



Adenovirus Genome in the Placenta: Association with Histological Chorioamnionitis and Preterm Birth

Effrosine A Tsekoura, Anastasia Konstantinidou, Sofia Papadopoulou, Stavros Athanassiou, Nicholas Spanakis, Dimitrios Kafetzis, Aris Antsaklis, Athanassios Tsakris

► To cite this version:

Effrosine A Tsekoura, Anastasia Konstantinidou, Sofia Papadopoulou, Stavros Athanassiou, Nicholas Spanakis, et al.. Adenovirus Genome in the Placenta: Association with Histological Chorioamnionitis and Preterm Birth. *Journal of Medical Virology*, 2010, 82 (8), pp.1379. 10.1002/jmv.21820 . hal-00552416

HAL Id: hal-00552416

<https://hal.science/hal-00552416>

Submitted on 6 Jan 2011

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Adenovirus Genome in the Placenta: Association with Histological Chorioamnionitis and Preterm Birth



Journal:	<i>Journal of Medical Virology</i>
Manuscript ID:	JMV-09-1419.R3
Wiley - Manuscript type:	Research Article
Date Submitted by the Author:	11-Mar-2010
Complete List of Authors:	Tsekoura, Effrosine; University of Athens, Attikon Hospital, Third Department of Paediatrics Konstantinidou, Anastasia; University of Athens, Medical School, Department of Pathology Papadopoulou, Sofia; University of Athens, Medical School, Department of Microbiology Athanassiou, Stavros; University of Athens, Medical School, First Department of Obstetrics and Gynaecology Spanakis, Nicholas; University of Athens, Medical School, Department of Microbiology Kafetzis, Dimitrios; University of Athens, Medical School, Second Department of Paediatrics Antsaklis, Aris; University of Athens, Medical School, First Department of Obstetrics and Gynaecology Tsakris, Athanassios; University of Athens, Medical School, Department of Microbiology
Keywords:	adenovirus , histologic chorioamnionitis, placenta, preterm birth , viral infection



1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

**Adenovirus Genome in the Placenta: Association with Histological
Chorioamnionitis and Preterm Birth**

**Effrossine A. Tsekoura,^{1,2} Anastasia Konstantinidou,³ Sofia Papadopoulou,¹
Stavros Athanassiou,⁴ Nicholas Spanakis,¹ Dimitrios Kafetzis,⁵ Aris Antsaklis,⁴
and Athanassios Tsakris^{1*}**

¹Department of Microbiology, Medical School, University of Athens, Athens, Greece

²Third Department of Paediatrics, University of Athens, Attikon Hospital, Athens, Greece

³Department of Pathology, Medical School, University of Athens, Athens, Greece

*⁴First Department of Obstetrics and Gynaecology, University of Athens, Alexandra Maternity
Hospital, Athens, Greece*

*⁵Second Department of Paediatrics, University of Athens, P. & A. Kyriakou Children's Hospital,
Athens, Greece*

Short title: Adenovirus in the placenta and preterm birth

*Correspondence to: Athanasios Tsakris, Department of Microbiology, Medical School,
University of Athens, 115 27 Athens, Greece. E-mail: atsakris@med.uoa.gr

Adenovirus is isolated frequently from the amniotic fluid and has been implicated in severe neonatal infections. A case control study was carried out to examine the association of detection of adenovirus in placentas with preterm birth and histological chorioamnionitis. Placentas from preterm and full term deliveries were collected prospectively. Preterm cases were divided into three subgroups according to the gestational age. PCR was carried out on placental tissues for the detection of adenovirus genome. Placentas were evaluated histologically for the presence of chorioamnionitis. Chi-square and odds ratios (OR) were used to determine if detection of adenovirus is associated with preterm birth and histological evidence of inflammation. Seventy-one preterm and 122 full term placentas were studied. Adenovirus genome was detected in 29 (40.8%) of preterm cases and in 25 (20.5%) of the full term controls (OR =2.6; 95% CI, 1.4-5.1; $P=0.002$). Detection of adenovirus in preterm placentas was significantly higher compared to full term particularly in the lower gestational age. Detection of adenovirus in placenta followed the seasonal variation of adenovirus infections. Thirty-seven preterm and 21 full term placentas were also selected for paraffin inclusion and histological examination. Chorioamnionitis was present more frequently in preterm adenovirus-positive placentas compared to preterm adenovirus-negative placentas (75% vs 36%; $P=0.03$) as well as compared to term adenovirus-positive placentas (75% vs 19%; $P=0.003$). This study demonstrates that adenovirus infection of the placenta is associated strongly with histological chorioamnionitis and preterm birth.

39

Keywords: adenovirus; histological chorioamnionitis; placenta; preterm birth; viral infection

INTRODUCTION

Since Knox and Hoerner [1950] first recorded an association between infection and preterm birth, there has been extensive evidence associating inflammation and infection to spontaneous preterm birth, especially when this occurs at less than 28 weeks of gestation [Goldenberg et al., 2003; Iams, 2003, Steer, 2005; Romero et al., 2007]. Various bacterial and viral agents have been linked to the pathogenesis of preterm birth. Bacteria recovered from the amniotic cavity following preterm delivery include *Gardnerella vaginalis*, *Neisseria gonorrhoeae*, *Chlamydia trachomatis*, *Mycoplasma hominis*, *Ureaplasma urealyticum*, group B *Streptococcus*, *Bacteroides* spp., and *Trichomonas vaginalis* [Goldenberg et al., 2003; Romero et al., 2007]. Despite the fact that viral infection during pregnancy may prove to be devastating, evidence based etiology on the role which specific viral infections may play in preterm birth still remains rare. A particularly interesting finding is that adenovirus, a double-stranded DNA virus associated commonly with conjunctivitis and upper respiratory tract infections, is detected more frequently in amniotic fluid samples in comparison to other viruses such as cytomegalovirus, herpes simplex virus, enteroviruses and parvovirus [Van den Veyer et al., 1998; Baschat et al., 2003a]. Congenital adenovirus infections have indeed been associated with a variety of adverse effects on the fetus ranging from complications such as pleural effusion, echogenic liver foci to more severe complications including myocarditis and central nervous system anomalies [Meyer et al., 1985; Baschat et al., 2003b].

In several case reports intrauterine adenovirus infections in preterm and full term neonates have been reported to result in a fatal outcome [Towbin et al., 1994; Montone et al., 1995; Ranucci-Weiss et al., 1998]. Perinataly-acquired systemic adenovirus infections in neonates have also been described [Abbondanzo et al., 1989; Matsuoka et al., 1990; Abzug et al., 1991; Pinto et al., 1992; Rieger-Fackeldey et al., 2000]. Adenovirus has also been detected in tracheal aspirates from preterm neonates, within the first week of life, thus suggesting a possible mother to neonate vertical transmission [Couroukli et al., 2000; Prosch et al., 2002].

Several adenovirus serotypes are known to exhibit tropism for the genital tract, causing cervical infection in women and urethritis in men [Lavery et al., 1977; Swenson et al., 1995; Arnebrg et al., 1997; Bradshaw et al., 2002; Bradshaw et al., 2006]. Finally the expression of coxsackie–adenovirus receptor (CAR) in the villous trophoblast cells, which is a prerequisite for adenoviral infection of the placenta, has been demonstrated recently [Koi et al., 2001a; Koi et al., 2001b].

In spite of all these observations a definitive association between adenovirus placental infection and an adverse reproductive outcome has not been established. The purpose of this study was to investigate in a case-control study, the detection of adenovirus in placentas in conjunction with preterm birth and histological evidence of inflammation.

MATERIALS AND METHODS

Study design

During a twelve month period (1 January 2005 – 31 December 2005) all placental specimens from spontaneous preterm deliveries were collected prospectively and consecutively, at the time of delivery at the Alexandra University Hospital, Athens, Greece. Inclusion criteria consisted of women who carried singleton pregnancies and delivered with a preterm birth secondary to premature rupture of membranes, premature labour, or cervical insufficiency. Pregnancies with congenital abnormalities of the fetus were excluded. Preterm cases were divided in three subgroups according to gestational age (subgroup A: early preterm \leq 29wks; subgroup B: intermediate preterm=30-33 wks; subgroup C: late preterm=34-36 wks). Gestational age was calculated by dates and by an ultrasound scan. For each preterm case, one or two full term specimens (\geq 37 weeks gestational age) were also collected consecutively at the same approximate day with preterm placentas ($<$ 37 wks gestational age). For both groups various demographic characteristics, including maternal age, race, obstetric history, estimated gestational

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

age at delivery and fetal sex, were collected by the primary investigator (ET). The study obtained ethics committee approval and informed consent by all patients.

Placenta samples

A total of 193 placenta samples, including part of the chorionic plate and chorionic villi, were stored at -80°C for adenovirus genome detection. Before DNA extraction placenta samples were washed in order to remove maternal blood from the intervillous space. In 58 cases, the peripheral membranes were also sampled, then fixed in a 10% buffer formalin dilution and embedded in paraffin for pathological examination.

Placenta DNA extraction and polymerase chain reaction (PCR)

PCR was used for the detection of adenovirus genome in the placenta tissue. Placental DNA extraction and PCR analysis were blinded to the gestational age of the sample. DNA extraction on both case and control tissues was carried out using the Wizard Genomic DNA Purification Kit (Promega Corporation, Madison, Wisconsin, USA) according to the protocol provided by the manufacturer. In order to verify that adequate amounts of DNA were extracted from all samples, a control PCR amplification of the beta-actin gene was performed on all specimens. Adenovirus genome detection were performed using primers (hexAA1885: 5'-GCC GCA GTG GTC TTA CAT GCA CAT C-3'; hexAA1913: 5'-CAG CAC GCC GCG GAT GTC AAA GT-3') and cycling parameters as describe previously [Allard et al., 1990]. With this method DNA of almost all human adenovirus types derived from a wide range of clinical samples can be detected. All specimens were tested both neat and diluted by 10⁻¹ in order to verify the absence of inhibition. Positive and negative controls were included in all PCR experiments and genetic material was extracted twice from each sample. In order to confirm the specificity of PCR products, the expected 308-bp adenovirus DNA amplicons were purified by a Qiaex gel extraction kit (Qiagen, Chatsworth, California, USA) and used as templates for sequencing of both strands with the ABI PRISM 377 DNA sequence analyzer (Perkin Elmer, Applied Biosystems Division, Foster City, California, USA).

Pathological examination

Thirty-seven preterm birth and 21 control placentas which contained part of the peripheral membranes were collected for histological examination of chorioamnionitis. Samples from the peripheral membranes stained with routine hematoxylin-eosin were blinded and evaluated by light microscopy to diagnose histological chorioamnionitis. The presence of neutrophils in the membranes (chorion and/or amnion) was used to define chorioamnionitis.

Statistical analysis

Variables entering the study were gestational age (preterm versus full term placentas), maternal age, the presence of adenovirus genome (PCR positive versus PCR negative), seasonal distribution and the presence of chorioamnionitis (positive versus negative). Differences between subgroups were evaluated using chi-square test. A P value < 0.05 was considered as statistically significant. Odds ratios (OR) and their respective 95% confidence intervals (CI) were calculated in order to estimate the magnitude of association between adenovirus genome detection and gestational age. Logistic regression was used to adjust for possible confounders, such as maternal age, in the detection of adenovirus. A statistical analysis was carried out using the statistical package STATA 8.0.

RESULTS

Characteristics of samples

During the study period 233 placenta samples were collected prospectively (98 preterm and 135 full term samples). All spontaneous preterm deliveries were included. From preterm placenta samples 48 were classified as late preterm (subgroup A), 30 as intermediate preterm (subgroup B) and 24 as very preterm (subgroup C). Of those, 28 late preterm, 3 intermediate preterm and 13 full term samples were excluded from the study because the inclusion criteria were not fulfilled, or due to inadequate sampling or storage procedure. Thus, 193 placenta samples (71 preterm and 122 full term placentas) which met the criteria were evaluated. From preterm placenta samples 20

145 were late preterm (subgroup A), 27 intermediate preterm (subgroup B) and 24 very preterm
146 (subgroup C).

147 Mean maternal age at delivery was 29.7 ± 5.6 years. Mean maternal age at preterm deliveries
148 was 29.2 ± 5.4 years, while in the full term controls was 30.8 ± 5.8 years ($P > 0.05$). The two
149 groups had similar demographic features and between preterm and full term cases there were no
150 statistically significant differences as far as maternal age at birth, previous fetal loss or therapeutic
151 abortions, previous gestations, nationality of mothers and fetal sex ($P > 0.05$).

152 **Molecular assays**

153 Adenovirus genome was detected in 54 (28.0%) of the total samples. From the 71 placenta
154 tissues of preterm birth, 29 (40.8%) were found positive for adenovirus genome, whereas from
155 the 122 placenta tissues of full term controls only 25 (20.5%) samples were positive for
156 adenovirus by PCR ($P = 0.002$; Table 1). The probability of detecting adenovirus genome in
157 preterm cases was estimated to be 2.7 times higher compared to term controls (OR=2.7, 95% CI,
158 1.4-5.1). A similar association appeared when the subgroups of very and intermediate preterm
159 placentas were compared to full term controls ($P = 0.026$; OR =2.8; 95% CI, 1.1-7.0 and $P = 0.026$;
160 OR =2.7; 95% CI, 1.1-6.5, respectively; Table 1). In the subgroup of the late preterm however,
161 this association was no longer significant statistically ($P = 0.056$; OR=2.6, 95% CI= 0.9-7.0; Table
162 1). It is also noteworthy that adenovirus detection was equally distributed among the subgroups of
163 preterm placentas (Table 1).

164 Sequencing results of all adenovirus PCR-positive cases showed that the amplified sequences
165 corresponded to the expected adenovirus hexon genome. Comparative genetic variability revealed
166 that the amplified hexon gene sequences had similarities ranging from 97.7 to 100%. This
167 variability between sequences almost excluded contamination events and confirmed validity of
168 the study. Alignments of the amplified adenoviral sequences have also shown that most of the
169 sequences corresponded to human adenovirus types of species C (n=41), while smaller numbers
170 of them corresponded to human adenovirus of species B (n=8) and species A (n=5). The

171 distribution of the adenovirus species did not differ between cases and controls. Adenoviruses
172 that belonged to other species were not amplified.

173 Multivariate logistic regression used to adjust gestational age for maternal age, showed that
174 maternal age of full term or preterm cases did not affect significantly the detection of adenovirus
175 genome ($P=0.38$). Adenovirus infection, among both preterm birth cases and controls, was more
176 frequently detected during the spring months (March to June) following the seasonal distribution
177 of adenovirus respiratory infections (Figure 1).

178 Histopathology

179 Table 2 presents results of histological chorioamnionitis among 37 preterm and 21 term
180 placenta samples. A statistically significant difference in the presence of chorioamnionitis was
181 found between preterm and full term placentas (49% vs 19%; $P=0.025$). Within the group of
182 preterm placentas, histological chorioamnionitis was significantly more common among
183 adenovirus PCR-positive samples than among adenovirus-negative samples (75% vs 36%;
184 $P=0.026$). Also, within the total adenovirus PCR-positive placentas, chorioamnionitis was more
185 frequent in preterm compared to full term samples (75% vs 19%; $P=0.003$). However, among
186 adenovirus PCR-negative placentas, the pathological findings did not differ significantly between
187 preterm and full term samples (36% vs 20%; $P=0.488$).

189 DISCUSSION

190 In the present case-control matched study, the detection rate of adenovirus genome in tissues
191 from preterm placentas was compared with healthy full term cases. Although previous studies
192 indicated that adenovirus is related to adverse pregnancy outcomes [Van den Veyer et al., 1998],
193 this is the first prospective cohort study that documents an association between the presence of
194 adenovirus genome in the placenta and spontaneous preterm birth. Baschat et al. [2003] reported
195 an association between adenovirus isolation from the amniotic fluid and the presence of fetal
196 echogenic liver lesions or neural tube defects, suggesting that adenovirus can be a fetal pathogen

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

not recognized previously. Severe adenovirus infection in preterm neonates has been documented in several case reports. Abbondanzo et al. [1989] described fatal adenovirus pneumonia in a preterm neonate born at 25 weeks gestation. In this study the autopsy findings revealed that alveolar and bronchiolar lining cells contain frequently intranuclear inclusions and an overwhelming adenovirus infection was confirmed by *in situ* DNA hybridization. There is also evidence that viral acquisition from the mother, perhaps via the genital tract, is an important mode of adenovirus transmission [Abzug et al., 1991]. Molecular studies revealed that adenovirus serotypes 19 and 37 have a tropism for the genital tract [Arnerg et al., 1997]. In addition adenovirus is recognised as a rare cause of cervical infection or urethritis in men [Swenson et al., 1995; Lavery et al., 1977; Bradshaw et al., 2002; Bradshaw et al., 2006]. Couroucli et al. [2000] isolated adenovirus from the tracheal aspirates of preterm neonates within the first week of life suggesting a possible vertical transmission from the mother to the neonate. Despite all the previous evidence, the role of adenovirus in preterm birth has not been investigated substantially.

 In the present study, preterm placentas were almost three times more likely to host adenovirus genome. Stratification by gestational age to early, intermediate and late preterm revealed that the earlier the gestational age the stronger the extent of this association, whereas in late preterm cases the association between adenovirus detection and preterm birth was no longer significant. These results are in line with the consensus that the role of infection in the etiopathogenesis of preterm labor pertains to the lower gestational age [Goldenberg et al., 2003; Iams, 2003]. However, it should be pointed out that with the primer sequences that were used some exotic types of adenovirus might have been missed from either cases or controls. It is of note that the PCR protocol in the present study does not use degenerated primers that are able to amplify all adenovirus genotypes [Allard et al., 2001]. It should be also mentioned that some of our full term and preterm samples were excluded from the study mainly due to inadequate sampling or storage procedure. Since the exclusion of these samples was random and not related with the study group we believe that the elimination of these specimens did not influence our results.

The histological findings of the present survey showed that chorioamnionitis is associated with premature birth, as expected. This association remained significant in the subgroup of adenovirus PCR-positive samples, whereas it was no longer significant in the subgroup of adenovirus PCR-negative placentas. These findings reveal a meaningful association between adenovirus infection and histological chorioamnionitis, suggesting an etiological relation. Seropositivity data which would confirm this relation were not available in the present study. In a recent report [Arechavaleta-Velasco et al., 2008] adenovirus IgM antibodies were found more frequently in the serum of 24 women, who delivered preterm and sampled before 20 weeks of gestation. However, this study group was rather small and the sampling period was restricted for any safe conclusions to be drawn regarding the possible etiological role of adenovirus in preterm birth.

The finding of increased adenovirus detection between March and June, following the seasonal variation in respiratory adenovirus infections, supports previous similar observations in samples from amniotic fluid and tracheal aspirates of preterm neonates [Prosch et al., 2002; Baschat et al., 2003a]. In conclusion, this study documents an association between adenovirus insult and preterm labour, possibly by causing chorioamnionitis expanding the spectrum of infectious pathogens which are related traditionally to chorioamnionitis. Further studies are warranted to investigate the pathogenic mechanisms relating adenovirus infection during pregnancy with placental inflammation and premature birth.

No conflict of interest for all authors.

REFERENCES

244

245 Abbondanzo SL, English CK, Kagan E, McPherson RA. 1989. Fatal adenovirus pneumonia in a

246 newborn identified by electron microscopy and in situ hybridization. *Arch Pathol Lab Med* 113:

247 1349-1353.

248 Abzug MJ, Levin MJ. 1991. Neonatal adenovirus infection: four patients and review of the

249 literature. *Pediatrics* 87:890-896.

250 Allard A, Albinsson B, Wadell G. 2001. Rapid typing of human adenoviruses by a general PCR

251 combined with restriction endonuclease analysis. *J Clin Microbiol* 39:498-505.

252 Allard A, Girones R, Juto P, Wadell G. 1990. Polymerase chain reaction for detection of

253 adenoviruses in stool samples. *J Clin Microbiol* 28:2659-2667.

254 Arechavaleta-Velasco F, Gomez L, Ma Y, Zhao J, McGrath CM, Sammel MD, Nelson DB, Parry

255 S. 2008. Adverse reproductive outcomes in urban women with adeno-associated virus-2

256 infections in early pregnancy. *Hum Reprod* 23: 29-36.

257 Arnberg N, Mei Y, Wadell G. 1997. Fiber genes of adenoviruses with tropism for the eye and the

258 genital tract. *Virology* 227:239-244.

259 Baschat AA, Towbin J, Bowles NE, Harman CR, Weiner CP. 2003a. Prevalence of viral DNA in

260 amniotic fluid of low-risk pregnancies in the second trimester. *J Matern Fetal Neonatal Med*

261 13:381-384.

262 Baschat AA, Towbin J, Bowles NE, Harman CR, Weiner CP. 2003b. Is adenovirus a fetal

263 pathogen? *Am J Obstet Gynecol* 189:758-763.

264 Bradshaw CS, Denham IM, Fairley CK. 2002. Characteristics of adenovirus associated urethritis.

265 *Sex Transm Infect* 78:445-447.

266 Bradshaw CS, Tabrizi SN, Read TR, Garland SM, Hopkins CA, Moss LM, Fairley CK. 2006.

267 Etiologies of nongonococcal urethritis: bacteria, viruses, and the association with orogenital

268 exposure. *J Infect Dis* 193:336-345.

- 269 Couroucli XI, Welty SE, Ramsay PL, Wearden ME, Fuentes-Garcia FJ, Ni J, Jacobs TN, Towbin
270 JA, Bowles NE. 2000. Detection of microorganisms in the tracheal aspirates of preterm infants
271 by polymerase chain reaction: association of adenovirus infection with bronchopulmonary
272 dysplasia. *Pediatr Res* 47:225-232.
- 273 Goldenberg RL, Culhane JF. 2003. Infection as a cause of preterm birth. *Clin Perinatol* 30:677-
274 700.
- 275 Iams JD. 2003. The epidemiology of preterm birth. *Clin Perinatol* 30:651-664.
- 276 Knox IC, Hoerner JK. 1950. The role of infection in premature rupture of the membranes. *Am J*
277 *Obstet Gynecol* 59:190-194.
- 278 Koi H, Zhang J, Makrigiannakis A, Getsios S, MacCalman CD, Kopf GS, Strauss JF 3rd, Parry S.
279 2001a. Differential expression of the coxsackievirus and adenovirus receptor regulates adenovirus
280 infection of the placenta. *Biol Reprod* 64:1001-1009.
- 281 Koi H, Zhang J, Parry S. 2001b. The mechanisms of placental viral infection. *Ann N Y Acad Sci*
282 943:148-156.
- 283 Lavery CR, Russell P, Black J, Kappagoda N, Booth N. 1977. Adenovirus infection of the
284 cervix. *Acta Cytol* 21:114-117.
- 285 Matsuoka T, Naito T, Kubota Y, Morita Y, Takei J, Akiyama K, Hata J, Hasegawa A, Sata T,
286 Kurata T. 1990. Disseminated adenovirus (type 19) infection in a neonate. Rapid detection of
287 the infection by immunofluorescence. *Acta Paediatr Scand* 79:568-571.
- 288 Meyer K, Girgis N, McGravey V. 1985. Adenovirus associated with congenital pleural effusion. *J*
289 *Pediatr* 107:433-435.
- 290 Montone KT, Furth EE, Pietra GG, Gupta PK. 1995. Neonatal adenovirus infection: a case report
291 with in situ hybridization confirmation of ascending intrauterine infection. *Diagn Cytopathol*
292 12:341-344.
- 293 Pinto A, Beck R, Jadavji T. 1992. Fatal neonatal pneumonia caused by adenovirus type 35.
294 Report of one case and review of the literature. *Arch Pathol Lab Med* 116:95-99.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

295 Prosch S, Lienicke U, Priemer C, Flunker G, Seidel WF, Kruger DH, Wauer RR. 2002. Human
296 adenovirus and human cytomegalovirus infections in preterm newborns: no association with
297 bronchopulmonary dysplasia. *Pediatr Res* 52:219-224.

298 Ranucci-Weiss D, UerpaiojkitB, Bowles N, Towbin JA, Chan L. 1998. Intrauterine adenovirus
299 infection associated with fetal non-immune hydrops. *Prenat Diagn* 18:182-185.

300 Rieger-Fackeldey E, Aumeier S, Genzel-Boroviczeny O. 2000. Disseminated adenovirus
301 infection in two premature infants. *Infection* 28:237-239.

302 Romero R, Espinoza J, Goncalves LF, Kusanovic JP, Friel L, Hassan S. 2007. The role of
303 inflammation and infection in preterm birth. *Semin Reprod Med* 25:21-39.

304 Steer P. 2005. The epidemiology of preterm labor--a global perspective. *J Perinat Med* 33:273-
305 276.

306 Swenson PD, Lowens MS, Celum CL, Hierholzer JC. 1995. Adenovirus types 2, 8, and 37
307 associated with genital infections in patients attending a sexually transmitted disease clinic. *J*
308 *Clin Microbiol* 33:2728-2731.

309 Towbin JA, Griffin LD, Martin AB, Nelson S, Siu B, Ayres NA, Demmler G, Moise KJ Jr,
310 Zhang YH. 1994. Intrauterine adenoviral myocarditis presenting as nonimmune hydrops fetalis:
311 diagnosis by polymerase chain reaction. *Pediatr Infect Dis J* 13:144-150.

312 Van Den Veyver IB, Bowles N, Carpenter RJ Jr, Weiner CP, Yankowitz J, Moise KJ Jr,
313 Henderson J, Towbin JA. 1998. Detection of intrauterine viral infection using the polymerase
314 chain reaction. *Mol Genet Metab* 63:85-95.

Table 1. Adenovirus genome detection in all preterm and in preterm subgroups according to gestational age vs full term controls.

Gestational age (wks)	PCR-positive <i>n</i> (%)	PCR-negative <i>n</i> (%)	Total	<i>P</i>	Odds ratio	95% CI
Term ≥ 37	25 (20.5 %)	97 (79.5 %)	122			
Preterm < 37	29 (40.8 %)	42 (59.2 %)	71	0.002	2.7	1.4-5.1
Subgroup A: 34-36	8 (40.0%)	12 (60.0%)	20	0.056	2.6	0.9-7
Subgroup B: 30-33	11 (40.7%)	16 (59.3%)	27	0.026	2.7	1.1-6.5
Subgroup C: 29-22	10 (41.7%)	14 (58.3%)	24	0.026	2.8	1.1-7

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

349 Table 2. Occurrence of histological chorioamnionitis in placenta tissues among different study
350 groups.

	Histological choriamnionitis			
	Positive <i>n</i> (%)	Negative <i>n</i> (%)	Total <i>n</i>	χ^2 (<i>P</i> value)
Overall sample (<i>n</i> =58)				
Preterm	18 (49%)	19 (51%)	37 (64%)	
Full term	4 (19%)	17 (81%)	21 (36%)	4.99 (0.025)
Adenovirus PCR-positive group (<i>n</i> =28)				
Preterm	9 (75%)	3 (25%)	12 (43%)	
Full term	3 (19%)	13 (81%)	16 (57%)	8.86 (0.003)
Adenovirus PCR-negative group (<i>n</i> =30)				
Preterm	9 (36%)	16 (64%)	25 (83%)	
Full term	1 (20%)	4 (80%)	5 (17%)	0.48 (0.488)
Preterm group (<i>n</i> =37)				
Adenovirus PCR-positive	9 (75%)	3 (25%)	12 (32%)	
Adenovirus PCR-negative	9 (36%)	16 (64%)	25 (68%)	4.94 (0.026)

351
352
353

FIGURE LEGENDS

Fig 1. Bar graph depicting the seasonal distribution of adenovirus genome detection in preterm and term placentas.

