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Daily serum piperacillin monitoring is advisable in critically ill patients

Nicolas Blondiaux a,*, Frédéric Wallet a, Raphaël Favory b, Thierry Onimus b, Saad Nseir b, René J. Courcol a, Alain Durocher b, Micheline Roussel-Delvallez a

a Pôle de Microbiologie, Centre de Biologie-Pathologie, Centre Hospitalier Régional Universitaire (CHRU) de Lille, Boulevard du Pr. J. Leclercq, F-59037 Lille, France

b Service de Réanimation Médicale, Hôpital A. Calmette, Centre Hospitalier Régional Universitaire (CHRU) de Lille, Boulevard du Pr. J. Leclercq, F-59037 Lille, France

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* Corresponding author. Tel.: +33 3 20 44 54 80; fax: +33 3 20 44 48 95.

E-mail address: nicolas.blondiaux@chru-lille.fr (N. Blondiaux).
ABSTRACT
The aim of the present study was to evaluate the benefit of monitoring serum piperacillin concentrations in critically ill patients. This was an 11-month, prospective, observational study in a 30-bed Intensive Care Unit in a teaching hospital, involving 24 critically ill patients with evidence of bacterial sepsis. All patients received a 66 mg/kg intravenous bolus of piperacillin in combination with tazobactam (ratio 1:0.125) followed by continuous infusion of 200 mg/kg/24 h. The dosage was adjusted when the serum piperacillin concentration either fell below 4× the drug’s minimum inhibitory concentration (MIC) for the causative agent or exceeded the toxic threshold of 150 mg/L. With the initial regimen, serum piperacillin concentrations were within the therapeutic target range in only 50.0% of patients (n = 12). This proportion increased to 75.0% (18 patients) (P = 0.006) following dosage adjustment. For patients with low initial serum piperacillin concentrations (n = 8), the percentage of time during which the concentration remained above 4× MIC (%T>4× MIC) was 7.1 ± 5.9% before dosage adjustment and 27.3 ± 8.6% afterwards. In conclusion, in critically ill patients, monitoring and adjustment of serum piperacillin levels is required to prevent overdosing and might also help to correct underdosing, an important cause of antibiotic therapy failure.
1. Introduction

Broad-spectrum \(\beta\)-lactams are commonly used to treat severe infections (either alone or combined with other antibiotics), especially in the Intensive Care Unit (ICU) setting. The bactericidal effects of these antibiotics are mostly time-dependent. Laboratory and clinical evidence indicates that to achieve effective bactericidal levels, a serum concentration of at least \(4\times\) times the minimum inhibitory concentration (MIC) for the pathogen involved may be required [1,2]. A related determinant of \(\beta\)-lactam efficacy is the time during which the drug's serum concentration remains above the MIC threshold (\(T>\text{MIC}\)) [3]. Continuous infusion of these antimicrobial agents facilitates the optimisation of their pharmacodynamic profiles [4,5].

In ICU patients, it is more difficult to maintain stable serum antibiotic levels. Apart from the excretory impairments observed in these patients, several other phenomena can significantly alter drug distribution and/or elimination profiles [6]. Furthermore, ICU patients show great interindividual variability in their pharmacokinetic parameters [7], which can also change quickly in response to improvements in or worsening of the clinical condition. Monitoring of serum antibiotic concentrations and subsequent dosage adjustment can avoid underdosing or overdosing and limit the risk of therapeutic failure and toxicity.

Piperacillin/tazobactam (PIP/TAZ) is a \(\beta\)-lactam/\(\beta\)-lactamase inhibitor combination with activity both against Gram-negative and Gram-positive bacteria (including \(\beta\)-lactamase
producers). The broad spectrum of action of PIP/TAZ makes it an appropriate choice in the ICU.

Over the last decade, new modes of administration have significantly optimised the antimicrobial activity of PIP/TAZ. In several pharmacokinetic studies, continuous infusion of PIP/TAZ was superior to intermittent injection (despite the use of lower doses) [8,9] and also appeared to be a better pharmacoeconomic option [10]. Under these supposedly optimal pharmacokinetic conditions, the objective of this study was to assess the utility of daily serum piperacillin monitoring to achieve pharmacodynamic targets associated with optimal piperacillin activity.

2. Patients and methods

2.1. Subjects

Study subjects were adult patients admitted to the Department of Respiratory Emergencies and Intensive Care at Lille University Medical Center (Lille, France) with various pathologies (Table 1). The study’s main inclusion criterion was the presence of a community- or hospital-acquired infection requiring antibiotic therapy with PIP/TAZ. Patients with known hypersensitivity/intolerance to penicillins or a previous infection with PIP/TAZ-resistant bacteria were excluded from the study.
2.2. Antibiotic administration

All patients received PIP/TAZ via continuous infusion. The regimen consisted of a loading bolus of 66 mg/kg piperacillin and 8.25 mg/kg tazobactam (Tazocilline®, Wyeth Pharmaceuticals, Paris, France) injected over 30 min followed by constant-rate infusion of 200 mg/kg/24 h piperacillin and 25 mg/kg/24 h tazobactam. Although this dosage may not be adapted to patients weighing more than 120 kg, we did not use daily doses above 24/3 g of PIP/TAZ because they are not recommended by the manufacturer.
Furthermore, none of the enrolled patients weighed more than 105 kg. In patients with impaired renal function [creatinine clearance (CL<sub>Cr</sub>) ≤ 30 mL/min, calculated according to the Cockcroft and Gault formula], the maintenance dose was halved.

2.3. Target serum concentrations

Dosage adjustment was performed when the serum piperacillin concentration was below 4× MIC for the identified pathogen or >150 mg/L (defined as the toxicity threshold, based on empirical data). Above this concentration, the likelihood of neurological adverse events increased (personal data). When the pathogen was not known, the lower target value was defined as 4× the critical concentration above which all bacterial strains are considered to be resistant to piperacillin, i.e. 4 × 16 mg/L = 64 mg/L (according to guidelines issued by the French Microbiology Society’s Antibiotics Committee).
2.4. Analytical method

Serum piperacillin concentrations were determined by high-pressure liquid chromatography using a 322-A pump, an autoinjector, a 332 variable ultraviolet wavelength detector (all from Serlabo Technologies, Entraigues-sur-la-Sorgue, France) and a C18 Resolve column (4 µm, 3.9 x 150 mm; Waters Associates, Milford, MA). The mobile phase consisted of 0.02 M ammonium acetate buffer and acetonitrile in a 76:24 ratio (v/v) at pH 5.0. The flow rate was 1 mL/min. Pooled human serum was used to prepare standards, to check samples and to dilute serum samples, as required. The assay was linear over the range 1 mg/L to 200 mg/L. Only the free fraction of piperacillin was measured, as an initial step of protein precipitation with acetonitrile was carried out [11].

2.5. Blood sampling

Blood samples for pharmacokinetic analysis were obtained from a peripheral venous catheter placed in the arm not used for drug infusion. Blood was collected before starting drug administration (t = 0) and 1.5 h afterwards (t = 1.5) to ensure that the bolus injection had been completed. A sample was then taken each morning at steady state until the end of the course of treatment.

2.6. Adverse events

Convulsions, changes in white blood cell count, modification of CL_{Cr} and signs of cutaneous intolerance were monitored throughout the study.
2.7. Statistical analysis

A paired Student’s t-test was used to check for statistical significance.

3. Results and discussion

In total, 24 patients [22 males and 2 females; mean ± standard deviation (S.D.) age, 62.5 ± 14.3 years; mean ± S.D. weight, 78.7 ± 16.8 kg] were initially included. Patient characteristics and PIP/TAZ dosage management are presented in Table 1. Seven patients (2, 3, 8, 12, A, E and F; Table 1) displayed severe renal function impairments (CL\textsubscript{Cr} ≤ 30 mL/min) The infecting pathogen was only documented in 9 of the 24 cases (Table 1b) and included *Pseudomonas aeruginosa* (Patients A and B), *Escherichia coli* (Patients C and D), *Klebsiella pneumoniae* (Patients E and F), *Acinetobacter baumannii* (Patient G), meticillin-susceptible *Staphylococcus aureus* (Patient H) and *Enterobacter aerogenes* (Patient I). In these strains, the MIC of piperacillin (when combined with tazobactam) ranged from 4 mg/L to 16 mg/L (mean value 7.6 ± 5.1 mg/L).

The serum piperacillin concentration was monitored in 4 to 21 samples per patient (mean ± S.D., 10 ± 4.6). With the initial regimen, the serum piperacillin concentrations were above the previously defined target value (i.e. >4× MIC) in 12 (50.0%) of the patients (Patients 1–6, 9, C, D, G, H and I) (Table 1). This goal was achieved in six additional patients (Patients 7, 8, 10, A, E and F) following dosage adjustment (*P* = 0.006). In four patients (Patients 8, A, E and F), antibiotic levels were considered toxic
(>150 mg/L), thus the dosage was reduced and no adverse events were observed. Conversely, a dosage increase in two patients (Patients 7 and 10) of the eight patients with below-target serum piperacillin concentrations enabled achievement of the therapeutic value. Dosage adjustment was not efficient in the six remaining patients (Patients 11–15 and B). Despite many dosage changes (with up to 18 g of piperacillin per day in some cases), serum piperacillin concentrations >4× MIC were never achieved. Interestingly, all but one (Patient B) of these subjects had extensive cellulitis, which might have influenced the pharmacokinetics of the antibiotic [12]. In this clinical setting, subcutaneous inflammation might generate an additional distribution compartment and could be responsible for extracellular and extraplasmatic leakage of the antibiotic. This hypothesis is supported by the substantial increase in the volume of distribution of PIP/TAZ seen in burns victims compared with healthy individuals (indicating translesional diffusion of the antibiotic) [13]. Further investigation is now required to establish suitable antibiotic regimens in this setting; nevertheless, daily monitoring of serum piperacillin concentrations significantly increased achievement of the therapeutic target concentration.

By contrast to that shown for fluoroquinolones [14], the possibility of selecting resistant mutants at serum piperacillin concentrations <4× MIC is not supported by experimental data. However, several authors have suggested that prolonged percentage of time during which the concentration remains above 4× MIC (T>4× MIC), instead of T>MIC, is an important pharmacokinetic/pharmacodynamic (PK/PD) parameter that better predicts treatment outcome with β-lactams (especially for critically ill patients) [2,5,15]. Here,
serum piperacillin concentrations were monitored daily for between 3 days and 20 days (mean ± S.D., 9 ± 5.2 days). For the eight patients (Patients 7, 10–15 and B) with low initial serum piperacillin concentrations, the mean percentage $T>4\times$ MIC was $7.1 ± 5.9\%$ before antibiotic dosage adjustment and $27.3 ± 8.6\%$ afterwards (Fig. 1). This significant difference ($P = 0.03$) demonstrates the beneficial impact of regular serum piperacillin monitoring on these important therapeutic PK/PD parameters.

4. Conclusion

In critically ill patients, monitoring and adjustment of serum piperacillin levels are warranted to prevent overdosing and to help correct underdosing, an important cause of antibiotic therapy failure. Additional trials in a more homogeneous patient population are required to underpin these preliminary findings.

Funding
None.

Competing interests
None declared.

Ethical approval
This study was conducted in accordance with the Declaration of Helsinki, and written informed consent was obtained from all patients or their legal guardians.
References


Fig. 1. Impact of piperacillin/tazobactam dosage changes on pharmacokinetic/pharmacodynamic parameters. Change in $% T>4\times$ MIC for patients who received dosage adjustment due to below-target serum concentrations. Horizontal bars represent the mean $% T>4\times$ MIC. $% T>4\times$ MIC, percentage of time during which the concentration remained above $4\times$ the minimum inhibitory concentration.
Table 1

Characteristics of the 24 patients treated with piperacillin/tazobactam (PIP/TAZ)

(a) Characteristics of the 15 patients with non-documented infection (no pathogen isolated). The lower target value was defined as 4× the critical concentration above which all bacterial species are considered to be resistant to PIP, i.e. 4 × 16 mg/L = 64 mg/L (according to guidelines issued by the French Microbiology Society’s Antibiotics Committee). For these patients, target values are between 64 mg/L and 150 mg/L (toxicity threshold); otherwise dosage was changed.

<table>
<thead>
<tr>
<th>Patient no.</th>
<th>Age (years)</th>
<th>Weight (kg)</th>
<th>Gender</th>
<th>Renal function impairment</th>
<th>Indication for antibiotic therapy</th>
<th>Initial dosage (g/day)</th>
<th>Dosage change (g/day)</th>
<th>PIP concentration (mg/L) c</th>
<th>n × PIP MIC Before dosage adjustment</th>
<th>After dosage adjustment</th>
</tr>
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<tbody>
<tr>
<td>1</td>
<td>75</td>
<td>90</td>
<td>M</td>
<td>No</td>
<td>HAP</td>
<td>18/2.2</td>
<td>No</td>
<td>80.5 ± 2.1</td>
<td>–</td>
<td>5.03 ± 0.13</td>
</tr>
<tr>
<td>2</td>
<td>46</td>
<td>70</td>
<td>M</td>
<td>Yes</td>
<td>HAP</td>
<td>7/0.87</td>
<td>No</td>
<td>105.9 ± 3.8</td>
<td>–</td>
<td>6.62 ± 0.24</td>
</tr>
<tr>
<td>3</td>
<td>69</td>
<td>90</td>
<td>M</td>
<td>Yes</td>
<td>HAP</td>
<td>9/1.12</td>
<td>No</td>
<td>97.5 ± 22.5</td>
<td>–</td>
<td>6.10 ± 1.41</td>
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<tr>
<td>4</td>
<td>68</td>
<td>59</td>
<td>M</td>
<td>No</td>
<td>HAP</td>
<td>12/1.5</td>
<td>No</td>
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<td>–</td>
<td>5.96 ± 0.81</td>
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<td>Age</td>
<td>Gender</td>
<td>Distance</td>
<td>Diabetic</td>
<td>HAP</td>
<td>Notes</td>
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<td>MCA D</td>
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<tr>
<td>5</td>
<td>80</td>
<td>M</td>
<td>No</td>
<td>HAP</td>
<td>14/1.7</td>
<td>No</td>
<td>70.1 ± 5.4</td>
<td>–</td>
<td>4.38 ± 0.34</td>
<td>–</td>
</tr>
<tr>
<td>6</td>
<td>67</td>
<td>M</td>
<td>No</td>
<td>HAP</td>
<td>20/2</td>
<td>No</td>
<td>77.0 ± 8.4</td>
<td>–</td>
<td>4.81 ± 0.53</td>
<td>–</td>
</tr>
<tr>
<td>7</td>
<td>74</td>
<td>M</td>
<td>No</td>
<td>HAP</td>
<td>14/1.7</td>
<td>Yes: 16/2</td>
<td>3.3 ± 1.6</td>
<td>64.9 ± 4.3</td>
<td>0.20 ± 0.10</td>
<td>4.06 ± 0.27</td>
</tr>
<tr>
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<td>56</td>
<td>M</td>
<td>Yes</td>
<td>HAP</td>
<td>8/1</td>
<td>Yes: 4/0.5</td>
<td>320.6 ± 19.4</td>
<td>92.7 ± 54.9</td>
<td>20.03 ± 1.21</td>
<td>5.79 ± 3.43</td>
</tr>
<tr>
<td>9</td>
<td>81</td>
<td>F</td>
<td>No</td>
<td>Cellulitis</td>
<td>10/1.2</td>
<td>No</td>
<td>78.9 ± 12.2</td>
<td>–</td>
<td>4.93 ± 0.73</td>
<td>–</td>
</tr>
<tr>
<td>10</td>
<td>53</td>
<td>M</td>
<td>No</td>
<td>Cellulitis</td>
<td>18/2.2</td>
<td>Yes: 20/2.5</td>
<td>44.8 ± 9.2</td>
<td>66.7 ± 6.1</td>
<td>2.80 ± 0.58</td>
<td>4.17 ± 0.38</td>
</tr>
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<td>11</td>
<td>60</td>
<td>M</td>
<td>No</td>
<td>Cellulitis</td>
<td>14/1.7</td>
<td>Yes: 16/2, then 18/2.2</td>
<td>51.9 ± 18.0</td>
<td>55.3 ± 17.1</td>
<td>3.24 ± 1.13</td>
<td>3.46 ± 1.07</td>
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<td>61</td>
<td>M</td>
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<td>Cellulitis</td>
<td>7/0.87</td>
<td>Yes: 9/1.12</td>
<td>35.6 ± 0.6</td>
<td>39.8 ± 3.2</td>
<td>2.21 ± 0.04</td>
<td>2.49 ± 0.20</td>
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</table>
(b) Characteristics of the 9 patients with documented infection. The lower target value was defined as $4 \times$ the PIP MIC for the identified pathogen. For these patients target values are between $4 \times$ the PIP MIC and 150 mg/L

<table>
<thead>
<tr>
<th>Patient no.</th>
<th>Age (years)</th>
<th>Weight (kg)</th>
<th>Gender</th>
<th>Renal function impairment a</th>
<th>Indication for antibiotic therapy</th>
<th>Isolates/PIP MIC (mg/L)</th>
<th>Initial dosage (g/day)</th>
<th>Dose change (g/day)</th>
<th>PIP concentration (mg/L) Before dosage adjustment</th>
<th>n × PIP MIC Before dosage adjustment</th>
<th>PIP concentration (mg/L) After dosage adjustment</th>
<th>n × PIP MIC After dosage adjustment</th>
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<tr>
<td>13</td>
<td>58</td>
<td>80</td>
<td>M</td>
<td>No</td>
<td>Cellulitis 16/2</td>
<td>Yes: 18/2.2</td>
<td>54.3 ± 3.3</td>
<td>3.39 ± 0.21</td>
<td>3.69 ± 0.29</td>
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<tr>
<td>14</td>
<td>34</td>
<td>80</td>
<td>M</td>
<td>No</td>
<td>Cellulitis 16/2</td>
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<td>42.6 ± 5.3</td>
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<td>3.13 ± 0.39</td>
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<td>15</td>
<td>43</td>
<td>100</td>
<td>M</td>
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<td>Cellulitis 20/2.5</td>
<td>Yes: 24/3</td>
<td>25.9 ± 5.4</td>
<td>1.62 ± 0.34</td>
<td>1.91 ± 0.18</td>
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<tr>
<td>A</td>
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<td>M</td>
<td>Yes</td>
<td>HAP</td>
<td>P.a./4</td>
<td>9/1.1</td>
<td>Yes:</td>
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<td>109.9 ± 8.8</td>
<td>36.98 ± 25</td>
<td>27.48 ± 75</td>
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<td>50</td>
<td>F</td>
<td>No</td>
<td>HAP</td>
<td>P.a./16</td>
<td>10/1.25</td>
<td>Yes:</td>
<td>55.2 ± 12/1.12</td>
<td>46.5 ± 3.4</td>
<td>3.45 ± 5</td>
<td>2.91 ± 0.21</td>
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<td>C</td>
<td>71</td>
<td>105</td>
<td>M</td>
<td>No</td>
<td>HAP</td>
<td>E.c./4</td>
<td>20/2.5</td>
<td>No</td>
<td>102.0 ± 9.0</td>
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<td>25.50 ± –</td>
<td>–</td>
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<tr>
<td>D</td>
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<td>90</td>
<td>M</td>
<td>No</td>
<td>HAP</td>
<td>E.c./8</td>
<td>18/2.25</td>
<td>No</td>
<td>69.9 ± 26.5</td>
<td>–</td>
<td>8.74 ± –</td>
<td>–</td>
</tr>
<tr>
<td>E</td>
<td>69</td>
<td>90</td>
<td>M</td>
<td>Yes</td>
<td>HAP</td>
<td>K.p./4</td>
<td>9/1.25</td>
<td>Yes:</td>
<td>224.1 ± 4/0.5</td>
<td>54.6 ± 30.4</td>
<td>25.2 ± 17.6</td>
<td>22.72 ± 13.65</td>
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<td>80</td>
<td>M</td>
<td>Yes</td>
<td>Septic shock</td>
<td>K.p./8</td>
<td>8/1 6/0.7</td>
<td>Yes:</td>
<td>181.8 ± 34.5</td>
<td>140.8 ± 17.6</td>
<td>22.72 ± 11.51</td>
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<td>G</td>
<td>65</td>
<td>46</td>
<td>M</td>
<td>No</td>
<td>HAP</td>
<td>A.b./4</td>
<td>10/1.25</td>
<td>No</td>
<td>25.0 ± 2.8</td>
<td>–</td>
<td>6.26 ± –</td>
<td>–</td>
</tr>
<tr>
<td>H</td>
<td>80</td>
<td>72</td>
<td>M</td>
<td>No</td>
<td>HAP</td>
<td>S.a./4</td>
<td>14/1.75</td>
<td>No</td>
<td>87.5 ± 17.7</td>
<td>–</td>
<td>21.87 ± –</td>
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<td>I</td>
<td>64</td>
<td>80</td>
<td>M</td>
<td>No</td>
<td>sUTI</td>
<td>E.a./16</td>
<td>16/2</td>
<td>No</td>
<td>65.6 ± 2.4</td>
<td>–</td>
<td>4.10 ± –</td>
<td>–</td>
</tr>
</tbody>
</table>
HAP, hospital-acquired pneumonia; sUTI, severe urinary tract infection; P.a., *Pseudomonas aeruginosa*; E.c., *Escherichia coli*; K.p., *Klebsiella pneumoniae*; A.b., *Acinetobacter baumannii*; S.a., *Staphylococcus aureus*; E.a., *Enterobacter aerogenes*.

\(^{a}\) Creatinine clearance \(\leq 30\) mL/min, calculated according to the Cockcroft and Gault formula.

\(^{b}\) PIP/TAZ combination.

\(^{c}\) Serum PIP concentration (free fraction): mean \(\pm\) S.D. of 2 to 17 samples according to the patient.
$P = 0.03^*$

$T > 4$ MIC (%)