



HAL
open science

Combinatorial biomarker expression in breast cancer

Emad A. Rakha, Jorge S. Reis-Filho, Ian O. Ellis

► **To cite this version:**

Emad A. Rakha, Jorge S. Reis-Filho, Ian O. Ellis. Combinatorial biomarker expression in breast cancer. *Breast Cancer Research and Treatment*, 2010, 120 (2), pp.293-308. 10.1007/s10549-010-0746-x. hal-00535436

HAL Id: hal-00535436

<https://hal.science/hal-00535436>

Submitted on 11 Nov 2010

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Combinatorial biomarker expression in breast cancer

Emad A. Rakha · Jorge S. Reis-Filho ·
Ian O. Ellis

Received: 3 December 2009 / Accepted: 12 January 2010 / Published online: 28 January 2010
© Springer Science+Business Media, LLC. 2010

Abstract Current clinical management of breast cancer relies on the availability of robust clinicopathological variables and few well-defined biological markers. Recent microarray-based expression profiling studies have emphasised the importance of the molecular portraits of breast cancer and the possibility of classifying breast cancer into biologically and molecularly distinct groups. Subsequent large scale immunohistochemical studies have demonstrated that the added value of studying the molecular biomarker expression in combination rather than individually. Oestrogen (ER) and progesterone (PR) receptors and HER2 are currently used in routine pathological assessment of breast cancer. Additional biomarkers such as proliferation markers and ‘basal’ markers are likely to be included in the future. A better understanding of the prognostic and predictive value of combinatorial assessment of biomarker expression could lead to improved breast cancer management in routine clinical practice and would add to our knowledge concerning the variation in behaviour and response to therapy. Here, we review the evidence on the value of assessing biomarker expression in breast cancer individually and in combination and its relation to the recent molecular classification of breast cancer.

Keywords Breast cancer · Immunohistochemistry · ER · PR · HER2 · Basal markers · Combinatorial expression

Introduction

Breast cancer (BC) is a complex genetic disease characterised by the accumulation of multiple molecular alterations [1, 2]. Routine clinical management of BC relies on well-established clinicopathological factors. Although these factors show strong overall association with patients’ prognosis and outcome, it has become clear that patients with similar features may show distinct outcomes and vary in their response to therapy [3]. For example, it has been shown that approximately one-third of patients with early stage BC develop recurrence [4], whilst a similar proportion of node positive patients remain free of distant metastases [5]. In an attempt to improve BC classification and to stratify patients into well-defined prognostic categories that can be used in management decision, these well-established prognostic factors have been combined to constitute prognostic indices, such as Nottingham Prognostic Index, which provides prognostic significance better than any of its components individually [6, 7]. In order to improve prediction of response to specific agents and to aid tumour classification and overcome the inherent subjectivity involved in histopathology, molecular biomarkers have been introduced. Currently, only hormone receptors (HR), including oestrogen (ER) and progesterone (PR) receptors, and human epidermal growth factor receptor 2 (HER2) are assessed and used in routine clinical practice at least in most centres [8]. Although several additional biomarkers are extensively studied, and some have shown prognostic (estimation of outcome after surgery alone) and predictive (estimation of

E. A. Rakha (✉) · I. O. Ellis
Department of Histopathology, Nottingham University Hospitals
NHS Trust, Nottingham City Hospital, Hucknall Road,
Nottingham NG5 1PB, UK
e-mail: emadrakha@yahoo.com

J. S. Reis-Filho
Molecular Pathology Laboratory, The Breakthrough Breast
Cancer Research Centre, Institute of Cancer Research, Fulham
Road, London, UK

response to therapy) value in the research setting, only few biomarkers are likely to be included in routine clinical use, at least in some centres; these include basal markers and proliferation-related markers.

It has been estimated that all these traditional clinico-pathological and molecular factors, which presently form the basis for determining adjuvant therapy, assign these patients into risk groups at an approximate absolute specificity level of only 10% to achieve an acceptable degree of sensitivity [9]. Therefore, there is an increasing need to improve patients risk stratification and targeting of treatment to those who will truly benefit, thereby avoiding iatrogenic morbidity in those who will not. Most importantly, whilst most predictive markers developed to date have acceptable negative predictive values (i.e. they identify the population of patients who will not benefit from a given therapy), their positive predictive values (i.e. their ability to identify the patients that will certainly benefit from a regimen) are clearly suboptimal. For instance, complete lack of ER expression does identify a group of BC patients that do not benefit from endocrine therapies (i.e. optimal negative predictive value), however, only a fraction of patients whose tumours express ER will benefit from endocrine therapy (i.e. suboptimal positive predictive value) [10]. Improved understanding of the molecular features of BC and the identification of the key genes that underpin the molecular heterogeneity of BC may lead to better prediction of tumour behaviour and treatment response.

Assessment of HR and HER2 in BC provides prognostic and predictive information on response to endocrine therapy and anti-HER2 targeted therapy, respectively. However, the expression of these biomarkers overlap and the prognostic and predictive value of these markers in combination need to be well-defined. Currently, it is recognised that a set of biological markers, rather than a single one, seem to be important to differentiate between a high or low chance for a response to systemic therapy [11]. In addition, recent microarray-based gene expression profiling studies (GEP) have demonstrated that the importance of assessment of key biomarkers in combination, which is expected to improve our understanding of the biology and behaviour of BC and help to tailor treatment [12]. GEP has also indicated that genomic fingerprints may refine prediction of the course of disease and response to adjuvant interventions. Currently, several commercially available prognostic BC tests based on the expression of multiple genes (using transcriptome) are available, including Oncotype DX (21 genes; Genomic Health, Redwood City, California, USA) [13], MammaPrint (70 genes; Agendia BV, Amsterdam, the Netherlands) [14], Theros H/I (2 genes; AvariaDX, Carlsbad, California, USA) [15], and Theros breast cancer index (a combination of Theros H/I and the molecular grade index, AvariaDx, Carlsbad, California, USA) which represent the first

introduction of multigene assays into clinical application. Out of these technologies, only Oncotype DX has been included in the American Society of Clinical Oncology (ASCO) and National Cancer Centre Network (NCCN) guidelines for the management of BC patients [12].

Here, we present an overview of the significance of assessment of expression of biomarkers used in routine BC management and the added value of their combinatorial expression.

Hormone receptors

Oestrogen receptor

The oestrogen receptor (ER) was first identified in the 1960s and subsequent studies have provided the evidence that ER is important in the carcinogenic process, and its inhibition, through endocrine targeting, either directly using oestrogen agonists (Selective ER Modulators) or indirectly by blocking the conversion of androgens to oestrogen (e.g. aromatase inhibitors), forms the mainstay of BC endocrine therapy [9, 16–18]. Therefore, ER status has been used since the mid-1970s in the clinical management of BC both as an indicator of endocrine responsiveness and as a prognostic factor for early recurrence. It has also been reported that ER expression in BC is stable and phenotypic drift from primary to metastatic breast carcinoma is reported to be an exceedingly rare phenomenon [19]. In addition, recent GEP of BC has also indicated that ER is a major determinant of the molecular portraits of BC [20–23]. ER status currently forms part of the UK minimum data set for histopathology reporting of invasive BC and it is routinely determined using a standardised technique [8].

Oestrogen (ER)-positive tumours (ER+) comprise the majority of breast cancers, accounting for up to 75% of all cases. Up to 65% of tumours developing in women aged <50 years are ER+, whereas this figure increases to 80% in women >50 years [24]. Although ER+ tumours are generally well-differentiated, show other less aggressive primary tumour characteristics and are associated with better clinical outcome largely independent of other clinico-pathological variables after surgery [25, 26], long term survival studies have reported that ER status loses its predictive significance and that the long term outcome of ER+ and ER– tumours is not different [10]. In fact, ER status provides limited prognostic information; currently, the major clinical value of determining ER status is to assess the likelihood that a patient will respond to endocrine therapy, and are unlikely to gain additional benefit from adjuvant chemotherapy [27, 28]. Most reports have concluded that ER is probably the most powerful single predictive factor identified in BC [18, 23, 29, 30].

Although ER expression is an accurate negative predictor of response to hormonal treatment, i.e. ER– tumours are unlikely to respond to hormone therapy, it provides limited positive predictive information, given that only approximately 50% of patients with ER+ tumours respond to hormone treatment [18]. It is also documented that a small proportion of ER– cancers respond to hormonal therapy [31, 32]. These observations, in addition to the fact that ER+ tumours comprise a large proportion of BC, demonstrate that ER-positivity per se defines a heterogeneous group of tumours with respect to their risk factors, clinical behaviour and biology [12, 18, 33]. Unsupervised analysis of the transcriptome of BC has revealed that at the transcriptional level, ER+ and ER– tumours are fundamentally different [20, 22, 34]. Furthermore, the type, pattern and complexity of genetic aberrations appear to be different in ER+ and ER– disease [35–37], and it has been observed that the molecular pathways and networks driven by copy number aberrations appear to some extent to be determined by the ER status of a tumour [36, 38].

Progesterone receptor

Progesterone receptor (PR) is an oestrogen-regulated gene and its expression is therefore thought to indicate a functioning ER pathway [39–41].

Progesterone receptor (PR)+ tumours comprise 55–65% of BC. Multiple studies have provided evidence for the prognostic and predictive importance of PR assessment in BC [33, 42–47]. PR+ cancers have been shown to have a better prognosis than PR– tumours, and there are some data to suggest that PR status can help to predict response to hormone treatment, both in patients with metastatic disease [45] and in the adjuvant setting [32, 47–50]. However, it is also important to mention that some authors questioned the value of assessing PR status in BC [18, 51], and in the latest (2009) guidelines published by the National Institute for Health and Clinical Excellence (NICE) in the UK for early and locally advanced BC, it is recommended not to routinely assess PR status in patients with invasive BC (www.nice.org.uk/CG80). The argument was based mainly on the lack of evidence to support PR being of additional predictive over ER status with respect to response to endocrine therapy [10]. It has also been stated that PR positivity hardly exists amongst ER– tumours [51]. However, false ER negativity has been reported in routine practice [52] and strong PR positivity in an apparent ER– case may be an indicator of a false negative ER result.

Combinatorial expression of ER and PR

It is recognised that ER expression is used as the main determinant of response to hormone therapy in BC.

Approximately 40% of ER+ tumours are PR– [33]. Lack of PR expression in ER+ tumours may be a surrogate marker of aberrant growth factor signalling that could contribute to tamoxifen resistance and that ER+/PR– tumours are generally less responsive than ER+/PR+ tumours [33, 48, 53, 54], particularly for Tamoxifen in the metastatic setting [45, 46]. Although some studies have reported that up to 10% of ER– BC are PR+ (ER–/PR+) [55, 56], recent evidence has indicated that this percentage is much lower when more sensitive immunohistochemical detection methods for ER are used or when analysis of ER and PR mRNA levels by quantitative real-time PCR is used [52, 57–59]. The higher frequency of ER–/PR+ tumours in some studies may be due to a false-negative ER assay, very low level ER or to variant ERs not recognised by the antibody, but still capable of stimulating PR expression [60]. In a study of 155,175 women with known joint ER/PR receptor status using data from the NCI's SEER program in the United States, Dunnwald and colleagues [26] reported that the proportion of ER–/PR+ tumours declined over the study period (1990–2000). In a central immunohistochemical analysis of ER and PR from 6,291 patients enrolled in the BIG198 clinical trial, Viale et al. reported that 0.2% of patients displayed an ER–/PR+ profile [61]. In our hands, the percentage of ER–/PR+ tumours reported recently in our routine practice is between 1 and 2% of BC [62 and unpublished data].

When the combinatorial expression of ER and PR are considered, four subgroups are recognised: double HR+ (ER+/PR+), single HR+ (ER+/PR– and ER–/PR+) and double HR– (ER–/PR–). The double positive group, which comprises the majority of tumours (55–65%) [26, 33, 63], shows the best prognosis and a good response to hormonal therapy, and has been used as a feature of the Luminal A class in some of the recent GEP classification systems of BC [22, 23, 64]. It has been reported that 75–85% of tumours with ER+/PR+ phenotype respond to hormonal therapy, whereas less than 10% of ER–/PR– tumours respond [32, 65, 66]. Compared to other subtypes, the double HR+ tumours are also associated with older age, lower grade, smaller size and lower risk of mortality. Dunnwald et al. [41] demonstrated that the associations between ER/PR subgroups and mortality risk are independent of tumours stage, age or grade and that the magnitudes of these relative risks vary amongst different tumour stage and grade.

The double HR– group which comprises the second largest group (18–25%), are more likely to be of grade 3 (approx 85%) and associated with a higher recurrence rate, decreased overall survival and unresponsiveness to endocrine therapy [45, 46, 48, 63, 67–69]. It has also been reported that HR– status is the most important predictive marker concerning response to a preoperative taxane/

anthracycline-based regimen. However, despite the high pathologic complete response rate, survival of patients with this phenotype was reported in several studies to be shorter than for those with HR+ tumours [11]. However, some types of invasive carcinoma that are typically HR–, e.g. adenoid cystic carcinoma and secretory carcinoma, have an excellent prognosis with minimal regional recurrence [70, 71]. This, in addition to other evidence, points towards the heterogeneous nature of the HR– subgroup of BC [12, 72]. This group of tumours corresponds to the vast majority of basal-like, normal breast-like and HER2+ classes in the GEP molecular subtype classification [22, 23].

The significance of BC with a single HR+ phenotype that includes ER+/PR– tumours, which comprise the third largest group of BC (12–17%) [26, 33, 63], and ER–/PR+ (1–2%) is still poorly understood. These tumours may correspond to the Luminal B class in the GEP classification [21–23, 64] and may show frequent expression of other features characteristic of poor prognosis. Interestingly, these tumours are more often of high histological grade, large size, more likely to be aneuploid and show higher expression of proliferation-related genes, EGFR and HER2 than ER+/PR+ cancers [33, 54]. Clinical data regarding metastatic and adjuvant treatment responsiveness suggest that hormone therapy is less effective in the single HR+ tumours than in the ER+/PR+ class [45, 48, 73], with only about 40% responding to hormonal manipulation [32, 65]. However, one study reported that the response rate of ER+/PR– tumours to an aromatase inhibitor is similar to that of ER+/PR+ cancers [74]. In addition, some studies have demonstrated that both single HR+ groups are similar in that they both might have biological characteristics somewhere in between ER+/PR+ and ER–/PR– [33, 75]. Moreover, Dowsett et al. [32] have demonstrated that ER–/PR+ cancers can benefit from endocrine therapy in contrast to ER–/PR– tumours. They concluded that measurement of PR status in ER– patients defines a group of patients that benefit from tamoxifen, but would be excluded from tamoxifen therapy on the basis of ER status alone. As discussed by the authors [32], it is plausible that the ER–/PR+ tumours derive benefit from tamoxifen because they result from false negative ER assessment results.

In a different approach, instead of using positive and negative categories, Goldhirsch and colleagues [76] have used the level of expression of both ER and PR to predict response to endocrine therapy. They reported two categories of HR+ BC; those that express high levels of both ER and PR (ER and PR > 50%) and are highly endocrine responsive, and those that express low levels of either/both receptors (ER or PR < 50% and ER > 10%) and are incompletely endocrine responsive. A third group which shows negative expression for ER and PR (both < 10%) do not benefit from endocrine therapy. Stendahl et al. [43]

have reported that adjuvant tamoxifen improved survival for premenopausal patients with tumours showing >75% PR positivity at which point PR was also independently associated with favourable overall survival. Tumours with lower percentage of PR positivity showed that no similar effect, whilst a gradually increasing tamoxifen effect was observed in tumours with >10% ER+ nuclei. Based on their findings, they concluded that a fractioned rather than dichotomized immunohistochemical evaluation of both ER and PR should be implemented in clinical practice. Furthermore, a meta-analysis of tamoxifen trials showed that women with ER+ tumours derive significant benefit from 5 years of tamoxifen in reducing the odds of recurrence and death, and this benefit is directly proportional to the level of ER, with patients with higher tumour ER levels deriving the greatest benefit from therapy [18].

In summary, there is sufficient evidence to demonstrate that joint ER/PR assessment defines phenotypic groups that have different biological characteristics, including tumour size, grade, stage, patient's outcome and response to therapy. Breast cancers can be ranked from good to worse for ER+/PR+ to ER+/PR– to ER–/PR+ to ER–/PR– and that joint ER/PR expression identifies BC variants better than either independent ER or PR expression [26, 33, 45, 46, 48, 50, 53, 54, 63].

Most GEP studies [20–23] emphasise the importance of HR expression in BC and showed that HR+ tumours constitute a distinct group of tumours that are different from HR– BC or HER2 over-expressing tumours [12]. GEP support the existence of at least two luminal-like subclasses (A and B), and recent studies have implied that rather these differences more probably represent a biological continuum [12, 77, 78] which includes the double positive and single HR+ tumours and also relates to the level of expression of HR as well as other biomarkers within the HR+ tumour class. At one end of the ER+ spectrum, there are the so-called Luminal A tumours which are characterised by high levels of ER and downstream transcriptional targets of ER, other luminal associated markers in addition to low levels of expression of proliferation-related genes, whereas the Luminal B group is characterised by low to moderate expression of ER and other luminal specific genes, but is further distinguished by high expression of proliferation-related genes [22, 23, 79]. It has been reported that Luminal A tumours respond better to hormonal therapy, whilst Luminal B tumours are more often resistant to this therapeutic modality and may benefit from combined endocrine treatment and chemotherapy [80]. However, it is important to mention that to date, there is neither internationally accepted single definition for the luminal subgroups/classes or spectrum [12] nor has the use of ER and PR alone to define them been widely adopted and additional markers including HER2 [12, 64, 81] and

proliferation markers, e.g. ki-67 (MIB1) [80, 82] or genomic-grade index [83], have been adopted by some groups.

HER2

The clinical importance of amplification of *HER2* gene in BC was recognised in 1987 [84]. Numerous subsequent studies found that *HER2* gene amplification/protein over-expression is a predictor of poor prognosis and response for systemic chemotherapy [11, 85–88]. *HER2* protein expression and gene amplification occurs in 13–20% of invasive ductal BC and more than half (~55%) of these cases are HR– [84, 89]. *HER2* expression shows an inverse relationship with both ER and PR expression [90]. The prognostic impact of *HER2* positivity is higher in node-positive compared with node-negative patients. Following the development of a humanised monoclonal antibody against *HER2* (trastuzumab; Herceptin; Genentech, South San Francisco, CA, USA) and clinical trials demonstrating benefit of the use of anti-*HER2* agents in patients with *HER2*+ BC [91–93], the reasons for establishing the *HER2* status in routine clinical practice has changed, since it is a prerequisite for clinical use of trastuzumab in patients with *HER2*+ advanced disease [94] as well as in the adjuvant setting for *HER2*+ early stage BC [92]. In addition to trastuzumab, *HER2* continues to be an important target in the development of a variety of other new cancer therapies, which include small-molecule drugs directed at the internal tyrosine kinase portion of the *HER2* oncoprotein (i.e. lapatinib), and vaccines.

Gene expression profiling (GEP) studies have also demonstrated that *HER2* is one of the key markers in BC as a high proportion of *HER2*+ tumours cluster together in a class which is distinct from HR+/*HER2*– and HR– (basal and normal breast-like) tumours [22]. *HER2* status may also be predictive for other systemic therapies [86]. It has been reported that *HER2* positivity is associated with relative, but not absolute, resistance to endocrine therapies in general [95]. However, this effect may be specific to tamoxifen, but not to oestrogen depletion therapies, such as aromatase inhibitors [96, 97]. Similar to ER, the *HER2* status of BC narrows the pool of candidates eligible for *HER2*-directed therapies, but it does not definitively select those who will respond. Several studies have reported that *HER2* may be a predictive marker of response to anthracycline-based chemotherapy [98–101]. It has also been suggested that *HER2* positivity is predictive of better response to higher dose anthracycline-containing regimens compared with standard regimens [102, 103] and to taxane compared with non-taxane-containing regimens [104, 105], however, the predictive value of *HER2* remains a complex subject and further validation is still required [98].

Combinatorial expression of ER, PR and *HER2*

The results of GEP studies demonstrate that BC is composed of distinct molecular classes largely characterised by well-defined patterns of expression of HR, *HER2* in addition to few other key molecular variables, such as proliferation and basal cell type-related gene alterations [12]. Importantly, these molecular classes showed that potential prognostic and predictive utility. The results of these studies, in addition, the availability of treatment option have emphasised the importance of studying the molecular portraits of BC in concert. Therefore, several attempts to validate and translate these molecular classes into defined groups that can be identified in routine practice have been carried out. Most studies have used a combination of various immunohistochemical (IHC) markers including ER, PR and *HER2* with or without additional markers, such as basal marker and proliferation markers (see below), as IHC surrogates to define the molecular classes initially identified by GEP and to improve our understanding of the prognostic and predictive value of studying these markers in combination. It should be noted that there is a paucity of data on direct comparisons between GEP and immunohistochemical surrogates to define the molecular subgroup of a given case. Furthermore, the stability of some of the molecular subgroups as defined by GEP has also been called into question [106].

Most studies have considered ER or HR positivity regardless of expression of other markers as the most important feature for a tumour to be classified as of Luminal type, whilst HR- and *HER2*-negative (triple negative; TN) phenotype was used to define the basal-like class [64, 107–110]. HR+ luminal tumours which comprise the largest proportion of BC phenotypes are a heterogeneous group where molecular subtyping and consideration of expression of other markers could be of utmost importance and of clinical relevance. Several studies have classified HR+ tumours that are also *HER2*+ as the Luminal B subclass [64, 81, 107, 109, 111], which constitutes approximately 6% of HR+ tumours (3–11%) [64, 82, 112]. This approach is supported by the fact that some Luminal tumours identified in GEP express *HER2* and that HR+/*HER2*+ show poorer outcome than HR+/*HER2*– tumours. However, other authors have classified all *HER2*+ tumours in the *HER2* subclass independent of their HR status [108]. The later approach is also supported by the fact that HR+/*HER2*+ tumours are candidates to specific systemic therapy targeting *HER2*. Importantly, there is evidence to suggest that *HER2*-amplified cases have similar genetic changes [113] and outcome [64, 82, 114] regardless of their HR status.

It has also been reported that some forms of ER+ BC are resistant to chemotherapy [27, 28, 115] and that a

significant proportion of cases do not respond to hormone therapy. Therefore, the addition of other predictive biomarkers, such as HER2 and proliferation markers to ER and PR may help predict response to chemo and endocrine therapy in HR+ tumours regardless of the terminology of Luminal subtypes. This hypothesis was utilised in the construction of the gene set that constitutes the Oncotype DX assay [13]. Preclinical and clinical data suggest that HER2 overexpression confers intrinsic resistance to hormonal treatment in HR+ tumours. This in addition to the adverse prognostic effect of HER2 overexpression may indicate that patients with HR+/HER2+ BC might not derive a benefit from single-agent hormone therapy. Results from randomised clinical trials that combined hormone treatment with targeted anti-HER2 therapy in postmenopausal women with HR+/HER2+ advanced BC indicate that this novel dual-targeting strategy significantly improves outcomes compared with hormone therapy alone. Other studies also suggest that HR+/HER2+ BC might benefit more from anti-HER2 therapy plus chemotherapy [116]. Darb-Esfahani and colleagues [117] have reported that HR+/HER2+ tumours show a good response rate to neo-adjuvant chemotherapy and a favourable prognosis. HR+/HER2– tumours have a good prognosis irrespective of achievement of a pathological complete response, whereas patients with HR–/HER2– and HR–/HER2+ tumours show the worst prognosis, particularly if they do not achieve a pathological complete response. Hayes et al. [105] have also demonstrated that a difference in response to chemotherapy based on HER2 and HR where HER2+ tumours are associated with a benefit from the addition of paclitaxel after adjuvant treatment with doxorubicin plus cyclophosphamide in node-positive BC, whilst HER2–, ER+ tumours, may gain little benefit. Konecny et al. [95] found an inverse correlation between HER2 expression and the level of expression of ER and that in patients with HR+ tumours, HER2+ tumours had statistically significantly lower ER/PR levels than HER2– cancers. Therefore, they suggested that the relative resistance of HER2+/HR+ tumours to hormone therapy is due to reduced ER/PR expression or high proliferation rates rather than positivity of HER2. In fact, there is evidence to support the conclusion that lower ER, lower PR and positive HER2 are associated with lower responsiveness to any type of endocrine therapy and HR+ tumours overexpressing HER2, therefore, require the blockage of the HER2 pathway in addition to oestrogen deprivation.

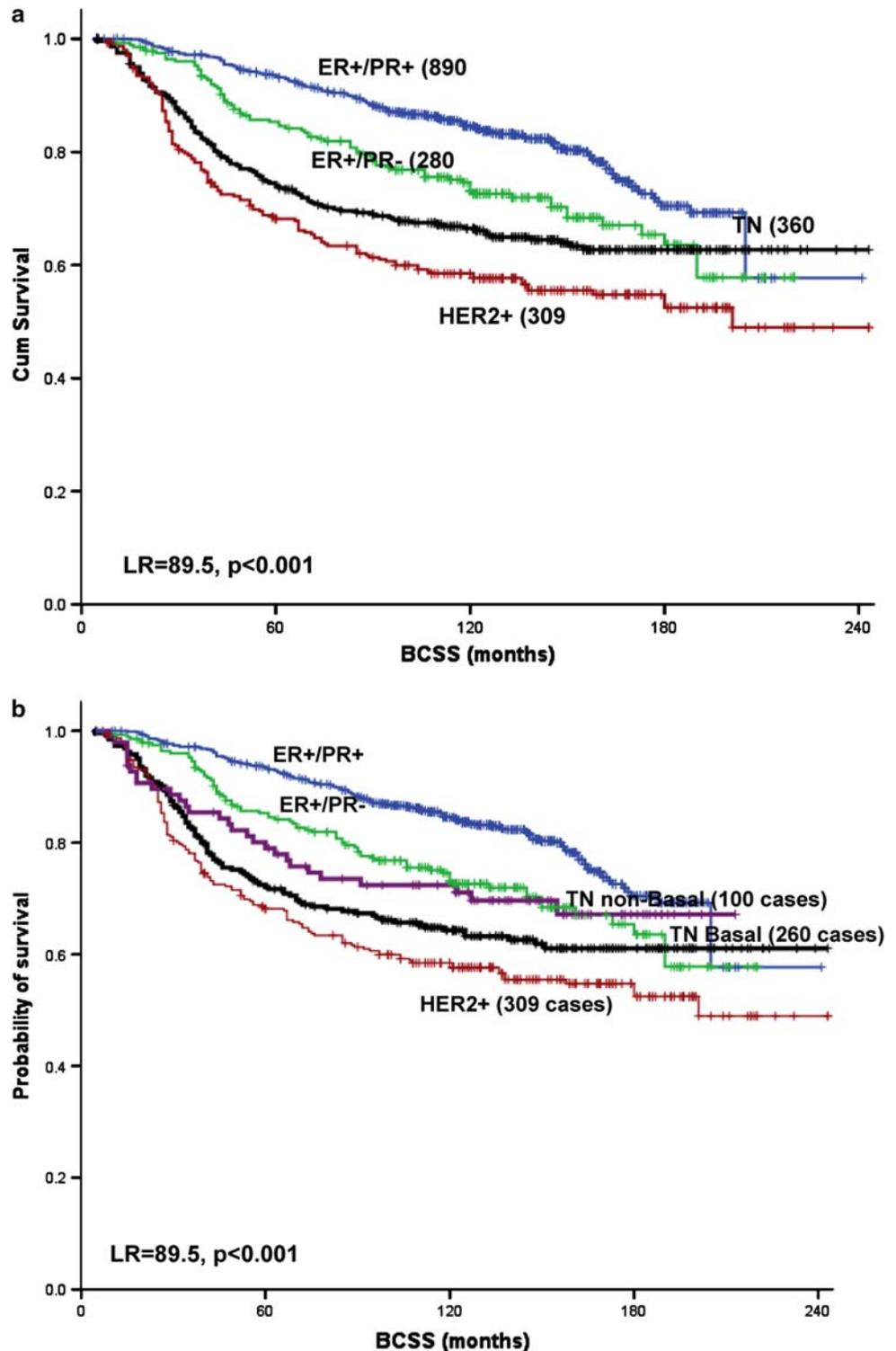
Current evidence indicate that ER+/PR+/HER2– tumours have the best prognosis and show the best response to hormone therapy. Single HR+ HER2– tumours show worse outcome and response to treatment whilst HR+ HER2+ tumours show the worst outcome and response to hormone therapy amongst the HR+/luminal

tumours [32, 118]. Although HR– tumours are usually poorly differentiated (i.e. 85% of HR– tumours are grade 3 in the Nottingham series), show aggressive behaviour and poor outcome and least likely to respond to hormone therapy, these tumours constitute a heterogeneous group that includes two main subclasses: HER+, and HER2– (TN) tumours. TN class can be further divided into basal-like (core basal phenotype) and TN non-basal (null phenotype) [33, 64, 107–110]. These subclasses show significant difference in behaviour and outcome (Fig. 1). The behaviour, outcome and response to therapy of these TN tumours are discussed in detail elsewhere [12, 119, 120]. Other classes that have been described amongst HR– tumours include the molecular apocrine subtype [12, 34], interferon-rich group and the Claudin low group [121].

Combinatorial expression and prediction of endocrine therapy resistance

Currently available antihormone-treatment strategies for ER+ BC consist of either targeting the ER itself through the use of HR selective ER modulators, such as tamoxifen, or by depriving cancer cells of their oestrogenic stimulus through (i) gonadal suppression (in premenopausal women) or (ii) aromatase inhibitors (in the postmenopausal setting). Although the absence of ER expression can predict lack of response to endocrine therapy, the predictive power of positive ER expression is limited by the phenomenon of hormone resistance. This resistance can be either de novo (present before hormone therapy) or acquired during the course of treatment. Two patterns of acquired resistance have also been documented; patients who initially respond and relapse quickly and patients with a sustained response followed by a late relapse. For example, it has been reported that approximately 50% of metastatic breast cancers will display de novo resistance to hormone therapies despite being HR+ and more than one-third of patients with endocrine-responsive, early stage BC, and almost all of those with metastatic disease, will develop hormone resistance during the course of their disease (acquired resistance) despite an initial response to the therapy resulting in disease relapse and progression [122, 123]. Several key mechanisms involved in the process of hormone resistance have been identified [124, 125]. There is, however, evidence that specific markers can be used to identify tumours that exhibit resistance to specific anti-hormone therapy agents. These markers include both conventional markers of endocrine responsiveness (ER and progesterone receptor, PR), receptor tyrosine kinases, such as the HER family of receptors and, in particular, HER2 and EGFR, CDK10 and the insulin-like growth factor-1 receptor (IGF1R), which might be through cross-talk with the ER itself [126–131]. Clinical evidence for the role of

Fig. 1 Correlation between biomarker expression (ER, PR, HER2 and basal marker (b) Ck5/6 and EGFR) and outcome (breast cancer specific survival; BCSS) in the Nottingham series [110, 136]. BCSSP = breast cancer specific survival TN = triple negative (ER/PR/HER2-) TN = triple negative basal (ER/PR/HER2- and Ck5/6 and/or EGFR positive) TN = triple negative non-basal (ER/PR/HER2/CK5/6/EGFR negative)



EGFR (HER1) and HER2 in tamoxifen resistance has come from neoadjuvant trials where EGFR and HER2+ patients have significantly greater response to aromatase inhibitors than to tamoxifen [96]. Tovey et al. [126] have demonstrated that EGFR/HER2/HER3-positivity and/or PR-negativity comprise a high-risk group within ER+ BC patients

that was more likely to show early relapse on tamoxifen. However, this predictive value of de novo resistance lost its significance after 3 years of tamoxifen. In the ATAC trial [50], PR- patients derived greater benefit from initial aromatase inhibitor treatment compared with tamoxifen. However, in the IES trial [132], PR status had no effect on

response when aromatase inhibitors were given as delayed treatment to patients who had been disease free on tamoxifen for 2–3 years. The recent results of the ATAC trial [118] demonstrate that quantitative expression of PR and HER2 status did not identify patients with differential relative benefit from anastrozole over tamoxifen and that time to recurrence is longer for anastrozole than for tamoxifen in all subgroups. The results of the BIG 1–98 clinical trial demonstrated that Letrozole improved disease-free survival compared with tamoxifen regardless of HER2 status [97, 133] or PR status [61]. Therefore, although initial data indicated that PR, EGFR and HER2 expression might be used to predict resistance to hormone therapy in the ER+ BC [48, 50, 126, 132], the evidence available suggest that PR and HER2 are prognostic rather than predictive markers, and that these biomarkers cannot be used to determine which patients would benefit most from aromatase inhibitors or tamoxifen [61, 97, 133, 134].

Basal markers

The potential poor outcome associated with the expression of high molecular weight or ‘basal’ Cks in BC has been known for over two decades [135], and this has been confirmed by numerous other studies that have also demonstrated that the poor prognosis of basal Cks is independent of the expression of HR or HER2 [112, 136–138]. However, the results of GEP studies, which showed that the majority of basal-like class of tumours are characterised by the TN phenotype in addition to expression of basal-associated markers, have emphasised the use of HR and HER2 expression in addition to basal markers to define a class of tumours that has a poor prognosis, lack the benefit of available targeted therapy and shows distinct molecular features (basal-like BC). In addition, it has been reported that there is a relationship between basal Cks expression and BRCA1-associated tumours and that basal Cks expression can predict BRCA1 status, and hence, may benefit from a similar therapeutic strategy [110, 139]. Tumours expressing basal Cks show specific pattern of distant metastasis [110]. The relationship between tumour size and proportion of lymph node positivity and outcome is also less clear amongst tumours that express basal markers [140, 141]. These findings may emphasise the importance of expression of basal markers in BC irrespective of controversial notion of their relationship to their potential cell of origin or their expression pattern in normal breast epithelial cells [142].

Although the number of basal IHC markers used to define basal-like tumours is large and expanding, the most widely used and generally accepted basal markers are Ck5/6 and EGFR; in addition to these markers, Ck17 and Ck14 have also been used in some studies [64, 108, 110, 112, 119, 143]. It has been demonstrated that using basal

markers (Ck5/6 and EGFR) in the TN tumours identifies a biologically and clinically distinct subgroup of TN tumours (core basal phenotype/basal-like tumours; 60–90%), which could justify their use in the TN tumours [110, 144, 145]. This is also supported by the findings of some studies which reported a difference in response to chemotherapy between basal-like tumours and TN non-basal tumours (five negative phenotype or the so-called normal-breast like class in GEP studies) [115].

A small percentage of HR+ tumours show basal markers expression (average 8% of HR+ (range 1–18%) [64, 112, 144]) prompting the question whether this group of patients belongs to the luminal-like or basal-like cancer subclass. We have observed that HR+ tumours with basal marker coexpression exhibit a poorer prognosis when compared to HR+ basal marker negative tumours (unpublished observation by EAR and IOE). However, it is not clear whether this effect is independent of level of HR expression or expression of proliferation markers or this could represent a possible link between Luminal B and basal-like BC as indicated in gene expression studies [22, 23]. Although a proportion of HER2+ tumours express basal markers, the number of these cases is small, and no difference in survival has been identified between HER2+/basal marker positive and HER2+/basal marker negative tumours, although it has been reported that HER+/basal marker positive BC less frequently respond to Herceptin [146], large scale study of HR+ and HER2+ tumours with basal markers expression is needed to clarify these points.

Proliferation markers

In early stage BC, it has been estimated that chemotherapy can achieve 20–30% improvement in disease-free survival and around 15% or greater increase in overall survival rates [147, 148]. However, de novo and acquired resistance to therapy is observed in a significant subset of patients, leading to subsequent disease progression [149]. In contrast to predictive factors for targeted therapy, predictive markers for chemosensitivity are less well-defined [150, 151]. Chemotherapy agents including CMF, taxanes and anthracycline-based chemotherapy affect cell division or DNA synthesis and function in some way. It is also now acknowledged that increased cell proliferation is a key determinant of clinical outcome in patients with BC [152, 153]. The most widely used proliferation marker in BC is Ki-67 (MIB1) which is present exclusively in cycling cells [154] and can be used to predict response to neoadjuvant [155, 156] or adjuvant [157] chemotherapy of BC and to neoadjuvant [158, 159] or adjuvant [160] endocrine therapy of ER+ tumours.

Ki-67 has been used in combination with other markers in BC to provide prognostic and predictive value [159,

[161]. In a recent study, Cheang et al. [82] used Ki-67 in addition to ER, PR and HER2 to define molecular classes of BC. They reported that Ki-67 and HER2 expression can divide HR+ tumours into three prognostically distinct classes; Luminal A, which is HER2– and Ki-67 proliferation index low, Luminal B, which is HER2– and Ki-67 proliferation index high and Luminal/HER2+ tumours that showed HER2+ and high proliferation. Using Ki-67 to divide HR+ HER– tumours into two subclasses, it was demonstrated that HR+ tumours, that are Ki-67 high, are associated with poor outcome regardless of systemic therapy [82]. In a different approach demonstrated that the importance of assessment of proliferation rate in HR+ tumours, Loi et al. [83] have used genomic grade index to classify HR+ tumours into two subtypes with difference in prognosis in both systemically untreated and tamoxifen-treated populations. It should be noted, however, that before proliferation assessment is introduced in clinical practice, methods (e.g. mitotic counting, Ki-67 and genomic grade) and cutpoints need to be standardised. Importantly, proliferation markers, akin to histological grade, are of limited value in the TN and HER2+ tumours [77], as the majority of these tumours are poorly differentiated with a high proliferation index.

Ellis et al. [158] have reported that in patients with ER+ BC, who have received neoadjuvant endocrine treatment, Ki-67 level and ER status were independently associated with both RFS and BCSS in multivariate analysis together with lymph node status and tumour size. Similar results were demonstrated in a retrospectively collected series of patients with ER+ tumours, who received adjuvant endocrine treatment [160].

Moreover, recent data presented at San Antonio Breast Cancer Symposium show that the standard immunohistochemical tests (ER, PR, HER2 and Ki-67), when used in combinatorial manner can provide similar information as expensive molecular assays such as the GHI Recurrence Score [162].

Additional markers

Other markers that have been used as a predictor of outcome and response to therapy include Topoisomerase II alpha expression and (TOP2A) gene amplification (molecular target for anthracyclines) [98, 150, 163], Bcl2 and p53 expression [164, 165]. TOP2A aberrations (amplification, deletion) are found in up to approximately 30–90% of HER2-amplified BC, and amplifications are more common than deletions. Although *TOP2A* amplification has been reported in cases devoid of *HER2* amplification, this is an infrequent (3–9%) event [163] and even rarer in studies where *HER2* and *TOP2A* copy numbers were determined by high-resolution microarray-based comparative genomic

hybridisation studies [37, 166]. Bcl2 is positively regulated by HR in BC and its expression has been reported to provide an independent predictor of BC outcome [164, 167]. Bcl2 has been reported to predict pathological response to a neoadjuvant anthracycline/docetaxel-based regimen [168]. In a previous study of Bcl2 and p53 expression in BC, we have demonstrated that a combination of both markers provides independent prognostic value with p53+/Bcl2– phenotype was independent predictor of a worse prognosis in multivariate analysis. Mauri et al. [169] used Bcl2 and p53 in HR+ tumours to predict outcome and response to therapy. They showed that ER+/p53+ phenotype was at higher risk of relapse/death as compared with ER+/p53– phenotype, whilst the worst prognosis was observed in ER–/p53+ tumours. Similar results were obtained when Bcl2 was combined with ER [169]. Yamashita et al. [170] have assessed the expression of p53, HER2 and Ki-67 in 506 invasive ductal carcinoma using IHC, and showed that the coexistence of HER2 and p53 expression is a strong prognostic marker in BC better than each marker individually. In another study, combination of Bcl2 and HER2 appears to be a useful in predicting prognosis in curatively resected stage III BC patients [167].

Multi-gene assays

Genomic prognostic tests are highly complex compared with more traditional tests used in routine practice (e.g. IHC). They require quantitative measurements of multiple candidate genes rather than the measurement of a single analyte. The Oncotype DX and MammaPrint assays are a prototype for an alternative type of genomic diagnostic test. Other multiparameter gene expression tools have also been developed [161, 171, 172]. For detailed reviews, the readers are referred to Sotiriou and Pusztai [173] and Weigelt et al. [12].

Oncotype DX

It was developed on the basis of a prospectively chosen 250-candidate gene set. Statistical analysis and modelling of these genes led to the selection of the 21 genes (16 cancer-related and 5 references) constituting the Oncotype DX assay panel to predict the likelihood of distant BC recurrence for individual patients. The expression levels of these genes are measured by using RT-PCR. A quantitative algorithm has been developed to produce a number between 0 and 100, the ‘recurrence score’ (RS) [13, 174]. RS is categorised into three risk strata: low (score < 18), intermediate (score > 18, but < 30) or high (score > 30). Oncotype DX is considered as a clinically validated, high-complexity, multianalyte RT-PCR genomic test that predicts the likelihood of BC recurrence in early stage,

node-negative, ER+ BC. Multiple studies have demonstrated that RS provides an accurate, reproducible measure of BC aggressiveness and therapeutic responsiveness [175–177]. In summary, Oncotype DX assay is prognostic for HR+, lymph node-negative patients. A low RS is predictive of tamoxifen benefit in HR+, node-negative cases. A high RS is predictive of chemotherapy benefit over hormonal therapy in HR+ patients. It should be noted, however, that the information provided by Oncotype DX is complementary to that provided by clinicopathological parameters, including tumour size, lymph node metastasis and even histological grade, as multivariable survival models demonstrate that all of these variables provide independent prognostic information [174].

MammaPrint

It is based on the 70-gene signature derived from the analysis of the microarray-based gene expression profiles of 78 retrospectively accrued young BC patients (<55 years), with tumours <5 cm and lymph node negative. The end point for test development was 5-year distant recurrence. Patients are classified by calculating the correlation coefficient between a patient's expression levels of the 70 genes and an average good-prognosis expression profile. Tumours are classified into good prognosis class if the correlation coefficient exceeds 0.4, and poor prognosis class if less [14, 178–180]. Interestingly, the 70-genes comprised those regulating cell cycle, invasion, metastasis, signal transduction and angiogenesis with omission of previously identified individual genes associated with outcome, e.g. ER, HER2 and cyclin D1. This supports the power of a collective genetic signature over individual genes. In fact, MammaPrint, Veridex (the 76 gene signature) and most of the first generation prognostic signatures mainly identify the poor prognostic highly proliferative ER+ BC and ER– cancers (regardless of proliferation [77]). Hence, their contribution for prognostication of breast cancers when other markers are used in conjunction is rather limited [181]. It is also important to mention that these multiparameter gene expression tools are not widely available, costly and as stated by Pusztai et al. [106] that there is substantially less experience with these emerging technologies than with the more established methods, the accuracy of which is often overestimated.

Combinatorial biomarker expression and pattern of survival

Consistent with the relevant biological and clinical role of these key molecular markers on BC behaviour, an important association with the pattern of survival is noted. First, HR+ tumours in general show distinct pattern of survival

that is different from HR– tumours. In HR+ tumours, the mortality rate appears constant overtime from diagnosis, whilst the rate for HR– tumours is high during the first 3–5 years and then declines showing a plateau curve of survival. However, not all HR+ or HR– tumours show the same patterns. HR+ tumours with high proliferation (Luminal B) show response to neoadjuvant chemotherapy and survival pattern similar to HR– tumours with early frequency of events and a later decline [82]. Results from multiple studies indicate that the prognosis of HR+/HER2+ show high mortality rate and poorer outcome during the first few years after diagnosis, but this difference decreases with time to and the two survival curves converge after 7–9 years. A similar pattern was observed for tumours expressing basal markers including HR+/basal markers positive when compared to HR+/basal negative and basal-like tumours when compared to TN basal markers negative. Furthermore, tumours with a high proliferation index behave in a similar way to HR– BC and basal markers positive tumours with early frequent events [64, 82, 112].

In conclusion, although biomarker expression in BC provides prognostic and predictive information, this can be improved by considering their combinatorial expression. ER, PR, HER2 and proliferation markers in addition to basal-associated markers, Bcl2, p53 and TOP2A are key molecular biomarkers in BC and provide prognostic and predictive value, however, their significance varies in the different molecular classes and in particular, in relation to HR status of BC. The results of GEP studies in BC have led to the understanding that BC is a genetically complex disease involving multiple molecular mechanisms and biological pathways that distinct molecular subtypes of BC may, in fact, constitute different diseases, and that proliferation is not only a prognostic marker but also a predictor of response to endocrine therapy and neoadjuvant chemotherapy [12]. Based on the current lines of evidence, the inclusion of a proliferation marker (i.e. mitotic counting and/or Ki-67 assessment) to the current panel of predictive markers (ER, PR and HER2) would be desirable. It should be noted, however, that standardisation of methods for proliferation assessment and the cut-offs for the different clinical contexts still need to be standardised before proliferation can be introduced in routine clinical practice.

The increasing number of treatment options has further increased our need to improve classification and clinical management for individual BC patients and emphasises the need for improved understanding of the significance and added value of combinatorial expression of key biomarkers in BC. Despite the requirement of stringent standardisation, quality control and the use of antibodies extensively validated [85, 134], IHC can provide excellent assessment of biomarker expression in BC. It should be emphasised that

IHC overcomes one of the most important limitations of GEP (i.e. the contamination of tumour samples with stromal cells and inflammatory infiltrate), as the distribution of the biomarker of interest can be determined in situ with direct morphological control. However, pathologists should strive for optimising pre-analytical and analytical parameters to ensure reproducible results.

Acknowledgement JSR-F is funded by Breakthrough Breast Cancer.

References

- Geyer FC, Lopez-Garcia MA, Lambros MB, Reis-Filho JS (2009) Genetic characterisation of breast cancer and implications for clinical management. *J Cell Mol Med* (Published online September 14). doi:10.1111/j.1582-4934.2009.00906.x
- Simpson PT, Reis-Filho JS, Gale T, Lakhani SR (2005) Molecular evolution of breast cancer. *J Pathol* 205:248–254
- Alizadeh AA, Ross DT, Perou CM, van de Rijn M (2001) Towards a novel classification of human malignancies based on gene expression patterns. *J Pathol* 195:41–52
- Early Breast Cancer Trialists' Collaborative Group (1998) Polychemotherapy for early breast cancer: an overview of the randomised trials. *Lancet* 352:930–942
- Feng Y, Sun B, Li X, Zhang L, Niu Y, Xiao C et al (2006) Differentially expressed genes between primary cancer and paired lymph node metastases predict clinical outcome of node-positive breast cancer patients. *Breast Cancer Res Treat* 103(3):319–329
- Balslev I, Axelsson CK, Zedeler K, Rasmussen BB, Carstensen B, Mouridsen HT (1994) The Nottingham Prognostic Index applied to 9,149 patients from the studies of the Danish Breast Cancer Cooperative Group (DBCG). *Breast Cancer Res Treat* 32:281–290
- D'Eredita G, Giardina C, Martellotta M, Natale T, Ferrarese F (2001) Prognostic factors in breast cancer: the predictive value of the Nottingham Prognostic Index in patients with a long-term follow-up that were treated in a single institution. *Eur J Cancer* 37:591–596
- Pathology reporting of breast disease. A Joint Document Incorporating the Third Edition of the NHS Breast Screening Programme's Guidelines for Pathology Reporting in Breast Cancer Screening and the Second Edition of The Royal College of Pathologists' Minimum Dataset for Breast Cancer Histopathology. Sheffield, January 2005
- Early Breast Cancer Trialists' Collaborative Group (1988) Effects of adjuvant tamoxifen and of cytotoxic therapy on mortality in early breast cancer: an overview of 61 randomized trials among 28,896 women. *N Engl J Med* 319:1681–1692
- Early Breast Cancer Trialists' Collaborative Group (2005) Effects of chemotherapy and hormonal therapy for early breast cancer on recurrence and 15-year survival: an overview of the randomised trials. *Lancet* 365:1687–1717
- Kaufmann M, von Minckwitz G, Bear HD, Buzdar A, McGale P, Bonnefoi H et al (2007) Recommendations from an international expert panel on the use of neoadjuvant (primary) systemic treatment of operable breast cancer: new perspectives 2006. *Ann Oncol* 18:1927–1934
- Weigelt B, Baehner FL, Reis-Filho JS (2010) The contribution of gene expression profiling to breast cancer classification, prognostication and prediction: a retrospective of the last decade. *J Pathol* 220:263–280
- Paik S, Shak S, Tang G, Kim C, Baker J, Cronin M et al (2004) A multigene assay to predict recurrence of tamoxifen-treated, node-negative breast cancer. *N Engl J Med* 351:2817–2826
- van 't Veer LJ, Dai H, van de Vijver MJ, He YD, Hart AA, Mao M et al (2002) Gene expression profiling predicts clinical outcome of breast cancer. *Nature* 415:530–536
- Ma XJ, Hilsenbeck SG, Wang W, Ding L, Sgroi DC, Bender RA et al (2006) The HOXB13:IL17BR expression index is a prognostic factor in early-stage breast cancer. *J Clin Oncol* 24:4611–4619
- Osborne CK (1998) Steroid hormone receptors in breast cancer management. *Breast Cancer Res Treat* 51:227–238
- Breast Cancer Trials Committee, Scottish Cancer Trials Office (MRC), Edinburgh (1987) Adjuvant tamoxifen in the management of operable breast cancer: the Scottish trial. *Lancet* 2:171–175
- Early Breast Cancer Trialists' Collaborative Group (1998) Tamoxifen for early breast cancer: an overview of the randomised trials. *Lancet* 351:1451–1467
- Robertson JF (1996) Oestrogen receptor: a stable phenotype in breast cancer. *Br J Cancer* 73:5–12
- Perou CM, Sorlie T, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA et al (2000) Molecular portraits of human breast tumours. *Nature* 406:747–752
- Sotiriou C, Neo SY, McShane LM, Korn EL, Long PM, Jazaeri A et al (2003) Breast cancer classification and prognosis based on gene expression profiles from a population-based study. *Proc Natl Acad Sci USA* 100:10393–10398
- Sorlie T, Perou CM, Tibshirani R, Aas T, Geisler S, Johnsen H et al (2001) Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. *Proc Natl Acad Sci USA* 98:10869–10874
- Sorlie T, Tibshirani R, Parker J, Hastie T, Marron JS, Nobel A et al (2003) Repeated observation of breast tumor subtypes in independent gene expression data sets. *Proc Natl Acad Sci USA* 100:8418–8423
- Anderson WF, Chatterjee N, Ershler WB, Brawley OW (2002) Estrogen receptor breast cancer phenotypes in the Surveillance, Epidemiology, and End Results database. *Breast Cancer Res Treat* 76:27–36
- Murphy LC, Watson P (2002) Steroid receptors in human breast tumorigenesis and breast cancer progression. *Biomed Pharmacother* 56:65–77
- Dunnwald LK, Rossing MA, Li CI (2007) Hormone receptor status, tumor characteristics, and prognosis: a prospective cohort of breast cancer patients. *Breast Cancer Res* 9:R6
- Poole CJ, Earl HM, Hiller L, Dunn JA, Bathers S, Grieve RJ et al (2006) Epirubicin and cyclophosphamide, methotrexate, and fluorouracil as adjuvant therapy for early breast cancer. *N Engl J Med* 355:1851–1862
- Colleoni M, Bonetti M, Coates AS, Castiglione-Gertsch M, Gelber RD, Price K et al (2000) Early start of adjuvant chemotherapy may improve treatment outcome for premenopausal breast cancer patients with tumors not expressing estrogen receptors. The International Breast Cancer Study Group. *J Clin Oncol* 18:584–590
- Oh DS, Troester MA, Usary J, Hu Z, He X, Fan C et al (2006) Estrogen-regulated genes predict survival in hormone receptor-positive breast cancers. *J Clin Oncol* 24:1656–1664
- Badve S, Nakshatri H (2009) Oestrogen-receptor-positive breast cancer: towards bridging histopathological and molecular classifications. *J Clin Pathol* 62:6–12
- Esserman LJ, Ozanne EM, Dowsett M, Slingerland JM (2005) Tamoxifen may prevent both ER+ and ER- breast cancers and select for ER- carcinogenesis: an alternative hypothesis. *Breast Cancer Res* 7:R1153–R1158

32. Dowsett M, Houghton J, Iden C, Salter J, Farnon J, A'Hern R et al (2006) Benefit from adjuvant tamoxifen therapy in primary breast cancer patients according oestrogen receptor, progesterone receptor, EGF receptor and HER2 status. *Ann Oncol* 17:818–826
33. Rakha EA, El-Sayed ME, Green AR, Paish EC, Powe DG, Gee J et al (2007) Biologic and clinical characteristics of breast cancer with single hormone receptor positive phenotype. *J Clin Oncol* 25:4772–4778
34. Farmer P, Bonnefoi H, Becette V, Tubiana-Hulin M, Fumoleau P, Larsimont D et al (2005) Identification of molecular apocrine breast tumours by microarray analysis. *Oncogene* 24:4660–4671
35. Chin K, DeVries S, Fridlyand J, Spellman PT, Roydasgupta R, Kuo WL et al (2006) Genomic and transcriptional aberrations linked to breast cancer pathophysiology. *Cancer Cell* 10:529–541
36. Nikolsky Y, Sviridov E, Yao J, Dosymbekov D, Ustyansky V, Kaznacheev V et al (2008) Genome-wide functional synergy between amplified and mutated genes in human breast cancer. *Cancer Res* 68:9532–9540
37. Natrajan R, Lambros MB, Rodriguez-Pinilla SM, Moreno-Bueno G, Tan DS, Marchio C et al (2009) Tiling path genomic profiling of grade 3 invasive ductal breast cancers. *Clin Cancer Res* 15:2711–2722
38. Natrajan R, Weigelt B, Mackay A, Geyer FC, Grigoriadis A, Tan DS et al (2009) An integrative genomic and transcriptomic analysis reveals molecular pathways and networks regulated by copy number aberrations in basal-like, HER2 and luminal cancers. *Breast Cancer Res Treat*. doi:10.1007/s10549-009-0501-3
39. Horwitz KB, Koseki Y, McGuire WL (1978) Estrogen control of progesterone receptor in human breast cancer: role of estradiol and antiestrogen. *Endocrinology* 103:1742–1751
40. Horwitz KB, McGuire WL (1975) Specific progesterone receptors in human breast cancer. *Steroids* 25:497–505
41. Lanari C, Lamb CA, Fabris VT, Helguero LA, Soldati R, Bottino MC et al (2009) The MPA mouse breast cancer model: evidence for a role of progesterone receptors in breast cancer. *Endocr Relat Cancer* 16:333–350
42. Colomer R, Beltran M, Dorcas J, Cortes-Funes H, Hornedo J, Valentin V et al (2005) It is not time to stop progesterone receptor testing in breast cancer. *J Clin Oncol* 23:3868–3869 (author reply 3869–3870)
43. Stendahl M, Ryden L, Nordenskjold B, Jonsson PE, Landberg G, Jirstrom K (2006) High progesterone receptor expression correlates to the effect of adjuvant tamoxifen in premenopausal breast cancer patients. *Clin Cancer Res* 12:4614–4618
44. Ryden L, Jonsson PE, Chebil G, Dufmats M, Ferno M, Jirstrom K et al (2005) Two years of adjuvant tamoxifen in premenopausal patients with breast cancer: a randomised, controlled trial with long-term follow-up. *Eur J Cancer* 41:256–264
45. Ravdin PM, Green S, Dorr TM, McGuire WL, Fabian C, Pugh RP et al (1992) Prognostic significance of progesterone receptor levels in estrogen receptor-positive patients with metastatic breast cancer treated with tamoxifen: results of a prospective Southwest Oncology Group study. *J Clin Oncol* 10:1284–1291
46. Elledge RM, Green S, Pugh R, Allred DC, Clark GM, Hill J et al (2000) Estrogen receptor (ER) and progesterone receptor (PgR), by ligand-binding assay compared with ER, PgR and pS2, by immuno-histochemistry in predicting response to tamoxifen in metastatic breast cancer: a Southwest Oncology Group Study. *Int J Cancer* 89:111–117
47. Regan MM, Viale G, Mastropasqua MG, Maiorano E, Golouh R, Carbone A et al (2006) Re-evaluating adjuvant breast cancer trials: assessing hormone receptor status by immunohistochemical versus extraction assays. *J Natl Cancer Inst* 98:1571–1581
48. Bardou VJ, Arpino G, Elledge RM, Osborne CK, Clark GM (2003) Progesterone receptor status significantly improves outcome prediction over estrogen receptor status alone for adjuvant endocrine therapy in two large breast cancer databases. *J Clin Oncol* 21:1973–1979
49. Hilsenbeck SG, Osborne CK (2006) Is there a role for adjuvant tamoxifen in progesterone receptor-positive breast cancer? An in silico clinical trial. *Clin Cancer Res* 12:1049s–1055s
50. Dowsett M, Cuzick J, Wale C, Howell T, Houghton J, Baum M (2005) Retrospective analysis of time to recurrence in the ATAC trial according to hormone receptor status: an hypothesis-generating study. *J Clin Oncol* 23:7512–7517
51. Olivotto IA, Truong PT, Speers CH, Bernstein V, Allan SJ, Kelly SJ et al (2004) Time to stop progesterone receptor testing in breast cancer management. *J Clin Oncol* 22:1769–1770
52. Allred DC (2008) Commentary: hormone receptor testing in breast cancer: a distress signal from Canada. *Oncologist* 13:1134–1136
53. Ferno M, Stal O, Baldetorp B, Hatschek T, Kallstrom AC, Malmstrom P et al (2000) Results of two or five years of adjuvant tamoxifen correlated to steroid receptor and S-phase levels. South Sweden Breast Cancer Group, and South-East Sweden Breast Cancer Group. *Breast Cancer Res Treat* 59:69–76
54. Arpino G, Weiss H, Lee AV, Schiff R, De Placido S, Osborne CK et al (2005) Estrogen receptor-positive, progesterone receptor-negative breast cancer: association with growth factor receptor expression and tamoxifen resistance. *J Natl Cancer Inst* 97:1254–1261
55. Yu KD, Di GH, Wu J, Lu JS, Shen KW, Liu GY et al (2008) Breast cancer patients with estrogen receptor-negative/progesterone receptor-positive tumors: being younger and getting less benefit from adjuvant tamoxifen treatment. *J Cancer Res Clin Oncol* 134:1347–1354
56. Bird PA, Hill AG, Houssami N (2008) Poor hormone receptor expression in East African breast cancer: evidence of a biologically different disease? *Ann Surg Oncol* 15:1983–1988
57. Rhodes A, Jasani B (2009) The oestrogen receptor-negative/progesterone receptor-positive breast tumour: a biological entity or a technical artefact? *J Clin Pathol* 62:95–96
58. De Maeyer L, Van Limbergen E, De Nys K, Moerman P, Pochet N, Hendrickx W et al (2008) Does estrogen receptor negative/progesterone receptor positive breast carcinoma exist? *J Clin Oncol* 26:335–336 (author reply 336–338)
59. de Cremoux P, Tran-Perennou C, Elie C, Boudou E, Barbaroux C, Poupon MF et al (2002) Quantitation of estradiol receptors alpha and beta and progesterone receptors in human breast tumors by real-time reverse transcription-polymerase chain reaction. Correlation with protein assays. *Biochem Pharmacol* 64:507–515
60. Di Fronzo G, Coradini D, Cappelletti V, Miodini P, Granata G, Schwartz M et al (1990) Hormone receptors and disease-free survival in breast cancer: impact of increasing threshold levels. *Anticancer Res* 10:1699–1705
61. Viale G, Regan MM, Maiorano E, Mastropasqua MG, Dell'Orto P, Rasmussen BB et al (2007) Prognostic and predictive value of centrally reviewed expression of estrogen and progesterone receptors in a randomized trial comparing letrozole and tamoxifen adjuvant therapy for postmenopausal early breast cancer: BIG 1–98. *J Clin Oncol* 25:3846–3852
62. Rakha EA, Ellis IO (2008) Does estrogen receptor-negative/progesterone receptor-positive breast carcinoma exist? In reply. *J Clin Oncol* 26:335–340
63. Anderson WF, Chu KC, Chatterjee N, Brawley O, Brinton LA (2001) Tumor variants by hormone receptor expression in white patients with node-negative breast cancer from the surveillance, epidemiology, and end results database. *J Clin Oncol* 19:18–27

64. Carey LA, Perou CM, Livasy CA, Dressler LG, Cowan D, Conway K et al (2006) Race, breast cancer subtypes, and survival in the Carolina Breast Cancer Study. *JAMA* 295:2492–2502
65. Kumar V, Abbas AK (2004) *Fausto N: the breast*, vol 3, 7th edn. Elsevier, Philadelphia, PA
66. Goussard J, Genot JY (1994) What can be now expected of the determination of estrogen and progesterone receptors in the treatment of breast cancers. *Bull Cancer* 81:22–28
67. Kinne DW, Butler JA, Kimmel M, Flehinger BJ, Menendez-Botet C, Schwartz M (1987) Estrogen receptor protein of breast cancer in patients with positive nodes. High recurrence rates in the postmenopausal estrogen receptor-negative groups. *Arch Surg* 122:1303–1306
68. Parl FF, Schmidt BP, Dupont WD, Wagner RK (1984) Prognostic significance of estrogen receptor status in breast cancer in relation to tumor stage, axillary node metastasis, and histopathologic grading. *Cancer* 54:2237–2242
69. Pichon MF, Broet P, Magdelenat H, Delarue JC, Spyrtos F, Basuyau JP et al (1996) Prognostic value of steroid receptors after long-term follow-up of 2257 operable breast cancers. *Br J Cancer* 73:1545–1551
70. Trendell-Smith NJ, Peston D, Shousha S (1999) Adenoid cystic carcinoma of the breast: a tumour commonly devoid of oestrogen receptors and related proteins. *Histopathology* 35:241–248
71. Rosen PP, Cranor ML (1991) Secretory carcinoma of the breast. *Arch Pathol Lab Med* 115:141–144
72. Weigelt B, Reis-Filho JS (2009) Histological and molecular types of breast cancer: is there a unifying taxonomy? *Nat Rev Clin Oncol* 6:718–730
73. Osborne CK, Yochmowitz MG, Knight WA III, McGuire WL (1980) The value of estrogen and progesterone receptors in the treatment of breast cancer. *Cancer* 46:2884–2888
74. Howell A, Cuzick J, Baum M, Buzdar A, Dowsett M, Forbes JF et al (2005) Results of the ATAC (Arimidex, Tamoxifen, Alone or in Combination) trial after completion of 5 years' adjuvant treatment for breast cancer. *Lancet* 365:60–62
75. Sundblad AS, Caprarulo L (1996) Immunohistochemical characteristics of mammary carcinomas with estrogen-negative and progesterone-positive receptors. *Medicina (B Aires)* 56:683–689
76. Goldhirsch A, Wood WC, Gelber RD, Coates AS, Thurlimann B, Senn HJ et al (2007) Progress and promise: highlights of the international expert consensus on the primary therapy of early breast cancer 2007. *Ann Oncol* 18:1133–1144
77. Wirapati P, Sotiriou C, Kunkel S, Farmer P, Pradervand S, Haibe-Kains B et al (2008) Meta-analysis of gene expression profiles in breast cancer: toward a unified understanding of breast cancer subtyping and prognosis signatures. *Breast Cancer Res* 10:R65
78. Desmedt C, Ruiz-Garcia E, Andre F (2008) Gene expression predictors in breast cancer: current status, limitations and perspectives. *Eur J Cancer* 44:2714–2720
79. Bhargava R, Dabbs DJ (2008) Luminal B breast tumors are not HER2 positive. *Breast Cancer Res* 10:404 (author reply 405)
80. Hugh J, Hanson J, Cheang MC, Nielsen TO, Perou CM, Dumontet C et al (2009) Breast cancer subtypes and response to docetaxel in node-positive breast cancer: use of an immunohistochemical definition in the BCIRG 001 trial. *J Clin Oncol* 27:1168–1176
81. Ihemelandu CU, Leffall LD, Dewitty RL, Naab TJ, Mezghebe HM, Makambi KH et al (2007) Molecular breast cancer subtypes in premenopausal african-american women, tumor biologic factors and clinical outcome. *Ann Surg Oncol* 14:2994–3003
82. Cheang MC, Chia SK, Voduc D, Gao D, Leung S, Snider J et al (2009) Ki67 index, HER2 status, and prognosis of patients with luminal B breast cancer. *J Natl Cancer Inst* 101:736–750
83. Loi S, Haibe-Kains B, Desmedt C, Lallemand F, Tutt AM, Gillet C et al (2007) Definition of clinically distinct molecular subtypes in estrogen receptor-positive breast carcinomas through genomic grade. *J Clin Oncol* 25:1239–1246
84. Slamon DJ, Clark GM, Wong SG, Levin WJ, Ullrich A, McGuire WL (1987) Human breast cancer: correlation of relapse and survival with amplification of the HER-2/neu oncogene. *Science* 235:177–182
85. Wolff AC, Hammond ME, Schwartz JN, Hagerty KL, Allred DC, Cote RJ et al (2007) American Society of Clinical Oncology/College of American Pathologists guideline recommendations for human epidermal growth factor receptor 2 testing in breast cancer. *J Clin Oncol* 25:118–145
86. Yamauchi H, Stearns V, Hayes DF (2001) When is a tumor marker ready for prime time? A case study of c-erbB-2 as a predictive factor in breast cancer. *J Clin Oncol* 19:2334–2356
87. Bartlett J, Mallon E, Cooke T (2003) The clinical evaluation of HER-2 status: which test to use? *J Pathol* 199:411–417
88. Chia S, Norris B, Speers C, Cheang M, Gilks B, Gown AM et al (2008) Human epidermal growth factor receptor 2 overexpression as a prognostic factor in a large tissue microarray series of node-negative breast cancers. *J Clin Oncol* 26:5697–5704
89. Dandachi N, Dietze O, Hauser-Kronberger C (2002) Chromogenic in situ hybridization: a novel approach to a practical and sensitive method for the detection of HER2 oncogene in archival human breast carcinoma. *Lab Invest* 82:1007–1014
90. Quenel N, Wafflard J, Bonichon F, de Mascarel I, Trojani M, Durand M et al (1995) The prognostic value of c-erbB2 in primary breast carcinomas: a study on 942 cases. *Breast Cancer Res Treat* 35:283–291
91. Press MF, Finn RS, Cameron D, Di Leo A, Geyer CE, Villalobos IE et al (2008) HER-2 gene amplification, HER-2 and epidermal growth factor receptor mRNA and protein expression, and lapatinib efficacy in women with metastatic breast cancer. *Clin Cancer Res* 14:7861–7870
92. Piccart-Gebhart MJ, Procter M, Leyland-Jones B, Goldhirsch A, Untch M, Smith I et al (2005) Trastuzumab after adjuvant chemotherapy in HER2-positive breast cancer. *N Engl J Med* 353:1659–1672
93. Ward S, Pilgrim H, Hind D (2009) Trastuzumab for the treatment of primary breast cancer in HER2-positive women: a single technology appraisal. *Health Technol Assess* 13(Suppl 1):1–6
94. Slamon DJ, Leyland-Jones B, Shak S, Fuchs H, Paton V, Bajamonde A et al (2001) Use of chemotherapy plus a monoclonal antibody against HER2 for metastatic breast cancer that overexpresses HER2. *N Engl J Med* 344:783–792
95. Konecny G, Pauletti G, Pegram M, Untch M, Dandekar S, Aguilar Z et al (2003) Quantitative association between HER-2/neu and steroid hormone receptors in hormone receptor-positive primary breast cancer. *J Natl Cancer Inst* 95:142–153
96. Ellis MJ, Coop A, Singh B, Mauriac L, Llombert-Cussac A, Janicke F et al (2001) Letrozole is more effective neoadjuvant endocrine therapy than tamoxifen for ErbB-1- and/or ErbB-2-positive, estrogen receptor-positive primary breast cancer: evidence from a phase III randomized trial. *J Clin Oncol* 19:3808–3816
97. Rasmussen BB, Regan MM, Lykkesfeldt AE, Dell'Orto P, Del Curto B, Henriksen KL et al (2008) Adjuvant letrozole versus tamoxifen according to centrally-assessed ERBB2 status for postmenopausal women with endocrine-responsive early breast cancer: supplementary results from the BIG 1–98 randomised trial. *Lancet Oncol* 9:23–28
98. Pritchard KI, Messersmith H, Elavathil L, Trudeau M, O'Malley F, Dhesy-Thind B (2008) HER-2 and topoisomerase II as predictors of response to chemotherapy. *J Clin Oncol* 26:736–744

99. Gianni L, Norton L, Wolmark N, Suter TM, Bonadonna G, Hortobagyi GN (2009) Role of anthracyclines in the treatment of early breast cancer. *J Clin Oncol* 27:4798–4808
100. Tubbs R, Barlow WE, Budd GT, Swain E, Porter P, Gown A et al (2009) Outcome of patients with early-stage breast cancer treated with doxorubicin-based adjuvant chemotherapy as a function of HER2 and TOP2A status. *J Clin Oncol* 27:3881–3886
101. Pritchard KI, Shepherd LE, O'Malley FP, Andrulis IL, Tu D, Bramwell VH et al (2006) HER2 and responsiveness of breast cancer to adjuvant chemotherapy. *N Engl J Med* 354:2103–2111
102. Wood WC, Budman DR, Korzun AH, Cooper MR, Younger J, Hart RD et al (1994) Dose and dose intensity of adjuvant chemotherapy for stage II, node-positive breast carcinoma. *N Engl J Med* 330:1253–1259
103. Thor AD, Berry DA, Budman DR, Muss HB, Kute T, Henderson IC et al (1998) erbB-2, p53, and efficacy of adjuvant therapy in lymph node-positive breast cancer. *J Natl Cancer Inst* 90:1346–1360
104. Konecny GE, Thomssen C, Luck HJ, Untch M, Wang HJ, Kuhn W et al (2004) Her-2/neu gene amplification and response to paclitaxel in patients with metastatic breast cancer. *J Natl Cancer Inst* 96:1141–1151
105. Hayes DF, Thor AD, Dressler LG, Weaver D, Edgerton S, Cowan D et al (2007) HER2 and response to paclitaxel in node-positive breast cancer. *N Engl J Med* 357:1496–1506
106. Puztai L, Mazouni C, Anderson K, Wu Y, Symmans WF (2006) Molecular classification of breast cancer: limitations and potential. *Oncologist* 11:868–877
107. Badve S, Turbin D, Thorat MA, Morimiya A, Nielsen TO, Perou CM et al (2007) FOXA1 expression in breast cancer—correlation with luminal subtype A and survival. *Clin Cancer Res* 13:4415–4421
108. Nielsen TO, Hsu FD, Jensen K, Cheang M, Karaca G, Hu ZY et al (2004) Immunohistochemical and clinical characterization of the basal-like subtype of invasive breast carcinoma. *Clin Cancer Res* 10:5367–5374
109. Matos I, Duffloth R, Alvarenga M, Zeferino LC, Schmitt F (2005) p63, cytokerin 5, and P-cadherin: three molecular markers to distinguish basal phenotype in breast carcinomas. *Virchows Arch* 447:688–694
110. Rakha EA, Elsheikh SE, Aleskandarany MA, Habashi HO, Green AR, Powe DG et al (2009) Triple-negative breast cancer: distinguishing between basal and nonbasal subtypes. *Clin Cancer Res* 15:2302–2310
111. Kurebayashi J, Moriya T, Ishida T, Hirakawa H, Kurosumi M, Akiyama F et al (2007) The prevalence of intrinsic subtypes and prognosis in breast cancer patients of different races. *Breast* 16:72–77
112. Rakha EA, El-Sayed ME, Green AR, Paish EC, Lee AH, Ellis IO (2007) Breast carcinoma with basal differentiation: a proposal for pathology definition based on basal cytokeratin expression. *Histopathology* 50:434–438
113. Marchiò C, Natrajan R, Shiu KK, Lambros MB, Rodriguez-Pinilla SM, Tan DS, Lord CJ, Hungermann D, Fenwick K, Tamber N, Mackay A, Palacios J, Sapino A, Buerger H, Ashworth A, Reis-Filho JS (2008) The genomic profile of HER2-amplified breast cancers: the influence of ER status. *J Pathol* 216:399–407
114. Pinto AE, Andre S, Pereira T, Nobrega S, Soares J (2001) C-erbB-2 oncoprotein overexpression identifies a subgroup of estrogen receptor positive (ER+) breast cancer patients with poor prognosis. *Ann Oncol* 12:525–533
115. Rouzier R, Perou CM, Symmans WF, Ibrahim N, Cristofanilli M, Anderson K et al (2005) Breast cancer molecular subtypes respond differently to preoperative chemotherapy. *Clin Cancer Res* 11:5678–5685
116. Prat A, Baselga J (2008) The role of hormonal therapy in the management of hormonal-receptor-positive breast cancer with co-expression of HER2. *Nat Clin Pract Oncol* 5:531–542
117. Darb-Esfahani S, Loibl S, Muller BM, Roller M, Denkert C, Komor M et al (2009) Identification of biology-based breast cancer types with distinct predictive and prognostic features: role of steroid hormone and HER2 receptor expression in patients treated with neoadjuvant anthracycline/taxane-based chemotherapy. *Breast Cancer Res* 11:R69
118. Dowsett M, Allred C, Knox J, Quinn E, Salter J, Wale C et al (2008) Relationship between quantitative estrogen and progesterone receptor expression and human epidermal growth factor receptor 2 (HER-2) status with recurrence in the Arimidex, Tamoxifen, Alone or in Combination trial. *J Clin Oncol* 26:1059–1065
119. Rakha EA, Reis-Filho JS, Ellis IO (2008) Basal-like breast cancer: a critical review. *J Clin Oncol* 26:2568–2581
120. Carey LA, Dees EC, Sawyer L, Gatti L, Moore DT, Collichio F et al (2007) The triple negative paradox: primary tumor chemosensitivity of breast cancer subtypes. *Clin Cancer Res* 13:2329–2334
121. Hennessy BT, Gonzalez-Angulo AM, Stenke-Hale K, Gilcrease MZ, Krishnamurthy S, Lee JS et al (2009) Characterization of a naturally occurring breast cancer subset enriched in epithelial-to-mesenchymal transition and stem cell characteristics. *Cancer Res* 69:4116–4124
122. Howell A, Bundred NJ, Cuzick J, Allred DC, Clarke R (2008) Response and resistance to the endocrine prevention of breast cancer. *Adv Exp Med Biol* 617:201–211
123. Conte P, Guarneri V, Bengala C (2007) Evolving nonendocrine therapeutic options for metastatic breast cancer: how adjuvant chemotherapy influences treatment. *Clin Breast Cancer* 7:841–849
124. Bachleitner-Hofmann T, Pichler-Gebhard B, Rudas M, Gnatt M, Taucher S, Kandioler D et al (2002) Pattern of hormone receptor status of secondary contralateral breast cancers in patients receiving adjuvant tamoxifen. *Clin Cancer Res* 8:3427–3432
125. Gutierrez MC, Detre S, Johnston S, Mohsin SK, Shou J, Allred DC et al (2005) Molecular changes in tamoxifen-resistant breast cancer: relationship between estrogen receptor, HER-2, and p38 mitogen-activated protein kinase. *J Clin Oncol* 23:2469–2476
126. Tovey S, Dunne B, Witton CJ, Forsyth A, Cooke TG, Bartlett JM (2005) Can molecular markers predict when to implement treatment with aromatase inhibitors in invasive breast cancer? *Clin Cancer Res* 11:4835–4842
127. Schiff R, Massarweh SA, Shou J, Bharwani L, Mohsin SK, Osborne CK (2004) Cross-talk between estrogen receptor and growth factor pathways as a molecular target for overcoming endocrine resistance. *Clin Cancer Res* 10:331S–336S
128. Benz CC, Scott GK, Sarup JC, Johnson RM, Tripathy D, Coronado E et al (1992) Estrogen-dependent, tamoxifen-resistant tumorigenic growth of MCF-7 cells transfected with HER2/neu. *Breast Cancer Res Treat* 24:85–95
129. Gee JM, Harper ME, Hutcheson IR, Madden TA, Barrow D, Knowlden JM et al (2003) The anti-epidermal growth factor receptor agent gefitinib (ZD1839/Iressa) improves anti-hormone response and prevents development of resistance in breast cancer in vitro. *Endocrinology* 144:5105–5117
130. Nicholson RI, Hutcheson IR, Hiscox SE, Knowlden JM, Giles M, Barrow D et al (2005) Growth factor signalling and resistance to selective oestrogen receptor modulators and pure anti-oestrogens: the use of anti-growth factor therapies to treat or

- delay endocrine resistance in breast cancer. *Endocr Relat Cancer* 12(Suppl 1):S29–S36
131. Iorns E, Turner NC, Elliott R, Syed N, Garrone O, Gasco M et al (2008) Identification of CDK10 as an important determinant of resistance to endocrine therapy for breast cancer. *Cancer Cell* 13:91–104
 132. Coombes RC, Hall E, Gibson LJ, Paridaens R, Jassem J, Delozier T et al (2004) A randomized trial of exemestane after two to three years of tamoxifen therapy in postmenopausal women with primary breast cancer. *N Engl J Med* 350:1081–1092
 133. Dixon MJ (2008) Role of ErbB2 in selection for adjuvant tamoxifen or aromatase inhibitors. *Womens Health (Lond Engl)* 4:229–231
 134. Allred DC, Carlson RW, Berry DA, Burstein HJ, Edge SB, Goldstein LJ et al (2009) NCCN Task Force Report: estrogen receptor and progesterone receptor testing in breast cancer by immunohistochemistry. *J Natl Compr Canc Netw* 7(Suppl 6):S1–S21
 135. Dairkee SH, Mayall BH, Smith H, Hackett A (1987) Monoclonal marker that predicts early recurrence of breast cancer. *Lancet* 1:514
 136. Rakha EA, Putti TC, Abd El-Rehim DM, Paish C, Green AR, Powe DG et al (2006) Morphological and immunophenotypic analysis of breast carcinomas with basal and myoepithelial differentiation. *J Pathol* 208:495–506
 137. Rakha EA, El-Rehim DA, Paish C, Green AR, Lee AH, Robertson JF et al (2006) Basal phenotype identifies a poor prognostic subgroup of breast cancer of clinical importance. *Eur J Cancer* 42:3149–3156
 138. van de Rijn M, Perou CM, Tibshirani R, Haas P, Kallioniemi O, Kononen J et al (2002) Expression of cytokeratins 17 and 5 identifies a group of breast carcinomas with poor clinical outcome. *Am J Pathol* 161:1991–1996
 139. Lakhani SR, Reis-Filho JS, Fulford L, Penault-Llorca F, van der Vijver M, Parry S et al (2005) Prediction of BRCA1 status in patients with breast cancer using estrogen receptor and basal phenotype. *Clin Cancer Res* 11:5175–5180
 140. Dent R, Hanna WM, Trudeau M, Rawlinson E, Sun P, Narod SA (2009) Time to disease recurrence in basal-type breast cancers: effects of tumor size and lymph node status. *Cancer* 115:4917–4923
 141. Foulkes WD, Grainge MJ, Rakha EA, Green AR, Ellis IO (2009) Tumor size is an unreliable predictor of prognosis in basal-like breast cancers and does not correlate closely with lymph node status. *Breast Cancer Res Treat* 117:199–204
 142. Gusterson B (2009) Do ‘basal-like’ breast cancers really exist? *Nat Rev Cancer* 9:128–134
 143. Rakha E, Reis-Filho JS (2009) Basal-like breast carcinoma: from expression profiling to routine practice. *Arch Pathol Lab Med* 133:860–868
 144. Cheang MC, Voduc D, Bajdik C, Leung S, McKinney S, Chia SK et al (2008) Basal-like breast cancer defined by five biomarkers has superior prognostic value than triple-negative phenotype. *Clin Cancer Res* 14:1368–1376
 145. Rakha EA, Tan DS, Foulkes WD, Ellis IO, Tutt A, Nielsen TO et al (2007) Are triple-negative tumours and basal-like breast cancer synonymous? *Breast Cancer Res* 9:404 (author reply 405)
 146. Harris LN, You F, Schnitt SJ, Witkiewicz A, Lu X, Sgroi D et al (2007) Predictors of resistance to preoperative trastuzumab and vinorelbine for HER2-positive early breast cancer. *Clin Cancer Res* 13:1198–1207
 147. Gonzalez-Angulo AM, Morales-Vasquez F, Hortobagyi GN (2007) Overview of resistance to systemic therapy in patients with breast cancer. *Adv Exp Med Biol* 608:1–22
 148. Bergh J, Jonsson PE, Glimelius B, Nygren P (2001) A systematic overview of chemotherapy effects in breast cancer. *Acta Oncol* 40:253–281
 149. Pusztai L, Hortobagyi GN (1998) High-dose chemotherapy: how resistant is breast cancer? *Drug Resist Updat* 1:62–72
 150. Arpino G, Ciocca DR, Weiss H, Allred DC, Daguette P, Vargas-Roig L et al (2005) Predictive value of apoptosis, proliferation, HER-2, and topoisomerase II α for anthracycline chemotherapy in locally advanced breast cancer. *Breast Cancer Res Treat* 92:69–75
 151. Yamashiro H, Toi M (2008) Update of evidence in chemotherapy for breast cancer. *Int J Clin Oncol* 13:3–7
 152. van Diest PJ, van der Wall E, Baak JPA (2004) Prognostic value of proliferation in invasive breast cancer: a review. *J Clin Pathol* 57:675–681
 153. Veronese SM, Gambacorta M, Gottardi O, Scanzi F, Ferrari M, Lampertico P (1993) Proliferation index as a prognostic marker in breast cancer. *Cancer* 71:3926–3931
 154. Colozza M, Azambuja E, Cardoso F, Sotiriou C, Larsimont D, Piccart MJ (2005) Proliferative markers as prognostic and predictive tools in early breast cancer: where are we now? *Ann Oncol* 16:1723–1739
 155. Vincent-Salomon A, Rousseau A, Jouve M, Beuzebec P, Sigal-Zafrani B, Freneaux P et al (2004) Proliferation markers predictive of the pathological response and disease outcome of patients with breast carcinomas treated by anthracycline-based preoperative chemotherapy. *Eur J Cancer* 40:1502–1508
 156. Miglietta L, Vanella P, Canobbio L, Parodi MA, Guglielmini P, Boccardo F (2009) Clinical and pathological response to primary chemotherapy in patients with locally advanced breast cancer grouped according to hormonal receptors, Her2 status, grading and Ki-67 proliferation index. *Anticancer Res* 29:1621–1625
 157. Aleskandarany MA, Green AR, Rakha EA, Mohammed RA, Elsheikh SE, Powe DG et al. (2009) Growth fraction as a predictor of response to chemotherapy in node negative breast cancer. *Int J Cancer*. doi:10.1002/ijc.24860
 158. Ellis MJ, Tao Y, Luo J, A'Hern R, Evans DB, Bhatnagar AS et al (2008) Outcome prediction for estrogen receptor-positive breast cancer based on postneoadjuvant endocrine therapy tumor characteristics. *J Natl Cancer Inst* 100:1380–1388
 159. Viale G, Regan MM, Mastropasqua MG, Maffini F, Maiorano E, Colleoni M et al (2008) Predictive value of tumor Ki-67 expression in two randomized trials of adjuvant chemoendocrine therapy for node-negative breast cancer. *J Natl Cancer Inst* 100:207–212
 160. Jacquemier J, Charafe-Jauffret E, Monville F, Esterni B, Extra JM, Houvenaghel G et al (2009) Association of GATA3, P53, Ki67 status and vascular peritumoral invasion are strongly prognostic in luminal breast cancer. *Breast Cancer Res* 11:R23
 161. Ring BZ, Seitz RS, Beck R, Shasteen WJ, Tarr SM, Cheang MC et al (2006) Novel prognostic immunohistochemical biomarker panel for estrogen receptor-positive breast cancer. *J Clin Oncol* 24:3039–3047
 162. Cuzick J, Dowsett M, Wale C, Salter J, Quinn E, Zabaglo L et al (2009) Prognostic value of a combined ER, PgR, Ki67, HER2 immunohistochemical (IHC4) score and comparison with the GHI recurrence score—results from TransATAC. San Antonio Breast Cancer Symposium, December 2009, Abstract No (74)
 163. Bouchalova K, Cizkova M, Cwiertka K, Trojanec R, Hajduch M (2009) Triple negative breast cancer—current status and prospective targeted treatment based on HER1 (EGFR), TOP2A and C-MYC gene assessment. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub* 153:13–17
 164. Callagy GM, Pharoah PD, Pinder SE, Hsu FD, Nielsen TO, Ragaz J et al (2006) Bcl-2 is a prognostic marker in breast

- cancer independently of the Nottingham Prognostic Index. *Clin Cancer Res* 12:2468–2475
165. Allred DC, Clark GM, Elledge R, Fuqua SA, Brown RW, Chamness GC et al (1993) Association of p53 protein expression with tumor cell proliferation rate and clinical outcome in node-negative breast cancer. *J Natl Cancer Inst* 85:200–206
 166. Arriola E, Marchio C, Tan DS, Drury SC, Lambros MB, Natrajan R et al (2008) Genomic analysis of the HER2/TOP2A amplicon in breast cancer and breast cancer cell lines. *Lab Invest* 88:491–503
 167. Lee KH, Im SA, Oh DY, Lee SH, Chie EK, Han W et al (2007) Prognostic significance of bcl-2 expression in stage III breast cancer patients who had received doxorubicin and cyclophosphamide followed by paclitaxel as adjuvant chemotherapy. *BMC Cancer* 7:63
 168. Ogston KN, Miller ID, Schofield AC, Spyrtantis A, Pavlidou E, Sarkar TK et al (2004) Can patients' likelihood of benefiting from primary chemotherapy for breast cancer be predicted before commencement of treatment? *Breast Cancer Res Treat* 86:181–189
 169. Mauri FA, Maisonneuve P, Caffo O, Veronese S, Aldovini D, Ferrero S et al (1999) Prognostic value of estrogen receptor status can be improved by combined evaluation of p53, Bcl2 and PgR expression: an immunohistochemical study on breast carcinoma with long-term follow-up. *Int J Oncol* 15:1137–1147
 170. Yamashita H, Nishio M, Toyama T, Sugiura H, Zhang Z, Kobayashi S et al (2004) Coexistence of HER2 over-expression and p53 protein accumulation is a strong prognostic molecular marker in breast cancer. *Breast Cancer Res* 6:R24–R30
 171. Wang Y, Klijn JG, Zhang Y, Sieuwerts AM, Look MP, Yang F et al (2005) Gene-expression profiles to predict distant metastasis of lymph-node-negative primary breast cancer. *Lancet* 365:671–679
 172. Lancashire LJ, Powe DG, Reis-Filho JS, Rakha E, Lemetre C, Weigelt B et al (2010) A validated gene expression profile for detecting clinical outcome in breast cancer using artificial neural networks. *Breast Cancer Res Treat* 120:83–93
 173. Sotiriou C, Puzstai L (2009) Gene-expression signatures in breast cancer. *N Engl J Med* 360:790–800
 174. Habel LA, Shak S, Jacobs MK, Capra A, Alexander C, Pho M et al (2006) A population-based study of tumor gene expression and risk of breast cancer death among lymph node-negative patients. *Breast Cancer Res* 8:R25
 175. Fan C, Oh DS, Wessels L, Weigelt B, Nuyten DS, Nobel AB et al (2006) Concordance among gene-expression-based predictors for breast cancer. *N Engl J Med* 355:560–569
 176. Esteva FJ, Sahin AA, Cristofanilli M, Coombes K, Lee SJ, Baker J et al (2005) Prognostic role of a multigene reverse transcriptase-PCR assay in patients with node-negative breast cancer not receiving adjuvant systemic therapy. *Clin Cancer Res* 11:3315–3319
 177. Chang JC, Makris A, Gutierrez MC, Hilsenbeck SG, Hackett JR, Jeong J et al (2008) Gene expression patterns in formalin-fixed, paraffin-embedded core biopsies predict docetaxel chemosensitivity in breast cancer patients. *Breast Cancer Res Treat* 108:233–240
 178. van de Vijver MJ, He YD, van't Veer LJ, Dai H, Hart AA, Voskuil DW et al (2002) A gene-expression signature as a predictor of survival in breast cancer. *N Engl J Med* 347:1999–2009
 179. Buyse M, Loi S, van't Veer L, Viale G, Delorenzi M, Glas AM et al (2006) Validation and clinical utility of a 70-gene prognostic signature for women with node-negative breast cancer. *J Natl Cancer Inst* 98:1183–1192
 180. Espinosa E, Vara JA, Redondo A, Sanchez JJ, Hardisson D, Zamora P et al (2005) Breast cancer prognosis determined by gene expression profiling: a quantitative reverse transcriptase polymerase chain reaction study. *J Clin Oncol* 23:7278–7285
 181. Dunkler D, Michiels S, Schemper M (2007) Gene expression profiling: does it add predictive accuracy to clinical characteristics in cancer prognosis? *Eur J Cancer* 43:745–751