



Mutation spectrum of EYS in Spanish patients with autosomal recessive retinitis pigmentosa

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**Mutation spectrum of *EYS* in Spanish patients with
autosomal recessive retinitis pigmentosa**

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Key Words:	EYS, Retinitis Pigmentosa, Spanish population, mutation, functional domain, recurrent mutation



review

MUTATION IN BRIEF

HUMAN MUTATION

Mutation spectrum of *EYS* in Spanish patients with autosomal recessive retinitis pigmentosa

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Short Title: Mutation spectrum of *EYS*.

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ABSTRACT: Retinitis pigmentosa (RP) is a heterogeneous group of inherited retinal dystrophies characterised ultimately by the loss of photoreceptor cells. We have recently identified a new gene (*EYS*) encoding an ortholog of *Drosophila* spacemaker (*spam*) as a commonly mutated gene in autosomal recessive RP. In the present study, we report the identification of 73 sequence variations in *EYS*, of which 28 are novel. Of these, 42.9% (12/28) are very likely pathogenic, 17.9% (5/28) are possibly pathogenic, whereas 39.3% (11/28) are SNPs. In addition, we have detected 3 pathogenic changes previously reported in other populations. We are also presenting the characterisation of *EYS* homologues in different species, and a detailed analysis of the *EYS* domains, with the identification of an interesting novel feature: a putative coiled-coil domain. Majority of the mutations in the arRP patients have been found within the domain structures of *EYS*. The minimum observed prevalence of distinct *EYS* mutations in our group of patients is of 15.9% (15/94), confirming a major involvement of *EYS* in the pathogenesis of arRP in the Spanish population. Along with the detection of three recurrent mutations in Caucasian population, our hypothesis of *EYS* being the first prevalent gene in arRP has been reinforced in the present study.

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KEY WORDS: *EYS*, Retinitis Pigmentosa, Spanish population, mutation, functional domain, recurrent mutation

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INTRODUCTION

Retinitis pigmentosa (RP) is a heterogeneous group of inherited retinal dystrophies featured by the loss of photoreceptor cells and clinically characterized by pigmentary deposits at mid periphery of the retina that are visible on fundus examination. Patients present with night blindness as the initial symptom, which is followed by the constriction of the visual field and progressive loss of visual acuity, leading to complete blindness after several decades [Hamel, 2006]. Prevalence of nonsyndromic RP is approximately 1 in 4000. The condition may segregate as an autosomal dominant, autosomal recessive, or an X-linked recessive trait [Humphries et al., 1990]. All genes identified to date are believed to account for roughly 50% of all retinal dystrophy cases [Pomares et al., 2007]. The autosomal recessive form of RP is the commonest worldwide, accounting for approximately 39% of cases in Spain [Ayuso et al., 1995]. To date, 29 loci have been reported being responsible for arRP, of which 25 genes have been identified (<http://www.sph.uth.tmc.edu/Retnet/>). However, all together the reported loci are responsible for only ~35-45% of the recessive RP cases, although none of them independently account for a substantial proportion of arRP (more than 10%) [Daiger et al., 2007; Hartong et al., 2006]. In contrast, the *RP25* locus, identified by our group in 1998 [Ruiz et al., 1998], was estimated to be linked to 27.7% of Spanish arRP families [Barragán et al., 2008]. Recently, we have identified a new gene encoding an ortholog of *Drosophila* spacemaker (spam) corresponding to *RP25* as a commonly mutated gene in arRP. The identification of six independent mutations, together with the presence of linked families from different ancestral origins, supports *EYS* (*Eyes Shut Homologue*, MIM# 612424) as one of the first major genes reported for arRP [Abd El-Aziz et al., 2008]. Spanning over 2 Mb within the *RP25* locus (6p12.1-6q15), *EYS* is the largest gene identified to be expressed in the human eye so far, and the fifth largest gene overall in the human genome. The longest isoform of *EYS* encodes a protein of 3165 amino acids whose function remains to be elucidated. Considering the evolutionary data and the known function of the only characterised homologue, *EYS* is likely to have a role in the modelling of retinal architecture [Zelhof et al., 2006]. The identification of the gene for *RP25* reveals what might be the genetic basis for a significant proportion of arRP cases and thus paves the way for genetic counselling, prenatal detection, and treatment. However, further characterisation of the novel *EYS* protein as well as an extended mutation spectrum of *EYS*-related arRP would be valuable to undertake.

In the present study based on 94 families, we report the identification of 73 sequence variations in *EYS*, of which 28 are novel. Of these novel changes, 42.9% (12/28) are very likely pathogenic, 17.9% (5/28) are possibly pathogenic, whereas 39.3% (11/28) are SNPs. In addition, we have detected 3 pathogenic changes previously reported in other populations. The estimated prevalence of distinct *EYS* mutations in our group of patients is of 15.9%, confirming the significant involvement of *EYS* in the pathogenesis of the arRP in the Spanish population. Besides, we present a detailed bioinformatic characterisation of *EYS* and its homologues, which would aid in the determination of the pathogenic nature of newly identified variations in *EYS*.

MATERIALS AND METHODS

Subjects and Clinical Data: Our current cohort of study comprises 94 unrelated Spanish families affected by arRP, all derived from the Ophthalmology Service of different Hospitals throughout Spain. The participating families conform to the phenotypic and inheritance patterns of arRP. A group of matching control individuals was also recruited. Informed consent was obtained from all participants in the study, in accordance with the tenets of the Declaration of Helsinki (Edinburgh, 2000). Clinical diagnosis was based on visual acuity, fundus photography, computerized testing of central and peripheral visual fields and electroretinography (ERG) findings. Clinical features of RP include initial hemeralopia, restriction of visual field, gradual increased bone spicule pigmentation and decrease of visual acuity, attenuation of retinal vessels, and waxy disc pallor.

Bioinformatic characterisation of EYS: Firstly, ExPASy ProtParam tool was employed to determine the physical and chemical parameters of *EYS*. Secondly, InterProScan program was used to search for known domains and functional sites within *EYS*. For a further characterisation of *EYS*, we used Coils and Secpred to analyse the secondary structure. SignalP 3.0 was utilised to predict the presence and localisation of signal peptide cleavage sites. The characterisation of *EYS* homologues was performed in different steps. Firstly, Blast analyses of the human *EYS* cDNA and encoded protein were run to detect annotated homologous proteins. However, only human and *Drosophila* were found to be completely annotated in the databases. Therefore, we employed the BLAT tool at the UCSC Genome Bioinformatics Site to identify and map sequences with high identity to the target sequences. Human and *Drosophila* *EYS* protein were used as the basis of this search. An *in-silico* splicing site

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characterisation of the positive genomic region ensued to build the homologous cDNA which was then translated into protein. Also, we identified those sequencing gaps within the genomic region of *EYS* homologues in different species. Comparison of protein homologues were performed using bl2seq (NCBI) and EMBOSS Pairwise Alignment Algorithms: Needle and Water (EBI) alignments. The fully characterised proteins were aligned using MUSCLE (MUltiple Sequence Comparison by Log-Expectation) program at EBI.
PCR-based direct genomic sequencing of EYS: Peripheral blood samples were collected from all subjects for genomic DNA purification using an automated DNA extractor (MagNA Pure LC Instrument, Roche Diagnostics, Switzerland). Forty-eight pairs of primers were designed using the Primer 3 Output program (<http://frodo.wi.mit.edu/primer3/>) in order to screen the forty coding exons, the three non coding exons, the intronic flanking sequences and the regulatory factor binding sites of *EYS* (Genbank Reference Sequence and Version FJ416331; GI: 212675237; Transcript Reference Sequence: NM_001142800.1). PCR conditions and primer sequences employed are available upon request. The amplified products were subsequently purified using an enzymatic procedure, according to manufacturer's recommendations (EXOSAP-IT®, USB Corporation) and sequenced with a ready reaction kit (BigDye Terminator Cycle FS Ready Reaction Kit; PE-Applied Biosystems, Foster City, CA). The fragments obtained were purified using fine columns (Sephadex G-501, Sigma-Aldrich Co.) and resolved on an automated sequencer (3730 DNA Analyzer, Applied Biosystems, USA). Finally, the data was analysed using Lasergene DNASTAR® software (DNASTAR, Inc). Nucleotide numbering reflects cDNA numbering with +1 corresponding to the A of the ATG translation initiation codon in the reference sequence, according to journal guidelines (www.hgvs.org/mutnomen). The initiation codon is codon 1. In order to evaluate the pathogenicity of the novel variants, we employed various softwares which analyse the potential role of a given variant on the function or structure of the encoded protein based on conservation and homology, physical properties of the amino acids, prediction of the protein disorder, or binding to transcription factor binding sites (TFBS) (Conseq: <http://conseq.tau.ac.il/>; PolyPhen (prediction of functional effect of human nsSNPs): <http://coot.embl.de/PolyPhen/>; SIFT (Sorting Intolerant From Tolerant): <http://blocks.fhcrc.org/sift/SIFT.html>; Disopred: <http://bioinf.cs.ucl.ac.uk/disopred/disopred.html> [Ramensky et al., 2002]. Besides, the tool DiANNA was employed for disulfide connectivity prediction when the variation affected a Cys residue, and NetPhos2.1 (NetPhos 2.0 Server: <http://www.cbs.dtu.dk/services/NetPhos/>) together with Diphos (Disorder-Enhanced Phosphorylation Sites Predictor: <http://core.ist.temple.edu/pred/pred.html>), to predict the alteration of phosphorylation [Blom et al., 1999]. In addition, intronic variants were evaluated for affecting any regulatory process at the transcriptional or splicing levels (TESS Transcription Element Search System: <http://www.cbil.upenn.edu/cgi-bin/tess/tess>; <http://www.fruitfly.org/seqtools/splice.html>; Splice SignalAnalysis: <http://www.ebi.ac.uk/asd-srv/wb.cgi>; Alternative Splicing DataBase: <http://hazelton.lbl.gov/~teplitski/alt/>; <http://www.fruitfly.org/cgi-bin/seq tools/splice.pl>; Splicing Element Annotation: <http://genes.mit.edu/aceScan2/index.html>; ESEfinder: <http://rulai.cshl.edu/tools/ESE2/>; Rescue-ESE: <http://genes.mit.edu/burgelab/rescue-ese/>; ACESCAN2 Web Server: <http://genes.mit.edu/aceScan2/index.html>; NetGene2 Server (<http://www.cbs.dtu.dk/services/NetGene2/>; http://www.ensembl.org/Homo_sapiens/generegulationview; <http://www.cisred.org/content/software>; <http://regRNA.mbc.nctu.edu.tw/html/about.html>. [Yeo et al., 2004; Matlin et al., 2005; Wang & Marin 2006; Fairbrother et al., 2002; Brunak et al., 1991].
GeneChip 6.0 data analysis (see Affymetrix Genotyping Console 2.1 user manual for details): When sample requirements were fulfilled, patients underwent Copy Number Variations (CNVs) analysis with Genechip 6.0 Affymetrix array. The CEL intensity files were loaded into Genotyping Console v2.1 (Affymetrix Inc.) for analysis. All samples passed the initial contrast QC metric (>0.4) that measures the ability of the intensity files to resolve SNPs into three genotyping clusters. Copy number data were generated by comparing intensities for both SNP and copy number probes *in silico* to the HapMap control provided by Affymetrix. The resulting log2 ratios were then analysed using a Hidden Markov Model (HMM) to generate copy number calls for each probe. The quality of the log2 data was assessed by the degree of variation, determined by the MAPD metric. MAPD is defined as the Median of the Absolute value of all Pairwise Differences between log2 ratios for a given chip. High MAPD >0.4 (using the HapMap control) is considered to be the cut-off at which copy numbers can no longer be accurately called. None of the samples included in this study had a MAPD >0.4. Using the copy number calls provided by Genotyping Console v2.1 as a guide, a more detailed analysis was performed by interrogation of the plots of log2 ratios paying particular attention to CNV regions called by the HMM. A minimum log2 ratio cut-off of +/-0.3 was used for autosomal CNVs. This excluded any false positive calls made by the HMM algorithm. Identified CNVs in the region of *EYS* were checked against the Database of Genomic Variants (DGV) in

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Genotyping Console v2.1. CNVs were excluded from further analysis if they matched a known CNV exactly, or if occurred in a region that did not include any of the coding exons of *EYS* [McCarroll et al., 2006; de Smith et al., 2007; Sharp et al., 2005]. The allelic difference and loss of heterozygosity plots generated from the difference in allele intensity for each SNP were analysed to investigate parental consanguinity and as an aid to the interpretation of potential mosaicism. Genotyping was performed using the Birdseed v.2 algorithm. All samples had call rates >97.5%.

RESULTS AND DISCUSSION

Molecular diagnosis of RP is a challenging task given the important genetic heterogeneity of these groups of diseases. For most genes, many different mutations with similar consequences are known, yet other mutations in the same gene may cause different diseases. Particularly, for recessive RP, which is the most prevalent form of the disease, estimated to comprise from 50 to 60% of all RP cases, 29 loci have been described as pathogenic. In aggregate, the known mutations in arRP genes cause about ~35-45% of all cases of this form of the disease [Daiger et al., 2007; Hartong et al., 2006]. Some authors suggest that 50-60% of all arRP associated causal loci have already been identified. However, mutations in individual genes do not account for a significant proportion of arRP cases.

EYS is the largest gene identified to be expressed in the human eye so far, and appears to be a frequent cause of arRP. Thus, *EYS* encoded protein EYS emerges as a relevant player in arRP pathogenesis. In previous studies, a total of 8 mutations had been identified [Abd El-Aziz et al., 2008; Collin et al., 2008] by a combination of different screening methods such as direct genomic sequencing, MLPA or CGH arrays. The domain structure of EYS has been predicted from the characterised sequence of *EYS* [Abd El-Aziz et al., 2008; Collin et al., 2008] as having several EGF-like and Laminin G domains.

Here we report on the molecular screening of *EYS* in a Spanish cohort of patients with arRP. Besides, we are presenting the characterisation of *EYS* homologues in different species, and a detailed analysis of the *EYS* domains, with the identification of an interesting novel feature.

Protein domains structure

We present here the identification of a putative coiled-coil structure, which is an interesting novel feature, in the central portion of the protein coincident with a region of Alpha helix overrepresentation (Fig. 1, Supplementary Figure S1). The insight into the functional repercussion of both the already known signal peptide domain and the novel coiled-coil domain reported here supports a structural role for this new protein, which would be secreted and polymerize into a scaffolding that would contribute to the human retinal architecture. This is consistent with the function of *Eys* in *Drosophila*, where it is secreted by photoreceptor cells [Husain et al., 2006] and is essential for the formation of the matrix-filled interrhabdomeral space. The signal peptide and its cleavage site consensus sequence located in the N-terminal region of *EYS* (Fig. 1, Fig. 2) may confer a secretory nature to the protein or result in an intracellular or cytoplasmatic location of the mature protein [Jarjanazi et al., 2008]. Remarkably, we have identified the consensus sequence for this feature in *EYS* homologues in a number of species such as orangutan, dog, horse, marmoset, monkey and chimpanzee (data available on request). Accordingly, *Drosophila* Spacemaker and other proteins which share several of *EYS* domains have been found to be secreted and to have a structural function, such as SCUBE [Yang et al., 2002] or CMG-2 [Bell et al., 2001]. The members of SCUBE gene family contain both a signal peptide domain and multiple EGF-like repeats. Interestingly, both *EYS* and SCUBE1 share homology with the same protein families, such as members of the fibrillin and Notch families among others. SCUBE1 and 2 are known to form oligomers and manifest a stable association with the cell surface in vascular endothelial cells [Yang et al., 2002]. CMG-2, containing a potential signal peptide, targets to the endoplasmic reticulum and shows affinity for the basement membrane matrix proteins, collagen type IV and laminin. Similarly, CMG-1, which encodes a protein with coiled-coil domains, was observed to target to an intracellular vesicular compartment and may play as well a structural role since it has been postulated that this gene may be implicated in the regulation of capillary formation in an *in-vitro* model of endothelial cell morphogenesis [Bell et al., 2001].

Furthermore, the fact that we have identified a putative coiled-coil domain within *EYS* reinforces the idea of *EYS* being a key player in the organisation of human retina. Coiled coils are important structural motifs involved in a

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variety of important interactions [Mason & Arndt, 2004]. Dystrophin, which resembles EYS in several aspects such as presenting a point mutation/deletion pattern of mutations in human disease and belonging to the group of five longest genes in the human genome, is known to play structural roles among others. Dystrophin also contains coiled-coil domains, which are responsible for the assembly of heterodimers of the so called Dystrophin glycoprotein complex [Sadoulet et al., 1997; Böhm et al., 2008]. In this regard, it is important to mention that *Drosophila* Spacemaker interacts with Prominin and the cell adhesion molecule Chaoptin to choreograph the partitioning of rhabdomeres into an open system, critically affecting retinal morphogenesis [Zelhof et al., 2006].

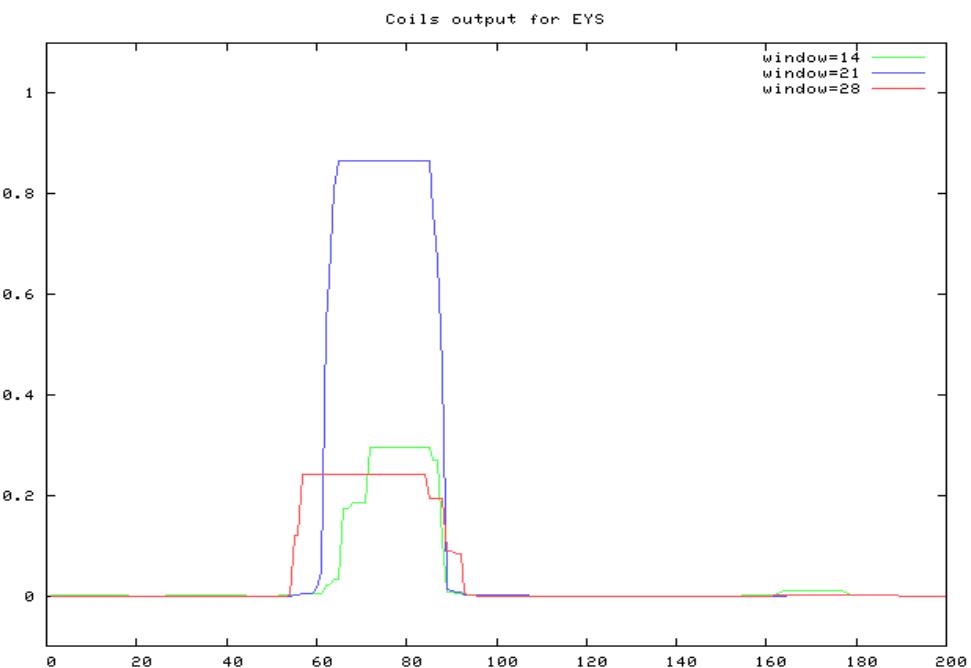


Figure 1. Coiled coil domain prediction in human EYS protein.

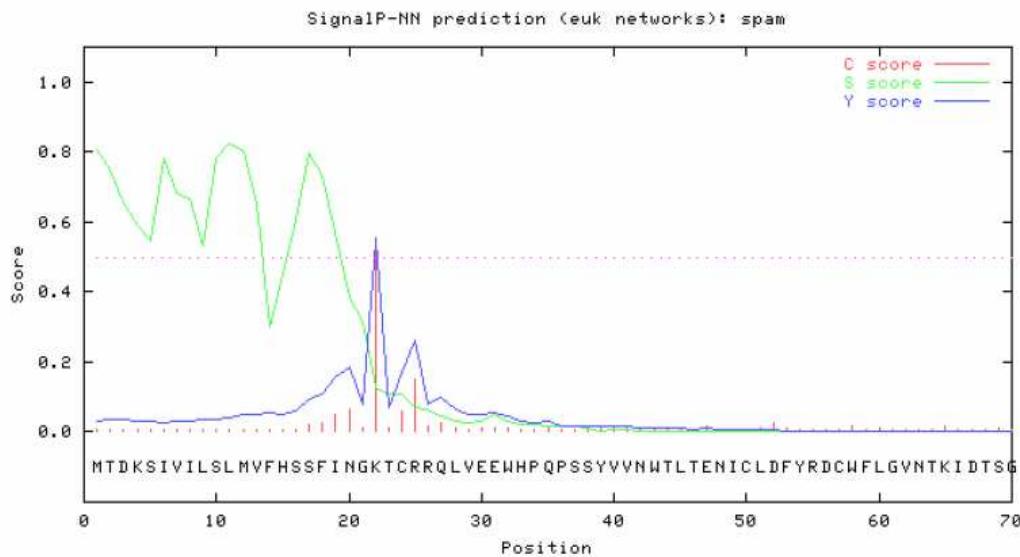


Figure 2. Signal peptide and cleavage site consensus sequence prediction in human EYS protein.

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Annotation of new homologues

With the aim of evaluating evolutionary conservation, we have performed the bioinformatic characterisation of EYS homologues in several species. Apart from *Drosophila Eyes Shut*, which was the first *Eys* gene to be annotated, and human EYS, only fragmented information was available of other EYS homologues. Here we report in detail the structures of Zebrafish, Chicken, Platypus, Opossum, Horse, Dog, Marmoset and Orang-utan homologues and pairwise comparisons with human EYS/EYS (Table 1, Supplementary Figure S1). However, EYS seems to be absent in some species such as those insects with a close rhabdom system or in mammals in mouse, rat and guinea pig, which represent two of the three major rodent clades [Abd El-Aziz et al., 2008].

It is noteworthy that the previously reported and newly identified functional domains described here are conserved throughout evolution. Consistently, as shown in Table 1, the more distant a species is from human, the lower the percentages of identity and similarity. Concerning the signal peptide in *Drosophila*, some studies report that spacemaker secretion would be upon interaction of *Eys* with a receptor, which could promote its spreading from the stalk to the rhabdomere to fill the interrhabdomeral space (IRS) [Husain et al., 2006]. Thus, there would be no need of a signal peptide for secretion from the photoreceptor cells.

Table 1. EYS homologues characterisation.

Specie	Chromosomal interval (bp)	Genomic Length (bp)	Genomic Identity %	Protein Identity %	Protein Similarity %
Drosophila	chr2L:2323799-2357874	34,076	NA	23.9	37.9
Zebrafish	chr13:37105545-37227793	122,249	NA	44.0	60.3
Chicken	chr3:87827859-88369092	541,234	71.9	33.1	40.3
Platypus	chr1:33486985-34313969	1,756,349	71.7	31.8	36.6
Opossum	chr2:312522958-314990462	2,467,505	71.9	35.1	40.5
Horse	chr20:56733300-58137441	4,247,273	80.5	61.4	67.1
Dog	chr12:30185993-31709408	1,701,147	81.9	62.8	68.4
Marmoset	Several contigs*	973,997	91.4	88.8	92.2
Orangutan	chr6:63945057-65776065	1,831,009	97.3	96.8	97.5
Human	chr6:64488454-66262024	1,773,571	100.0	100.0	100.0

* Available on request

NA=Not Available

Pathogenic nature of the identified changes.

In this study, 12 novel very likely pathogenic changes have been identified in 10 families. Of these 10 families, 5 present mutations in both alleles, whereas the remaining 5 have mutations in just one allele. The clearly pathogenic variants consisted of 6 truncating mutations, 1 in frame deletion of 300 nucleotides leading to a protein truncation of 100 aminoacids, 1 splice site mutation and 4 missense changes. Out of the 28 novel variations, we have also identified 5 possible pathogenic changes in 5 separate families. In addition, we have detected 3 pathogenic variations previously published in ours and other populations [Abd El-Aziz et al., 2008; Collin et al., 2008] (Table 2, Table 3, Figs. 3 and 4). As mentioned in the Methods section, the sequence variants were designated in accordance with the Human Genome Variation Society recommendations (<http://www.hgvs.org/mutnomen/>). All the patients with mutations had received a defined clinical diagnosis of RP with a recessive mode of inheritance and were Spanish. The variations were regarded as pathogenic changes as long as they met the criteria of pathogenicity, i.e absence in 200 control individuals and the segregation with the disease phenotype within the family (Fig. 4). Particularly, missense mutations were considered pathogenic according to their effect on functional EYS domains that they target, their evolutionary conservation and/or to the fact that they are found together with a second variant, especially if this is truncating.

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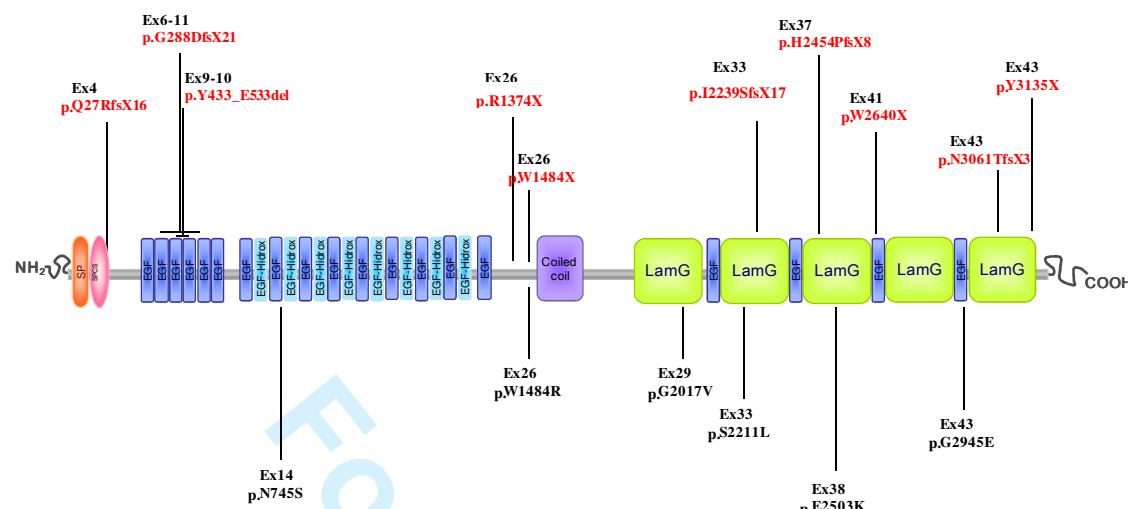


Figure 3. *EYS* mutation distribution along the domain structure of *EYS/EYS*. 5' UTR and splice site variations are not included in this depiction.

Table 2. Mutation spectrum of *EYS* gene in Spanish families.

Family ID	Nucleotide change	Predicted effect	Domains	Location in gene	Type of change	Reference of the variation
Families with novel very likely pathogenic changes and both alleles affected						
RP1052M	c.[1300-17039_1599+22208del] + [9178_9181delATAA]	p.Y433_E533del p.N3061TfsX3	EGF-like LamininG	Exons 9 and 10 Exon 43	Heterozygous Heterozygous	This study This study
RP1237M	c.[2234A>G] + [7919G>A]	p.N745S p.W2640X	EGF EGF	Exon 14 Exon 41	Heterozygous Heterozygous	This study This study and Abd El-Aziz et al., 2008
RP84B	c.6050G>T	p.G2017V	LamininG	Exon 29	Homozygous	This study
RP136B	c.[6632C>T] + [8834G>A]	p.S2211L p.G2945E	LamininG EGF	Exon 33 Exon 43	Heterozygous Heterozygous	This study This study
RP367B	c.[4120C>T] + [6424+1G>T]	p.R1374X Splice mutation	Close to EGF	Exon 26 Intron 31	Heterozygous Heterozygous	This study This study

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Families with single novel very likely pathogenic changes						
RP358B	c.7361delA	p.H2454PfsX8	LamininG	Exon 37	Heterozygous	This study
RP60	c.78_79dupGC	p.Q27RfsX16	Signal peptide cleavage site	Exon 4	Heterozygous	This study
RP180M	c.862-10671_1766+10020del	p.G288DfsX21	EGF	Exons 6-11	Heterozygous	This study
RP33	c.4451G>A	p.W1484X	Close to coiled-coil	Exon 26	Heterozygous	This study
RP81	c.2234A>G	p.N745S	EGF	Exon 14	Heterozygous	This study
Families with novel possible pathogenic changes						
VRP8	c.4450T>C	p.W1484R	Close to coiled-coil	Exon 26	Heterozygous	This study
RP383	c.7507G>A	p.E2503K	LamininG	Exon 38	Heterozygous	This study
RP228B	c.-462G>C	-	-	Exon 1 5'UTR	Homozygous	This study
RP107B	c.-204G>A	-	-	Exon 3 5'UTR	Heterozygous	This study
RP509M	c.-349G>T	-	-	Exon 2 5'UTR	Heterozygous	This study
Families with very likely pathogenic changes reported in other populations						
RP155B	c.6714delT	p.I2239SfsX17	LamininG	Exon 33	Homozygous	This study Collin et al., 2008.
RP194B	c.9405T>A	p.Y3135X	LamininG	Exon 43	Homozygous	This study Collin et al., 2008.

Genbank Reference Sequence and Version FJ416331; GI: 212675237; Transcript Reference Sequence: NM_001142800.1

Nucleotide numbering reflects cDNA numbering with +1 corresponding to the A of the ATG translation initiation codon in the reference sequence, according to journal guidelines (www.hgvs.org/mutnomen). The initiation codon is codon 1.

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3 Table 3. Novel variations identified by direct genomic sequencing of *EYS*

Gene_exon	Nucleotide Change	Predicted effect	Patients Frequency	- Controls frequency
<i>EYS_1</i>	c.-462G>C	-	2/188	0/400
<i>EYS_2</i>	c.-349G>T	-	1/188	0/400
<i>EYS_3</i>	c.-204G>A	-	2/188	0/400
<i>EYS_4</i>	c.748+209delA c.78_79dupGC	- p.Q27RfsX16	1/188 1/188	- 0/400
<i>EYS_6-11</i>	c.862-10671_1766+10020del	p.G288DfsX21	2/12	0/400
<i>EYS_9-10</i>	c.1300-17039_1599+22208del	p.Y433_E533del	2/12	0/400
<i>EYS_11</i>	c.1766+61A>G	-	7/188	-
<i>EYS_13</i>	c.2024-14_-13insT	-	13/188	-
<i>EYS_14</i>	c.2234A>G	p.N745S	2/188	0/400
<i>EYS_17+18</i>	c.2733T>C	p.N911N	1/188	-
<i>EYS_24</i>	c.3684+61T>A	-	2/188	-
<i>EYS_25</i>	c.3877+18_22delAGATA	-	10/188	68/400
<i>EYS_26</i>	c.4450T>C c.4451G>A c.4120C>T	p.W1484R p.W1484X p.R1374X	1/188 1/188 1/188	0/400 0/400 0/400
<i>EYS_29</i>	c.6050G>T c.5959A>C	p.G2017V T1987P	1/188 1/188	0/400 2/400
<i>EYS_30</i>	c.6119T>A	p.V2040D	1/188	2/400
<i>EYS_31</i>	c.6424+1G>T	-	1/188	0/400
<i>EYS_33</i>	c.6632C>T	p.S2211L	1/188	0/400
<i>EYS_37</i>	c.7361delA	p.H2454PfsX8	1/188	0/400
<i>EYS_38</i>	c.7507G>A c.7578+18C>T	p.E2503K -	1/188 2/188	0/400 6/400
<i>EYS_39</i>	c.7666A>T c.7723+64T>A	p.S2556C -	2/188 3/188	20/400 34/400
<i>EYS_43</i>	c.9178_9181delATAA c.8834G>A	p.N3061TfsX3 p.G2945E	1/188 1/188	0/400 0/400

46 Genbank Reference Sequence and Version FJ416331; GI: 212675237; Transcript Reference Sequence:
47 NM_001142800.148 Nucleotide numbering reflects cDNA numbering with +1 corresponding to the A of the ATG translation initiation
49 codon in the reference sequence, according to journal guidelines (www.hgvs.org/mutnomen). The initiation codon
50 is codon 1.

53 Families with novel very likely pathogenic changes and both alleles affected

54 As previously mentioned, 5 out of the 10 families bearing novel very likely pathogenic changes have both
55 alleles affected. In 4 of them, they occurred as compound heterozygotes and hence it is sufficient to explain the
56 recessive phenotype in their corresponding patients (Table 2, Fig. 4). In the case of family RP1052M, 1 frameshift
57 deletion of 4 nucleotides involving the loss of the last residues of the Laminin G domain in the C-terminal region
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of *EYS* together with an in frame deletion of 300 nucleotides leading to a protein truncation of 100 amino acid residues that disrupts one EGF-like domain were identified in the same patient. In family RP1237M, the truncating p.W2640X change and the missense p.N745S variation, also occurring in heterozygosity in RP81 family, are showing a recessive segregation through generations I and II. The same applies to p.S2211L and p.G2945E double heterozygotes in family RP136B (Fig. 4). These protein substitutions p.S2211L and p.G2945E are each transmitted by one of the progenitors. The altered residues are part of Laminin G and EGF domains, respectively. Turning to evolutionary conservation, both Serine and Glycine are present in these positions in *EYS* homologues characterised in this study. The new residue at amino acid position 2211 is of different polarity than Serine, and the substitution of Glycine to Glutamate in position 2945 introduces an acidic polarity in a previously hydrophobic position. Whereas the latter is not tolerated according to computational predictions, the former implies the loss of one phosphorylation site (NetPhos2.1). Finally, all affected members of family RP367B were compound heterozygous for a splice site and a nonsense mutation (c.6424+1G>T, p.R1374X) (Fig. 4). The c.6424+1G>T variation is predicted to lead to an abolishment of the donor splice site located at this position. It is known that splice sites mutations may disrupt protein function by diverse mechanisms such as exon skipping or the use of cryptic acceptor sites, presenting even multiple splice outcomes for a mutation in a given splice site [Takahara et al., 2002].

The fifth family of this group, RP84B, presents a homozygous coding variation, p.G2017V, which alters a residue that lies within a Laminin G domain of the protein and it is predicted not to be tolerated by SIFT. Family segregation of this variation shows a transmission pattern compatible with the recessive trait of the disease (Fig. 4).

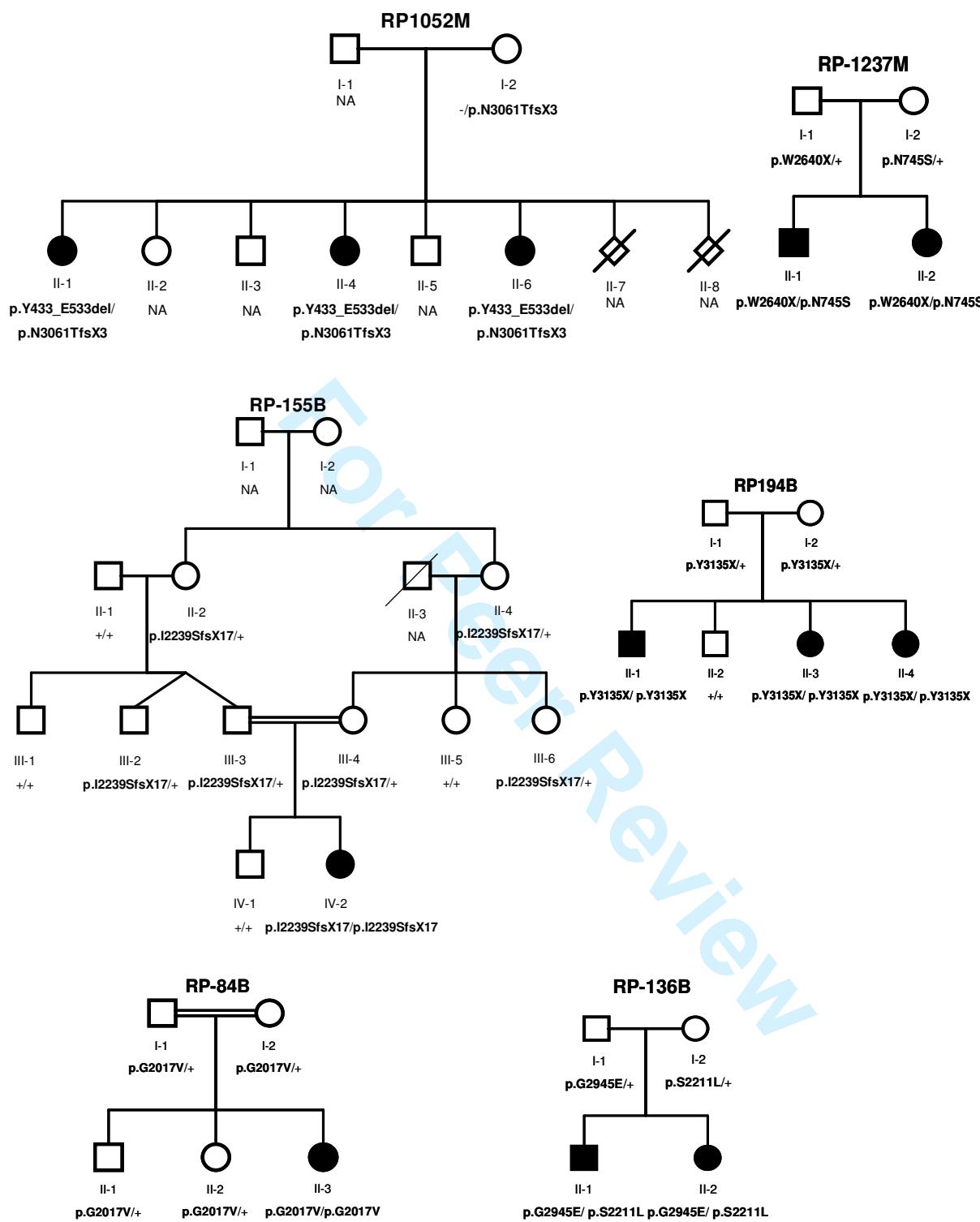
Families with single novel very likely pathogenic changes

The rest of the families comprising the group with novel very likely pathogenic changes present only single mutations. This is the case in families RP358B, RP60, RP180M, RP33 and RP81, with p.H2454PfsX8, p.Q27RfsX16, p.G288DfsX21, p.W1484X and p.N745S mutations, respectively (Table 2, Fig. 4).

Families with novel possible pathogenic changes

The group of families with possibly pathogenic variations is composed of 2 families bearing missense variations which did not appear in 200 control individuals and affect important domains of the protein (VRP8 and RP383, Table 2 and Fig. 4), and 3 additional families with variations in the 5' UTR segment of *EYS* (RP228B, RP107B and RP509M, Table 2 and Fig. 4). Of these, transversion c.-462G>C deserves special interest as it has been identified in a homozygous state in the proband of the consanguineous family RP228B. The hypothesized pathogenic potential for this change would ensue from its position in regulatory sequences important for protein translation [Scheper et al., 2007]. Moreover, family segregation is compatible with disease.

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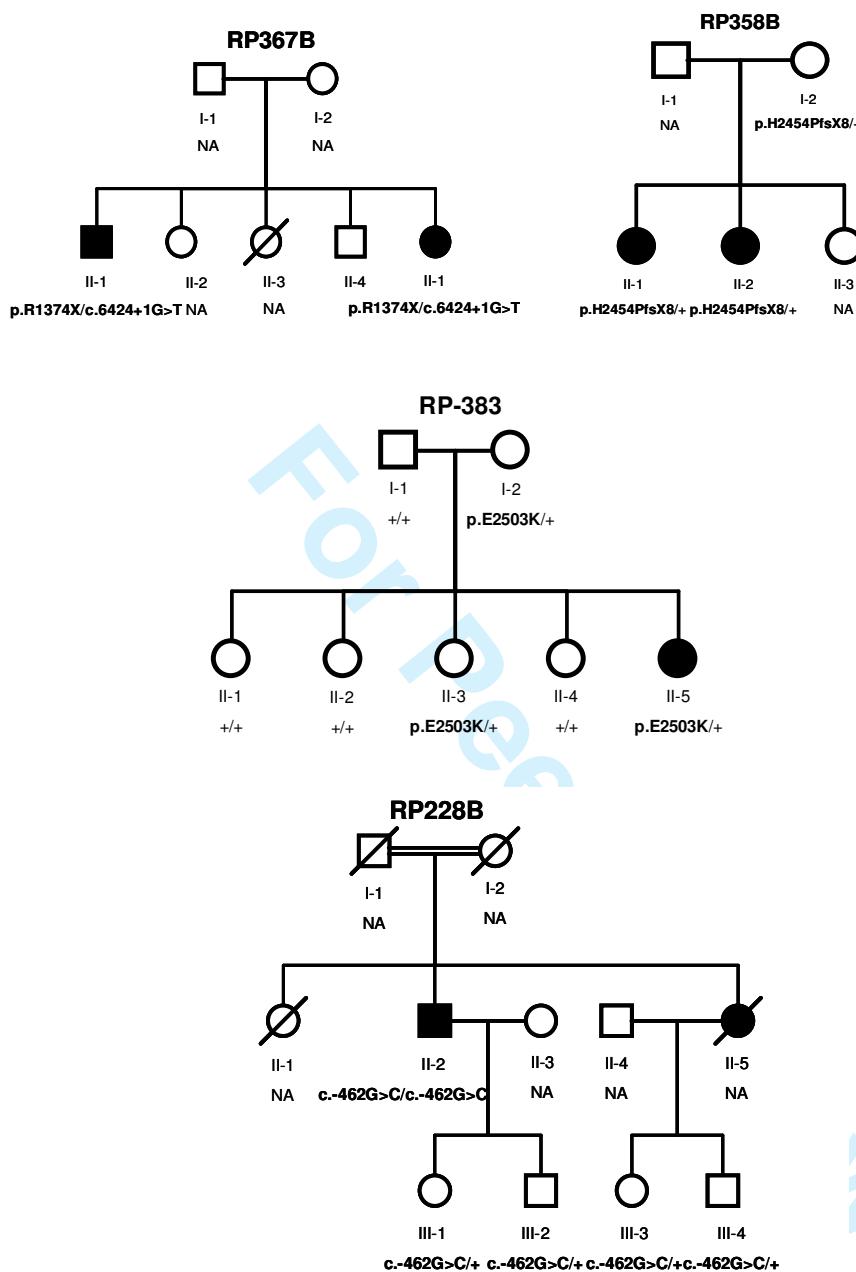
Mutation spectrum of *EYS* in Spanish patients with autosomal recessive retinitis pigmentosa 13

Figure 4. Family segregation of variations identified in the studied arRP families. Below the individuals, genotypes are presented for each change detected to segregate with the RP. For example, p.G2017V/p.G2017V represents homozygous mutants; p.G2017V/+ indicates heterozygous carriers, +/- indicates individuals carrying two wild-type alleles, whereas p.R1374X/c.6424+1G>T represents individuals presenting both mutations as compound heterozygous. NA means non available DNA sample.

Considering only the most likely pathogenic variations (truncating, stop, and frameshift), a prevalence estimate of 9.6% of distinct *EYS* variants in the Spanish arRP population can be drawn. Additionally, if the very likely pathogenic changes are included in the prevalence estimation, the figure could rise up to 15.9%. In an additional

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study performed in a separate population from United Kingdom, we have recently published eleven other novel mutations within *EYS* with probable allele frequency of 11% [Abd El-Aziz et al., 2010].

Furthermore, it is worth mentioning that many of the domains that feature *EYS* protein are targeted by pathogenic variations. Yet, different pathogenic mechanisms are postulated depending on the nature of the variation. Probably, in the case of variations generating a premature stop codon, most of these altered mRNA transcripts will be lost through Nonsense Mediated Decay (NMD) [Frischmeyer et al., 2002].

With regard to the coding mutations, there is not a clear clustering of both the previously reported mutations and those identified in this study to indicate a common target in the primary sequence of the protein. Although the distribution of these mutations implicated in arRP reveals 15 affected exons, a trend for alterations in residues of the C-terminal region containing alternating EGF-Laminin G domains of the protein is observed. Particularly, 4 out of the 6 missense variations are found in the Laminin G and EGF domains of the second half of the protein (Fig. 3). This is consistent with the hypothesis presented by other groups that the C-terminal region would be crucial for the function of the protein [Collin et al., 2008], and with the homology results presented in the current study, which outline the C-terminal region as one of the highly conserved intervals. Accordingly, the homology analysis presented here also reveals the high degree of evolutionary conservation of all the domains presenting altered residues by the mutations identified in arRP patients (Supplementary Figure S1).

Interestingly, mutations p.I2239SfsX17, p.Y3135X and p.W2640X had been previously reported as disease causing in 2 Dutch and 1 Spanish family respectively [Abd El-Aziz et al., 2008; Collin et al., 2008]. We have performed an extensive haplotype analysis and based on all available marker data we get different genotype information associated with the mutant alleles. Therefore, it is very likely that they are recurring mutations. Identifying recurrent mutations in Caucasian and especially specific populations such as the Spanish one provides an essential source for the molecular and clinical diagnosis of such a heterogeneous disease. Furthermore, this fact reinforces our hypothesis that *EYS* is the first prevalent gene in arRP [Abd El-Aziz et al., 2008].

The identification of 6 missense variations within the disease related changes in *EYS* differs from the mainly deletion/truncation mutations reported in previous studies ([Abd El-Aziz et al., 2008; Collin et al., 2008]. Nonetheless, the identification of missense mutations in arRP patients have already been reported [Sun et al., 1997; Molday et al., 2000], suggesting a refinement of the model based on the observation that some missense alleles might behave as true null allele at the functional level and may be responsible for severe impairment of protein function. Furthermore, majority of missense mutations reported here are located in functional conserved *EYS* domains and are more prevalent in the domains of the second half of the protein, thereby indicating a pathologic role for such variants (Fig. 3, Supplementary Figure S1).

It is noteworthy that a significant proportion of families in our study with an *EYS* mutation had only one identified mutation. Interestingly, structural variations have been found in 2 out of the 6 assessed patients. Besides, in the previous paper we had identified 2 large deletions which are not detectable by direct genomic sequencing [Abd El-Aziz et al., 2008] and this may explain why in a proportion of patients the second mutation remained unidentified as reported in the current work. Additional experiments consisting of Copy Number Variations (CNVs) or MLPA (Multiplex Ligation-dependent Probe Amplification) analysis would be useful to rule out long heterozygous deletions.

The identification of distinct mutations in *EYS* reveals a probable mutation frequency of 15.9% in the Spanish arRP population. Along with the detection of three recurrent mutations in Caucasian population, our hypothesis of *EYS* being the first prevalent gene in arRP has been reinforced in the present study.

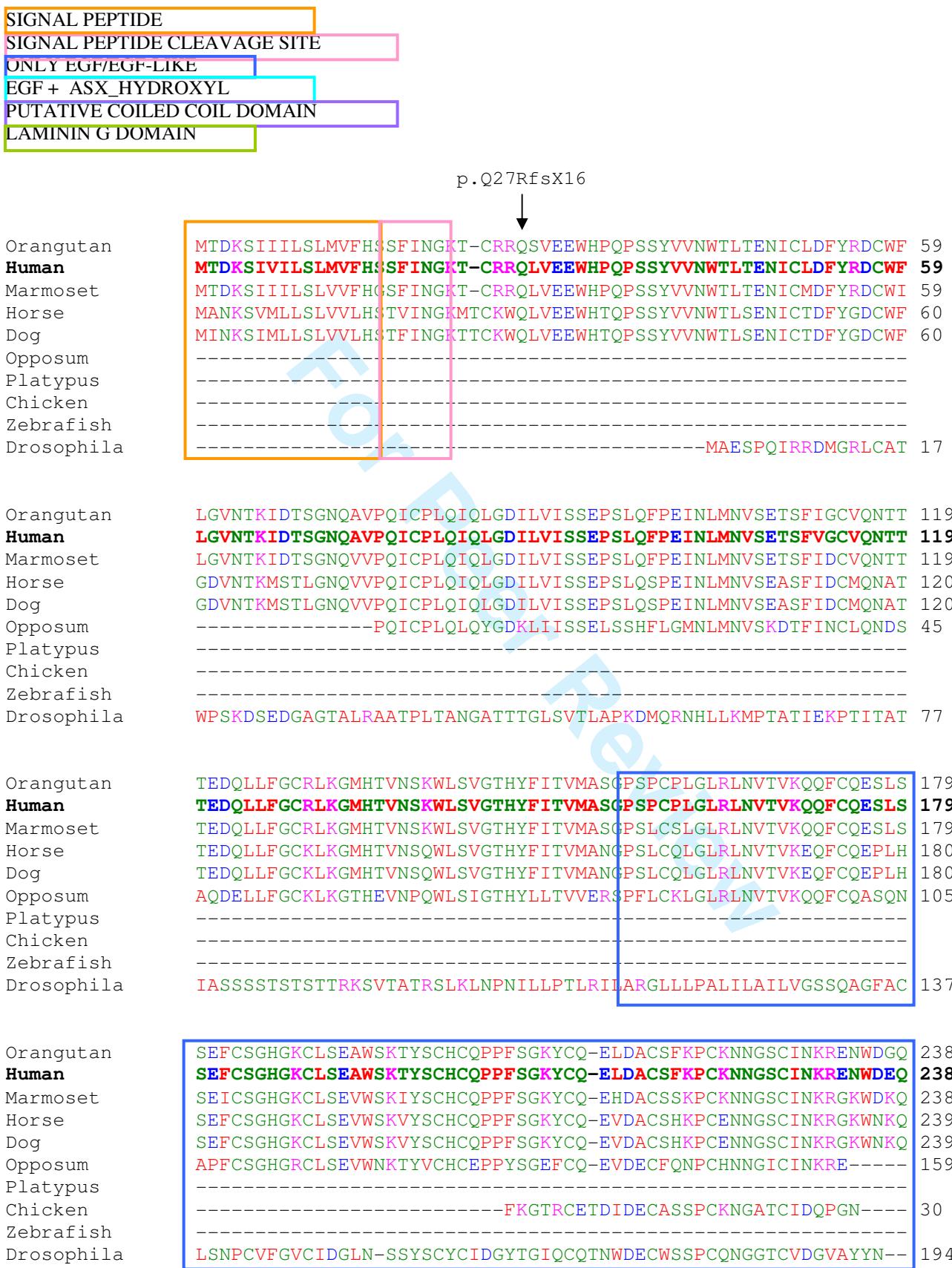
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REFERENCES

- Abd El-Aziz MM, Barragan I, O'Driscoll CA, Goodstadt L, Prigmore E, Borrego S, Mena M, Piersas JI, El-Ashry MF, Safieh LA, Shah A, Cheetham ME, Carter NP, Chakarova C, Ponting CP, Bhattacharya SS, Antinolo G. 2008. EYS, encoding an ortholog of Drosophila spacemaker, is mutated in autosomal recessive retinitis pigmentosa. *Nat Genet* 40:1285-1287.
- Abd El-Aziz MM, O'Driscoll CA, Kaye RS, Barragan I, El-Ashry MF, Borrego S, Antinolo G, Pang CP, Webster A, Bhattacharya SS. 2010. Identification of Novel Mutations in the ortholog of Drosophila eyes shut Gene (EYS) Causing Autosomal Recessive Retinitis Pigmentosa. *Invest Ophthalmol Vis Sci, in press*.
- Ayuso C, Garcia-Sandoval B, Najera C, Valverde D, Carballo M, Antinolo G. 1995. Retinitis pigmentosa in Spain. The Spanish multicentric and multidisciplinary group for research into retinitis Pigmentosa. *Clin Genet* 48:120-122.
- Barragán I, Abd El-Aziz MM, Borrego S, El-Ashry MF, O'Driscoll C, Bhattacharya SS, Antinolo G. Linkage validation of RP25 Using the 10K genechip array and further refinement of the locus by new linked families. 2008. *Ann Hum Genet* 72:454-462.
- Bell SE, Mavila A, Salazar R, Bayless KJ, Kanagala S, Maxwell SA, Davis GE. 2001. Differential gene expression during capillary morphogenesis in 3D collagen matrices: regulated expression of genes involved in basement membrane matrix assembly, cell cycle progression, cellular differentiation and G-protein signaling. *J Cell Sci* 114:2755-2773.
- Blom N, Gammeltoft S, Brunak S. 1999. Sequence- and structure-based prediction of eukaryotic protein phosphorylation sites. *J Mol Biol* 294:1351-1362.
- Böhm S, Jin H, Hughes SM, Roberts RG, Hinits Y. 2008. Dystrobrevin and dystrophin family gene expression in zebrafish. *Gene Expr Patterns* 8:71-78.
- Brunak S, Engelbrecht J, Knudsen S. 1991. Prediction of Human mRNA Donor and Acceptor Sites from the DNA Sequence. *J Mol Biol* 220:49-65.
- Collin RW, Littink KW, Klevering BJ, van den Born LI, Koenekoop RK, Zonneveld MN, Blokland EA, Strom TM, Hoyng CB, den Hollander AJ, Cremers FP. 2008. Identification of a 2 Mb human ortholog of Drosophila eyes shut/spacemaker that is mutated in patients with retinitis pigmentosa. *Am J Hum Genet* 83:594-603.
- Daiger SP, Bowne SJ, Sullivan LS. 2007. Perspective on Genes and Mutations Causing Retinitis Pigmentosa. *Arch Ophthalmol* 125:151-158.
- Fairbrother WG, Yeh RF, Sharp PA, Burge CB. 2002. Predictive identification of exonic splicing enhancers in human genes. *Science* 297:1007-1013.
- Frischmeyer PA, van Hoof A, O'Donnell K, Guerrero AL, Parker R, Dietz HC. 2002. An mRNA surveillance mechanism that eliminates transcripts lacking termination codons. *Science* 22:2258-2261.
- Hamel C. Retinitis pigmentosa. 2006. *Orphanet J Rare Dis* 1:40.
- Hartong DT, Berson EL, Dryja TP. 2006. Retinitis pigmentosa. *Lancet* 368:1795-1809.
- Humphries P, Farrar GJ, Kenna P, McWilliam P. 1990. Retinitis pigmentosa: genetic mapping in X-linked and autosomal forms of the disease. *Clin Genet* 38:1-13.
- Husain N, Pellikka M, Hong H, Klimentova T, Choe KM, Clandinin TR, Tepass U. 2006. The agrin/perlecan-related protein eyes shut is essential for epithelial lumen formation in the Drosophila retina. *Dev Cell* 11:483-93.
- Jarjanazi H, Savas S, Pabalan N, Dennis JW, Ozcelik H. 2008. Biological implications of SNPs in signal peptide domains of human proteins. *Proteins* 70:394-403.
- Mason JM, Arndt KM. 2004. Coiled coil domains: stability, specificity, and biological implications. *ChemBioChem* 5:170-176.
- Matlin AJ, Clark F, Smith CW. 2005. Understanding alternative splicing: towards a cellular code. *Nat Rev Mol Cell Biol* 6:386-398.

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- 1
2
3 - Molday LL, Rabin AR, Molday RS. 2000. ABCR expression in foveal cone photoreceptors and its role in Stargardt macular
4 dystrophy. *Am J Ophthalmol* 130:689.
- 5
6 - Pomares E, Marfany G, Brión MJ, Carracedo A, Gonzalez-Duarte R. 2007. Novel high-throughput SNP genotyping
7 cosegregation analysis for genetic diagnosis of autosomal recessive retinitis pigmentosa and Leber congenital amaurosis.
8 *Hum Mutat* 28:511–516.
- 9
10 - Ramensky V, Bork P, Sunyaev S. 2002. Human non-synonymous SNPs: server and survey. *Nucleic Acids Res* 30:3894–3900.
- 11
12 - Ruiz A, Borrego S, Marcos I, Antiñolo G. 1998. A major locus for autosomal recessive retinitis pigmentosa on 6q, determined
13 by homozygosity mapping of chromosomal regions that contain gamma-aminobutyric acid-receptor clusters. *Am J Hum
Genet* 62:1452–1459.
- 14
15 - Sadoulet-Puccio HM, Rajala M, Kunkel LM. 1997. Dystrobrevin and dystrophin: an interaction through coiled-coil motifs.
16 *Proc Natl Acad Sci U S A* 94:12413–12418.
- 17
18 - Scheper GC, van der Knaap MS, Proud CG. 2007. Translation matters: protein synthesis defects in inherited disease. *Nat Rev
Genet* 8:711–723.
- 19
20 - Sun H, Nathans J. 1997. Stargardt's ABCR is localized to the disc membrane of retinal rod outer segments. *Nat Genet* 17:15–
16.
- 21
22 - Takahara K, Schwarze U, Imamura Y, Hoffman GG, Toriello H, Smith LT, Byers PH, Greenspan DS. 2002. Order of intron
23 removal influences multiple splice outcomes, including a two-exon skip, in a COL5A1 acceptor-site mutation that results in
24 abnormal pro-alpha1(V) N-propeptides and Ehlers-Danlos syndrome type I. *Am J Hum Genet* 71:451–465.
- 25
26 - Wang M, Marin A. 2006. Characterization and prediction of alternative splice sites. *Gene* 366:219–227.
- 27
28 - Yang RB, Ng CK, Wasserman SM, Colman SD, Shenoy S, Mehraban F, Komuves LG, Tomlinson JE, Topper JN. 2002.
29 Identification of a novel family of cell-surface proteins expressed in human vascular endothelium. *J Biol Chem* 277:46364–
46373.
- 30
31 - Yeo G, Hoon S, Venkatesh B, Burge CB. 2004. Variation in sequence and organization of splicing regulatory elements in
32 vertebrate genes. *Proc Natl Acad Sci U S A* 101:15700–15705.
- 33
34 - Zelhof AC, Hardy RW, Becker A, Zuker CS. 2006. *Nature* 443:696–699.
- 35
36 - McCarroll SA, Hadnott TN, Perry GH, Sabeti PC, Zody MC, Barrett JC, Dallaire S, Gabriel SB, Lee C, Daly MJ, Altshuler
37 DM, International HapMap Consortium. Common deletion polymorphisms in the human genome. 2006. *Nat Genet* 38:86–
92.
- 38
39 - Sharp AJ, Locke DP, McGrath SD, Cheng Z, Bailey JA, Vallente RU, Pertz LM, Clark RA, Schwartz S, Segraves R, Oseroff
40 VV, Albertson DG, Pinkel D, Eichler EE. 2005. Segmental duplications and copy-number variation in the human genome.
41 *Am J Hum Genet* 77:78–88.
- 42
43 - de Smith AJ, Tselenko A, Sampas N, Scheffer A, Yamada NA, Tsang P, Ben-Dor A, Yakhini Z, Ellis RJ, Bruhn L, Laderman
44 S, Froguel P, Blakemore AI. 2007. Array CGH analysis of copy number variation identifies 1284 new genes variant in
45 healthy white males: implications for association studies of complex diseases. *Hum Mol Genet* 16:2783–2794.
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4	Orangutan	GYECVCHPPFTGKNCSEIIIGQCQPHVCFHGNCSNITSNSFICECDEQFSGPFCEMSTKPC	298
5	Human	AYECVCHPPFTGKNCSEIIIGQCQPHVCFHGNCSNITSNSFICECDEQFSGPFCEVSAKPC	298
6	Marmoset	GYECVCHPPFTGKNCSEIIIGQCQPHICFHGNCSNITSNSFICECDERFKGPFCEVSTKPC	298
7	Horse	GYECICHPPFTGKNCSEIIIDQCQPYVCFHGNYSNITTSNSFICECDEPFSG-----	289
8	Dog	GYECICHPPFTGKNCSEIIIDQCQPYVCFHGNYSNITTSNSFICECDEPFSG-----	289
9	Opposum	-----FTGRNCEEVIDYCR-----	173
10	Platypus	-----	
11	Chicken	-----YFCQCMAPFKFVN-----	43
12	Zebrafish	-----	
13	Drosophila	-----CTCPPEGFGSNCEENVDECMSNPCQNGGLCRDRTNGYICTCQPGYLG-----	241
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16	Orangutan	VSLLCWKRGICPNSSSAYTYECPKGSSSQNGETDVSECSLVPQNGTDCIQLISNDVMCIC	358
17	Human	VSLLFWKRGICPNSSSAYTYECPKGSSSQNGETDVSEFLVPQNGTDCIKISNDVMCIC	358
18	Marmoset	VSLLCWKRGICPNSSSAYTCECPKRSPSQNDEIDVNDCSLIPQNGTDCIKISNDVMCIC	358
19	Horse	-----	
20	Dog	-----	
21	Opposum	-----	
22	Platypus	-----	
23	Chicken	-----	
24	Zebrafish	-----	
25	Drosophila	-----	
26			
27	Orangutan	SPIFTDLLCKSIQTSCESFSLRNNATCKWEKDYHCSCISGFTEKNCEKAIDHCRLLSIN	418
28	Human	SPIFTDLLCKSIQTSCESFPLRNNATCKCEKDYPYCSCISGFTEKNCEKAIDHCKLLSIN	418
29	Marmoset	SPIFTDILCRSIQTSYEFPLKNTTICKCEKEYHC-----KNCEKVIDHCRLLSIN	410
30	Horse	-----KNCEKVIDHCRLLCVN-----	305
31	Dog	-----R-CSYYLGRIDRFCIL-----	304
32	Opposum	-----LLSIN-----	178
33	Platypus	-----	
34	Chicken	-----	
35	Zebrafish	-----	
36	Drosophila	-----SHCELDVAV-----	250
37			
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39	Orangutan	CLNEEWCFNIIGRF-YVCIPGCPKNPCWFLKNVYLIHQHLCYCGVTFHGICQDKGPAQFE	477
40	Human	CLNEEWCFNIIGRFKYVCIPGCTKNPCWFLKNVYLIHQHLCYCGVTFHGICQDKGPAQFE	478
41	Marmoset	CLNEEWCFNIIGRF-YVCIAGCKINPCWLVKNVYSISIHQHPCYYGVTFCRGICQDKGPAQFQ	469
42	Horse	CQNEGWCFCNIIIGRF-----	319
43	Dog	DVED-----	308
44	Opposum	CLNEGLCFNIIGGF-----	192
45	Platypus	-----	
46	Chicken	-----	
47	Zebrafish	-----	
48	Drosophila	CETGTGAR-----	258
49			
50			
51	Orangutan	YVWQLGFLAGSEGEKCQGVIDAYFFLAANCTEDAIKVNDPEDYNSSCRFPREGTKEICANG	537
52	Human	YVWQLGFLAGSEGEKCQGVIDAYFFLAANCTEDATYVNDPEDDNSSCWFPHEGTKEICANG	538
53	Marmoset	CVWQLGFTGSEGEKCQRVIDVYFFLAANCTEDAIYVNMPEDINNSCWFPCEGTKEICANG	529
54	Horse	-----	
55	Dog	-----	
56	Opposum	-----	
57	Platypus	-----	
58			
59			
60			

Mutation spectrum of *EYS* in Spanish patients with autosomal recessive retinitis pigmentosa 19

1
2
3 Chicken
4 Zebrafish
5 Drosophila
6
7

8 Orangutan	CSCLSEEDSQEYLYLCFLRWAGNMYLENTDDQENEQHEAICKDEINRPRCSCSLSYIG	597
9 Human	CSCLSEEDSQEYRYLCFLRWAGNMYLENTDDQENEQHEAVCKDEINRPRCSCSLSYIG	598
10 Marmoset	CSSFRKEDSQEQYQCLCFLRWADKYLENTDDQENEYQHEAICKDEINRPRCSCSLSCSG	589
11 Horse	-----	
12 Dog	-----	
13 Opposum	-----	
14 Platypus	-----	
15 Chicken	-----	
16 Zebrafish	-----	
17 Drosophila	-----	

19 Orangutan	RLCVVNVDYCLGNQSISVHGLCLALSHKCNCISLQRYERNICEIDTEDCKSVSCKNGTTS	657
20 Human	RLCVVNVDYCLGNHSISVHGLCLALSHNCNCNSGILQRYERNICEIDTEDCKSASCKNGTTS	658
21 Marmoset	RLCVVNVGYCLENQSISGHGLCLAHLDNCNCSELQRYERNICEIDTEDCKSVSCKNGTTS	649
22 Horse	-----	
23 Dog	RVCLGNESISVHGLCLVRLHNCNCSCILQRYERNICEIETEDCKSVPCKNGTTG	372
24 Opposum	-----	
25 Platypus	CLGNESISVHGLCLVRLHNCNCSCILQRYERNICEIETEDCKSVPCKNGTTG	359
26 Chicken	-----	
27 Zebrafish	-----	
28 Drosophila	-----	

31 Orangutan	IHLRGYFFCKCVPFGK---GTQREIDIDECA SHPCKNGATCIDQPGNYFCQ	712
32 Human	THLRGYFFRKCVPFGK---GTQCEIDIDECA SHPCKNGATCIDQPGNYFCQ	713
33 Marmoset	IHLRGYVFCKCVPFGKGFE GTRCKIDVDECASHPCKNGATCTDQPGNYLCQ	709
34 Horse	-----	
35 Dog	IHSSGYFFCKCVPFGFTG-----	392
36 Opposum	IHSSGYFFCKCVPFGKGT---RSETDTDERASHPFKNGATCADQPGNYFCQ	414
37 Platypus	VAPFS---V	193
38 Chicken	-----	
39 Zebrafish	-----	
40 Drosophila	CQI GGE CIE GP	269

42 p.N745S
43
44

45 Orangutan	VDGF SCLGNPGYVGIRCEQDIDDCILNACEHNSTCKDLHLSYQCVC	772
46 Human	VDGF SCLCNPGYVGIRCEQDIDDCILNACEHNSTCKDLHLSYQCVC	773
47 Marmoset	VDGF SCLCNPCCAGVRCEQDIDDCILNACEHNSTCKELHLSFQCVC	769
48 Horse	LSGWEGNFSEQESN-----	432
49 Dog	AVGF SCLCSAACVGLRCEQDIDDCNLNACEHTSACKDLYL-----	454
50 Opposum	AVGF SCLCSAACVGLRCEQDIDDCNLNACEHTSACKDLYL-----	234
51 Platypus	VDGF SCLCNPGYAGLRCEQTIDDCITHACENN STCQDLHLR-----	82
52 Chicken	--GFYCLCNPGYAGLTCEQDIDDCINNACEHNSTCVDLHLV-----	17
53 Zebrafish	MCEDTSENGCLFIISVR-----	311
54 Drosophila	GLEFTCDCPAGWHGRICQEEINECASSPCQNGGV	

20 <Barragán et al.>

1							
2							
3	Orangutan	ECKMNPCKNNSTCIDL Y KSYRCECTSGWTGQNCSEEINECSDPCMNGGLCHESTIPGQF					832
4	Human	ECKMNPCKNNSTCTDLYKSYRCECTSGWTGQNCSEEINECSDPCMNGGLCHESTIPGQF					833
5	Marmoset	ECEMNPKNNSTCIDL Y KSYRCECTSGWTGQNCSEEINECDCPCMNGGLCHESTIPGQF					829
6	Horse	-----CECTSGWTGQNCSEEINECSDPCLNGALCHESTIPGQF					471
7	Dog	-----CECTSGWTGQNCSEEINECSDPCLNGALCHESTIPGQF					493
8	Opposum	-----CGCTPGWTGQNCSEEINECDASPCKNGAICQESTVPGQF					273
9	Platypus	-----RCICKPEWTGQNCSEELNKCDSNPCMNGATCFNSAVPGKV					40
10	Chicken	-----CLCTAGWTGPDCSEDINECDSEPCLNAGATCYESVKQQF					121
11	Zebrafish	-----SAKSPASQSQWHEPPSTSSTHSPTL---LITVETPPGEW					53
12	Drosophila	-----ACACPMGYTGINCEEILICADNPQCQNNALCLMEEG--VP		*	*	*	349
13							
14							
15	Orangutan	VCLCPPLYTGRFCHQRYNPCDLLHNPCRNNSTC--LALVDGNQHCICREEFEKGKNEIDV					890
16	Human	VCLCPPLYTGFCHQRYNLC DLLHNPCRNNSTC--LALVDANQHCICREEFEKGKNEIDV					891
17	Marmoset	VCLCPPLYTGQFCHQRYNPCCELLNNPCCRNNSTC--LALVDGNQHCICR-EFEGKHCEIDT					886
18	Horse	-----LNDPCRNNATC-LTLDGQRYCVCR-----					508
19	Dog	VCLCPPFTGKFC-----LNDPCRNNATC-LTLDGQRYCVCR-----					539
20	Opposum	VCLCPPFTGFFCQQIYNPCDMAYNLCIINNSTC-LITVDGNSNCVCR-----					319
21	Platypus	ECLCPPLYTGAFCHHNSCTFHKPCINNSTC-LTRADGNTEFICF-----					86
22	Chicken	VCICPPFYTGDFCHQRFSPELPYNPCINNSTC-LAQADGNPMCICK-----					167
23	Zebrafish	VRVLSPQTQPAVCPQG-----ICLNGGTCPVSLPSGASSFFCD-----					92
24	Drosophila	TCYCVPDYHGEKCEFYDECQLG-PRCMNGGVC-----IDGVDTFSCS-----		*	*	*	391
25				*	*	*	
26							
27	Orangutan	KECLFLSCQDYGICEDMVNNFRCICRPGFSGSLCEIEINECSSEPCKNNGTCVDLTNRFF					950
28	Human	KDCLFLSCQDYGICEDMVNNFRCICRPGFSGSLCEIEINECSSEPCKNNGTCVDLTNRFF					951
29	Marmoset	NECLFLPCQGYGICEAIVNNFRCICRPGFSGSLCEIEINECSSEPCKNNGTCVDLTNRFF					946
30	Horse	-----ERCVORPGFSGPLCEIETNECSSKPKCKNNGTCVDLTN-----					545
31	Dog	-----ERCVORPGFSGPLCEIETNECSSKPKCKNNGTCVDLTN-----					576
32	Opposum	-----CTOSPGFSGSQCEIEINECYSTPCKNNGTCVDLINR-----					355
33	Platypus	-----IGARCETDIDECDSFPCKNRANCIDQPG-----					114
34	Chicken	-----TGASVSMAS-----					176
35	Zebrafish	-----PPLLTGMLCECLMVGEESLCDNYTAPATQSPPR-----					
36	Drosophila						425
37							
38	Orangutan	CNCEPGYHGPFCLELDVNKCKISPCLDEENCVYRTDGYNCLCAPGYTGINCEINLDECLSE					1010
39	Human	CNCEPEYHGPFCLELDVNKCKISPCLDEENCVYRTDGYNCLCAPGYTGINCEINLDECLSE					1011
40	Marmoset	CNCEPGYHGPLCLEDINECKTSPCLDEENCVYRADGYNCLCAPGYTGINCEINLDECLSE					1006
41	Horse	-----RCLSK-----					550
42	Dog	-----RCLSK-----					581
43	Opposum	-----CEVNINECLSE-----					366
44	Platypus	-----					
45	Chicken	-----					
46	Zebrafish	-----					
47	Drosophila	-----RTTTSTMA-----					434
48							
49							
50	Orangutan	PCLHDGVCIDGINHYTCDC K SGFFGTHCETNANDCLSNPCLHGRCTELINEYPCSCDADG					1070
51	Human	PCLHDGVCIDGINHYTCDCKSGFFGTHCETNANDCLSNPCLHGRCTELINEYPCSCDADG					1071
52	Marmoset	PCLHDGVCIDGINHYTCDC K SGFFGTHCETNANDCLSNPCLHGRCTELINEYPCSCDADG-----					1049
53	Horse	PCLHDGAWTDGVNHYTCDC K SGSGFLGTHCETNANDCLSNPCLHGR-----					593
54	Dog	PCLHDGAWTDGVNHYTCDC K DGFIGTHCETNANDCLSNPCLHGR-----					625
55	Opposum	PCLNDGICADGIISYYTCY K DGFIGTHCETNADAACLSDPCLHG-----					409
56	Platypus	-----NYFCQCVAFK-----VVVDGFYCLCN-----					135
57	Chicken	-----					
58							
59							
60							

Mutation spectrum of EYS in Spanish patients with autosomal recessive retinitis pigmentosa 21

Zebrafish	PPTVRPVTPPETVSPSRASEEVEIIVVTTSAPAEVVTSLSPS	478
Drosophila		
Orangutan	TSTOCKIKINIDCTSIPCMNEGFCQKS TSTOCKIKINIDCTSIPCMNEGFCQKS	1130
Human	SAHGF <small>C</small> IICP SAHGF <small>C</small> IICP	1131
Marmoset	NLGLGPSIPCMNEGFCQKSANGFTC NLGLGPSIPCMNEGFCQKSANGFTC	1102
Horse		
Dog	IDCTSVSCLNEGICQKSVHGVTC IDCTSVSCLNEGICQKSVHGVTC	676
Opposum		
Platypus		
Chicken		
Zebrafish		
Drosophila		
Orangutan	LNGGICVDGP <small>G</small> HFTDCR-CLPGFSGQFCE LNGGICVDGP <small>G</small> HFTDCR-CLPGFSGQFCE	1189
Human	ININECSSSPCLHGADCEDHINGYVCKCQPG ININECSSSPCLHGADCEDHINGYVCKCQPG	1190
Marmoset	LNGGICVDGP <small>G</small> D <small>F</small> RRCLPGFSGQFCDININ LNGGICVDGP <small>G</small> D <small>F</small> RRCLPGFSGQFCDININ	1162
Horse	R-CLPGFSGQFCEININECSSSPCLNGAN R-CLPGFSGQFCEININECSSSPCLNGAN	636
Dog	CLPGFSGQFCEININECSSSPCLNGAN CLPGFSGQFCEININECSSSPCLNGAN	735
Opposum	EDHINGYICKCQRG EDHINGYICKCQRG	453
Platypus	RRCP EGFSGNFCEVN RRCP EGFSGNFCEVN	165
Chicken	PGYAGLRC <small>D</small> QDIDDCIINTCDHN PGYAGLRC <small>D</small> QDIDDCIINTCDHN	197
Zebrafish	KDLH LGHYVRWRLMNAYQDPAKTMG	112
Drosophila	CPLHFTGRLCEQDITVFSPR CPLHFTGRLCEQDITVFSPR	520
Orangutan	SSSSSEEGSVEIKPTVAPPESGSHSIS SSSSSEEGSVEIKPTVAPPESGSHSIS	
Human	VEQTTAVPAQPE VEQTTAVPAQPE	
Marmoset	:	
Horse		
Dog		
Opposum		
Platypus		
Chicken		
Zebrafish		
Drosophila		
Orangutan	WSGHHCEKELECI WSGHHCENELECI	1249
Human	PNSCVHELCMENE PNSCVHELCMENE	1250
Marmoset	PGFMTCSIGLLCGDEIRRITCLTPIF PGFMTCSIGLLCGDEIRRITCLTPIF	1222
Horse	CLCTPGFMTRSIGLLC CLCTPGFMTRSIGLLC	658
Dog	DDEIRRISCLPPVF DDEIRRISCLPPVF	757
Opposum		
Platypus		
Chicken		
Zebrafish		
Drosophila		
Orangutan	PESEQEPESKPHPESES	537
Human		
Marmoset		
Horse		
Dog		
Opposum		
Platypus		
Chicken		
Zebrafish		
Drosophila		
Orangutan	QRTDPISTQTYTVPPSETLVSSFPSIKAT QRTDPISTQTYTI PPSETLVSSFPSIKAT	1309
Human	TRPAIMDTYPVDQGP TRPAIMDTYPVDQGP	1310
Marmoset	QRTDAIVTQIYAVPPSETLVSSFPSVKAT ARTDTISTQTHTVP PAPATSVHNFPRTGAPRLWTTMDTYPVDQGP	1282
Horse	TRPAIMDTYPVDQGP ARTDTISTQTHTVP PAPATSVHNFP	718
Dog	QRTDAIVTQIYAVPPSETLVSSFPSVKAT ARTDTISTQTHTVP PAPATSVHNFP	807
Opposum		
Platypus		
Chicken		
Zebrafish		
Drosophila		
Orangutan	ASESETETEEEII PGTTARPPTSRS SSSEESP SIFT	597
Human	TLPLPGKPQTSAS SESSGEVVT	
Marmoset		
Horse		
Orangutan	AALRISTPLESYLLEELIVTREL ATLRISTPLESYLQL	1369
Human	SAKHGLLSSAD SAKHSLSSAD	1370
Marmoset	VSSRFLNFGIHDPAQIVQDKTSVSH VSSRFLNFGIRDP AQIVQDKTSVSH	1342
Horse	SSSRTDVSSSRFLNFGIHDPAQTD RFLNFGVPGPAQVVWGKTSVPH	778

22 <Barragán et al.>

1		
2		
3	Dog	AALGTGISFERYLLKHVIAAKELLAKHSLPSSTDVSSSRFLNFGVPGPAQVVWGKTSVPH 867
4	Opposum	-----
5	Platypus	-----
6	Chicken	-----
7	Zebrafish	-----
8	Drosophila	SEYYTTVPHFEVSGSKESGSEEVTTVRPTAAPSITISVDITSSGSSSSSESVEVFVTPP 657
9		
10		p.R1374X
11		↓
12		
13	Orangutan	MPIRTSAATLGFFFDPDRARTPFIMSSVMSDFIFPTQSLLFENYQTVASSATPTTSVIRS 1429
14	Human	MPIRTSAATLGFFFDPDRARTPFIMSSLMSDFIFPTQSLLFENCQTVALSATPTTSVIRS 1430
15	Marmoset	MPIQTSAAATLGFFLSDRRAARTPFIMSSLMTDFISPTQSLLFENYQTVASSATIMTSVIRS 1402
16	Horse	LPIQASAATPRFFFDRGERT-----SSMTDFIFPTQSLLFESDRSVASSATTMSSVISG 833
17	Dog	LPIQASAATPRFFFDRGERT-----SSMTDFIFPTQSLLFESDRSVASSATTMSSVISG 922
18	Opposum	-----IFPTQVSSLKNHQTISLSATDMSSAISS 481
19	Platypus	-----LPVKAIILKQVTTSSSVIHKGSTQAP 192
20	Chicken	-----LVWTSYRFVLLDFLDSFVFKNKLMSVIHHL 226
21	Zebrafish	-----FDGNSFLELPSLTSLFQSDTYFPS---- 136
22	Drosophila	APVFVQRVTTIETSISIDYVTPTPLPETTTPRVVPVPRPTFAPEPPLDVETTASTHHLW 717
23		:
24		p.W1484X
25		p.W1484R
26		↓
27	Orangutan	IPGADIELNRQSLLSRGFLLTAASISATPVVSRGQAQEDIEEYSADSLISRREHWRLLSPS 1489
28	Human	IPGADIELNRQSLLSRGFLLIAASISATPVVSRGQAQEDIEEYSADSLISRREHWRLLSPS 1490
29	Marmoset	IPGADIKNRHSLLSRGFLLTAASISATPVVSRRAQEDIEEYSAVSLISRKEHWRLLSSS 1462
30	Horse	ILGADVELNRHSLLSHGFLLKASTGAPPVVSMDGAQEGIEEYSAVSLISRREYWRLLSSS 893
31	Dog	ILGADVELNRHSLLSHGFLLKASTGAPPVVSMDGAQEGIEEYSAVSLISRREYWRLLSSS 982
32	Opposum	IPGDEIELNSHSFLSHRFLL-----QEDTREYSAFSLSS----- 515
33	Platypus	LLS-----EYSIMAVAS----- 204
34	Chicken	VYMVQP----- 232
35	Zebrafish	-----
36	Drosophila	TEVPTTAAPFTTEYPAEVLIITHTRTSAGRFTTVQPPAGVTTSPTEDSSVELPTPHTPQI 777
37		
38		
39	Orangutan	MSPIFFPAKIIISKQVTILNSSALHRFGTKAFNPSEYQAITEASSNQRQLTNIKSQAADSLR 1549
40	Human	MSPIFFPAKVIISKQVTILNSSALHRFSTKAFNPSEYQAITEASSNQRQLTNIKSQAADSLR 1550
41	Marmoset	MSPIFFPAKIIISKQVTILNSSAVHRFDTKAFIPSEYQVITEASSNQRQLTNIKSQAADSLR 1522
42	Horse	MPPISPAPKVIISKQVAIVNSSLHRFTTQDSIPSEYQVITEASSNQRQLTNIKSQSADSL 953
43	Dog	MPPISPAPKVIISKQVAIVNSSLHRFTTQDSIPSEYQVITEASSNQRQLTNIKSQSADSL 1042
44	Opposum	-----QPADSL 522
45	Platypus	-----SKGRLTNIKSQSADSL 221
46	Chicken	-----VKIMSMG 239
47	Zebrafish	-----RSSEDKR 143
48	Drosophila	VVTILDNEVIPSLITTGSPPTTHHHHHHPHHEAEGTTLQPLEEDEHHHHHHDEFVTPP 837
49		
50		
51	Orangutan	ELSQTCACTCSMTEIKSSREFSDQVLHSKQSHFYETFWMNSAILASWCALMGAQITSGHS 1609
52	Human	ELSQTCACTCSMTEIKSSREFSDQVLHSKQSHFYETFWMNSAILASWYALMGAQITSGHS 1610
53	Marmoset	ELSQICTTCSMTEIKSSHEFSDQVSHSKQSHFYETFWMNSAILASWYALMGAQITSGHS 1582
54	Horse	ELSQTCATCSMTEIKSSHEFSDQVLHSKQSHFYETFWMNSELASWYALMRTQITSGHS 1013
55	Dog	ELSQTCATCSMTEIKSSHEFSDQVLHSKQSHFYETFWMNSELASWYALMRTQITSGHS 1102
56	Opposum	ELSQTCVTCMTEIKPKSSDECSVQALHSKQSQFYEPFWMNSAILSSWYLTGATVITSGHS 582
57	Platypus	ELIQTCACTCSMTEIKPSDEFSHQVLHSKQYQFYETFWMNSAILTSWYTLMRITSGHS 281
58		
59		
60		

Mutation spectrum of EYS in Spanish patients with autosomal recessive retinitis pigmentosa 23

1							
2							
3	Chicken	TLVSVKKSNIFFIKLFDFVFPDQVLHSKQSPFYEAFWMNSAILNSWYALMGATAITSAYL	299				
4	Zebrafish	ILYLTMK-----RTPHGSLLYCREQDLGERFLHVFLQNAVARLGCAGAHILTA	194				
5	Drosophila	QPVEITTGHPPLQTEDLIGVQEPAVVTESPFAPAETTVVPVVVPATIAPLGTAPPATPA	897				
6		:					
7							
8	Orangutan	FSSATEITPSVAFTEVPSSLFPSKKSAKRTILSSSLEESITLSSNLDVNLCLDKTCLSIVP	1669				
9	Human	FSSATEITPSVAFTEVPSSLFPSKKSAKRTILSSSLEESITLSSNLDVNLCLDKTCLSIVP	1670				
10	Marmoset	FSSATEITPSVAFTEVPSSLFPSKMSAKRTILSASLEESITLSSNLDVNLCLDKTCLSIVP	1642				
11	Horse	FSSATEIMPSVAFMEVSSSFPSKKSTKRRIISTPSVEDSIALSTNL DANLC LDKTRLSIVP	1073				
12	Dog	FSSATEIMPSVAFMEVSSSFPSKKSTKRRIISTPSVEDSIALSTNL DANLC LDKTRLSIVP	1162				
13	Opposum	FSSVTEITPSVEFTELSSPFSFKK-----	606				
14	Platypus	FSPATEITSSVEFTELSSSFLPKMS-----	306				
15	Chicken	FSSSSRITSSVEFTIEHPHIPLQK-----	323				
16	Zebrafish	VAAQNIRIDSLVAITVRYALPSQN-----	218				
17	Drosophila	PVPPATTPPPSPPSLATEPTLPPVTLPPVTQPPPPIPPTPPSTQS AQLLPPPTS	957				
18		:					
19							
20	Orangutan	SQTISSDLMNSDLTSKMTTDELSVSANILKLLKIRQYGITMGPTEEVLNQDSLLDMEKSKG	1729				
21	Human	SQTISSDLMNSDLTSKMTTDELSSENILKLLKIRQYGITMGPTEEVLNQESLLDMEKSKG	1730				
22	Marmoset	SQTMSLDLMNSDLTSQPTNDQLSVSENILKLLQIRQYGITMGPTEELNQDSLLDMEKCKG	1702				
23	Horse	SQTVSSDPLLNSDLTSELT-EDLSVSENILKLLKIGQYGITMGPTEEVLNQDNLLAVHESKG	1132				
24	Dog	SQTVSSDPLLNSDLTSELT-EDLSVSENILKLLKIGQYGITMGPTEEVLNQDNLLAVHESKG	1221				
25	Opposum	-----					
26	Platypus	-----					
27	Chicken	-----					
28	Zebrafish	-----					
29	Drosophila	AINVYTTPDGPPTASQTKPSVTESEEVEGTNTVSTGGRGSGGVPEEKAGDVDCIKLGCY	1017				
30							
31	Orangutan	SHTPFKLHPSDSSLDLFE NLQIYPDVTLKTYSEI THANDFKNTLPPLTGSVPDFSEVTTN	1789				
32	Human	SHTLFKLHPSDSSLDLFE NLQIYPDVTLKTYSEI THANDFKNNLPPLTGSVPDFSEVTTN	1790				
33	Marmoset	SHTLFKLHPCDSSLDLQLNLQSHPDVTLRTYSEIIHAN DLKNNLPPLTGSIPDFSEVSTN	1762				
34	Horse	SHKQLKLHTSDRSLDLFE NLPLSH-----PLKNNLPPYMDSRSDLSEVTSN	1177				
35	Dog	SHKQLKLHTSDRSLDLFE NLLKQ-----EVR-----	1247				
36	Opposum	-----					
37	Platypus	-----					
38	Chicken	-----					
39	Zebrafish	-----					
40	Drosophila	NGGTCVTTSEG SRCVC RFD RQGP-----	1040				
41							
42	Orangutan	VAFYT VSATPALS I QTSSSMSVIRPDWPF TDYMTSLKKEVKTSSEWSKWE LQPSVQYQE	1849				
43	Human	VAFYT VSATPALS I QTSSSMSVIRPDWPF TDYMTSLKKEVKTSSEWSKWE LQPSVQYQE	1850				
44	Marmoset	VAFYT VSATPALS I QTSS-MSVTRPEWPDTDYVNALKDIKTSSEWSKWE LQPSVQYQE	1821				
45	Horse	VAFYT VSAT-----QSLPVQTSTSSEWSKWE LQPSVHDWE	1212				
46	Dog	-----TSSEWSKWE LQPSVHDWE	1265				
47	Opposum	-----					
48	Platypus	-----					
49	Chicken	-----					
50	Zebrafish	-----					
51	Drosophila	-----					
52							
53							
54	Orangutan	FPTASWHLPFTRSLT LSSLES ILAPQQLTISDFSCVRYYGDSYLEFQNVVLPQNNI SLE	1909				
55	Human	FPTASRHL PFTRSLT LSSLES ILAPQR LMISDFSCVRYYGDSYLEFQNVVALNPQNNI SLE	1910				
56	Marmoset	FPTASRHL PFTRSFTLSSLES ILAPQQLMISDFSCVRYYGDSYLEFQNVLLNPQNNI YLE	1881				
57							
58							
59							
60							

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1						
2						
3	Horse	SPAASQTPAITRSLTLPSLESIPAPRQLMI	SDFTCVCYGGDSYLEFQDVFLNPQNNISLE		1272	
4	Dog	SPAASQTPAITRSLTLPSLESIPAPRQLMI	SDFTCVCYGGDSYLEFQDVFLNPQNNISLE		1325	
5	Opposum		SDFSYMHQYFGDSYLKERVLLPLQNSFSVE		636	
6	Platypus					
7	Chicken		IHTLIGLSNFGEQFLHLYLVEGRP	---TVR	350	
8	Zebrafish					
9	Drosophila		LCELPIIIRNAAFSGDSYVSHRIYKDIGGHESLDAVLPMHSQL		1083	
10						
11						
12	Orangutan	FQTFSYYGLLLYVKQDSNLVDGFFIQLFIENSTLIK	PGEAKFKSINTTIRVDDGQK		1964	
13	Human	FQTFSYYGLLLYVKQDSNLVDGFFIQLFIENGTLIK	YHFYCPGEAKFKSINTTIRVDDGQK	1970		
14	Marmoset	FQTFSYYGLLLYVKQDSNLVDGLSIQLIFIENGTLIK	PGEAKLKSINTTIRVDDGQK		1941	
15	Horse	FQTSSSYGLLLYMQQDSNSIDGFVTQLIFIENGTLIK	KYHFFCAGEAKLKNINTTIKVDDGQK		1332	
16	Dog	FQTSSSYGLLLYMQQDSNSIDGFVTQLIFIENGTLIK			1360	
17	Opposum	FQTFSNHGLLFIKHDTISMGKFFIQLFIEGGTLK			671	
18	Platypus		NFGEQFLHVYIVDGRLR		323	
19	Chicken	FSCGNQNILTSGNQTISKGIFIPIIIMLP			381	
20	Zebrafish					
21	Drosophila	KVRTRATNGLIMLAAAQGTKGGHYMALFLQKGLMQFQFSCG			1124	
22						
23						
24						
25	Orangutan	YTLLIR-QELYPCKAELTILGRNTQTCESISHVLGKPLPKSGSVFIGGFPDLHGKIQMPV			2023	
26	Human	YTLLIR-QELDPCNAELTILGRNTQTICESINHV	LGKPLPKSGSVFIGGFPDLHGKIQMPV	2029		
27	Marmoset	YTLLIRRQELDPCNAELTILGRNTQTSESINHV	LGLPKPRPKSGSVFIGGFPDLHGKIQP-I		2000	
28	Horse	YALLIRRQELDPCAEALTILGRTMKASESISHV	VSGRSLPESGSIFIGGFPDLHG-----		1386	
29	Dog	RQELDPCAEALTILGRTMKASESISHV	VSGRSLPESGSIFIGGFPDLHG-----		1408	
30	Opposum					
31	Platypus					
32	Chicken					
33	Zebrafish					
34	Drosophila					
35						
36						
37	Orangutan	PVKNF TGCIEVIEINNWRSFIPS KAVKNYHINNCRSQGFMSP	TASFVDVSDVTQGVDTM		2083	
38	Human	PVKNF TGCIEVIEINNWRSFIPS KAVKNYHINNCRSQGFMSP	TASFVDVSDVTQGVDTM	2089		
39	Marmoset	PVKNF TGCIEVIEINNWRSFIPS KAVKNYHIDNCRSQGFMSP	TASFVDVSDVTQGVDTM		2060	
40	Horse	PVENFTGCIEVIELNNWRSFIPS KAVKKIHVENC	RSQDSPLSAASAFVAPSGVTEGVAST		1446	
41	Dog	PVENFTGCIEVIELNNWRSFIPS KAVKKIHVENC	RSQDSPLSAASAFVAPSGVTEGVAST		1468	
42	Opposum					
43	Platypus					
44	Chicken					
45	Zebrafish					
46	Drosophila				LQTMLLSEL	1133
47						
48						
49	Orangutan	WTSVSPSVAAPSVCQQDVCHNGGTCHPIFLSRGIVSFQCDCPLHFTGRFCE	KDASLFFPS		2143	
50	Human	WTSVSPSVAAPSVCQQDVCHNGGTCHAIFLSSGIVSFQCDCPLHFTGRFCE	KDAGLFFPS	2149		
51	Marmoset	WTSITPSAAVPSVCQEDLCHNGGTCHPIFPSSGVVSFQCDCPLYFTGRFCE	KDAGLFFPS		2120	
52	Horse	WTSLSAPPAAAPSVCQGA	VCHNGGTCHPVFLSSGAFSFQCDCPLHFTGRFCE	-DAGLCFPS	1505	
53	Dog	WTSLSAPPAAAPSVCQGA	VCHNGGTCHPVFLSSGAFSFQCDCPLHFTGRFCE	-DAGLCFPS	1527	
54	Opposum				711	
55	Platypus				352	
56	Chicken				412	
57	Zebrafish				246	
58						
59						
60						

Mutation spectrum of EYS in Spanish patients with autosomal recessive retinitis pigmentosa 25

1	Drosophila	ETPVNTGHEITIRAEELDFSRNYTHCNASLLVNDTLAMSGDQPTWLKLLPPR-----	1184
2		. . : .	
3	Orangutan	FNGNSYLELPFLKFVLEKEHNRTVTIYLTIKTNSLNGTILYSNGNNFGKQFLHLFLVEGR	2203
4	Human	FNGNSYLELPFLKFVLEKEHNRTVTIYLTIKTNSLNGTILYSNGNNCGKQFLHLFLVEGR	2209
5	Marmoset	FNGNSYLELPFLNSVLEKEHNRTVTIYLTIKTNSLNGTVLYSNGNDFGKQFLHLFLVEGR	2180
6	Horse	FNGNSYLELPFLKSVLEKEHNRTVTIYLTIKTNTLNGTILYS---FGQQFLHLFLLEGR	1561
7	Dog	FNGNSYLELPFLKSVLEKEHNRTVTIYLTIKTNTLNGTILYS---FGQQFLHLFLLEGR	1584
8	Opposum	-----	
9	Platypus	-----	
10	Chicken	-----	
11	Zebrafish	-----	
12	Drosophila	-----	
13			
14			
15			
16			
17			
18			
19			
20	Orangutan	p.S2211L	2263
21	Human	p.I2239SfsX17	2269
22	Marmoset	PSVKYGCNSQNILTVA NSINTNAFTPI TIRYTT PGVSPGV VVCMIEMTADG KPPVQKK	2240
23	Horse	PSVKYGCNSQNILTVA NSINTNAFTPI TIRYTT PGVSPGV VVCMIEMTADG KSPAQKK	1594
24	Dog	PSVKYGCNSQNILTVA NSINTNAFIP IPI TIRY TMPVGSPGV ICMIE MTADG KSPAQKK	1644
25	Opposum	-----	
26	Platypus	-----	
27	Chicken	-----	
28	Zebrafish	-----	
29	Drosophila	-----	
30			
31	Orangutan	DTEISQASQAYFESMFLGHIPANVQ-----IHKKSGP VYGFRC GICLDLQ VNNEEFF IIIDEA	2319
32	Human	DTEISHASQAYFESMFLGHIPANVQ-----IHKKAGP VYGFRC GICLDLQ VNNEEFF IIIDEA	2325
33	Marmoset	DTKISHASQAYFESMFLGHIPANVQ-----IHKKAGP VYGF KGCICLDLQ VNNEEFF IIIDEA	2296
34	Horse	-----QAYFESMFLGHIPANVQ-----IHKKAGP VYGF KGCICLDLQ VNNEEFF IIIDEA	1642
35	Dog	DTETPHASQAYFESMFLGHIPANVQ-----IHKKAGP VYGF KGCICLDLQ VNNEEFF IIIDEA	1700
36	Opposum	-----KFGSIFLGHIPAKVK-----VHENMG QTYGY RGCICREF QVNNKEL FFIIDEA	757
37	Platypus	-----IRSQANLGS MFLGNIPAN-----GFRGC CIREMQV NNKELFF IIDEA	392
38	Chicken	-----QITFGSTFLGNVPV HKE-----VPECAGQ IRGY KGCIRDF QVNNKEL FFIIDDA	460
39	Zebrafish	-----EVVFGPTFLGGFPSV LE-----LHHNSGN VSGF IGCICRELQ MGSKELYVV GEA	294
40	Drosophila	---LHTPEAILNTWLH LGGAPQAPIGLI IELPPAQSGSGFT GCLHTLR INGQAREIFG DA	1241
41		.* * * * : * : * : : : : : : : * : * : : : : : : : : : :	
42			
43	Orangutan	RHGKNIENC HVPWCAHHLCRNN NGTCIS-----DNE NLFCECPRLYS	2360
44	Human	RHGKNIENC HVPWCAHHLCRNN NGTCIS-----DNE NLFCECPRLYS	2366
45	Marmoset	RRGKNIENC HVPWCAHHPC CHNN NGTCIS-----DSE NLFC CECPRLYS	2337
46	Horse	LRGRNIENC HVPWCAHHLC CHNN NGTCIS-----DSE NWFC CECPRLSS	1683
47	Dog	LRGRNIENC HVPWCAHHLC CHNN NGTCIS-----DSE NWFC CECPRLSS	1741
48	Opposum	LSGKNIENC NVPVC NYHPC RN GGTCIS-----DTEN WLCE CLQLYS	798
49	Platypus	LSGRNIENC NIPVCDY HPC RN GGTCIS-----NTEN WFCE CPGLYS	433
50	Chicken	LGGRNV ENCN VPICDY HPC RN GGTC TRS-----DAE NWFC CECP KLYS	502
51	Zebrafish	IRGQNIQNC DAAVCQH QP CR NGGT CISLN PPV PLPN LCFL RK LHQ QSD AES WF CAC PSL YS	354
52	Drosophila	LDGFGI TECG SLAC LSS PCR NGAAC IKI ETN-----DL DENGE KA E WK CK CPT GYM	1293
53		* . : : * * : * : * : * : : * . * . * *	
54			
55	Orangutan	GKLCQFASC ENN PC GNG ATC VP KSG TDIV CLCP YGR SGPL CTD-----AIN ITQPR FSGT	2415
56	Human	GKLCQFASC ENN PC GNG ATC VP KSG TDIV CLCP YGR SGPL CTD-----AIN ITQPR FSGT	2421
57	Marmoset	GKLCQFASC ENN PC GNG ATC VP KSG TDIV CLCP YGR SGPL CTD-----AIN IIQPR FSGT	2392
58			
59			
60			

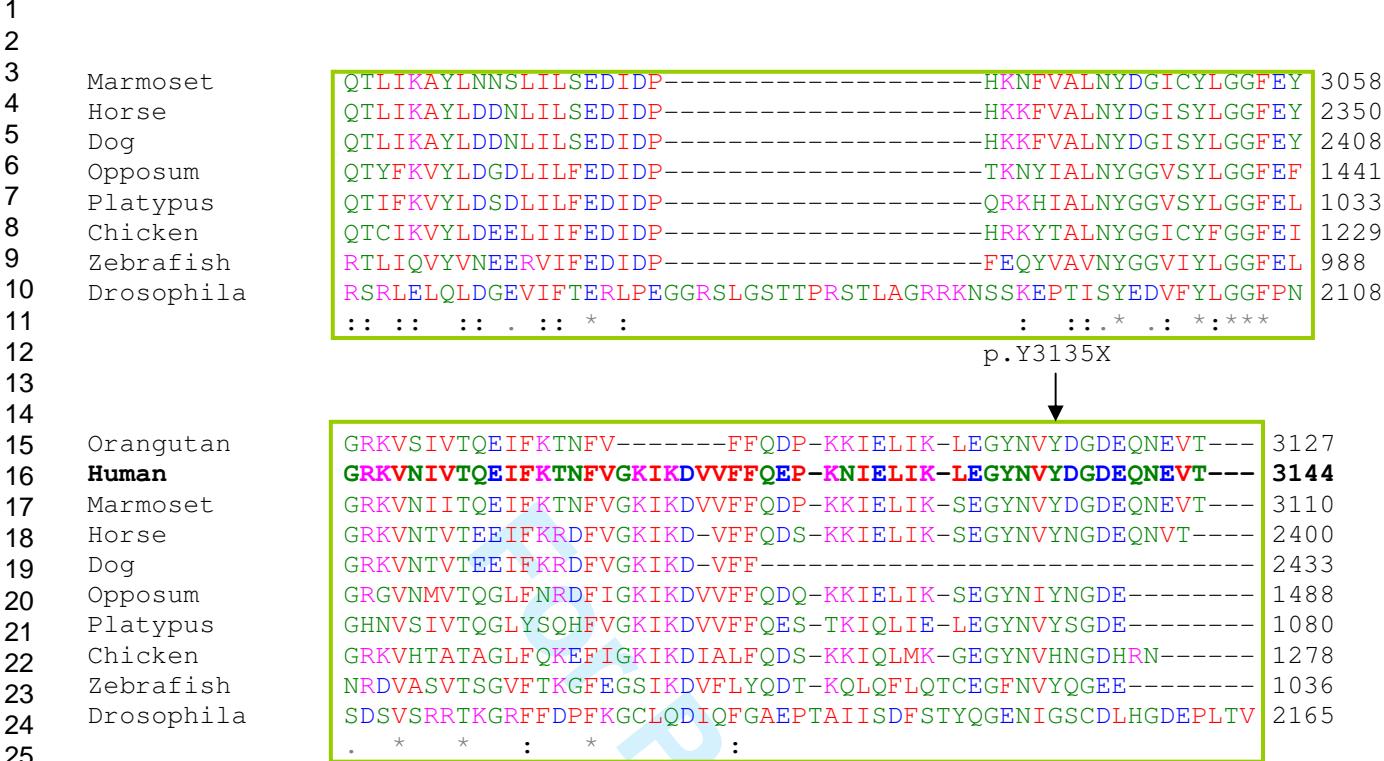
26 <Barragán et al.>

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2							
3	Horse	GKLCQFATCENNPGNGATCVPKSGTEIVCLCPYGRSGVLCTD		AINITQPSFSGT	1738		
4	Dog	GKLCQFATCENNPGNGATCVPKSGTEIVCLCPYGRSGVLCTD		AINITQPSFSGT	1796		
5	Opposum	GKLCQFATCENNPGNGATCVPKSNRDAVCLCPYGRSGVLCTD		AINITKPSFSGT	853		
6	Platypus	GKLCQFSSCEKNPGCGYATCFPKSNQDAVCLCPYGRGILCND		AITITRPSFSST	488		
7	Chicken	GRLCQFMTCDESPNPGNGATCFPKSRQDVVCLCPYGRSGILCND		VVNISQPSFSGT	557		
8	Zebrafish	GKLCQFTACERNPCARGATCVPQTQLEAACLPYGRQGLLDEGRSRQAINITRPKFSGL		414			
9	Drosophila	GPTCEISVCEDEDNPQCYGGTCVQFPGSGYLCLCPLGHGHYCEHN		LEVALPSFSG-	1347		
10		*	*	:	*	:	*
11		.	*	.	*	.	*
12		*****	*	:	*	*	*
13		p.H2454PfsX8					
14	Orangutan	DAFGYTSFLAYSRISDISFHYEFLHKFQLANNHSALQNNLIFTTGQKGHLNGDDFLAVG			2475		
15	Human	DAFGYTSFLAYSRISDISFHYEFLHKFQLANNHSALQNNLIFTTGQKGHLNGDDFLAVG			2481		
16	Marmoset	DAFGYTSFLAYSRISDISFHYEFLHKFQLANNHSALQNNLIFTTGQKGHLNGDDFLAVG			2452		
17	Horse	DAFGYTSFLAYSRVPDIFGFDYEFHVTFLQANNHSALQNNLIFTTGQKGHLNGDDFLAVG			1798		
18	Dog	DAFGYTSFLAYSRVPDIFGFDYEFHVTFLQANNHSALQNNLIFTTGQKGHLNGDDFLAVG			1856		
19	Opposum	DSFGFTSFLAYSRIPDISFYFEFLHKFQLANNNSALQDNLIIFTTGQKGQGLNGDDFLAVG			913		
20	Platypus	DAFGYTSFLAYSRIPDISSYYEFLRKFQLAANISALQNNLIFTTGQKGHRNGDDFLAVG			538		
21	Chicken	DVFGYTSFLAYSTIPDITFYEFHLKFQLNNHHSALQDNLIIFTTGQKGQGLNGDDFLVLG			617		
22	Zebrafish	DEFGYSSYVAYPSIPSTGHFYEFHLKLTFANNASALRNNNLFSGQKGQGLSGDDFFALG			474		
23	Drosophila	SVNLSSVAYTVP--IPLEYSLELSFKILP-QTMSQISLLAFFGQSGYHDEKSDHLAVS			1404		
24		*	:	:	*	*	*
25		:	:	*
26		p.E2503K					
27	Orangutan	LLNGSVVSYNLGSGIASIRSEP-----LNLSLGVHTVHLGKFFQ---LKVDHHKNKSII			2527		
28	Human	LLNGSVVSYNLGSGIASIRSEP-----LNLSLGVHTVHLGKFFQEGWLKVDDHKNKSII			2536		
29	Marmoset	LLNGSVVSYNLGSGIASLRSEP-----LDLSLGVHTVHLGKFFQEGWLKVDDHKNKSII			2507		
30	Horse	LRDGRVVSYNLGSGIASVSSDP-----LDRSLGIHAVRLGRFLQMGLWKVDDHKNKSIV			1853		
31	Dog	LRDGRVVSYNLGSGIASVSSDP-----LDRSLGIHAVRLGRFLQMGLWKVDDHKNKSIV			1911		
32	Opposum	LRNGCLVSYNLGSGTANLHSDP-----LNLSLRVHVVLGRSFQGTGLWKVDDQKNKSIT			968		
33	Platypus	-----VDNQKNKSIT			548		
34	Chicken	LCDGRVVSYNLGSGTATIISK-----LDLTNIHVIHLGRYLQKGWLKVDDQKNKTTT			672		
35	Zebrafish	VRNRGIVHKYNLGSGLATIISDR-----LNPRINIHTVHFGRYLKTGWLKVNGQKRTG			529		
36	Drosophila	FIQGYIMLTWNLGAGPRRIFTQKPIDFRLDAPRVPYEIKVGRIGRQAWLSVDGKFNTGR			1464		
37		*:..: . :					
38							
39	Orangutan	APGRLAGLNVFSQFYVGGYSEYTPDLLPNGADFKNGFQGCIFTLQVRTEKDGHFRGLGNP			2587		
40	Human	APGRLVGLNVFSQFYVGGYSEYTPDLLPNGADFKNGFQGCIFTLQVRTEKDGHFRGLGNP			2596		
41	Marmoset	APGRLVGLNVFSQFYVGGYSEYTPDLLPNGADFKNGFQGCIFTLQVRTEKDGHFRGLGNP			2567		
42	Horse	APGRLVGLNVFSQFYVGGYSEYTPELLNGSEFKNGFQG-----			1892		
43	Dog	APGRLVGLNVFSQFYVGGYSEYTPELLNGSEFKNGFQG-----			1950		
44	Opposum	SPGKLVGLNVFSQFYVGGYSEYTPELLPNESKFQNGFQG-----			1007		
45	Platypus	SPGRLVGLNVFSQFYVGGYSEYTPELLNGSDFKNGFQG-----			587		
46	Chicken	SPGRLVGLNVFSQFYVGGYSEYTPELLPKGSRFKNGFQGCIFTDVFQVRTNMNQEFKSPGTP			732		
47	Zebrafish	SPGPLMGLNTFSQLYIGGYEEYTPELLPPGSRFQNSFQGCIFTDMLFRTRQDGKFHALGGP			589		
48	Drosophila	SPGSRSMDVLPILYLGGEIANFNTLPHDLPLHSGFQGCIFYDVLQLKAG---QVTVPLQ			1520		
49		:** : : .. : *: **: . : * : : : ***					
50							
51							
52	Orangutan	EGHPNAGRSGVGQCHASPCSLMKCGNGGTCIESGTSV-CNCTGWKGAFCETEVSTCDPEH			2646		
53	Human	EGHPNAGRSGVGQCHASPCSLMKCGNGGTCIESGTSVYCNCCTGWKGSFCTETVSTCDPEH			2656		
54	Marmoset	EGHPNAGRSGVGQCHASPCSLMKCGNGGTCIES-----GCNCTGWKGAFCETEVSTCDPEH			2623		
55	Horse	-----CDCPSGWKGAFCTEMVSTCDPEH			1915		
56	Dog	-----CDCPSGWKGAFCTEMVSTCDPEH			1973		
57							

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Opposum	CIFDVQVRTRKDHFRSLGDPEG	1030
Platypus	CDCVLGKGMFCTETVSICDQEHH	610
Chicken	EGHPNSGRSVGQC KASPCSLIKCRNGGKCMESG-STVCNCLAGWKGAFCTEMVTVC DPEH	791
Zebrafish	DIRPLSGRVNVGQC GVNPCSLVFCHNGGTCVDSGSSVYCQCVFGWKGALCSEKVSFCDAE H	649
Drosophila	ETRGVGRGVGQC GTRCHRACQHDGACLQHGATFTCICQEGWYGPLCAQPTN PCDSFN	1580
Orangutan	DPPHHCSRGATCISLPHGYTCPLGTTGIYCE QALSISDPSFRSNELSWMS	2698
Human	DPPHHCSRGATCISLPHGYTCPLGTTGIYCEQALSISDPSFRSNELSWMS	2708
Marmoset	DPPHNCSRGATCISLPHGYTCPLGTTGIYCE QALSISDASFRSNELSWMS	2674
Horse	DPPHNCSKGATCVPLPHGYTCRCPLGTTGIYCE QALSISDASFRSHELSWMS	1966
Dog	DPPHNCSKGATCVPLPHGYTCRCPLGTTGIYCE QALSISDPSFRSHELSWMS	2024
Opposum	HPNAGRSGV-QCEV FPCSLIKCQNGGTC I QALSISDPSFRSHDSSWMS	1076
Platypus	DPPHQCRPGATCVPLTIGY TCPLGTTGIHCE QALSISDPSFRRNESSWMS	661
Chicken	DPPHLCKQGGTC CVPLPNGYMCHCPLGTSGTYCEQDIS ISDPSFRSNKSSWMS	843
Zebrafish	IPPPFCARGSTCVPLSDGYTCQC PLGSAGLHCQQAITISDPFFSGNQSSWMS	701
Drosophila	N---KCYEDATCVPLVNGYE CDCPVGRTGKNCEE VIRSLSDVSLTGRRSY LAVRW LYD	1637
Orangutan	-F	2699
Human	-F	2709
Marmoset	-F	2675
Horse	-F	1967
Dog	-F	2025
Opposum	-F	1077
Platypus	-F	662
Chicken	-F	844
Zebrafish	-F	702
Drosophila	GGDKLGA KRSQMVSYRNFTKKLMPPKITT PSHFM KLLNEVEK Q RSFSPVPLMGS KSF	1697
Orangutan	ASFHV RKKTHIQLQFQPLAADGILFYAAQHLKAQSG --DFLCISLVNSSVQLRYNLGDRT	2757
Human	ASFHVRKKTHIQLQFQPLAADGILFYAAQHLKAQSG--DFLCISLVNSSVQLRYNLGDRT	2767
Marmoset	ASF RVKRKKTHIQLQFQPLAADGILFYAAQHLKAQSG --DFLCISLANGSVQLRYNLGDRT	2733
Horse	SSF RIRKRTHIQLQFRPLSADGILFYVAQNLKAQSG --DFLCISLVNGSVQLRYNLGDRT	2025
Dog	SSF RIRKRTHIQLQFRPLSADGILFYVAQNLKAQSG --DFLCISLVNGSVQLRYNLGDRT	2083
Opposum	APF YIRQKMHIQLQFQALSTDGILFYTAQHLHSRSG --DFLCISLTRGSVQLRYNLGDRT	1135
Platypus	APF HIRHKTHIQLQFQPLSADGILFYTAQHLSTHSG --DFLCISL LANGYVQLRYNLGDRT	720
Chicken	APF YIRHKTHIQLQFQPLSDPGDILFYTAQRLGTQSG --DFLCISLVNGFIQLRYNLGDKT	902
Zebrafish	PPINIR RTHVQLQFQTLSPEGILFYTAQHLSTHSG --DFLSISLSAGFLQLRYNLGNQT	760
Drosophila	EEHHRVQFFFIEFQLRPLSERGLLLYFGTLNNNQDKKIGFV SLSQGGVVEFRI SGPSNH	1757
Orangutan	IILETLQKVTINGSTWHIIKAGR VGAEGYL DLDG INVTEKAST KMSSL DTNTDFY IGGV S	2817
Human	IILETLQKVTINGSTWHIIKAGRVGAEGYLDLDGINVTEKASTKMSSLDTNTDFYIGGVS	2827
Marmoset	IILETLQKVTTNGSTWHVIKAGR VGAEGYL DLDG INVTEKAST KMSSL DTNTDFY IGGV S	2793
Horse	IILETLQKVNMNGSTWHVIKAGR VGAEGYL DLDG KTVTEKAKAEMNSL DTNTDFYIGGV S	2085
Dog	IILETLQKVNMNGSTWHVIKAGR VGAEGYL DLDG KTVTEKAKAEMNSL DTNTDFYIGGV S	2143
Opposum	VILQSLQK---WHTIKAGR VGNEGYLELDG INVTKGATAGM RALDT STDFYIGGV S	1188
Platypus	VILQSFQKVDTSGDA WL IKAGR HGT EGYL TLDGT NTQKATGRMTVLD NTDFY VGGV S	780
Chicken	IVLQAVQKVADGQTWHV LKVGRVGNEG YV DLDG IN ITHTAS AGMNVL DTH DFYVGGV S	962
Zebrafish	IVLQSPKE LDVTGVRWHTV KAGR EGNSGFL IVDG ESVTRNSSEG STTLDVG ANIF IGGIS	820
Drosophila	VTVVRSVRMLAIG-EWH KIKMAQRGRWLT LWVE EGSASSALAPSAEV LPD SL LYIGGLK	1816

1			
2			
3			
4	Orangutan	SLNLVNPMAIENEPEVGFQGCIRQVIINNQELQLTEFGAKGGSNVGDCDGTACGYNTRNG	2877
5	Human	SLNLVNPMAIENEPEVGFQGCIRQVIINNQELQLTEFGAKGGSNVGDCDGTACGYNTRNG	2887
6	Marmoset	SLNLINPMAIENEPEVGFQGCIREVIIINNQELQLTESGAKGGSNVGDCDGTACGYNICRNG	2853
7	Horse	SLNLVNPMAIANEPEVGFQGCIREVIIINNQELQLTELGAKGGSNVGDCDGTACGYNCRNR	2145
8	Dog	SLNLVNPMAIANEPEVGFQGCIREVIIINNQELQLTELGAKGGSNVGDCDGTACGYNCRNR	2203
9	Opposum	SLSLVNPMAIENEPEVGFNGCVREILINGRELKLTEAGAKRGSNVGDCDGTPCGYKVCENK	1248
10	Platypus	SLDSVNSMAVENDPVGFDFGCVREIFINNRELKLTDKGAKDGLIGDCDGTACGYTVCKNK	840
11	Chicken	SLNLVNSMATENEPTGFSGCIREIVINDKELKLTVTDPKGGANIGDCDGTVCGVSKNN	1022
12	Zebrafish	SLNTVSIDAVEKELVGFTGGIREVVVNGQELELTETGALDGANVGDWDTACGYKVCKNG	880
13	Drosophila	DVKSLPHNAISGFPIFRGCVRGLVSGTRIVLNETNIVESRNIRDGCDGTACGGDSCESG	1876
14		... : * * * : * : . . . : * . . . * : * *** * * * .	
15			p.G2945E
16			↓
17	Orangutan	GECTVNGTTF-SCRCLPDWAGNTCNQS VYCLNNLCLHQSLCIPN-QSFSYSCLCTLG WVG	2935
18	Human	GECTVNGTTF-SCRCLPDWAGNTCNQS VCLNNLCLHQSLCIPD-QSFSYSCLCTLG WVG	2945
19	Marmoset	GKCTVNGTTF-SCRCLPDWAGNTCNQSAYCLNNLCLHQSLCIPD-QSFSYSCLCTLG WVG	2911
20	Horse	GECVVNGTTF-SCQCSPPWAGNTCEQSAYCLNNLCLHQSLCVPD-QSSSYRCLCTLG WEG	2203
21	Dog	GECVVNGTTF-SCQCSPPWAGNTCEQSAYCLNNLCLHQSLCVPD-QSSSYRCLCTLG WEG	2261
22	Opposum	GQCRAQGSKF-SCKCLQPWIGKRCEESANCRRNNLCLHHSRCIPV-QPAAYICLCPLG WVG	1306
23	Platypus	GECILHHTNF-SCKCTPGWAGNTCEQS MNCLNNKCQHQSLCIPD-NTFSYSCACPLG WVG	898
24	Chicken	GTCQVESSGF-SCSCPQGWIGNTCEESVHCLHNRCRSQALCIPQALLSYTCVCPLG WSG	1081
25	Zebrafish	GHCHPSAG-	888
26	Drosophila	GHCWLDEKLQPHCICPEYAKGDRCEYSETCKLIPCKNNGRCLRS---GRCSCPNG WGG	1931
27		* *	
28			
29	Orangutan	RYCENKTSF-STAKFMGN SYIKYIDPNYRMRNLQ-----FTTISLN FSTTKTEG	2983
30	Human	RYCENKTSF-STAKFMGN SYIKYIDPNYRMRNLQ-----FTTISLN FSTTKTEG	2993
31	Marmoset	RYCENKTSF-STAKFMGN SYIKYIDPNYRMRNLH-----FTTISLN FSTTKTEG	2959
32	Horse	RYCENKISF-STAKFMGN SYIKYIDPNYRMRNHH-----FTTVSLNFSTTETEG	2251
33	Dog	RYCENKISF-STAKFMGN SYIKYIDPNYRMRNHH-----FTTVSLNFSTTETEG	2309
34	Opposum	RYCDNETSF-ITAKFVGNSYIKYIDPNYEKRDLR-----FTAVSLNFSTTETEG	1354
35	Platypus	GYCETEILF-LIAFKQGNSYIKHTDPNYGKRNLH-----FTTVSLNFSTTETEG	946
36	Chicken	KHCDSKISF-FTAKFVGNSYIKYIDPLYGKRDLQ-----YSRISLNFTTNQIEG	1129
37	Zebrafish	-----DG-----	890
38	Drosophila	FYCEIAMSKPTTPSFRGNSYLILPPPRIPMKDKRRGP SLYVRPREAIQVSLNFSTIEPDG	1991
39		: *	
40			
41			
42	Orangutan	LIVWMGTAQNEENDFLAIGLHNQTLKIAVN LGERISVPMSYNNGT FCCNK-WHHVVVIQN	3042
43	Human	LIVWMGIAQNEENDFLAIGLHNQTLKIAVN LGERISVPMSYNNGT FCCNK-WHHVVVIQN	3052
44	Marmoset	LIVWMGIAQNEENDFLAIGLHNQTLKIAVN LGERISVPMSYNNGT FCCNK-WHHVVVIQN	3018
45	Horse	LIVWIGKAQNEENDFLAIGLHNQSLKIAVN LGESISVPVIYNSNGT FCCNK-WHHVVIVSQN	2310
46	Dog	LIVWIGKAQNEENDFLAIGLHNQSLKIAVN LGESISVPVIYNSNGT FCCNK-WHHVVIVSQN	2368
47	Opposum	LILWMGKAEHEENDFLAIGVHNRTLKVMVN LGERISVP-----WHHVKVVQN	1401
48	Platypus	LILWMGKAEHEENDFLAIGLNSGILKVVVNLGNGLSVP-----WHYITVAQN	993
49	Chicken	LMVWLGKAEDEDNDFLAIGLANGRLKVVINLGERISVPMIHSKDSICTDERWHFVTVIQN	1189
50	Zebrafish	LIFWMGKAESEDDDHlavglQDGylkisvnlgERTALPLVYQN-SFCCNY-WNYLSITHN	948
51	Drosophila	LLLWS---EHERSKFLGLGLEAGHLKLASNLGSTNDTVRAPASGFIADGAWHWTISVLLD	2048
52		* : . * : * . . * : * : ** : * . * : : :	
53		p.N3061TfsX3	
54		↓	
55	Orangutan	QTLIKAYVNNSLILSEDIDP-----HKNFVALNYDGICYLGGFEY	3082
56	Human	QTLIKAYINNSLILSEDIDP-----HKNFVALNYDGICYLGGFEY	3092
57			
58			
59			
60			

Mutation spectrum of *EYS* in Spanish patients with autosomal recessive retinitis pigmentosa 29

Supplementary Figure S1. *EYS* homologues protein alignment and domain distribution with identified variations.
5' UTR and splice site variations are not included in this figure.

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