Echinacea. Effective doses, however, are rather high in most cases (*Albizzia*
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9 *julibrissin*: 100-200 mg/kg in the rat plus-maze, Kim et al., 2004; *Centella asiatica*: 500
10
11 mg/kg in the rat plus-maze, Wijeweera et al., 2006; *Eschsholzia californica*: 25 mg/kg in
12
13 the mouse light/dark test, Rolland et al., 1991; *Hypericum perforatum*: 100-200 mg/kg in
14
15 the mouse T-maze, Flausino et al., 2002; *Passiflora species*: 300-800 mg/kg in the rats
16
17 plus-maze, Reginatto et al., 2006; *Valeriana officinalis*: above 100 mg/kg, various tests,
18
19 Oliva et al., 2004). In other cases, the effective dose-range was as narrow as with our
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21 Extracts No. 1 and 5 (Hasenohrl et al., 1998). More promising effects were obtained
22
23 with kava kava (*Piper mysticum*) that proved to be clinically effective (Pittler and
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25 Ernst, 2003). Later research, however, revealed that this plant has major side effects
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27 (Christl et al., 2009; Teschke et al., 2008).
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34 Evidence accumulated over many decades demonstrates that Echinacea preparations
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36 have an excellent safety profile (Barnes et al., 2005; Birt et al., 2008; Blumenthal and Busse,
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38 1998; Borchers et al., 2000; Izzo and Ernst, 2001; Tesch, 2003). In addition, the doses that
39
40 showed anxiolytic effects in the present study (3-7 mg/kg) were about one order of magnitude
41
42 lower than those that were efficient in laboratory models of traditional indications (30-130
43
44 mg/kg; Abouelella et al., 2007; Zhai et al., 2007). This comparison suggests that human
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46 anxiolytic doses would be similarly low, which further decreases the risks associated with the
47
48 potential use of Echinacea preparations for the treatment of anxiety. Taken together, these
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50 considerations suggest that from the point of view of safety, Echinacea preparations are
51
52 excellent candidates for the alternative treatment of anxiety. On the other hand, the
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54 remarkable anxiolytic effects demonstrated here suggest that certain Echinacea preparations
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56 are excellent alternative anxiolytics from the point of view of efficacy as well.
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References

- Abouelella AM, Shahein YE, Tawfik SS, Zahran AM. 2007. Phytotherapeutic effects of *Echinacea purpurea* in gamma-irradiated mice. *J Vet Sci* **8**: 341-351.
- Astin JA. 1998. Why patients use alternative medicine: results of a national study. *JAMA* **279**: 1548-1553.
- Barnes J, Anderson LA, Gibbons S, Phillipson JD. 2005. *Echinacea* species (*Echinacea angustifolia* (DC.) Hell., *Echinacea pallida* (Nutt.) Nutt., *Echinacea purpurea* (L.) Moench): a review of their chemistry, pharmacology and clinical properties. *J Pharm Pharmacol* **57**: 929-954.
- Barrett B, Marchand L, Scheder J, Plane MB, Maberry R, Appelbaum D, Rakel D, Rabago D. 2003. Themes of holism, empowerment, access, and legitimacy define complementary, alternative, and integrative medicine in relation to conventional biomedicine. *J Altern Complement Med* **9**: 937-947.
- Bauer R, Remiger P. 1989. TLC and HPLC Analysis of Alkamides in *Echinacea* Drugs. *Planta Med* **55**: 367-371.
- Ben-Arye E, Frenkel M, Klein A, Scharf M. 2008. Attitudes toward integration of complementary and alternative medicine in primary care: perspectives of patients, physicians and complementary practitioners. *Patient Educ Couns* **70**: 395-402.
- Biber A, Franck-Karl G, Waimer F, Riegert U, Wiget R. 2009. Analytical characterisation of homoeopathic mother tinctures. *Pharmeur Sci Notes* **2009(1)** :1-4.
- Birt DF, Widrlechner MP, Lalone CA, Wu L, Bae J, Solco AK, Kraus GA, Murphy PA, Wurtele ES, Leng Q, Hebert SC, Maury WJ, Price JP. 2008. *Echinacea* in infection. *Am J Clin Nutr* **87**: 488S-492S.
- Blumenthal M, Busse WR. 1998. *The complete German Commission E monographs, Therapeutic guide to herbal medicines*. Austin, Texas American Botanical Council: Boston.
- Borchers AT, Keen CL, Stern JS, Gershwin ME. 2000. Inflammation and Native American medicine: the role of botanicals. *Am J Clin Nutr* **72**: 339-347.
- Butterweck V, Wall A, Liefländer-Wulf U, Winterhoff H, Nahrstedt A. 1997. Effects of the total extract and fractions of *Hypericum perforatum* in animal assays for antidepressant activity. *Pharmacopsychiatry* **30 Suppl 2**:117-124.
- Calabrese EJ, Baldwin LA. 2001. U-shaped dose-responses in biology, toxicology, and public health. *Annu Rev Public Health* **22**:15-33.
- Calabrese EJ. 2008. Hormesis and medicine. *Br J Clin Pharmacol* **66**: 594-617.

1
2
3 Christl SU, Seifert A, Seeler D. 2009. Toxic hepatitis after consumption of traditional
4 kava preparation. *J Travel Med* **16**: 55-56.

5
6
7 Davies C, Steinberg H. 1984. A biphasic effect of chlordiazepoxide on animal
8 locomotor activity. *Neurosci Lett* **46**: 347-351.

9
10 Dilhuydy JM. 2003. Patients' attraction to complementary and alternative medicine
11 (CAM): a reality which physicians can neither ignore nor deny. *Bull Cancer* **90**: 623-628.

12
13
14 Fernandes C, File SE. 1999. Dizocilpine does not prevent the development of tolerance
15 to the anxiolytic effects of diazepam in rats. *Brain Res* **815**: 431-434.

16
17
18 File SE, Cheeta S, Akanezi C. 2001. Diazepam and nicotine increase social interaction
19 in gerbils: a test for anxiolytic action. *Brain Res* **888**: 311-313.

20
21
22 File SE, Hyde JRG. 1978. Can social interaction be used to measure anxiety? *Br J*
23 *Pharmacol* **62**: 19-24.

24
25
26 File SE, Johnston AL. 1989. Lack of effects of 5HT₃ receptor antagonists in the social
27 interaction and elevated plus-maze tests of anxiety in the rat. *Psychopharmacology* **99**: 248-
28 251.

29
30
31 File SE, Pellow S. 1985. The effects of triazolobenzodiazepines in two animal tests of
32 anxiety and in the holeboard. *Br J Pharmacol* **86**: 729-735.

33
34
35 Flausino OA Jr, Zangrossi H Jr, Salgado JV, Viana MB. 2002. Effects of acute and
36 chronic treatment with *Hypericum perforatum* L. (LI 160) on different anxiety-related
37 responses in rats. *Pharmacol Biochem Behav* **71**: 251-257.

38
39
40 Freund TF. 2003. Interneuron Diversity series: Rhythm and mood in perisomatic
41 inhibition. *Trends Neurosci* **26**: 489-495.

42
43
44 Guy AP, Gardner CR. 1985. Pharmacological characterization of a modified social
45 interaction model of anxiety in the rat. *Neuropsychobiology* **13**, 194-200.

46
47
48 Haller J, Bakos N. 2002. Stress-induced social avoidance: a new model of stress-
49 induced anxiety? *Physiol Behav* **77**: 327-332.

50
51
52 Haller J, Halász J, Makara GB. 2000. Housing conditions and the anxiolytic efficacy of
53 buspirone: the relationship between main and side effects. *Behav Pharmacol* **11**: 403-412.

54
55
56 Haller J, Leveleki C, Baranyi J, Mikics E, Bakos N. 2003. Stress, social avoidance and
57 anxiolytics: a potential model of stress-induced anxiety. *Behav Pharmacol* **14**: 439-446.

58
59
60 Haller J, Varga B, Ledent C, Freund TF. 2004. CB1 cannabinoid receptors mediate
anxiolytic effects: convergent genetic and pharmacological evidence with CB1-specific
agents. *Behav Pharmacol* **15**: 299-304.

- 1
2
3 Hasenohrl RU, Topic B, Frisch C, Häcker R, Mattern CM, Huston JP. 1998.
4
5 Dissociation between anxiolytic and hypnestic effects for combined extracts of zingiber
6 officinale and ginkgo biloba, as opposed to diazepam. *Pharmacol Biochem Behav* **59**: 527-
7
8 535.
9
10 Izzo AA, Ernst E. 2001. Interactions between herbal medicines and prescribed drugs: a
11
12 systematic review. *Drugs* **61**: 2163-2175.
13
14 Kapteyn J, Goldsbrough B, Simon E. 2002. Genetic relationships and diversity of
15
16 commercially relevant Echinacea species. *Theor Appl Genet* **105**: 369-376.
17
18 Kim WK, Jung JW, Ahn NY, Oh HR, Lee BK, Oh JK, Cheong JH, Chun HS, Ryu JH.
19
20 2004. Anxiolytic-like effects of extracts from Albizzia julibrissin bark in the elevated plus-
21
22 maze in rats. *Life Sci* **75**: 2787-2795.
23
24 Konishi Y, Hitomi Y, Yoshida M, Yoshioka E. 2005. Pharmacokinetic study of caffeic
25
26 and rosmarinic acids in rats after oral administration. *J Agric Food Chem* **53**: 4740-4746.
27
28 Leveleki C, Sziray N, Levay G, Barsvari B, Soproni K, Mikics E, Haller J. 2006.
29
30 Pharmacological evaluation of the stress-induced social avoidance model of anxiety. *Brain*
31
32 *Res Bull* **69**: 153-160.
33
34 Martin JR, Oettinger R, Driscoll P, Buzzi R, Bättig K. 1982. Effects of
35
36 chlordiazepoxide and imipramine on maze patrolling within two different maze
37
38 configurations by psychogenetically selected lines of rats. *Psychopharmacology* **78**: 58-62.
39
40 Mazzola C, Medalie J, Scherma M, Panlilio LV, Solinas M, Tanda G, Drago F, Cadet
41
42 JL, Goldberg SR, Yasar S. 2009. Fatty acid amide hydrolase (FAAH) inhibition enhances
43
44 memory acquisition through activation of PPAR-alpha nuclear receptors. *Learn Mem* **16**:
45
46 332-337.
47
48 McElroy JF, Fleming RL, Feldman RS. 1985. A comparison between chlordiazepoxide
49
50 and CL 218,872--a synthetic nonbenzodiazepine ligand for benzodiazepine receptors on
51
52 spontaneous locomotor activity in rats. *Psychopharmacology* **85**: 224-226.
53
54 Melchart D, Linde K, Worku F, Sarkady L, Holzmann M, Jurcic K, Wagner H. 1995.
55
56 Results of five randomized studies on the immunomodulatory activity of preparations of
57
58 Echinacea. *J Altern Complement Med* **1**: 145-160.
59
60 Oliva I, Gonzalez-Trujano ME, Arrieta J, Enciso-Rodriguez R, Navarrete A. 2004.
Neuropharmacological profile of hydroalcohol extract of Valeriana edulis ssp. procera roots
in mice. *Phytother Res* **18**: 290-296.
Patel JB, Malick JB. 1982. Pharmacological properties of trazolam: a new non-
benzodiazepine anxiolytic agent. *Eur J Pharmacol* **78**: 323-333.

- 1
2
3 Pellati F, Benvenuti S, Melegari M, Lasseigne T. 2005. Variability in the composition
4 of anti-oxidant compounds in Echinacea species by HPLC. *Phytochem Anal* **16**: 77-85.
5
6
7 Pellow S, Chopin P, File SE, Briley M. 1985. Validation of open: closed arm entries in
8 an elevated plus-maze as a measure of anxiety in the rat. *J Neurosci Methods* **14**: 149-167.
9
10 Percival SS. 2000. Use of Echinacea in medicine. *Biochem Pharm* **60**:155-158.
11
12 Pereira P, Tysca D, Oliveira P, da Silva Brum LF, Picada JN, Ardenghi P. 2005.
13 Neurobehavioral and genotoxic aspects of rosmarinic acid. *Pharmacol Res* **52**: 199-203.
14
15 Perry NB, Burgess EJ, Glennie VL. 2001. Echinacea standardization: analytical
16 methods for phenolic compounds and typical levels in medicinal species. *J Agric Food Chem*
17 **49**: 1702-1706.
18
19
20
21 Piomelli D, Tarzia G, Duranti A, Tontini A, Mor M, Compton TR, Dasse O, Monaghan
22 EP, Parrott JA, Putman D. 2006. Pharmacological profile of the selective FAAH inhibitor
23 KDS-4103 (URB597). *CNS Drug Rev* **12**: 21-38.
24
25
26 Pittler MH, Ernst E. 2003. Kava extract for treating anxiety. *Cochrane Database Syst*
27 *Rev* (1):CD003383.
28
29
30 Reginatto FH, De-Paris F, Petry RD, Quevedo J, Ortega GG, Gosmann G, Schenkel
31 EP. 2006. Evaluation of anxiolytic activity of spray dried powders of two South Brazilian
32 *Passiflora* species. *Phytother Res* **20**: 348-351.
33
34
35 Rolland A, Fleurentin J, Lanhers MC, Younos C, Misslin R, Mortier F, Pelt JM. 1991.
36 Behavioural effects of the American traditional plant *Eschscholzia californica*: sedative and
37 anxiolytic properties. *Planta Med* **57**: 212-216.
38
39
40 Starowicz K, Cristino L, Di Marzo V. 2008. TRPV1 receptors in the central nervous
41 system: potential for previously unforeseen therapeutic applications. *Curr Pharm Des* **14**: 42-
42 54.
43
44
45
46 Takeda H, Tsuji M, Miyamoto J, Masuya J, Imori M, Matsumiya T. 2003. Caffeic acid
47 produces antidepressive- and/or anxiolytic-like effects through indirect modulation of the
48 alpha 1A-adrenoceptor system in mice. *Neuroreport* **14**: 1067-1070.
49
50
51 Tamta H, Pugh ND, Balachandran P, Moraes R, Sumiyanto J, Pasco DS. 2008.
52 Variability in in vitro macrophage activation by commercially diverse bulk echinacea plant
53 material is predominantly due to bacterial lipoproteins and lipopolysaccharides. *J Agric Food*
54 *Chem* **56**: 10552-10556.
55
56
57
58 Tesch BJ. 2003. Herbs commonly used by women: an evidence-based review. *Am J*
59 *Obstet Gynecol* **188**: S44-S55.
60

1
2
3 Teschke R, Schwarzenboeck A, Hennermann KH. 2008. Kava hepatotoxicity: a clinical
4 survey and critical analysis of 26 suspected cases. *Eur J Gastroenterol Hepatol* **20**: 1182-
5 1193.
6
7

8 Treit D, Pinel JP, Fibiger HC. 1981. Conditioned defensive burying: a new paradigm
9 for the study of anxiolytic agents. *Pharmacol Biochem Behav* **15**: 619-626.
10
11

12 Wijeweera P, Arnason JT, Koszycki D, Merali Z. 2006. Evaluation of anxiolytic
13 properties of Gotukola (*Centella asiatica*) extracts and asiaticoside in rat behavioral models.
14 *Phytomedicine* **13**: 668-676.
15
16

17 Witkin JM, Tzavara ET, Nomikos GG. 2005. A role for cannabinoid CB1 receptors in
18 mood and anxiety disorders. *Behav Pharmacol* **16**: 315-331.
19
20

21 Woelkart K, Frye RF, Derendorf H, Bauer R, Butterweck V. 2009. Pharmacokinetics
22 and Tissue Distribution of Dodeca-2E,4E,8E,10E/Z-tetraenoic Acid Isobutylamides after
23 Oral Administration in Rats. *Planta Med* Apr 27. [Epub ahead of print] DOI: 10.1055/s-0029-
24 1185631.
25
26
27

28 Woelkart K, Xu W, Pei Y, Makriyannis A, Picone RP, Bauer R. 2005. The
29 endocannabinoid system as a target for alkamides from *Echinacea angustifolia* roots. *Planta*
30 *Med.*; 71:701-705.
31
32

33 Wu L, Dixon PM, Nikolau BJ, Kraus GA, Widrlechner MP, Wurtele ES. 2009.
34 Metabolic profiling of echinacea genotypes and a test of alternative taxonomic treatments.
35 *Planta Med.*; 75:178-183.
36
37
38

39 Zhai Z, Liu Y, Wu L, Senchina DS, Wurtele ES, Murphy PA, Kohut ML, Cunnick JE.
40 2007. Enhancement of innate and adaptive immune functions by multiple *Echinacea* species.
41 *J Med Food.*; 10:423-434.
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Table 1. The effects of extract No I in the elevated plus-maze.

Group	Closed entries ±SEM	% open entries ±SEM	% time in open arm ±SEM
vehicle	7,40 ±0,61	28,18 ±2,04	9,53 ±1,44
<i>E_I</i> 0.05 mg/kg	7,30 ±0,63	32,59 ±3,26	12,17 ±1,78
<i>E_I</i> 1 mg/kg	8,80 ±1,16	27,17 ±2,21	12,42 ±2,33
<i>E_I</i> 1.5 mg/kg	8,67 ±0,53	37,62 ±2,08	21,41 * ±4,30
<i>E_I</i> 2 mg/kg	8,30 ±0,91	33,33 ±3,14	17,09 * ±1,99
<i>E_I</i> 3 mg/kg	<i>10,80</i> + ±0,80	30,20 ±4,36	12,23 ±3,32
<i>E_I</i> 6 mg/kg	9,30 ±0,68	27,43 ±1,83	10,91 ±1,58
<i>H</i> (6,80)	<i>12,83</i>	<i>10,62</i>	<i>16,10</i>
<i>p</i>	<i>0,05</i>	<i>0,1</i>	<i>0,01</i>

E_I, extract No. I; *H*, Kruskal-Wallis coefficient; *Post hoc comparisons*:*, significantly different from control (bolded); +, marginal difference from control (italicized).

Table 2. The effect of Extracts No II-V in the elevated plus-maze.

<i>Treatment</i>	Closed entries ±SEM	% open entries ±SEM	%time in open arm ±SEM
<i>Control</i>	6.72 ±0.91	21.46 ±4.15	6.07 ±1.36
<i>E_{II} 1mg/kg</i>	7.75 ±1.28	19.03 ±4.95	6.53 ±2.67
<i>E_{II} 3mg/kg</i>	6.80 ±1.25	18.75 ±7.89	3.08 ±1.01
<i>E_{II} 4mg/kg</i>	5.90 ±1.03	19.19 ±5.56	5.43 ±2.15
<i>E_{II} 5mg/kg</i>	7.00 ±1.02	22.76 ±5.49	8.92 ±3.18
<i>H(4,56)</i>	1.82	1.23	2.11
<i>p</i>	0.8	0.9	0.7
<i>control</i>	7.50 ±0.78	24.69 ±2.67	7.58 ±1.29
<i>E_{III} 1 mg/kg</i>	8.00 ±0.89	29.88 ±4.67	13.38 ±3.12
<i>E_{III} 2 mg/kg</i>	5.92 ±1.43	26.34 ±8.52	4.81 ±1.25
<i>E_{III} 3 mg/kg</i>	8.20 ±0.63	23.19 ±3.73	10.08 ±2.65
<i>E_{III} 4 mg/kg</i>	7.89 ±1.03	23.58 ±5.09	8.26 ±3.38
<i>H(4,55)</i>	2.10	3.55	6.26
<i>p</i>	0.7	0.5	0.2
<i>control</i>	7.33 ±0.71	11.24 ±3.35	2.18 ±0.57
<i>E_{IV} 1 mg/kg</i>	7.92 ±0.92	8.46 ±3.57	2.37 ±1.30
<i>E_{IV} 2 mg/kg</i>	6.50 ±1.10	10.00 ±3.22	2.67 ±0.84
<i>E_{IV} 3 mg/kg</i>	7.92 ±0.92	18.54 ±4.38	6.93 ⁺ ±2.08
<i>E_{IV} 4 mg/kg</i>	7.75 ±0.73	23.93 [*] ±4.12	10.43 [*] ±3.49
<i>H(4, 60)</i>	2.36	10.32	10.54
<i>p</i> <	0.7	0.04	0.03
<i>Control</i>	8,94 ±0,82	32,85 ±3,93	18,69 ±3,43
<i>E_V 0.5 mg/kg</i>	8,00 ±0,85	34,94 ±4,84	18,64 ±4,43
<i>E_V 1 mg/kg</i>	9,00 ±0,87	41,97 ±4,70	35,18 [*] ±5,72
<i>E_V 1.5 mg/kg</i>	9,56 ±0,81	40,47 ±2,66	27,66 ±3,46
<i>E_V 2 mg/kg</i>	8,63 ±1,04	37,63 ±3,25	25,81 ±3,90
<i>H(4,80)</i>	1,65	3,16	9,42
<i>p</i>	0,80	0,50	0,05

E_{I-V}, extract No. I-V; *H*, Kruskal-Wallis coefficient; *Post hoc comparisons*:*, significantly different from control (bolded); ⁺, marginal difference from control (italicized).

Legend for figures

Fig. 1. The effects of extract No 4 on anxiety-like behaviors and locomotion. E-IV, the Echinacea extract that showed promising anxiolytic effects in the screening experiments (see Table 2); *, significantly different from control in post-hoc comparisons ($p < 0.05$ at least); ⁺, marginally different from control ($0.1 > p > 0.05$).

Fig. 2. The effects of chlordiazepoxide in three anxiety tests. CDP, chlordiazepoxide; *, significantly different from control in post-hoc comparisons ($p < 0.05$ at least); ⁺, marginally different from control ($0.1 > p > 0.05$).

Fig. 3. HPLC chromatograms of the 5 extracts tested. The insert (upper right hand chromatogram) shows that Extracts No. 2 and 3 did not lack alkamides, but this is not visible at Y-axis scales appropriate to accommodate the much higher levels seen in the other three extracts. **1**, undeca-2E,4Z-diene-8,10-diynoic acid isobutylamide; **2**, undeca-2Z,4E-diene-8,10-diynoic acid isobutylamide; **3**, dodeca-2E,4Z-diene-8,10-diynoic acid isobutylamide; **4**, undeca-2E,4Z-diene-8,10-diynoic acid 2-methylbutylamide; **5**, dodeca-2E,4Z,10E-triene-8-ynoic acid isobutylamide, **6**, trideca-2E,7Z-diene-10,12-diinsav isobutylamide; **7**, dodeca-2E,4Z-diene-8,10-diynoic acid 2-methylbutylamide; **8**, dodeca-2E,4E,8Z,10E-tetraenoic acid isobutylamide; **9**, dodeca-2E,4E,8Z,10Z-tetraenoic acid isobutylamide; **11**, dodeca-2E,4E-dienoic acid; **18**, pentadeca-2E,9Z-diene-12,14-diynoic acid isobutylamide.

Haller et al., Figure 1

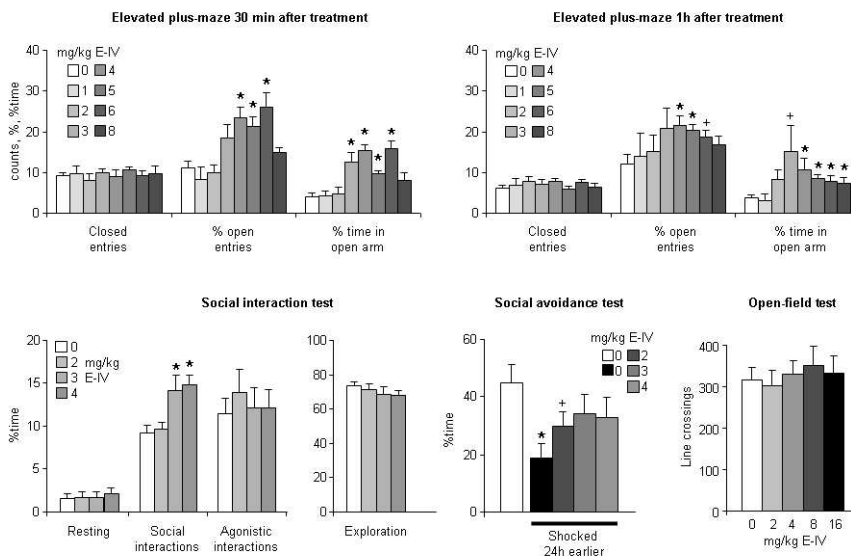


Figure 1
275x190mm (96 x 96 DPI)

Review

Haller et al., Figure 2

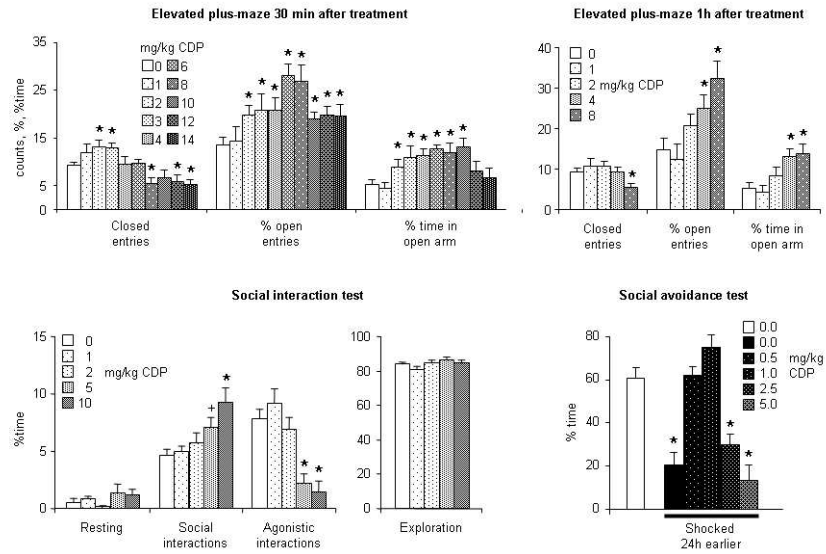
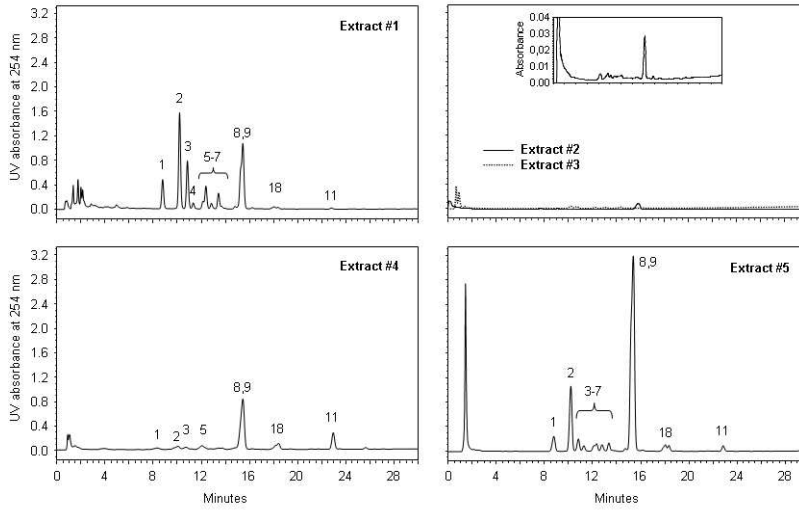


Figure 2
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275x190mm (96 x 96 DPI)

Review