Licking behaviour induces partial anthelmintic efficacy of ivermectin pour-on formulation in untreated cattle.

Alain Bousquet-Mélou, Philippe Jacquiet, Hervé Hoste, Julien Clément, Jean-Paul Bergeaud, Michel Alvinerie, Pierre-Louis Toutain

To cite this version:

Licking behaviour induces partial anthelmintic efficacy of ivermectin pour-on formulation in untreated cattle

Alain Bousquet-Mélou a,d, *, Philippe Jacquiet b,d, Hervé Hoste b,d, Julien Clément b,d, Jean-Paul Bergeaud b,d, Michel Alvinerie c,d and Pierre-Louis Toutain a,d

a UMR181 Physiopathologie et Toxicologie Expérimentales, INRA, ENVT, Ecole Nationale Vétérinaire de Toulouse, France
b UMR1225 Interactions Hôtes Agents Pathogènes, INRA, ENVT, Ecole Nationale Vétérinaire de Toulouse, France
c UR66 Pharmacologie-Toxicologie, INRA, Toulouse, France
d Université de Toulouse, Toulouse, France.

*Corresponding author: Ecole Nationale Vétérinaire de Toulouse, 23 chemin des Capelles, BP 87614, 31076 Toulouse cedex 03, France
Tel: +33 561 193 925; Fax: +33 561 193 917; email: a.bousquet-melou@envt.fr
ABSTRACT

Licking behaviour in cattle has been reported to account for the disposition of topically administered macrocyclic lactones. However, its impact on anthelmintic efficacy remains to be established. Therefore, we evaluated the impact of ivermectin exchange between cattle on the reduction in the faecal egg count (FEC) after pour-on administration in a group of 10 heifers experimentally infected with O. ostertagi and C. oncophora. Four treated (500 µg/kg, pour-on) and six untreated animals were put together after treatment and plasma and faecal exposure to ivermectin as well as the FECs were evaluated before and over 40 days after pour-on. Ivermectin was detected in plasma and faeces of the six untreated heifers, with maximal exposures 2-3-fold lower than the minimal exposures in treated animals. The interindividual variability of exposure was very high in untreated, a 10-fold difference between the upper and lower range limits, compared to treated heifers, where there was only a 2-fold difference. Anthelmintic efficacy, expressed as an average reduction of the FECs over the experimental period, was maximal in the treated group. In untreated heifers, anthelmintic efficacies ranged from zero to maximal efficacy, with intermediary values between 30 and 80%. The use of a classical pharmacodynamic model demonstrated a clear relationship between exposure and efficacy and enabled us to define the critical plasma or faecal ivermectin concentrations delimiting an exposure window associated with partial anthelmintic efficacy. This range of ivermectin plasma concentrations [0.1–1ng/mL] could be considered as a potential selection window for anthelmintic resistance. Finally, our results show that macrocyclic lactone exchange between cattle after pour-on administration, resulting from natural grooming behaviour, can significantly impact on anthelmintic efficacy. This raises several issues such as the design of comparative clinical trials and the occurrence of partial
efficacy which is considered a risk factor for the development of anthelmintic resistance.

**Key words:** Macrocyclic lactones; Ivermectin; topical; allo-grooming; Anthelmintic resistance; Cattle
1. Introduction

Macrocyclic lactones (MLs) such as ivermectin (IVM), doramectin (DOR), eprinomectin (EPR) and moxidectin (MOX) are administered topically to conveniently treat different parasitic infections in cattle. The efficacy of pour-on (PO) formulations depends on systemic (blood) exposure for all internal parasites. We have reported that after a topical administration of DOR or IVM (500 µg/kg) the range of individual exposures was very large, with values of the areas under the plasma concentration-time curves (AUC) showing up to a 3.6-fold variation for IVM and up to a 2.2-fold variation for DOR, indicating a poor inter-individual reproducibility of this route of administration (Gayrard et al., 1999). Low and variable individual exposures after PO administration was also reported for MOX in calves (Sallovitz et al., 2002) and for EPR in dairy cows (Alvinerie et al., 1999). In addition, in a parallel trial involving two groups of 20 cattle dosed by PO (500 µg/kg) or subcutaneously (SQ, 200 µg/kg) with either DOR or IVM (Toutain et al., 1997), we showed that the SQ administration was associated with not only a higher systemic exposure for both products but also a much lower inter-animal variability than PO (coefficients of variation of 3.1% and 4.7% vs. 25% and 37% for SQ vs. PO formulations of DOR and IVM respectively). These results indicated that the PO formulation was responsible for the poor reproducibility of plasma drug exposure. This phenomenon was better understood when it was demonstrated that the actual disposition of MLs poured on the backs of cattle was largely influenced by both self-and allo-grooming and that consequently a large fraction of the MLs was actually orally and erratically ingested by the animal itself (Laffont et al., 2001; Laffont et al., 2003) or exchanged with another animal in the herd (Bousquet-Mélou et al., 2004) rather than directly absorbed throughout the skin. The consequences of this largely overlooked “behavioural clearance”
mechanism for topically administered drugs in cattle are manifold, including inconsistence of drug efficacy, unexpected residue levels (Imperiale et al., 2009), contamination of the environment (Herd et al., 1996), or the design of clinical and bioequivalence trials (Barber and Alvinerie, 2003). It is often quoted that underdosing and subtherapeutic ML exposure are factors favouring reduced anthelmintic efficacies in the field (El-Abdellati et al., 2010), which can in turn favour the emergence of anthelmintic resistance (Smith et al., 1999) for which there is experimental evidence (Van Zeveren et al., 2007). As drugs poured on the back of treated animals can easily be exchanged between animals of a herd by allo-licking we hypothesized that some animals could easily be underexposed to MLs especially if only some animals within a herd are treated. Under these conditions, undesirable subtherapeutic concentrations could be anticipated in both treated and untreated animals.

In the present study using an experimental infection, we addressed the question of the occurrence of a possible underexposure window in a group of young cattle for which only a few of the animals were treated with an IVM PO dose. More precisely, we sought to establish a range of plasma and faecal IVM concentrations associated with partial anthelmintic efficacy.
2. Materials and methods

2.1. Experimental Animals

Ten heifers, nine Prim Holstein and one Limousine, which were 95-225-days old at the beginning of the trial, were first de-wormed with an oral dose of oxfendazole (5mg/kg) (Synanthic®, Merial, France). Faecal egg counts (FEC) were performed according to Raynaud (Raynaud, 1970) to check the efficacy of the oxfendazole drench. All the animals were negative at the time of the future L3 challenge. The heifers were housed together in a small yard (no grass) to prevent re-infestation. They were fed with a commercial concentrate diet (2 kg per day) and hay ad libitum.

2.2. Experimental infection, treatment and sampling

Three weeks after the de-worming, each heifer was orally infected with 20,000 L3 of *Ostertagia ostertagi* and 20,000 L3 of *Cooperia oncophora* (adapted from Vercruysse et al., 2000)). Four out of the ten animals were randomly selected to be treated 35 days after the experimental infection with an IVM PO dose (500µg/kg, Ivomec® bovine pour-on, Merial France). IVM was gently poured down the dorsal middle-line according to the manufacturer’s recommendations. The six other untreated cattle were considered as controls and remained permanently housed with the group of the four treated animals. Before treatments, the mean and standard deviation of egg excretion in the treated and untreated groups were similar: 538 ± 448 and 503 ± 392 eggs per gram (EPG), respectively. After the IVM treatment, efficacy was assessed by daily FEC measurements over two weeks and then three times weekly up until five weeks after the IVM treatment. Blood samples (n=21 per animal) were regularly collected from the jugular vein before (control value) and up to
day 36 post-IVM administration. Faecal samples (n=26 per animal) were obtained directly from the rectum between 08:00 am and 09:00 am. They were regularly collected before (4 control samples) and after IVM administration up to 41 days after IVM administration.

2.3. Analytical procedures

IVM (22, 23-dihydroavermectin B1a) concentrations in plasma and faeces were measured using a HPLC technique as previously described (Alvinerie et al., 1987). The limits of quantification of IVM were 0.05 ng/mL for plasma and 0.5 ng/g for wet faeces. Accuracy and precision (intra-assay variation) expressed as relative standard deviation were less than 8 and 6%, respectively.

2.4. FEC and efficacy measurements

FECs were carried out using the modified McMaster method (Raynaud, 1970) with a sensitivity of 7 eggs per gram of faeces. The time development of individual efficacy was assessed using daily FECs. The percentage (from 0 to 100%) of reduction from the control FEC (geometric mean of the four control measures) was calculated daily using the following equation:

\[
FEC_{\text{reduction}(\%)}(\text{day } i) = 100 \times \left(1 - \frac{FEC_{\text{day } i}}{FEC_{\text{control}}}\right) \text{ Equation 1}
\]

where \(FEC_{\text{day } i}\) is the FEC at day \(i\) post-IVM administration.

When the FEC after IVM administration was higher than control values, its value was arbitrarily fixed at the control value. The area under the \(FEC_{\text{reduction}(\%)}(\text{day } i)\) versus time curve (AUC\(_{FEC_{\text{reduction}(\%)}[0-\text{day } i]}\) was calculated using the trapezoidal rule and then the efficacy at a given day (\(\text{day } i\)) was expressed as the AUC\(_{FEC_{\text{reduction}(\%)}[0-\text{day } i]}\) divided
by the time interval \([0-\text{day}_i]\):

\[
\text{Efficacy}(\%)_{\text{day}_i} = \frac{AUC_{\text{FEC reduction}(\%)[0-\text{day}_i]}}{\text{Time}[0-\text{day}_i]}
\]

Equation 2

\text{Efficacy}(\%)_{\text{day}_i} \text{ corresponds to the average } \text{FEC}_{\text{reduction}(\%)} \text{ over the time interval } [0-\text{day}_i]. \text{ Expressing efficacy this way gives at each day, not the result observed on that day, but an average efficacy from day 0 to } \text{day}_i. \text{ The advantages of this method were obtaining a smooth curve that avoided the spurious rebound in some daily FECs, and taking into account the more or less rapid time development of efficacy associated with the unpredictable beginning of IVM exposure in untreated animals.}

2.5. Pharmacokinetic measurements and pharmacodynamic modelling

Pharmacokinetic analyses were performed using WinNonlin Professional version 5.2 (Pharsight Corporation, Cary, NC, USA). Areas under the plasma or faecal IVM concentration curves (AUC\textsubscript{IVM}) were obtained by the trapezoidal rule from time 0 to the last measurable concentration (Non-compartmental module of WinNonlin).

The exposure-efficacy relationship was described using the following sigmoid Emax model:

\[
\text{Efficacy}(\%)_{\text{day}_{30}} = \frac{E_{\text{max}} \times (AUC_{\text{IVM}})^n}{(AUC_{\text{IVM}})^{50} + (AUC_{\text{IVM}})^n}
\]

Equation 3

where \text{Efficacy}(\%)_{\text{day}_{30}} \text{ is the individual efficacy at day 30 calculated using Equation 2, } E_{\text{max}} \text{ is the maximal estimated Efficacy(\%), AUC}_{\text{IVM}} \text{ is the plasma or faecal IVM AUC as obtained for each individual, } n \text{ is the Hill coefficient and } (AUC_{\text{IVM}})^{50} \text{ is the plasma or faecal IVM AUC giving 50\% of Emax. Descriptive statistics were obtained with WinNonlin and the results are reported as means (arithmetic or geometric) and SD.}
3. Results

3.1. Plasma and faecal exposure to IVM in treated and untreated heifers

Figs 1 and 2 show the semi-logarithmic plot of individual plasma and faecal exposures in the ten heifers. Visual inspection of the graphs indicates that all the cattle (treated and untreated) were exposed to IVM, but that treated cattle were always more exposed to IVM than untreated ones. Mean and extreme $\text{AUC}_{\text{IVM}}$ values are given in Table 1.

Table 1 shows that the mean plasma $\text{AUC}_{\text{IVM}}$ of untreated cattle was 9.3% of the mean plasma $\text{AUC}_{\text{IVM}}$ of treated cattle. However, on an individual basis, the highest value for an untreated heifer reached 36.5% of the lowest value for the treated cattle.

Faecal IVM exposure confirmed that the six untreated cattle ingested by allo-licking some of the IVM poured on the back of the four treated animals. The mean faecal $\text{AUC}_{\text{IVM}}$ of untreated cattle represented 14.8% of the mean faecal $\text{AUC}_{\text{IVM}}$ of treated cattle but the most exposed of the untreated heifers reached 53.6% of the value of the least exposed treated cattle. It is worth to note that the faecal excretion of IVM at detectable levels was shorter in the untreated heifers than in the treated ones.

3.2. FEC reduction in treated and untreated heifers

The time development of IVM anthelmintic activity from day 1 to day 30 post-IVM administration is shown in Fig. 3. Individual FEC expressed as percent of control values clearly shows partial and highly variable FEC reductions in the untreated group whereas FEC reductions were almost total in the treated group (Fig. 3, panel A). It should be noticed that FEC reduction was also total for one untreated animal (open squares). In untreated animals, $\text{Efficacy}(\%)$ ranged from 0% (one heifer) to
approximately 95% (one heifer), with intermediary values from around 30% to 80% in the four other untreated animals (Fig. 3, panel B). It should be noted that the heifer exhibiting no efficacy (Fig. 3, star symbols) was the one for which plasma and faecal exposures were the lowest (2.1 ng*day/mL and 89.5 ng*day/g), while the highest efficacies among untreated cattle were observed for the two heifers exhibiting the highest plasma and faecal IVM exposures (Fig. 3, open square and triangle).

In treated animals, $Efficacy(\%)$ was about 95% at day 30, but the time development of $Efficacy(\%)$ showed some variability. This variability can be exemplified when considering the efficacy level of 80%, which was achieved in the four treated and two untreated animals: the times to reach this level were 3 days for three heifers (two treated, one untreated) and 9 days for the three others (two treated, one untreated).

3.3. Modelling of the exposure – efficacy relationship

In order to better characterize the relationship between efficacy and IVM exposure, individual IVM efficacies at day 30 were plotted against the corresponding plasma or faecal $AUC_{IVM}$ (Fig. 4). The observed data were fitted using the sigmoid Emax model described by Equation 3, and the estimated parameters of the fitting are presented in Table 2. This model allowed us to compute the plasma or faecal $AUC_{IVM}$ corresponding to selected levels of $Efficacy(\%)$, that are presented in Table 3. By dividing these $AUC_{IVM}$ by a standard duration of exposure of 21 days, we computed the corresponding average plasma or faecal concentrations associated with the selected levels of $Efficacy(\%)$. Inspection of Table 3 indicates that an average plasma IVM concentration over 1.2 ng/mL maintained for 21 days was fully efficacious ($Efficacy(\%)_{day30}>90\%$), while lower concentrations between 0.1 and 0.6 ng/mL were associated with efficacies ranging from 20% to 80%.
4. Discussion

The present experiment confirms that the natural grooming behaviour of cattle accounts for the pharmacokinetic disposition of topical MLs and that allo-licking allows significant exchanges of IVM between animals in a group (Bousquet-Mélou et al., 2004). More importantly, we report here the consequences of this phenomenon in terms of anthelmintic efficacy, showing that an untreated animal may be partially or totally cured from an experimental parasitic challenge. This raises several issues related to the design of comparative clinical trials and to the possible drug underexposure, a claimed risk factor favouring the emergence of resistance.

The design of the present experiment was selected to represent a worst-case scenario for ML treatment namely a situation where only a part (40%) of a herd is treated. Due to the apportionment of the total administered dose \( (4 \times 500 \, \mu g/kg) \) between the different members of the group, a possible underexposure was anticipated in both treated and untreated animals. In the present experiment, all treated animals were in fact sufficiently exposed to obtain maximal IVM efficacy, despite the fact that they shared part of their dose with their untreated congeners as a result of allo-licking activity. It should be pointed out here that expressing efficacy as an average \( FEC_{\text{reduction}}(\%) \) over a time period (Equation 2) led to maximal values at 30 days lower than 100\%, even if daily FEC reductions could reach 100\% during the last days (data not shown). This was due to the integration in the calculation of the progressive increase of FEC reduction during the first days post-administration (Fig.3). Moreover, taking into account the time development of the reduction in the FECs highlighted its clear slowing down in half of the treated cattle (time to reach 80\% efficacy increasing from 3 days to 9 days), which might be attributed to the fact that IVM loss by allo-licking occurred, mainly during the first days post-administration.
The average plasma IVM exposure values (AUC_{IVM}) of treated animals (121±43 ng.day/mL) was similar to those reported in a trial (Gayrard et al., 1999) in which all the animals were treated (115±43 ng.day/mL), and in a trial (Bousquet-Mélou et al., 2004) where two treated cattle were pooled with six non-treated congeners (81±27 ng.day/mL). In addition, by using the sigmoid Emax model to describe the exposure-efficacy relationship, we evaluated the critical plasma IVM exposure corresponding to 90% efficacy to 25 ng.day/mL (Table 3). This value is much lower than those we observed both in the treated cattle of the present study (78-181 ng.day/mL, Table 1) and in a previous experiment where two out of eight cattle had been treated (61-100 ng.day/mL, (Bousquet-Mélou et al., 2004). These results indicate that it is unlikely that under field conditions allo-licking will be responsible for a loss of IVM anthelmintic efficacy in treated cattle. In contrast, plasma IVM exposure in untreated cattle ranged from 2.1 to 28.4 ng.day/mL in the present experiment, in agreement with the range of plasma IVM exposures [3-43 ng.day/mL] observed in the six untreated cattle from the same previous experiment (Bousquet-Mélou et al., 2004). Such plasma IVM exposures cover the range from 10-20% to maximal efficacy (Table 3). Thus the population at risk of underexposures leading to partial anthelmintic efficacy appears to be the untreated rather than the treated animals. This merits attention when considering the potential factors responsible for reduced anthelmintic efficacies observed in the field, which as recently pointed out (El-Abdellati et al., 2010), should not be systematically attributed to resistant parasites. However, as it has been shown that an exposure of 14 ng.day/mL was able to initiate a process of resistance selection (Van Zeveren et al., 2007), our results are also of relevance to the discussion concerning the potential emergence of anthelmintic
Many comparative clinical trials have been carried out to compare, in the same setting, different topical formulations or to compare topical versus non-topical formulations. Such trials can be severely biased if the two groups of animals are not totally separated. For example, a slow release bolus of fenbendazole was compared to a PO formulation of doramectin and a control non-treated group (Houffschmitt et al., 2003). After treatment, all the cattle of the three groups were turned out on the same pasture. It was shown that the bolus performed better in terms of FEC reduction, with no significant difference between the control and the doramectin groups at day 56 (mean±SD of FECs were 10±21% of pre-treatment FECs for the fenbendazole group, 70±111% for the PO doramectin group and 80±95% for the control group). Considering the results of the present experiment, these data could be interpreted as follows: 1) the bolus group was fully exposed to fenbendazole and likely to a fraction of the doramectin doses, thus contributing to its better activity, 2) the anthelmintic activity in the doramectin group was declining at day 56 as the claim warrant efficacy for a lower period, 3) the control group was exposed to doramectin thanks to allo-licking activity in some animals, thus promoting a partial activity in these animals and contributing to the large inter-individual variability of FECs and to a mean not different from the doramectin group. Barber and Alvinerie (2003) drew attention to this possible cross-contamination during a comparative clinical trial, where individuals from the treated groups (four MLs administered as PO) and from the control group, were put together in the same paddocks. They showed that 7 days after treatment, the faeces of 80% of the animals in the control group contained detectable amounts of at least one ML, with 55% of the animals having two or more MLs. At the same time, they observed that post-treatment FECs were reduced in the
Emergence of anthelmintic resistance is another issue for PO formulations because underexposure of worms is often quoted as one of the risk factors (Smith et al., 1999), even if “underexposure” was classically referring to under-dosing of treated animals rather than the “unexpected dosing” of untreated animals. In the present experiment, we assessed the range of plasma IVM concentrations associated with a partial efficacy, i.e. a situation putatively able to develop a selective pressure by eliminating the most susceptible subpopulations of worms. In our conditions of worm load, we estimated this possible selection window between 0.1 and 1 ng/mL of plasma IVM concentrations. Above 1 ng/mL, efficacy tends to be maximal (EC₉₀=1.2 ng/mL) and under 0.1 ng/mL (EC₂₀=0.114 ng/mL) anthelmintic efficacy is probably not sufficient to eliminate all susceptible worms. The question of what is the most dangerous exposure level to promote anthelmintic resistance, i.e. to promote selection for parasites possessing genes that confer survival fitness, was addressed using a mathematical model and it was shown that no simple recommendation could be made to reduce the selection pressure for anthelmintic resistance (Smith et al., 1999). Indeed, the extent of the selection window could not be a simple drug property but more probably depends on a complex interaction between drug exposure and the genetic status of the initial worm population.

Nevertheless, the most important recent concept in preventing the development of anthelmintic resistance does not seem to be related to dosing rate but rather to the concept of refugia, i.e. a proportion of the worm population that is not exposed to the drug (Kaplan, 2004). In this context, our results question the use of pour-on formulations of anthelmintic drugs for the development of targeted selective treatment in cattle as a strategy for maintaining refugia (Hoglund et al., 2009; Gaba...
et al., 2010). Indeed, such strategy implies the co-habitation of treated and untreated animals, which is precisely the situation at risk for the occurrence of underexposures consecutive to licking-driven drug transfer in the untreated ones.

To conclude, the present experiment confirms that the disposition of macrocyclic lactones administered as PO formulations is influenced by the social behaviour of cattle, explaining a poor inter-individual reproducibility of plasma exposure. In terms of risk of underexposure of worms to the drugs, the worst situation seems when only a proportion of the animals within a herd are treated, leading the untreated animals to have plasma drug exposures associated with a partial anthelmintic efficacy and corresponding to a possible anthelmintic resistance selection window.
Acknowledgments

The authors wish to express their thanks to Pr. J. Vercruysse from Ghent University who kindly provided the infective larvae of both parasite species. The authors also thank J.F. Sutra for technical assistance.
References


Legends of the figures

**Fig. 1.** Semi-logarithmic plot of IVM plasma concentration-time profiles in four treated (filled symbol) and six non-treated (open symbol) heifers over a 40-day period. The four treated heifers received a single 500 µg/kg topical administration of IVM. Similar slopes for the plasma terminal phases were observed in treated and untreated animals.

**Fig. 2.** Semi-logarithmic plot of IVM faecal concentration-time profiles in four treated (filled symbol) and six untreated (open symbol) heifers over a 40-day period. The four treated heifers received a single 500 µg/kg topical administration of IVM. Similar slopes for the faecal terminal phase were observed in treated and untreated animals.

**Fig. 3.** Time development of IVM anthelmintic activity in ten heifers including four treated animals (filled symbols) and six untreated animals (open symbols). Panel A: individual FEC expressed as percent of pre-treatment values are presented over a 30 days period. Panel B: at each day, efficacy (from 0 to 100%) represents the average cumulated FEC\textsubscript{red}(%) from time 0 (the time of IVM administration) to that day.

**Fig. 4.** Average IVM efficacy over 30 days versus plasma (panel A) or faecal (panel B) IVM exposure (AUC\textsubscript{IVM} from time 0 to the last quantifiable plasma concentration). Efficacy was assessed from daily faecal egg counts (FECs) and was expressed as a percentage (from 0 to 100%) corresponding to the average efficacy over the first 30 days post IVM administration.
Table 1
Descriptive statistics for plasma and faecal IVM exposure in a group of ten heifers of which four were treated with a pour-on dose of IVM (500 µg/kg) and six were untreated but housed in the same yard.

<table>
<thead>
<tr>
<th></th>
<th>Plasma exposure</th>
<th>Faeces exposure</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>ng*day/mL</td>
<td>ng*day/g</td>
</tr>
<tr>
<td><strong>Mean (SD)</strong></td>
<td><strong>Min-Max</strong></td>
<td><strong>Min-Max</strong></td>
</tr>
<tr>
<td><strong>CV%</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>11.3 (11.1)</td>
<td>365 (340)</td>
</tr>
<tr>
<td></td>
<td>2.1 - 28.4</td>
<td>89.5 - 899</td>
</tr>
<tr>
<td></td>
<td>98.4</td>
<td>93.2</td>
</tr>
<tr>
<td>Treated</td>
<td>121 (43.0)</td>
<td>2458 (1063)</td>
</tr>
<tr>
<td></td>
<td>77.8 - 181</td>
<td>1676 - 3998</td>
</tr>
<tr>
<td></td>
<td>35.4</td>
<td>43.2</td>
</tr>
</tbody>
</table>

Exposures are expressed as area under the concentration curve computed from time 0 to the last measured concentration.
Table 2
Pharmacodynamic parameters of a sigmoid Emax model describing the exposure versus efficacy relationship.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Units</th>
<th>Plasma</th>
<th>Faeces</th>
</tr>
</thead>
<tbody>
<tr>
<td>Emax</td>
<td>(%)</td>
<td>93.3</td>
<td>95.5</td>
</tr>
<tr>
<td>AUC$_{50}$</td>
<td>ng*days/mL (plasma)</td>
<td>4.56</td>
<td>173.6</td>
</tr>
<tr>
<td></td>
<td>or ng*days/g (faeces)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Gamma</td>
<td>No unit</td>
<td>1.94</td>
<td>1.64</td>
</tr>
</tbody>
</table>

Emax is the predicted maximal efficacy. The estimated Emax was not 100% due to the way efficacy was expressed (see Material and Methods). AUC$_{50}$ is the AUC corresponding to Emax/2. Gamma is the slope of the exposure versus efficacy relationship.
Table 3
Average plasma or faecal concentrations of IVM corresponding to different percentages of anthelmintic efficacy.

<table>
<thead>
<tr>
<th>Efficacy (%)</th>
<th>Plasma exposure</th>
<th>Faecal exposure</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>AUC (ng*day/mL)</td>
<td>Concentration (ng/mL)</td>
</tr>
<tr>
<td>10</td>
<td>1.6</td>
<td>0.076</td>
</tr>
<tr>
<td>20</td>
<td>2.4</td>
<td>0.114</td>
</tr>
<tr>
<td>50</td>
<td>5.0</td>
<td>0.238</td>
</tr>
<tr>
<td>80</td>
<td>11.6</td>
<td>0.552</td>
</tr>
<tr>
<td>90</td>
<td>25.0</td>
<td>1.190</td>
</tr>
</tbody>
</table>

The concentrations were obtained from the corresponding (plasma or faeces) AUC predicted by the sigmoid Emax model, scaled by a standard duration of 21 days.
Fig. 1.
Fig. 2.
Fig. 3.

A

0 5 10 15 20 25 30
Time (day)

FEC (%)

B

0 5 10 15 20 25 30
Time (day)

Efficacy (%)
Fig. 4.