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Doxorubicin Loaded Magnetic Polymersomes: Theranostic Nanocarriers for MR Imaging and Magneto-Chemotherapy

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ABSTRACT: Hydrophobically modified maghemite (γ-Fe₂O₃) nanoparticles were encapsulated within the membrane of poly(trimethylene carbonate)-b-poly(L-glutamic acid) (PTMC-b-PGA) block copolymer vesicles using a nanoprecipitation process. This formation method gives a simple access to highly magnetic nanoparticles (MNPs) (loaded up to 70 wt %) together with a good control over the vesicles size (100 to 400 nm). The simultaneous loading of maghemite nanoparticles and doxorubicin was also achieved by nanoprecipitation. The deformation of the vesicle membrane under an applied magnetic field has been evidenced by small angle neutron scattering. These superparamagnetic hybrid self-assemblies display enhanced contrast properties that open potential applications for Magnetic Resonance Imaging. They can also be guided in a magnetic field gradient. The feasibility of controlled drug release by radio-frequency magnetic hyperthermia was demonstrated in the case of encapsulated doxorubicin molecules, showing the viability of the concept of magneto-chemotherapy. These magnetic polymersomes can be used as efficient multifunctional nano-carriers for combined therapy and imaging.

KEYWORDS: block copolymer vesicles, polymersome, nanoprecipitation, superparamagnetic iron oxide nanoparticles, maghemite, magnetic hyperthermia, magneto-chemotherapy, multifunctional, MRI contrast agent, doxorubicin, theranostics

Over the past decades, nanopolymeric therapeutics has proven to improve the effectiveness of cancer treatments in animal experiments.1-4 During this period, progresses in modern polymer (physico)-chemistry have enabled the design of polymeric carriers with ever higher levels of complexity featuring addressable chemically reactive functions, defined chain architectures, and controlled morphologies and sizes. When applied to the field of drug delivery, these features allow achieving and combining several desirable properties such as high drug loading content, controlled release, increased circulation half-life and targeting of pathological areas or specific cell receptors. Polymer nanomedicines have the potential to increase the shelf life of chemotherapeutics before administration and to improve their efficacy after administration.5-7 A direct consequence of the latter is the reduction in the dosing concentration and frequency of administration of the drug, hence the minimization of toxic side effects on healthy tissues, which are currently a major problem in chemotherapy. Among the different classes of polymeric nanomedicines, block copolymer vesicles also termed polymersomes offer an attractive structure for drug delivery applications.8-12 This block copolymer self-assembly in a closed bilayer has fostered a considerable attention since both hydrophilic and hydrophobic drugs can be loaded either in the aqueous lumen or in the membrane core respectively,13, 14 thanks to a thick membrane that imparts long-term stability to the object. After drug loading, vesicle disruption inducing drug release can be either triggered by an environmental stimulus, such as pH, temperature, light, or oxidation,15-17 or can be the consequence of polymer hydrolytic or enzymatic degradation.18, 19 Even though a wide variety of polymer nanocarriers for drug delivery has shown efficient entrapment and controlled release of drugs in vitro, the evaluation of their biodistribution in vivo has become possible by non invasive methods. To address this issue, one strategy consists in incorporating imaging probes together with the drugs into the polymer nanoparticles. These dual polymer nanocarriers for simultaneous cancer imaging and treatment open the field to “theranostic nanomedicines”, combining diagnostic and
therapeutic components in an all-in-one nanoparticle, \(^{20, 21}\) Imaging probes to be loaded can belong to different families such as visible\(^{22, 23}\) and NIR fluorochromes, \(^{24}\) radiotracers\(^{-27}\) or inorganic nanoparticles such as quantum dots,\(^{28-30}\) gold nanoparticles,\(^{31, 32}\) or magnetic nanoparticles (MNPs).\(^{33, 36}\) Among the different MNPs, the so-called “ultra-small superparamagnetic iron oxide” (USPIO) particles are synthetic \(\gamma\)-Fe\(_2\)O\(_3\) or \(\text{Fe}_3\text{O}_4\) nanometric grains in a perfectly dispersed state (i.e. unclustered individual nanoparticles).\(^{37}\) As described in numerous review articles\(^{38-43}\), iron oxide MNPs also called USPIOs are commonly used as negative contrast-enhancing agents in MRI, enabling high spatial resolution acquisition, three-dimensional, non-invasive imaging of the human body. Hydrophilic “stealth” polymers are often employed to enhance the stability and biocompatibility of the MNPs in vivo by hindering their further aggregation and by an anti-fouling effect against proteins of the immune system called opsonins.\(^{42, 44}\) Besides the MRI contrast enhancement properties attributed to their ability to distort strongly the magnetic field lines,\(^{45}\) USPIOs can be used to kill cancer cells by their heating effect in radio-frequency magnetic fields. Hyperthermia (or thermal ablation) is identified as a promising approach in cancer therapy, particularly in combination with chemo- or radiotherapy.\(^{47}\) A promising hyperthermia route for treating deep tumors consists in concentrating MNPs around and inside the tumor site and increasing the temperature locally through conversion into heat of the energy from an external alternating magnetic field in the range of radio frequencies 100 kHz – 1 MHz. This magnetic hyperthermia led to an intense research activity both on the optimization of the conditions of treatment (power, concentration, geometrical parameters...)\(^{46-48}\) and on the characteristics of the USPIOs themselves (chemical nature, distribution of sizes...).\(^{39, 49-57}\)

For this purpose of obtaining multifunctional drug vectors, hydrophilic USPIOs have been loaded at first in the aqueous compartment of liposomes.\(^{58, 59}\) Under a permanent magnetic field, magnetic liposomes deform into elongated ellipsoids, as it was evidenced for giant unilamellar vesicles.\(^{60-62}\) Interesting studies dealt with much smaller magneeto-liposomes analogous in sizes to the pegylated lipid vectors of the DOXIL\(^{74}\) formulation of the anticancer drug doxorubicin hydrochloride (DOX). To combine magnetism and thermo-sensitivity, DOX was encapsulated into magnetic vesicles with a lipid membrane initially in the gel state and becoming fluid at a temperature reachable by magnetic hyperthermia.\(^{63, 64}\) The application of a RF magnetic field led to massive release of encapsulated DOX since the magnetic hyperthermia was sufficient to reach locally the main chain phase transition temperature of the bilayer, thereby increasing the membrane permeability.\(^{53}\) Recently, an analogous study with small hydrophobic USPIOs embedded in the membrane of liposomes evidenced the release triggered by a RF magnetic field of a fluorescent dye used as a model of hydrophobic drug.\(^{65}\) The possibility to target a solid tumor by using magneeto-liposomes and an extracorporeal magnetic field to guide them has also been reported.\(^{64, 65}\) Despite the tremendous results obtained with liposomes, the morphology of lipid/MNP systems strongly varies with MNP and lipid concentrations.\(^{66}\) They also suffer from the classical issue of instability associated with lipid bilayers,\(^{67}\) which incite to use of polymersomes as an alternative to liposomes. We have shown for the first time that hydrophobic USPIOs can be loaded into polymersome membranes of PB-b-PGA and that a reversible variation of the membrane thickness can be induced by the application of a magnetic field.\(^{71, 72}\) Later, Förster et al. induced the bridging of adjacent bilayers and formed multi-lamellar hybrid polymersomes by incorporating hydrophobic USPIOs into PI-b-PEO bilayers at a feed weight ratio up to 20 % sufficiently large to guide the vesicles by a magnetic field gradient.\(^{73}\)

In the present article, we describe a convenient procedure to prepare well-defined magnetic polymersomes featuring a hydrophobic internal membrane core made of the biodegradable block poly(trimethylene carbonate) (PTMC) and a polypeptide biocompatible corona of poly(L-glutamic acid) (PGA). Having synthesized USPIOs with the appropriate characteristics (size and hydrophobic coating), those were embedded together with the efficient antitumor drug doxorubicin hydrochloride (DOX) into the membrane of dual-loaded vesicles by one-step nanoprecipitation. This process allowed reaching quantitative loading contents and controlling the final sizes with low polydispersity. The two-dimensional confinement of USPIOs inside the vesicular membrane was evidenced by small angle neutron and light scattering techniques and observed by atomic force and transmission electron microscopy. The magnetic membrane of the PTMC-b-PGA polymersomes was shown to be reversibly deformable under a permanent magnetic field. The release of DOX under local hyperthermia conditions induced by an oscillating RF magnetic field was also evidenced as a proof of concept of magneto-chemotherapy with magnetic polymersomes.

**RESULTS AND DISCUSSION**

**Characteristics of the USPIOs**

After synthesis\(^{75}\), size fractionation\(^{76}\) and surfactant coating\(^{77}\), the radius of gyration and the hydrodynamic radius of the USPIO nanoparticles used in this work were \(R_g^{\text{USPIO}}=3.05_{-0.06}^{+0.07}\) nm and \(R_h^{\text{USPIO}}=4.7_{-0.07}^{+0.07}\) nm as measured respectively by SANS and DLS. The ratio \(R_h^{\text{USPIO}}/R_g^{\text{USPIO}}=0.65\) is not far from the theoretical value 0.775 for dense spherical particles,\(^{78, 79}\) the gap being reasonably ascribed to the contribution of the surfactant layer to \(R_h^{\text{USPIO}}\) only. Those sizes are in good agreement with the distribution measured by VSM (Supporting Information, \(S_d\))\(^{80}\) described by a Log-normal law of median diameter \(D_m^{\text{USPIO}}=6.5\) nm, width \(\sigma=0.22\) and weight averaged diameter \(D_w=7.5\) nm. Concerning the magnetic hyperthermia capability, a specific loss power (SLP) of 14 W/g was reported for USPIOs synthesized by the same route and of analogous distribution of diameters \((D_m^{\text{USPIO}}=6.7\) nm, \(\sigma=0.20, D_w=7.7\) nm) but at larger frequency \((f=700\) kHz) and much higher field intensity \(H=24.8\) kA/m.\(^{81}\) Using its expected variation with these parameters \((\gamma\text{-Fe}_2\text{O}_3)\), we estimate a SLP value of 0.07 W/g in the conditions of biocompatible RF field used in this work \((f=500\) kHz and \(H=2.12\) kA/m).\(^{82}\)

**Characteristics, structure and stability of USPIO-loaded vesicles**

In a previous study, the conditions of nanoprecipitation with the PTMC-b-PGA block copolymer were varied: choice of THF or DMSO as good solvent of the blocks, (order in water or reverse) and duration of the addition.\(^{83}\) The influence of each experimental parameter was rationalized in order to finely tune the sizes and PDI of the vesicles. In addition, the PTMC block was shown to be semi-crystalline with an apparent melting temperature in vesicles near 34°C (lower that value at 37°C in the bulk) that has a strong influence on the size of the vesicles and on their interactions.\(^{84}\)

In the present study, we checked that incorporating inorganic nanoparticles at the nanoprecipitation step did not affect the self-assembly process of the diblock copolymer and that vesicles morphology were well retained. The conditions were selected according to the low PDI obtained, the compatibility of the organic solvent and of the obtained vesicles’ sizes with \(\text{in vivo}\) applications. The copolymer was first dissolved in DMSO with or without \(\gamma\)-Fe\(_2\)O\(_3\) USPIOs. Then water was added (up to 90 % of the final volume) at a controlled flow rate to trigger self-assembly. As the flow rate strongly influences the final size of the vesicles,\(^{32}\) we
considered two sets of conditions: an almost instantaneous addition (5 seconds) leading to small vesicles ($R_{H}=45-67$ nm) denoted WDi and a 15 minutes-addition leading to larger ones ($R_{H}=187-202$ nm) denoted WD15. The characteristics of nanoparticles’ dispersions prepared by either one or the other of these conditions at increasing USPIO feed weight ratios (FWR) are shown in Table 1.

Table 1. Size and polydispersity index (PDI) of nanoparticles’ dispersions prepared with increasing feed weight ratios of USPIO relatively to copolymer. Vesicles were prepared by nanoprecipitation in DMSO by adding water either in 5 s (WDi) or in 15 min (WD15). The image is a macroscopic view of samples.

<table>
<thead>
<tr>
<th>Sample code</th>
<th>FWR (%)</th>
<th>$R_{H}$ (nm)</th>
<th>PDI</th>
</tr>
</thead>
<tbody>
<tr>
<td>WDi-0</td>
<td>0</td>
<td>67</td>
<td>0.07</td>
</tr>
<tr>
<td>WDi-20</td>
<td>20</td>
<td>50</td>
<td>0.14</td>
</tr>
<tr>
<td>WDi-35</td>
<td>35</td>
<td>45</td>
<td>0.16</td>
</tr>
<tr>
<td>WDi-50</td>
<td>50</td>
<td>47</td>
<td>0.16</td>
</tr>
<tr>
<td>WDi-70</td>
<td>70</td>
<td>52</td>
<td>0.18</td>
</tr>
<tr>
<td>WD15-0</td>
<td>0</td>
<td>202</td>
<td>0.05</td>
</tr>
<tr>
<td>WD15-20</td>
<td>20</td>
<td>196</td>
<td>0.09</td>
</tr>
<tr>
<td>WD15-35</td>
<td>35</td>
<td>195</td>
<td>0.20</td>
</tr>
<tr>
<td>WD15-50</td>
<td>50</td>
<td>187</td>
<td>0.22</td>
</tr>
</tbody>
</table>

The vesicles were found homogeneous in sizes, as observed by small PDI values in DLS. The loading of vesicles by USPIOs progressively increases the PDI (while remaining low) and slightly decreases $R_{H}$. This hydrodynamic size decrease (more pronounced for WDi than for WD15) and slight broadening of the sizes’ distribution (for both nanoprecipitation speeds) are ascribed to a larger hydrophobic effect when the copolymer is combined with USPIOs coated by surfactants, and thus to a larger driving force for a faster self-assembling process. No aggregation occurred below a critical USPIO FWR. Beyond this threshold value, the hydrophobic USPIOs began to aggregate during nanoprecipitation forming ill-defined macroscopic clusters that rapidly migrated to the vial walls when approaching a magnet. This maximum FWR was respectively 50 wt % for WD15 and 70 wt % for WDi. The larger threshold FWR with WDi vesicles compared to WD15 ones can be ascribed to a much faster kinetics of formation, thereby minimizing the probability of USPIOs’ clustering before the completion of co-assembly with the copolymer. The maximum loading content of USPIOs in the membrane of the WD15-70 vesicles corresponds to a local volume fraction $\Phi_{USPIO}=12.1\%$. Interestingly, this is close to the reported value of 11% for the insertion of USPIOs in bilayers of polystyrene-\text{-}b\text{-}polyacrylate, whereas larger volume fractions e.g. 21% lead to a morphological transition into micelles via nanoparticles’ clustering. Fully dispersed and stable suspensions were observed below and up to these maximum USPIO FWR values. In these conditions, the shelf life is longer than several months at room temperature. Static light scattering (SLS) measurements conducted on the WDi-50 sample strongly suggested a vesicular morphology. By drawing the Berry plot over a scattering angular range from 50° to 150° and a concentration range from 0.2 to 1 mg/mL, we obtained the $z$-averaged radii of gyration ($R_{G(z)}$) allowing to calculate the ratio $p=R_{G(z)}/R_{H}$. While vesicles are characterized by $p$ values close to 1, $p$ values around 0.775 are expected for spherical micelles. Fully dispersed and stable suspensions had a $p$ value of 1.02 in good agreement with vesicular self-assemblies.

A further insight to the exact morphology of USPIO-loaded PTMC$_{24}$-\text{-}b\text{-}PGA$_{19}$ particles in either WDi or WD15 conditions was brought by small angle neutron scattering (SANS) experiments. Figure 1 represents the intra-aggregate structure factor $S_{\text{intra}}(q)$ of the USPIOs measured by SANS in a solvent mixture matching the neutron scattering length density of the copolymer. The shape of $S_{\text{intra}}(q)$ reflects both the interactions between the USPIOs inside the object and the overall shape of their aggregates in the attractive regime. In the small q-regime, $S_{\text{intra}}(q)$ followed a power law with a slope approximately -2 typical of flat samples, supporting a vesicle--type morphology. In this q-region (Kratky-Porod regime), the thickness of the USPIO layer in the vesicle membrane can be calculated from the slope of $\ln[q^{2}S_{\text{intra}}(q)]$ plotted vs. $q^{2}$ which is $-\phi^{2}/2$. From the experimental data, we obtained respectively $\phi=13$ nm and $\phi=10$ nm for samples WD15-50 and WDi-70. The vesicular membrane thus contains no more than one or two layers of magnetic colloids. More precisely, the data were properly fitted using a hollow sphere form factor with respectively an internal radius $R=130$ nm and shell thickness $\delta=12$ nm for WD15-50, and $R=45$ nm lumen radius with membrane thickness $\delta=9$ nm for WDi-70. These radii deduced from SANS fits agree pretty well with the hydrodynamic radii on Table 1 measured by DLS. At large wave-vectors, the $q^{2}$ scaling law is typical of the Porod’s regime expected for nanoparticles with a smooth interface. Moreover, $S_{\text{intra}}(q)$ presents a correlation peak around $8\times10^{2}$ Å$^{-4}$ (see Figure 1), associated to a most probable USPIO inter-particle distance $d_{\max}=2\pi q_{\max}=7.8$ nm. Considering their weight-average diameter $D_{w}=7.5$ nm, we deduce that the USPIOs are closely packed inside the vesicular membrane for both WD15-50 and WDi-70 samples. The SANS curve of WD15-50 vesicles in D$_{2}$O where the neutron scattering contrast of the USPIOs is almost matched also exhibits this correlation peak (Supporting Information S-1), as explained by the close-packed structure of holes in the copolymer membrane confining the USPIOs at a high local volume fraction.
Figure 1. SANS curves of PTMC<sub>24</sub>-b-PGA<sub>19</sub> vesicles WD15-50 (A) and WDi-70 (B), centrifuged then dispersed at 10 mg/mL in a mixture H<sub>2</sub>O:D<sub>2</sub>O (65.6/34.4 v/v). Experimental intra-aggregate structure factors \( S_{\text{intra}}(q) \) of USPIOs are plotted as open circles. The solid lines represent the simulated form factors respectively for hollow spheres of mean radius \( R = 130 \) nm (PDI=0.17) with membrane thickness \( \delta = 12 \) nm (PDI=0.3) for WD15-50 (A), \( R = 45 \) nm (PDI=0.35) with \( \delta = 9 \) nm (PDI=0.3) for WDi-70 (B).

Figure 2. TEM images of USPIO-loaded vesicles prepared by nanoprecipitation. (A) Low magnification picture of WD15-50 vesicles (scale bar 1 μm); (B) Close-up view of a WD15-50 vesicle containing ~1500 USPIOs as measured by image analysis (scale bar 300 nm); (C) WDi-70 vesicles spreading on the substrate, which enables counting ~190 USPIOs on the left and ~220 USPIOs on the right (scale bar 100 nm); (D) Image of negatively stained WDi-50 vesicles, showing a group of vesicles laying intact on the carbon substrate (scale bar 50 nm); (E) Cryo-TEM image showing homogeneously dispersed WDi-50 vesicles (scale bar 200 nm). Inset: close-up view of two vesicles showing a mantle of respectively ~80 and ~110 close-packed USPIOs with some uncovered areas (scale bar 50 nm).
To summarize our SANS results, we found membrane thicknesses equal to $12.5\pm0.5$ nm and $9.5\pm0.5$ nm (either by scaling law or by form factor fitting) for WD15-50 and WDi-70 vesicles respectively. Due to the chosen H$_2$O/D$_2$O solvent matching the copolymer scattering, these values represent the thickness of the USPIOs’ layer only. In D$_2$O solvent were the neutron scattering signal originates both from the magnetic contrast of iron oxide and the nuclear contrast of the copolymer, we measured a total membrane thickness $29.1\pm0.6$ nm from Kratky-Porod’s plots of the data reported in the ESI file (figure S-c) for WDi vesicles independently of their iron oxide content (from 0 to 50% FWR), in accordance with the value $30\pm2$ nm reported for the total membrane thickness of non magnetic vesicles made of PTMC$_{24}$-b-PGA$_{12}$ with a similar PTMC block of molar mass $M_n=2750$ g/mol. The measurement by SANS of the hydrophobic bilayer thickness for WDi vesicles well compares to the value $\delta=9.6$ nm measured by cryo-TEM for polymersomes made of poly(ethylene oxide)-b-polybutadiene (noted EO$_{26}$-BD$_{46}$ or OB2) with a total molar mass of 3600 g/mol and a hydrophilic fraction of 28%, thus a hydrophobic block mass of 2600 g/mol close to the one of PTMC here. The 25% increase of hydrophobic thickness for WD15 vesicles is ascribed to the swelling of vesicles’ membranes by the incorporation of USPIOs, which was presumably not possible for WDi ones due to their much higher curvature.

The WD15-50 and WDi-70 samples were further observed by TEM (Figure 2) and AFM (Figure 3) to confirm the vesicular morphology. TEM images mainly show the arrangement of the USPIOs because of the low electron scattering density of the copolymer compared to iron oxide. For both nanoprecipitation conditions (WD15 and WDi), hollow structures made of a close-packed arrangement of USPIOs were observed. The diameters measured on the TEM images 2B and 2C are around 750 nm and 150 nm respectively for WD15-50 and WDi-70 vesicles, which is larger than two times their hydrodynamic radii reported on Table 1 (374 nm and 104 nm respectively). This apparent discrepancy is ascribed to the total spreading of the vesicles onto the carbon substrate. The drying step during sample preparation and the strong wetting on substrates presumably induced the rupture of membranes, which explains the presence of fragments as well as not entirely closed structures. Unlike images A, B and C of Figure 2 that were obtained by spraying the samples onto the grids, image D originates from a more gentle protocol combined with staining (see Experimental) that led to vesicles sitting intact on the substrate. Both images D and E (cryoTEM) show apparent diameters much closer to the light scattering results, undoubtedly confirming the proposed structure. However we chose to show images A, B and C in spite of the spreading effect, because the flattening of the membrane onto the substrate enables to count the USPIOs per vesicle much easier than with the projection of intact spherical vesicles (D and E).

The WDi vesicles were also observed by AFM with and without the presence of 50 wt % USPIO. AFM phase images of empty vesicles (WDi-0) showed spherical vesicles, which aqeous interior leaked out due to drying and strong adsorption onto the freshly cleaved mica surface. When USPIOs were incorporated into the membrane (WDi-50), those presented multiple bright spots. The contrast of phase AFM pictures being proportional to the surface toughness, we identify those bright spots with the hard inorganic USPIOs embedded within the soft polymer matrix and spatially distributed over the vesicular surface as large patches. The average thicknesses of membranes spread on mica were analyzed on the AFM height images. These profiles revealed that the presence of USPIOs increase the thickness from 8 nm to 15 nm, the difference being very close to the weight average inorganic diameter $D_w=7.5$ nm. If the vesicles were adhering intact on the mica substrate, simply deflated by soft drying conditions, one would expect to measure an inorganic thickness equivalent to two layers of

![Figure 3](image-url). Tapping Mode™ AFM phase and height images of 1×1 μm surfaces of PTMC$_{24}$-b-PGA$_{19}$ vesicles prepared by nanoprecipitation WDi without magnetic nanoparticles (upper panel) and with 50 wt % USPIO WDi-50 (lower panel). The average heights are measured on the right by cross-sections.
WD15 samples and with respectively USPIOs in membrane and their number per vesicle. The migration of the WD15-50 vesicles under a controlled magnetic field gradient was also used to estimate differently the number of confined USPIOs averaged on a large population of vesicles. The assessment of the ability of magnetic polymersomes to be attracted and concentrated at a specific location in vivo is also particularly relevant. Compared to magnetophoresis experiments with objects of sizes around 10 µm such as giant liposomes or biological cells, a supplemental difficulty arose from the low value of the Peclet’s hydrodynamic number, which means that the magnetophoretic motion of the vesicles was not significantly larger than their Brownian motion (see the videos supplied as ESI). Usually, magnetophoretic measurements with suchcolloidal particles prone to thermal agitation are done by measuring light absorption profiles as a function of time and space. In the present work, we chose alternatively a statistical method to infer the average drift velocity \( V_{\text{drift}} \) and the diffusion constant \( D_{\text{vesicle}} \) by following a large number of individual trajectories, as once described in a study of Brownian colloids in a liquid crystal.

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The Brownian motion appears isotropic with a unique translation diffusion constant $D_{dx}$=1.11 μm²/s. The Stokes-Einstein’s formula gives a hydrodynamic radius deduced by video-microscopy $R_{hd}$=196 nm comparable to the value obtained by DLS. Due to an imperfect alignment of the magnetic field gradient with the x axis, we had to extract both coordinates of the magnetophoretic motion to calculate the total drift velocity:

$$\vec{V}_{drift} = \sqrt{V_x^2 + V_y^2} = 1.09 \mu m/s$$

(2)

This experimental value is compared to the theoretical estimate obtained by balancing the forces acting on a spherical magnetic vesicle at steady state in an external magnetic field gradient, which are the magnetophoretic force $F_{M}$, and the viscous drag, $F_{v}$, acting against it. The two forces are given by:

$$F_{M} = m \frac{dB}{dz}$$

and $$F_{v} = 6 \pi \eta R_{hd} V_{drift}$$

(3) and (4)

where $m$ is the magnetic moment of the vesicle, $\frac{dB}{dz}$ is the gradient of the magnetic field, $\eta$ is the viscosity of the solvent, $R_{hd}$ is the hydrodynamic radius of the vesicle, and $V_{drift}$ is the velocity of the particle. From the exact balance of the magnetic and the viscous forces, we calculate a theoretical magnetophoretic velocity $V_{drift}$=0.5 μm/s for the WD15-50 vesicles under a field gradient $\frac{dB}{dz}$=18.5 T/m. The factor around one-half between the expected drift velocity and the value $V_{drift}$=1.09 μm/s measured experimentally cannot be ascribed to the statistical noise because the uncertainties of the average displacements were estimated at 0.4% and 1.6% for the histograms at $r$ and $2 \pi t$ time steps containing respectively 64519 and 3920 data points. As a tentative explanation, we know from a reported work on giant magnetic liposomes that the drift coefficient is enhanced compared to Stokes’ formula (4) if the vesicles were deformed by the field into high aspect ratio ellipsoids during their migration. Another correction compared to the drag coefficient of a solid sphere originates also from the viscous dissipation inside the fluid magnetic membrane, for instance if it was subjected to a “caterpillar” or a “crawling” motion. In addition to these pure hydrodynamic effects, the measured $V_{drift}$ higher than its expected value might be explained by an underestimate of the numbers of USPIOs per vesicle appearing in Table 2. This would occur for example in the case of a non negligible amount of “blank vesicles” that were undetected but increased the average LC of iron oxide inside the magnetic vesicles above the USPIO/copolymer ratio (FWR) used for nanoprecipitation.

Apart from estimating the magnetic payload of the vesicles, the magnetophoresis experiment is also relevant to estimate their efficiency for magnetic guiding both in vivo and in vitro. Their magnetophoretic mobility in the vicinity of a strong NdFeB magnet is indeed of the same order of magnitude than values $\geq 1 \mu m/s$ reported by studies that evidenced the enhanced uptake of magnetic nanocarriers by cell cultures under field gradients. For in vivo experiments, it was hypothesized that the accumulation of magnetic colloids injected in the main blood stream at a specific region under magnetic field requires that their migration is faster than the blood velocity in the smallest vessels alimenting the tumor. However, the guiding of magnetic stealth liposomes injected in the caudal vein of mice by a strong permanent magnet applied directly on a solid tumor was evidenced, even though their drift velocity was 10 μm/s only. A model experiment consisting in attracting clusters of MNPs of sub-micron diameters (330 nm) by a permanent magnet while they were circulating in a flow loop showed that they were efficiently deposited at the surface of the capillary near the magnet even with a stream velocity as high as 1 cm/s. Therefore we believe that the USPIO loaded vesicles WD15-35 or WD15-50 are good candidates for such magnetic targeting applications, whereas the WDi vesicles might be too small and contain an insufficient number of USPIOs.

The magnetic response of PTMC$_{24}$-b-PGA$_{19}$ vesicles with their membrane filled by 50 wt % (WD15-50) or 70 wt % USPIO (WD1-70) was also studied by anisotropic SANS under an applied magnetic field. Vesicles were dispersed in light water (H$_2$O) in order to work in almost pure nuclear contrast conditions under field. The magnetic contrast of the γ-Fe$_2$O$_3$ USPIOs in H$_2$O being much lower than the nuclear contrast, the anisotropy of the SANS signal was not simply due to magnetization but reflects the spatial organization of the USPIOs and their possible rearrangement under magnetic field. The SANS patterns of WD15-50 vesicles are shown on Figure 5 at increasing field intensities up to 1 T.

The scattering patterns became clearly anisotropic when a magnetic field was applied to the sample. The lines of intensity in the $3 \times 10^{-3} - 3 \times 10^{-2}$ Å$^{-1}$ q-range were elliptical, elongated perpendicularly to the field direction. One possible scenario compatible with this asymmetry consists in the deformation of the hollow spheres formed by the USPIOs into either oblate or prolate ellipsoids symmetric by rotation along the field direction. However, one should keep in mind that the observed q-range corresponds to the length scale of the membrane thickness rather than to the whole size and shape of the vesicles.

In order to study this shape anisotropy more quantitatively, the scattering patterns were averaged in angular sectors around two directions parallel (ι) and perpendicular (χ) to the magnetic field. Examples of the resulting intensity curves are plotted on Figure S-2 (Supporting Information S-b). By comparing the difference $q_{\perp}$ - $q_{\parallel}$ in these two directions relatively to the wave vector $q_{\parallel}$ obtained by an isotropic averaging at the same intensity value, an anisotropy factor could be calculated for each sample and each magnetic field intensity (Table 3).

![Figure 5](http://pubs.acs.org/doi/abs/10.1021/nn102762f)

Figure 5. Anisotropic SANS patterns of WD15-50 in H$_2$O in the q range $3 \times 10^{-3} - 3 \times 10^{-2}$ Å$^{-1}$ under a magnetic field (horizontal) of intensity $B=0$ T (left), $B=0.1$ T (middle) and $B=1$ T (right). Each color corresponds to an iso-intensity range.
Table 3. Anisotropy factor of WD15-50 and WDi-70 vesicles calculated from anisotropic averaging of their SANS patterns at 6 cm⁻¹ iso-intensity under increasing magnetic field intensities.

<table>
<thead>
<tr>
<th>B (T)</th>
<th>(q² - q²) / q² (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>WD15-50</td>
<td>WDi-70</td>
</tr>
<tr>
<td>0.1</td>
<td>11.3</td>
</tr>
<tr>
<td>0.6</td>
<td>24.4</td>
</tr>
<tr>
<td>1</td>
<td>26.8</td>
</tr>
</tbody>
</table>

The calculated anisotropy factors confirmed the increase of membrane anisotropy with the applied magnetic field already visible on the SANS patterns. The anisotropy factor increased mainly between 0.1 and 0.6 T for both vesicular dispersions and remained almost constant up to 1 T. The plateau value reached at 0.6 T is ascribed to the saturation of the magnetic moment of a vesicle above B ≈ 0.7 T as observed on the magnetization curve (Figure S-5). It is worth noticing that the anisotropy parameter is smaller for WDi-70 than for WD15-50 vesicles, as explained by the number of USPIOs per vesicle (12 times smaller for WDi-70 than for WD15-50) and by the vesicle size (4 times smaller). Presumably due to their smaller size associated with a higher membrane curvature, WDi-70 vesicles are less prone to magnetic deformation than the much larger WD15-50 ones.

USPIO-loaded vesicles as contrast agents for MRI

The efficiency of MRI contrast agents based on USPIO is usually assessed by measuring the T₁ (longitudinal) and T₂ (transverse) relaxation times of the proton spins relaxations. Then the relaxation rates 1/T₁ and 1/T₂ are plotted versus total iron concentration in mM and the resulting slopes (s⁻¹mM⁻¹) called respectively r₁ and r₂ relaxivities can be used to compare different samples of USPIO differing by their size, dispersity, local concentration, aggregation state or any other parameter like the confinement in either a hydrophilic or a hydrophobic environment. In particular the encapsulation of USPIOs in hydrophobic polymers hampers the diffusion of water protons in the vicinity of USPIOs, which results in poor T₁ contrast enhancement, so that we can infer the same effect in our systems where the USPIOs are buried within a hydrophobic membrane. In addition, r₁ decreases rapidly as a function of the applied magnetic field (i.e. the Larmor’s resonance frequency) while r₂ reaches a plateau value due to the so-called “secular term” in its theoretical expression. The slopes give the relaxivity r₂ when increasing the size of USPIOs.

To evidence the effect of USPIO-loaded vesicles on T₁-weighted MR imaging, Figure 6-A shows MR images of wells containing USPIO-loaded vesicles. The results show that USPIO-loaded vesicles increase the signal intensity of the suspensions when compared to control. These findings are consistent with previous reports showing that the presence of iron oxide nanoparticles in aqueous solutions increases the relaxation rates of water protons.

Figure 6. (A) Transverse relaxation rates (1/T₂; s⁻¹) as a function of iron concentration (mM) for PTMC-b-PGA vesicles (WDi) loaded with 20, 35, 50 and 70 USPIO wt %. The slopes give the r₂ values, respectively 81±1, 134±2, 173±7 and 182±4 s⁻¹mM⁻¹; (B) T₁-weighted MRI images extracted from T₁ measurements experiment (4.7 T; multiple spin-echo 2D imaging sequence; Tₑ=10000 ms; inter echo-time, 5 ms; number of echo images, 256; FOV, 50×50 mm; matrix, 128×128; slice thickness, 1 mm) of WDi-70 vesicles at different dilution factors. The table gives the molar concentrations of iron ions, the total weight concentrations and the equivalent molar concentrations of vesicles.

To evidence the effect of USPIO-loaded vesicles on T₁-weighted MR imaging, Figure 6-B shows MR images of wells...
containing increasing concentrations of WDi-70 vesicles. A remarkable darkening (i.e., negative contrast enhancement) appeared even at low vesicle concentration. The MRI detection limit, defined as the copolymer concentration at which the MRI signal intensity decreases to 50% of that of pure water, was measured at 6.7 μg/mL for WDi-70 vesicles. Since the molar mass of the vesicles measured by SLS is 1.182×10^7 g/mol (Supporting Information S-i), the above sensitivity limit corresponds to a vesicle concentration of approximately 0.57 nM, which is one order of magnitude lower than the 5 nM reported for magnetic micelles, and, to our knowledge, the lowest value ever reported. For applications such as the evaluation of the bio-distribution or the targeting efficiency of a drug conveyed in nano-carriers, the concentration of ferric ions may not be the most relevant parameter for the radiologists. Therefore the r1 and r2 relaxivities are also expressed in Table 4 according to the concentration of vesicles in nM to facilitate the comparison with other nano-particular contrast agents. Unlike the relaxivities per ferric ion which saturate, their values per WDi vesicle increase monotonously with the magnetic FWR inside the membrane, at an almost constant hydrodynamic size.

Table 4. Longitudinal (r1) and transverse (r2) relaxivities of USPIO-loaded WDi vesicles when used as contrast agents for MRI at 4.7 T, deduced from the linear fits of the relaxation rates 1/T1 and 1/T2 versus molar concentrations both of ferric ions (in mM) or of vesicles (in nM). The number of Fe3+ per vesicle is the product of the number of USPIOs per vesicle (Table 2) by 8200 Fe3+ per USPIO on average.

<table>
<thead>
<tr>
<th>Sample code</th>
<th>RH (nm)</th>
<th>N^Fe/vesicle</th>
<th>r1 Fe^3+ (s^-1mM^-1)</th>
<th>r1 vesicle (s^-1nM^-1)</th>
<th>r2 Fe^3+ (s^-1mM^-1)</th>
<th>r2 vesicle (s^-1nM^-1)</th>
</tr>
</thead>
<tbody>
<tr>
<td>WDi-20</td>
<td>50</td>
<td>4.5×10^5</td>
<td>2.8±0.02</td>
<td>1.3±0.01</td>
<td>81±1</td>
<td>37±0.4</td>
</tr>
<tr>
<td>WDi-35</td>
<td>45</td>
<td>6.2×10^5</td>
<td>3.6±0.08</td>
<td>2.2±0.05</td>
<td>134±2</td>
<td>83±1.2</td>
</tr>
<tr>
<td>WDi-50</td>
<td>47</td>
<td>9.0×10^5</td>
<td>3.6±0.2</td>
<td>3.3±0.2</td>
<td>173±7</td>
<td>156±6</td>
</tr>
<tr>
<td>WDi-70</td>
<td>52</td>
<td>1.6×10^6</td>
<td>3.5±0.1</td>
<td>5.5±0.2</td>
<td>182±4</td>
<td>283±7</td>
</tr>
</tbody>
</table>

Doxorubicin loading and release by macroscopic heating

To determine the feasibility of magnetically controlled drug release, a dual loading of USPIOs and of doxorubicin was carried out. The nanoprecipitation was performed at pH 10.5 in order to deprotonate the DOX (pKa~8.3), thus maximizing the loading content (LC=34% without size variation at 50% FWR) and extending the release duration as described in a previous work. For each vesicular dispersion, the USPIO feed weight ratio (FWR) was fixed at a value lower than the maximum USPIO loading (namely 50 wt % for WDi and 35 wt % for WD15) so that space was left in the membrane for DOX entrapment. The DOX FWR in the nanoprecipitation mixture was then progressively increased. A DOX FWR of 20% was selected for both vesicular types since a drug loading at this level did not alter the self-assembly of the vesicles significantly: Table 5 shows indeed a moderate variation of their hydrodynamic size (RH decreases by 16% for WD15 and increases by 8% for WDi) and an unchanged surface charge. A larger 30% DOX FWR can be sustained by WD-15 vesicles without any size change, but for the smaller WDi-50 vesicles it leads to a two-fold size increase, presumably due to their larger curvature energy already invoked to explained their lower deformability under static magnetic field.

Table 5. Doxorubicin feed weight ratio (FWR), hydrodynamic size, polydispersity index and ζ potential of WD15-35 and WDi-50 vesicles. The pictures show the corresponding sample tubes.
and its maximal value being only

final edited and published work see http://pubs.acs.org/doi/abs/10.1021/nn102762f

membrane. The loading content (LC) and loading efficiency (LE) of DOX were determined by spectrophotometry. Values obtained for both vesicular dispersions with or without USPIOs are gathered in Table 6. A DOX LC around 10 wt% was found unaffected, which excludes the precipitation of the USPIOs and of the drug onto the hydrophilic chains and proves their embedment deeply inside the vesicular PTMC-b-PGA membrane. Finally, the colloidal stability of the WDi vesicles tested in MEM cell culture medium with fetal bovine serum (10% v/v FBS), and no change in size was observed for 24 hours.

Table 6. Influence of USPIO feed weight ratio on the DOX loading content and efficiency into WDi and WD15 vesicles. The FWR is in wt % relatively to copolymer in DMSO before nanoprecipitation. The LC is measured by spectrophotometry after nanoprecipitation and dialysis. The LE is the yield LC/FWR.

<table>
<thead>
<tr>
<th>Sample Code</th>
<th>USPIO FWR (%)</th>
<th>DOX FWR (%)</th>
<th>DOX LC (%)</th>
<th>DOX LE (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>WD15</td>
<td>0</td>
<td>152</td>
<td>0.15</td>
<td>-39.6</td>
</tr>
<tr>
<td></td>
<td>20</td>
<td>124</td>
<td>0.23</td>
<td>-39.3</td>
</tr>
<tr>
<td></td>
<td>30</td>
<td>128</td>
<td>0.16</td>
<td>nd*</td>
</tr>
<tr>
<td>WDi-50</td>
<td>0</td>
<td>56.5</td>
<td>0.22</td>
<td>-40.8</td>
</tr>
<tr>
<td></td>
<td>20</td>
<td>61</td>
<td>0.15</td>
<td>-42.0</td>
</tr>
<tr>
<td></td>
<td>30</td>
<td>137</td>
<td>0.20</td>
<td>nd*</td>
</tr>
</tbody>
</table>

*not determined

After nanoprecipitation with dual-loading in DOX and USPIOs, an extensive dialysis against a large volume (4 L) of Tris buffer (pH 7.4, 30°C, ionic strength 150 mM) during 4h allowed to reduce the pH back to 7.4 and to completely remove the unbound drug and DMSO. As for the colloidal stability of these dual loaded vesicles, their \( \zeta \) potential remained strongly negative (~ -40 mV). Therefore the corona of PGA chains was unaffected, which excludes the precipitation of the USPIOs and of the drug onto the hydrophilic chains and proves their embedment deeply inside the vesicular PTMC-b-PGA membrane. The loading content (LC) and loading efficiency (LE) of DOX were determined by spectrophotometry. Values obtained for both vesicular dispersions with or without USPIOs are gathered in Table 6. A DOX LC around 10 wt% was found in all cases, independently of the presence of USPIOs in the membrane. Finally, the colloidal stability of the WDi vesicles tested in MEM cell culture medium with fetal bovine serum (10% v/v FBS), and no change in size was observed for 24 hours.

Influence of USPIO feed weight ratio on the DOX loading content and efficiency into WDi and WD15 vesicles. The FWR is in wt % relatively to copolymer in DMSO before nanoprecipitation. The LC is measured by spectrophotometry after nanoprecipitation and dialysis. The LE is the yield LC/FWR.

<table>
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<th>Sample code</th>
<th>USPIO FWR (%)</th>
<th>DOX FWR (%)</th>
<th>DOX LC (%)</th>
<th>DOX LE (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>WD15</td>
<td>0</td>
<td>20</td>
<td>12.5</td>
<td>74</td>
</tr>
<tr>
<td></td>
<td>35</td>
<td>20</td>
<td>9</td>
<td>52</td>
</tr>
<tr>
<td>WDi-50</td>
<td>0</td>
<td>20</td>
<td>9.5</td>
<td>47.5</td>
</tr>
<tr>
<td></td>
<td>50</td>
<td>20</td>
<td>12</td>
<td>70</td>
</tr>
</tbody>
</table>

Comparing precisely the DOX loading efficiency between dual loaded vesicles and non magnetic ones, we observe that LE decreases by 22% for WD15 vesicles, whereas it increases by 22.5 % for WDi ones. As a result, the insertion of USPIOs and DOX appears competitive in the case of the larger WD15 vesicles, certainly due to a lack of space in the membrane (the difference between \( \Phi_{\text{emb}} \) and its maximal value being only 2.5 %, see Table 2). On the opposite, the incorporation seems cooperative for the smaller WDi vesicles. Such synergetic effect of dual loading has already been mentioned for copolymer micelles, for which the LC of DOX could be enhanced from 3 to 12 wt % by the co-encapsulation with hydrophobic USPIOs.36

Subsequently, in vitro release studies from the several prepared vesicular dispersions were monitored in various conditions by comparing the absorbance at \( \lambda_{\text{max}}=485 \text{ nm} \) with the DOX absorbance calibration curve (after background correction). The release kinetics in vitro at 37 °C of WDi-50 and WD15-35 vesicles fed with 20 wt % DOX appear almost similar. As seen on Figure 7 indeed, a plateau at around 50 wt % of released DOX was reached in both cases after one day. As stated in a previous work on the physicochemical conditions to optimize the loading and release of DOX with PTMC-b-PGA vesicles (but for a DOX LC of 34 wt % 3 times larger than in the present work and without USPIOs),118 temperature has a strong influence on the kinetics as well as on the amount of drug released: the plateau values at 5°C, 20°C, 37°C and 45°C were found respectively equal to 5%, 30%, 60% and 85% of the initial DOX load in the vesicles. This temperature sensitivity is presumably due to the semi-crystalline nature of the PTMC blocks inside membranes evidenced once by microcalorimetry,83 and in this work by birefringence measurement (see Supporting Information S-e).

On Figure 7, only 15 % of DOX was released after 6 hours at 23°C compared to 45 % released after the same time at 37 °C thus above the melting temperature of PTMC in the membrane of vesicles.

Doxorubicin release by magnetic hyperthermia

Having in mind this thermo-sensitivity of the release rate of DOX in vitro from dual-loaded PTMC-b-PGA vesicles, we studied the effect of an excitation by an oscillating magnetic field of the USPIOs confined in the membranes. Submitted to a strong radio-frequency field, USPIOs are known to dissipate heat originating from friction losses of their magnetic dipoles according to two different relaxation modes: Néel’s relaxation originating in the flips of each dipole between the “easy axes” of the crystalline structure and the Brownian rotational diffusion of the USPIO grains in the solvent of viscosity \( \eta \). According to a commonly accepted model,50 the specific loss power under a
field of frequency $f_{RF}$ and mean intensity $H_0$ expressed in W/g writes:

$$S_{LP}(f_{RF}, H_0) = \frac{\pi f_{RF}^2}{d^2} \chi''(f_{RF}) H_0^3$$

with

$$\chi''(f) = \frac{m_S}{3k_B T} \frac{f_{RF} \tau_{eff}}{1 + (f_{RF} \tau_{eff})^2}$$

Here $\chi''(f_{RF})$ is the loss term of the dynamic susceptibility of an USPIO with specific magnetization $m_S$, mass density $d$ and volume $V_{USPIO}$. The effective relaxation time $\tau_{eff}$ corresponds to the fastest mode between the two mechanisms participating to thermal dissipation. Both of them can be expressed as functions of the particle volume:

$$\tau_{Néel} = \tau_0 \exp(K_{USPIO}/k_B T)$$

and

$$\tau_{Brown} = 3\eta V_{USPIO}/k_B T$$

Although it does not take into account the possible variation of relaxation times with the magnetic field intensity, this model correctly describes the strong dependence of the $S_{LP}$ with the size distribution of a suspension of USPIOs and gives an optimal diameter about 14-15 nm. When USPIOs are confined in a viscous environment as in lipid compartments inside biological cells (endosomes), the Brownian relaxation mode can be neglected.

Figure 8 displays the kinetic profiles at constant temperature 23 °C; ●: $B=0$ T; ■: AC magnetic field ($f_{RF}=500$ kHz, $B_0=2.65$ mT).

Figure 8. Influence of a RF oscillating magnetic field on the in vitro release kinetics of WDi-50 vesicles at constant bath temperature (23 °C); ●: $B=0$ T; ■: AC magnetic field ($f_{RF}=500$ kHz, $B_0=2.65$ mT).

CONCLUSIONS

In the present study, the formation of new hybrid vesicular self-assemblies from the biodegradable PTMC-b-PGA copolymers and hydrophobically coated $\gamma$-Fe$_2$O$_3$ nanoparticles has been investigated. Hybrid vesicles have been obtained by one-step nanoprecipitation, leading to high loading content of magnetic nanoparticles (up to 70 wt %) in the membrane together with a good control over vesicles’ size and dispersity. The vesicular morphology was elucidated by combining light and neutron scattering techniques together with electronic and atomic force microscopy. These magnetic vesicles exhibited a long-term colloidal stability and showed suitable properties for biomedical applications: being guided by an external magnetic field gradient created by a small permanent magnet, they also showed an important contrast enhancement in Magnetic Resonance Imaging with a particularly low (sub-nanomolar) detection limit. Dual encapsulation of magnetic nanoparticles with doxorubicin in the biodegradable vesicular matrix is very promising as a versatile method to prepare multifunctional drug nanocarriers. The drug release rate could indeed be enhanced twice under the application of a RF oscillating magnetic field producing a local hyperthermia at the scale of the membranes.

The well-known hyperthermia effect of USPIOs was utilized here in a softer and gentler manner of action on the polymersomes’ membrane permeability than by thermoablation, which is based on the melting temperature of a semi-crystalline polycarbonate block. In future studies, we will enhance the RF-triggered release effect by using USPIOs with larger diameters (e.g. by a factor 2), which are known to exhibit much higher specific loss powers (~100 W/g or more). Apart from a higher thermal dissipation acting on the membrane fluidity and hence on the diffusion constants, those larger USPIOs will be partially ferrimagnetic, i.e. with a magnetic anisotropy energy $E_0\sim k_B T$. This might introduce another mechanism of membrane permeation, by direct rotation of the grains at the frequency of the oscillating magnetic field. Such a mechanism would be reminiscent of the “molecular drill” effect predicted long ago for lipid bilayers under mechanical stress by adsorption onto a corrugated surface.

To summarize, by exhibiting biocompatibility of the polymeric matrix, ease of preparation, contrast enhancement in MRI and triggered release under RF oscillating field, those hybrid vesicles are good candidates for the magneto-chemotherapeutic treatment of cancer. This work evidenced for the first time the concept of multi-functional polymersomes to combine imaging and therapy, opening new avenues to improve cancer treatments and to understand their mechanisms. The impact of such theranostic systems on tumor regression is currently under investigation.
EXPERIMENTAL DETAILS

Materials and syntheses
Polymer, drug and buffers. PTMC$_{24}$-b-PGA$_{12}$ diblock copolymer was synthesized by ring-opening polymerization (ROP) of γ-Benzyl-L-glutamate N-carboxyanhydride initiated by an amino functionalized PTMC macroinitiator upon a previously published method. All the experiments were conducted on a PTMC$_{24}$-b-PGA$_{12}$ (M$_{n}$ ~ 4900 g/mol) block copolymer which presents a hydrophilic weight fraction of 50 wt % and a molar mass dispersity of 1.15. The solvent for nonprecipitation (DMSO) was used without prior purification. Doxorubicin hydrochloride (CAS: 25316-40-9) was supplied by Discovery Fine Chemicals (Wilmington, UK). DOX was reconstituted in DMSO, stored at 5 °C and used within one month. Sodium chloride, Tris-HCl and Tris base were provided by Sigma.

Iron oxide nanoparticles. Superparamagnetic nanoparticles of maghemite (γ-Fe$_2$O$_3$), also called USPIOs, were synthesized by alkaline coprecipitation of iron(II) and iron(III) salts and sorted according to their size by fractionated phase separations. Briefly, the ionic strength was increased to screen the electrostatic interactions between the nanoparticles and obtain successive fractions of narrower size distribution, as measured all along the sorting process by vibrating sample magnetometry (VSM) and on the final sample by scattering techniques (SLS, DLS and SANS). For dispersion in CH$_2$Cl$_2$, the surface of the nanoparticles was grafted by the anionic surfactant Beycostat NB09 (CECA, Arkema group, France) used to disperse manganic pigments in aromatic and chlorinated solvents (but insoluble in aliphatic solvents), which is a mixture of mono- and diesters of phosphoric acid. The grafting procedure (30 min at 60°C, 20 mol% relatively to iron) was previously described.

Preparation of empty, USPIO loaded and DOX/USPIO dual-loaded vesicles
Carbonate buffer (pH 10.5, 50 mM, 4.5 mL) was added onto PTMC$_{24}$-b-PGA$_{12}$ (5 mg) dissolved in DMSO (0.5 mL) under magnetic stirring (1000 rpm) in a plastic tube (1.5 cm diameter) leading to a homogeneous dispersion of vesicles. A syringe pump controlled the water flow rate during injection. Two addition durations (5 seconds and 15 minutes respectively) of water solution into DMSO were used in order to tune the final average vesicle size. The resulting samples were respectively called WDi (for “instantaneous”) and WD15. The organic solvent was then removed by extensive dialysis against 4 L Tris buffer replaced at least twice (10 mM Tris, pH 7.4, 25 °C ionic strength 150 mM).

USPIO loading was performed at different feed weight ratios (FWR) (w$_{USPIO}$/w$_{copolymer}$) using the same nonprecipitation method. A negligible volume of USPIO suspension in CH$_2$Cl$_2$ (e.g. V$_{CH2Cl2}$/V$_{org}$=0.55% for w$_{USPIO}$/w$_{PTMC24-b-PGA12}$=50%) was added into the DMSO/copolymer solution prior to the addition of water. For DOX/USPIO dual-loaded vesicles, doxorubicin hydrochloride was at first solubilized in the DMSO/copolymer solution at 2 mg/mL before mixing with the USPIOs. After water addition, organic solvent and free DOX were removed by dialysis for 4h with a membrane of 3500 g/mol MWCO against 4 L Tris buffer (10 mM Tris, pH 7.4, 30 °C, ionic strength 150 mM). The doxorubicin loading content (LC) was determined after vesicle rupture using sonication in a mixture containing 80 % of water. This solvent mixture induced the aggregation of USPIOs that were then separated by centrifugation (1h, 10000 rpm). Then the titration of DOX was performed from the UV absorbance at $\lambda_{max}$=485 nm using the known value for doxorubicin in a DMSO/Tris buffer (80/20 v/v) mixture as calibration (see Supporting Information S-g).

Experimental Methods

Dynamic light scattering (DLS) and static light scattering (SLS) were performed using an ALV Laser goniometer, which consisted of a 35 mW HeNe linear polarized laser with a wavelength of 632.8 nm and an ALV-5000/EP Multiple Tau Digital correlator with 125 ns initial sampling time. The samples were kept at constant temperature (25 °C) during all the experiments. The accessible scattering angle range ranged from 30° up to 150°. However, most of the dynamic measurements were carried out at 90°. Aliquots of the samples (1 mL in a 10 mm diameter cylindrical glass cell) were immersed in a filtered toluene bath. The data acquisition was done with the ALV-Correlator Control software and the counting time for DLS was fixed for each sample at 30 s. To perform light scattering in static mode, the differential refractive index increment $dn/dc$ of PTMC$_{24}$-b-PGA$_{12}$ vesicles in buffer was measured over a concentration range from 0.2 to 1 mg/mL by means of a differential refractometer (Wyatt Optilab rEX) operating at a wavelength of 658 nm and at 25°C. A $dn/dc$ value of 0.345 ± 0.01 mL/g was obtained for WDi vesicles loaded with 50 wt % USPIOs, which is larger than the value $dn/dc$=0.085±0.012 mL/g reported for pure USPIOs coated with the same Beycostat surfactant. The mean hydrodynamic radii and polydispersity indexes (PDI) were determined using the second 0.9 order cumulant analysis.

Isotopic Small Angle Neutron Scattering (SANS) measurements were performed on the PAXY spectrometer of the Laboratoire Léon Brillouin (CEA-Saclay, France) equipped with a two dimension detector made of 128×128 cells. We used two configurations: the first one with a sample-to-detector distance of D=6.7 m and a neutron wavelength of $\lambda$=10 Å to cover a q range of 2.5×10$^{-3}$ – 2.5×10$^{-2}$ Å$^{-1}$; the second one with D=2 m and $\lambda$=6 Å to cover a q range of 2×10$^{-2}$ – 0.2 Å$^{-1}$. Full angular averaging of the detector cells at constant q was realized for the scattering patterns with the PASINET software available at www-llb.cea.fr.

The samples were prepared by nanoprecipitation, centrifuged and redispersed in the desired mixture of hydrogenated and deuterated solvents at a final concentration of 10 mg/mL. Three solvents were used in order to match the scattering length densities of the various components of the loaded magnetic polymersomes and to focus the contrast on selected features (see Supporting Information S-a). The magnetic scattering length density of the USPIOs estimated from the magnetization at saturation $M_s$ and the volume of the nanoparticles was $\rho_{mag}$=10$^{-6}$cm$^{-2}$. One solvent was pure H$_2$O, which allowed observing mainly the nuclear scattering of USPIO but also in a reduced way the copolymer signal. Pure D$_2$O almost matched the nuclear signal of the USPIOs: this scattering intensity revealed the fluctuation of the polymeric membrane together with the magnetic scattering of the USPIOs. Finally, the use of a H$_2$O/D$_2$O (65.6/34.4 v/v) mixture matching the copolymer scattering length density enabled to focus on the nuclear scattering of the USPIOs only. The calculated contrast of neutrons scattering-length densities between γ-Fe$_2$O$_3$ and this H$_2$O/D$_2$O mixture was $\Delta \rho$=5×10$^{10}$ cm$^{-2}$. SANS measurements were done in 5 mm thick quartz cuvettes for D$_2$O or 1 mm thick ones for H$_2$O and H$_2$O/D$_2$O solvents to minimize the incoherent scattering. All the scattered intensity curves were corrected from the incoherent background of their proper solvents. They have been also normalized by the incoherent signal delivered by a 1 mm gap water sample in order to account for the efficiency of the detector cells. Absolute values of the scattering intensity, $I(q)$ in cm$^{-2}$, were obtained from the direct determination of the number of neutrons in the incident beam and the detector cell solid angle.

Here we mainly discuss the SANS signal obtained with USPIO loaded polymersomes’ suspensions in the H$_2$O/D$_2$O mixture, which matches the copolymer. Following a method used for other kinds of nanocomposites made from colloids or
micelles.\textsuperscript{86, 87} The SANS curves of the USPIO loaded vesicles were divided by the volume fraction $\phi_{USPIO}$ and by the form factor of the USPIO nanoparticles measured independently on a dilute solution. This procedure yields the intra-aggregate structure factors $S_{\text{agg}}(q)$ of the USPIO nanoparticles, which tell us about their spatial arrangement into aggregates of a given geometry (micellar, vesicular, fractal...). The calculated form factor of hollow shells took into account their radius, membrane thickness, dispersity and the experimental resolution of the spectrometer.\textsuperscript{88}

Anisotropic SANS measurements. The sample was placed between the poles of an electromagnet producing a homogeneous magnetic field at the sample position, as checked by a Hall probe (Walker scientific). The solvent used was pure water which does not match the nuclear scattering length density of the copolymer but is insuring a negligible magnetic scattering of iron oxide ($\Delta \rho = 1.4\times10^{-10}$ cm$^{-2}$ between H$_2$O and the magnetic scattering length density of iron oxide). Nevertheless, the nuclear contrast of the USPIO ($\Delta \rho = 7.5\times10^{-10}$ cm$^{-2}$) remained still three times larger than the one of the copolymer ($\Delta \rho = 2.5\times10^{-10}$ cm$^{-2}$). The scattering intensity being proportional to the square of the contrast, we can neglect the contribution arising both from the copolymer and from the magnetic moments of the USPIOs in the total scattered intensity. An anisotropic analysis was applied to the scattering patterns obtained under magnetic field. To obtain anisotropic curves with a good statistics, the intensity on the 2D-detector was averaged in angular sectors either $[-30^\circ; 30^\circ]$ along the field direction where the scattered intensity was weaker, and thus called $I(q)$, or $[-15^\circ; 15^\circ]$ around the perpendicular direction and denoted $I^\perp(q)$.

Magnetization measurements. The magnetization curves of the maghemite USPIOs and of the USPIO-loaded vesicles were determined using a home-made vibrating sample magnetometer (VSM) under an applied magnetic field up to 0.93 Tesla. From the shape of the magnetization $M(H)$, the size distribution of the magnetic cores was obtained by convolving the first order Langevin’s law of paramagnetism $\mu_0 M(t) = -\mu_0 H_{\text{mag}} \phi_{\text{USPIO}} \mu_0 M_{\text{core}}(t)$, or $[-15^\circ; 15^\circ]$ around the perpendicular direction and denoted $I^\perp(q)$.\textsuperscript{89}

Magnetic birefringence. The setup that has been described previously\textsuperscript{90} was improved for temperature control. Briefly, it consisted in an electromagnet used to magnetically induce a macroscopic birefringence in a magnetic colloidal made of birefringent magnetic nanoparticles (or made of nano-objects filled with such MNPs). This induced birefringence was then measured by sending a linearly polarized He/Ne laser beam (10 mW) though the birefringent sample and analyzing the transmitted light with a second polarizer and a photodiode. A photo-electric modulator and a lock-in amplifier were used to increase the setup sensitivity, the resulting AC and DC signals being related respectively to the levels of birefringence and dichroism under the applied magnetic field. To perform measurements at various controlled temperatures, the glass cell containing the sample was put in a specifically designed copper cell, which temperature was regulated using a Pt100 temperature probe and Peltier devices connected to a current source and externally controlled by a PC using NI LabVIEW.

Magnetophoresis. A magnetophoretic experiment consists in measuring the constant velocity reached by magnetic objects in a magnetic field of increasing intensity (spatial gradient), applying on them a magnetic force balanced by a viscous drag one.\textsuperscript{62, 92, 93, 95, 97, 104} In our case, a drop of an aqueous vesicle solution was placed between a glass slide, a 200 µm spacer and a cover slip to prevent evaporation and convection. This cell was mounted on the stage of an inverted optical microscope (Leica DM-IL). A strong NdFeB magnet of 22 mm diameter and 10 mm thickness (Aimants Calamit, France) was held by a clip 6 mm away from the centre of the focus plane of the microscope. Bright field optical microscopy images taken with a 40X objective were recorded with a digital camera (Infinity3-1U, Lumenera, Ottawa, Ontario, Canada) enabling pixel-binning to enhance the recording rate. The magnetophoretic trajectories of about 280 vesicles exhibiting a biased thermal motion toward the magnet were recorded at video rate (24 frames/s). Three sequences (each containing 240 frames of 800x600 pixels) were analyzed offline using the “ParticleTracker” plug-in developed by the MOSAIC group at ETH Zürich for the free image processing software ImageJ with the following parameters: Kernel radius = 6, Cutoff radius = 0.0, Percentile = 0.6, Displacement = 5.0, Linkrange = 120.\textsuperscript{105} Each stack of 240 frames (10 s duration) necessitated a computing time of 16 min with a 64-bit desktop PC with 4Gb RAM. For theoretical calculations of the number of USPIOs per vesicle from the average drift velocity, a magnetic field gradient $dB/dz = 18.5$ mT/mm and an average magnetic flux density $B_H = 374$ mT were used as reported for an identical magnet.\textsuperscript{93}

MRI Relaxometry. For different USPIO-vesicle formulations, $T_1$ and $T_2$ relaxivities were measured at 4.7 T ($\gamma_{spin} = 200$ MHz) on a research MRI system (Bruker Biospec 47/50, Ettlingen, Germany) at 20°C. The transverse $T_2$ measurements were acquired using a multiple spin-echo 2D imaging sequence ($T_E=1000$ ms; inter-echo-time, 5 ms; number of echo images, 256; FOV: 50×50 mm; matrix: 128×128; slice thickness, 1 mm). The longitudinal relaxation times $T_1$ were obtained using an inversion-recovery 2D imaging sequence (increment of inversion delay: 34 ms with 456 increments) followed by a RARE imaging sequence (RARE Factor: 8; $T_E/T_2^{\text{eff}}$: 10 000/7.7 ms; FOV: 50×50 mm; matrix: 128×128; slice thickness: 1 mm). The relaxivity values $r_1$ and $r_2$ were calculated by linear fits of the relaxation rates $1/T_1$ and $1/T_2$ (s$^{-1}$) vs. iron concentration (mM) or vesicle concentration (nM).

Iron titration. The total iron concentration (mol/L) was determined by atomic absorption spectroscopy (AAS) with a Perkin-Elmer Analyst 100 apparatus after degrading the USPIO-loaded vesicles in boiling HCl (35%). The volume fraction of iron oxide was deduced from the molar mass (159.7 g/mol) and mass density (5.1 g/cm$^3$) of γ-Fe$_2$O$_3$, i.e. numerically $\phi_{USPIO}$ (% v/v) = 1.577 [Fe] (mol/L).

Electrophoretic mobility. Empty and loaded vesicles were analyzed with a ZetaSizer NanoZS (Malvern Instruments, Worcestershire, UK). The electrophoretic mobility ($\mu$) was converted into zeta potential ($\zeta$) using Smoluchowski’s approximation, which is valid since the vesicles are all much larger than the Debye length $\kappa_0$ of the buffers ($\kappa_0D_p > 1$). All the measurements were performed at 25°C and the data were at least the average of triplicate values.

Transmission electron microscopy. TEM images were recorded on a Hitachi H7650 microscope working at 80 kV equipped with a GATAN Orius 11 Megapixel camera. Samples were prepared by spraying a 1 mg/mL solution of the vesicles onto a copper grid coated with carbon (200 mesh) using a homemade spray tool.

TEM with negative staining. USPIO-loaded polymer vesicles (0.04 mg/mL in water) were adsorbed on a carbon-coated EM grid and negatively stained with 1% uranyl acetate. TEM was performed with a CM120 (FEI) microscope.

Cryo-TEM imaging. USPIO-loaded polymer vesicles (2 mg/mL of inca) were deposited on an EM grid coated with a 24 perforated carbon film. After draining the excess liquid with a filter paper, grids were quickly plunged into liquid ethane and mounted onto a Gatan 626 cryoholder. TEM was performed with a Tecnai F20 (FEI) microscope operated at 200 kV. The images were recorded with a 5 Megapixel USC1000-SSCCD camera (Gatan).
Atomic force microscopy. AFM images were recorded in air with a Nanoscope IIIa microscope operating in dry Tapping-mode. The probes were commercially available silicon tips with a spring constant of 42 N/m, a resonance frequency of 285 kHz and a typical radius of curvature in the 10-12 nm range. Freshly cleaved mica was used as sample substrate materials. For the observation of empty and USPIO loaded vesicles, sample solutions in water at concentrations of 0.01 mg/mL and 0.1 mg/mL respectively were deposited on the substrate (20 µL) and dried under vacuum at 40 °C for 12 hours.

In vitro DOX release. The required quantity of drug-loaded vesicles was poured into a dialysis tubing (Spectra/Por Float-A-Lyzer, 50 000 g/mol MWCO, 10 mm diameter, 10 mL volume). The dialysis membrane filled with 5 mL of DOX loaded polymersomes was introduced into a bath of 50 mL buffer (10 mM Tris, pH 7.4, ionic strength 150 mM). At each sampling point, sink conditions were maintained by replacing 2 mL of the outer medium reservoir by fresh buffer. Because of the known sensitivity of DOX to degradation, the amount of released drug was calculated by the difference between the initial drug content and the drug remaining at each sampling point in the suspension of vesicles. More precisely, a spectrophotometric measurement at λ_{max}= 485 nm was performed on an aliquot taken inside the dialysis bag. To take into account absorption by iron oxide and turbidity, the DOX concentration was calculated from the measured absorbance using a calibration curve in water after subtracting the absorbance value of similar USPIO-loaded vesicles. Another method consisted in redispersing the vesicles inside the aliquots into individual components (USPIOs, molecular DOX and copolymer unimers) in a DMSO:Tris (80:20) mixture before measuring the absorbance. Their calibration curves being provided as Supporting Information S-g, both methods led to comparable results, attesting the reliability of the measurements.

In vitro DOX release under an oscillating RF magnetic field. We used a RF generator built at the ICMCB laboratory in Pessac, France. An alternating magnetic field with \( f_{R}=500 \text{ kHz} \) frequency and mean field intensity \( H_{0}=2.12 \text{ kA/m} \) (induction \( B_{0}=\mu H_{0}=2.65 \text{ mT} \) ) was generated by a 28-turn pancake coil (20 cm height) cooled by a water circulation. The frequency was adjusted by a Celes inductor C97104 (Celem Passive Components, Israel). The electrical current was provided by a wave generator (FlI202, Française d’Instrumentsation, France) connected to a power amplifier (AR Worldwide 800A3, 10kHz–3MHz, EMV, France). The vesicles were prepared as usual then diluted by a factor 2. A dialysis bag filled with half of the dispersion (4 mL) was placed inside a plastic cylindrical vessel filled with 50 mL Tris buffer, fitting inside the coil of the above described setup. The release profile of the other half was performed in the same conditions of volumes, vessels and ambient temperature of the room (23°C), but kept far away from the magnetic field as a control experiment of release without RF field. In particular, both reservoirs of release medium were not stirred to avoid any parasitic heating due to the presence of a magnetic bar inside the RF magnetic field.

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91. The release of unbound USPIOs in water was indeed very unlikely because of their strong hydrophobic surface covered by the Beycostat™ surfactant. Taken several days after the preparation of the vesicles, the TEM images never exhibited individual nanoparticles or clusters that did not look as the hollow structures depicted on Figure 2. After attracting all the vesicles toward a strong magnet like during a magnetophoresis experiment, the solvent became colorless. Even with a sensitive analytical technique such as AAS, no traces of magnetic nanoparticles could be detected outside the membranes. This absence of detectable iron in the supernatant (i.e. at a concentration below 10⁻⁵ mol/L) combined with the VSM results strongly supports the fact that, concerning the USPIOs, the FWR (in the solvent mixture) and the LC (after nanoprecipitation and dialysis) are equal.


101. In our case, the histograms of elementary motions were built in both directions of the (x,y) plane of focus (Figure 5) for a total number of 64519 steps at a time delay τ =1/24s and 3920 steps at 2τ. Those curves were fitted with Gaussian laws centered respectively around VμSPIO and 2VμSPIO with standard deviations $\sigma_{x}(\tau) = (2\sigma_{x})_{\tau}$ and $\sigma_{y}(\tau) = (2\sigma_{y})_{\tau}$.

102. The magnetic moment per nanoparticle $\mu_{SPIO}$ per vesicle. The magnetic moment per nanoparticle $\mu_{SPIO}$ is simply the product of the specific magnetization $m_{S}$ by the USPIOs. For the mean magnetic field intensity $B_{0}$=174 mT in this magnetophoretic experiment, the USPIO-loaded vesicle magnetization $M$ corresponds to 55 % of its saturation value $M_{s}$ ($M_{s}(\tau)$=0.55, as seen on Figure S-4).


The relaxivity was measured at 0.47T but the value can be safely extrapolated at 9.4T according to the saturation of $r_2$ as a function of the field well known for USPIOs.

116. Roch, A.; Gossuin, Y.; Muller, R. N.; Gillis, P., Superparamagnetic colloid suspensions: Water magnetic relaxation and clustering. *J. Magn. Magn. Mater.* 2005, 293, 532–539. For USPIOs of magnetic core diameter 6.4 nm very close to ours, these authors localized the plateau of $r_2$ for clusters’ radii ranging from 33 nm to 132.5 nm due to the exit of the domain of validity of the Outer Sphere Diffusion model, where $r_2$ varies like the square of the number of USPIOs and the inverse of the cluster size, and the crossover to the Static Dephasing Regime characterized by a constant $r_2$. Therefore all the WDI samples which have hydrodynamic radii around 50 nm correspond to the onset of this SDR model, which might explain why $r_2$ starts to increase, but with a slower variation than a parabolic law, and then reaches a plateau for the two most highly loaded samples.


122. Taking into account dilution effects, the overall iron oxide concentration in the sample is indeed 0.175 g/L. Using 0.07 W/g as the SLP value of the USPIOs under such a RF magnetic field and inferring adiabatic conditions, we expect a heat production of 0.18 J per hour. The mass of water to heat up being ~4 g, we expected a temperature increase of only 0.01°C/h.


Supporting Information for

Doxorubicin Loaded Magnetic Polymersomes: Theranostic Nanocarriers for MR Imaging and Magneto-Chemotherapy

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S-a. Contrasts and treatments of the SANS data of γ-Fe₂O₃ USPIOs embedded within the membrane of poly(trimethylene carbonate)-b-poly(L-glutamic acid) (PTMC-b-PGA) vesicles.

The scattering length in neutron scattering is quantifying the strength of the interaction of a neutron wave with a given nucleus. Thus every element is characterized by a particular scattering length. The overall scattering length of a molecule \( b \) is defined as the sum of the scattering lengths of the different elements present in the molecule. Scattering length densities of the solvents, copolymer and maghemite related to their molar volumes are presented in Figure S-1.

\[
\delta^2 \propto (b - b_0)^2
\]  

\textbf{Figure S-1.} Scale of the scattering lengths densities of solvents, block copolymer and maghemite.

For magnetic molecules, a magnetic scattering length density corresponding to the interaction between the magnetic moment carried by neutrons and magnetic induction due to magnetic moments carried by the molecules has to be considered. It can be calculated knowing the number of Bohr magnetons per unit volume of the magnetic molecule (numerically \( \sim 38 \mu_B/nm^3 \) for the USPIOs used in this study according to VSM) and the scattering length of the moment of one Bohr magneton, \( 2.7 \times 10^{-13} \text{ cm/} \mu_B \).

When a particular entity (molecule, micelle...) is immersed in a solvent, its nuclear contrast factor \( \delta \) is defined as the square of the difference between the scattering length density of the entity \( b \) and that of the solvent \( b_0 \):

\[
\delta^2 \propto (b - b_0)^2
\]  

(S-1)
The scattered intensity by an ensemble of symmetric objects (as micelles, spheres) in solution is defined for each scattering vector \( q \) as:

\[
I(q) \propto \delta^2 \Phi [P(q)S(q)]
\]

(S-2)

where \( \Phi \) is the volume fraction of the objects in the solution, \( P(q) \) the form factor of the objects and \( S(q) \) their structure factor (revealing the interaction between different them).

In our case where USPIOs are embedded in block copolymer vesicles in solution, the scattered signal is complex since multiple interactions exist between different \( \gamma \)-Fe\(_2\)O\(_3\) nanoparticles, different copolymer chains but also cross-interactions between nanoparticles and copolymers. By omitting the signal due to magnetic fluctuations, the scattered intensity of such a system can be written as the sum of complex partial structure factors weighted by the corresponding contrasts:

\[
I(q) \propto \delta^2_{\gamma-Fe_2O_3} \Phi_{\gamma-Fe_2O_3} [P_{\gamma-Fe_2O_3}(q)S_{\text{intra}}(q)] + \delta^2_{\text{copo}} \Phi_{\text{copo}} S_{\text{copo}}(q) + \delta^2_{\gamma-Fe_2O_3} \delta_{\text{copo}} S_{\text{copo}/\gamma-Fe_2O_3}(q)
\]

(S-3)

This equation can be simplified when the solvent is matching the scattering length density of the copolymer or of the USPIOs. For instance, by using as a solvent a particular mixture of H\(_2\)O/D\(_2\)O such as \( \delta_{\text{copo}} = (b_{\text{copo}} - b_{\text{H}_2\text{O}/\text{D}_2\text{O}}) = 0 \), it is possible to remove all the contributions of the copolymer.

Equation (S-3) thus becomes:

\[
I(q) \propto \delta^2_{\gamma-Fe_2O_3} \Phi_{\gamma-Fe_2O_3} [P_{\gamma-Fe_2O_3}(q)S_{\text{intra}}(q)]
\]

(S-4)

The structure factor between \( \gamma \)-Fe\(_2\)O\(_3\) nanoparticles is now defined as:

\[
S_{\text{intra}}(q) = \frac{I(q)}{\Phi_{\gamma-Fe_2O_3} P_{\gamma-Fe_2O_3}(q) \delta^2_{\gamma-Fe_2O_3}}
\]

(S-5)

where \( P_{\gamma-Fe_2O_3}(q) \) is the form factor of \( \gamma \)-Fe\(_2\)O\(_3\) nanoparticles. The latter can simply be obtained from the scattering of a dilute solution of USPIOs (Equation (S-2) with \( S(q)=1 \)).

In the case of an assembly of magnetic objects, Equation (S-3) is even more complex since the organization of magnetic moments carried by the \( \gamma \)-Fe\(_2\)O\(_3\) nanoparticles gives rise to an additive signal.
with a magnetic contrast. By using several contrast matching conditions, it will be possible to better describe the organization of such complex structures.

**S-b. Shape anisotropy of the vesicles and variation of their membrane thickness under magnetic field**

In order to study the deformation of the vesicles under a constant magnetic field, the anisotropic SANS patterns were averaged in angular sectors either [-30°; 30°] along the field direction or [-15°; 15°] around the perpendicular direction, as represented by a mask on Figure S-2-A. The resulting curves \( I_{//}(q) \) and \( I_{\bot}(q) \) were compared by two different methods. At first, we considered a given intensity (e.g. 6 cm\(^{-1}\)) and we calculated an anisotropic factor \((q_{\bot} - q_{//})/q_0(\%)\) (Figure S-2-B).

**Figure S-2.** Masks used to average an anisotropic SANS pattern under a horizontal magnetic field in two perpendicular angular sectors (A). Intensity curves \( I_{//}(q) \) and \( I_{\bot}(q) \) obtained in the \( q \) range \( 5 \times 10^{-3} - 3 \times 10^{2} \) Å\(^{-1}\) (B). For a given intensity (horizontal line here at 6 cm\(^{-1}\)), corresponding wave vectors \( q_{//} \) and \( q_{\bot} \) were extracted and compared to the value \( q_0 \) that would be obtained by an isotropic averaging.
Then for each direction, we calculated a membrane thickness respectively $\delta^//$ and $\delta^\perp$ by plotting Kratky-Porod’s asymptotic law $\ln[q^2 I(q)] \sim -q^2\delta^2/12$ in the low-$q$ regime. We recall that the solvent is pure H$_2$O, allowing to observe mainly the nuclear scattering of the USPIOs. Consequently, the membrane thickness calculated here is the inorganic layer, which does not take into account the outer leaflet of the copolymer bilayer. Thus variations of $I(q)$ in both directions translate a reorganization of the USPIOs in the vesicle assembly.

![Graph showing membrane thickness variation](image)

**Figure S-3.** Variation of membrane thicknesses $\delta^//$ and $\delta^\perp$ of WD15-50 vesicles in light water calculated in Kratky-Porod’s regime (A). Sketch representing the possible deformation of the vesicle into an ellipsoid elongated along the field, combined with the migration of the USPIOs from the magnetic poles towards the equator in order to minimize their magnetic dipolar energy (B).

The variation of $\delta^//$ and $\delta^\perp$ as a function of the magnetic field intensity plotted on Figure S-3-A can be interpreted in several manners. One possibility consists in the stretching of the membrane near the magnetic poles (i.e. the portions of the shell almost perpendicular to the magnetic field) due to an overall deformation of the vesicle into an elongated (prolate) ellipsoid. Almost equivalently, the apparent decrease of membrane thickness $\delta^//$ could also signify that the USPIOs move away from the magnetic poles, where dipolar repulsions between them are the strongest. In the meantime, the apparent thickness of the remaining parts of the membrane and especially near the equator $\delta^\perp$ increases (up to 18 nm at 0.6 T). In this model, the USPIOs would concentrate in the regions of the membrane with a normal perpendicular to the field, where their dipolar interactions can be attractive. A
combination of the two interpretations (ellipsoidal deformation and migration of the USPIOs in the fluid membrane) is sketched on the Figure S-3-B. It is mainly in the field interval 0.1 – 0.6 T that the membrane thicknesses $\delta^\parallel$ and $\delta^\perp$ together with the anisotropy factor $(q^\perp - q^\parallel) / q^0$ at 6 cm$^{-1}$ vary, before remaining almost constant up to 1 T. The plateau values reached between 0.6 T and 1 T are likely due to the saturation of the magnetic moment of the vesicle above $B \approx 0.7$ T as measured by VSM (Figure S-4).

**S-c. SANS study of magnetic vesicles in D$_2$O.**

Pure D$_2$O almost matches the nuclear signal of USPIOs, so that the scattering intensity reveals the fluctuation of the polymeric membrane together with the magnetic scattering of iron oxide. A correlation bump progressively appears on Figure S-4 with increasing FWR of USPIOs. The associated correlation distance (similar as the one observed in the solvent H$_2$O/D$_2$O mixture matching the copolymer) is thought to translate the correlation between the holes left by the USPIOs in the polymer membrane.

![Figure S-4. SANS curves of PTMC$_{24}$-$b$-PGA$_{19}$ vesicles prepared under WDi conditions with increasing USPIO loading centrifuged then dispersed in D$_2$O at a concentration of 10 mg/mL.](image-url)
S-d Magnetization of USPIO-loaded vesicles

The magnetization curve of vesicles loaded with 50 wt% USPIO (WD15-50) was measured by VSM and compared to the starting $\gamma$-Fe$_2$O$_3$ USPIO dispersion in CH$_2$Cl$_2$ (Figure S-5). The absence of hysteresis in the experimental data confirmed the superparamagnetic behavior of the USPIOs embedded in the hydrophobic PTMC membranes of the vesicles. The slight magnetization decrease for magnetic vesicles compared to bare USPIOs (ferrofluid) was thought to arise from a much smaller signal/noise ratio due to the overall iron concentration during analysis which is about 200 times lower for USPIO-vesicles. Both USPIO ferrofluid and USPIO-loaded vesicles magnetization curves were fitted according to Langevin’s law of paramagnetism, each USPIO being a giant magnetic monodomain with an average magnetic dipole of 8200 $\mu$B, which is also the approximate number of Fe$^{3+}$ ions per USPIO (see below).

![Figure S-5. Vibrating Sample Magnetometry measurements. Magnetization ($M$) of USPIOs dispersed in CH$_2$Cl$_2$ (squares) and of vesicles loaded with 50 wt % USPIOs (WD15-50) normalized by the value of magnetization at saturation ($M_s$) for magnetic field intensities up to 0.93T (increasing and decreasing). On the magnetization curve of WD15-50 vesicles, the plateau value $M_s/\Phi_m$ allowed to determine a concentration of USPIOs 0.43 g/L very close to the expected one 0.429 g/L, taking into account the USPIO FWR and the dilution effect during dialysis. Langevin’s fits are given in plain line. On those curves, the Langevin’s law was convolved with a Log-normal probability law of median diameter $D_{mag}^{USPIO} = 6.3$ nm and width $\sigma=0.22$, defined as the standard deviation of the distribution $\ln(D/D_{mag}^{USPIO})$. Due to the tail of this distribution law towards large diameters, the weight averaged
diameter $D_w = \langle D^3 \rangle / \langle D^3 \rangle$ was shifted to a larger value compared to the median value $D_{mag}^{USPIO}$. Simple statistics gives $D_w = D_{mag}^{USPIO} \times \exp(3.5\sigma^2) = 7.5 \text{ nm}$, which is intermediate between the values determined by scattering experiments $2R_G^{USPIO} = 6.1 \pm 0.12 \text{ nm}$ and $2R_H^{USPIO} = 9.4 \pm 0.14 \text{ nm}$. The weight average volume needed in the magnetophoresis experiment is thus $V_{USPIO} = \pi D_w^3 / 6 = 220 \text{ nm}^3$. From the mass density 5.1 g/cm$^3$ of maghemite, a molar mass $M_w^{USPIO} = 670 \text{ kg/mol}$ was calculated. This estimate compares well with the experimental value $M_w^{USPIO} = 675 \text{ kg/mol}$ measured by a Zimm-plot of a series of dilutions of the USPIOs in toluene at several concentrations and scattering angles from 50° to 130°. The molar mass of $\gamma$-Fe$_2$O$_3$ being equal to 159.7 g/mol, each USPIO contains on average 8400 Fe$^{3+}$ ions. Inferring a typical value of the anisotropy constant of maghemite $K_a \approx 10^4 \text{ J/m}^3$, the magneto-crystalline energy $E_a = K_a V_{USPIO}$ was estimated around 0.5$k_B T$. Therefore these magnetic nanoparticles exhibit a pure superparamagnetic behavior, with no ferromagnetic contribution originating from the tail of the Log-normal distribution towards the largest diameters. As for their specific magnetization, we found $m_S = 2.8 \times 10^5 \text{ A/m}$. For comparison with the literature in CGS units, it corresponds to $m_S = 3520 \text{ Oe}$ and $m_S / 4\pi = 280 \text{ emu/cm}^3$ or 55 emu/g. This value is about 30% less than for bulk maghemite ($m_S = 4.2 \times 10^5 \text{ A/m}$), this discrepancy being ascribed to the magnetic disorder of iron spins at nanoparticles’ surface.

**S-e. Magnetic birefringence of USPIO-loaded vesicles**

![Magnetic birefringence curve](image1)

**Figure S-6.** A. Birefringence curve as a function of the applied magnetic field for WD15-15 polymersomes at 8mg/mL inside a glass cuvette of 3 mm light path at 19°C and 64°C. B. Raw birefringence signal at constant magnetic field $H=2100 \text{ Oe}$ during a cooling ramp from 64°C to 19°C.
A suspension of WD15-15 vesicles was measured with the experimental setup designed to measure the birefringence as a function of magnetic field intensity and at regulated temperature. While the curve at 64°C can be adjusted with a 2nd order Langevin’s function (with a parabolic variation at low field and saturating at large fields at a plateau value), the birefringence appears much higher at 19°C, which cannot be accounted simply by a lower thermal agitation of the USPIOs. The raw optical signal on the photodiode is also plotted at a constant magnetic field intensity (0.21 Tesla) while decreasing slowly the sample temperature at 1°C/min. The observed peak that compares well with \( T_{\text{melting}} = 36\)°C measured for PTMC-b-PGA vesicles by micro-Differential Scanning Calorimetry in a previous work corresponds to a burst of birefringence thought to arise from crystalline spherulites of PTMC inside the membranes.

**S-g. Calibration curves for DOX titration by spectrophotometry**

**Figure S-7.** A. Calibration curves of the absorbance at 485 nm vs. doxorubicin concentration either on intact vesicles in Tris buffer (blue square) or on disrupted vesicles (red circles) in DMSO/Tris buffer (80:20 v/v). B. Absorption spectra of PTMC-b-PGA vesicles loaded at pH 10.5 with increasing amounts of doxorubicin (FWR values) after dialysis at pH 7.4. Once normalized by the DOX concentration, the curves can be superposed and compared to the spectra of free doxorubicin in its protonated (red circles) and deprotonated (blue square) forms, respectively solubilised in Tris pH 7.4 and carbonate pH 10.5 buffers. The shoulder around 600 nm typical of the alkaline DOX progressively disappeared from the vesicles’ curves when FWR increases, as expected from the dilution effect of weak acids in solution.
The concentration of doxorubicin was determined by two different methods allowing a cross check. Firstly, absorbance at $\lambda_{\text{max}} = 485$ nm was measured on aliquots of vesicles in the release buffer after each sampling point. Secondly, the dual loaded USPIO/DOX vesicles were broken by sonication in a mixture of DMSO and Tris buffer (80:20 v/v) before measuring the absorbance at 485 nm.

**S-h. Other Atomic Force Microscopy Images**

Figure S-8. AFM Tapping Mode™ phase images of vesicles deposited on mica. Images show pure PTMC$_{24}$-b-PGA$_{19}$ vesicles (WDi-0) on the left and USPIO-loaded ones (WDi-50) on the right.
The discrepancy between the diameters around 200 nm appearing on the AFM images and the hydrodynamic sizes (134 nm for WDi-0 and 94 nm for WDi-50) is ascribed to the total spreading of the vesicles onto the mica substrate that we describe by the following sketch:

![Sketch of vesicles spreading on mica substrate](image)

The same spreading effect is invoked to explain the larger diameters measured on some of the TEM images (those of samples deposited by spraying) compared to hydrodynamic sizes (Figure 2A, B and C).

**S-i. Static Light Scattering**

![Zimm plot of WDi-50 vesicles’ suspensions](image)

**Figure S-9.** Zimm plot of WDi-50 vesicles’ suspensions at five different concentrations ranging from 0.2 to 1 mg/ml. The extrapolated intercept leads to a molar mass $1.182 \times 10^7$ g/mol.