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R. Sobrino, M.C. Arnal, D.F. Luco, C. Gortázar. Prevalence of antibodies against canine distemper virus and canine parvovirus among foxes and wolves from Spain. Veterinary Microbiology, 2007, 126 (1-3), pp.251. 10.1016/j.vetmic.2007.06.014 . hal-00532281

## HAL Id: hal-00532281 https://hal.science/hal-00532281

Submitted on 4 Nov 2010

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## Accepted Manuscript

Title: Prevalence of antibodies against canine distemper virus and canine parvovirus among foxes and wolves from Spain

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PII:	S0378-1135(07)00312-4
DOI:	doi:10.1016/j.vetmic.2007.06.014
Reference:	VETMIC 3735
To appear in:	VETMIC
Received date:	29-3-2007
Revised date:	12-6-2007
Accepted date:	18-6-2007



Please cite this article as: Sobrino, R., Arnal, M.C., Luco, D.F., Gortázar, C., Prevalence of antibodies against canine distemper virus and canine parvovirus among foxes and wolves from Spain, *Veterinary Microbiology* (2007), doi:10.1016/j.vetmic.2007.06.014

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### Veterinary Microbiology

#### Short Communication

Prevalence of antibodies against canine distemper virus and canine parvovirus

- 2 among foxes and wolves from Spain.
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### 8 ABSTRACT

9 Viral diseases can influence the population dynamics of wild carnivores and can have 10 effects on carnivore conservation. Hence, a serologic survey was conducted in an 11 opportunistic sample of 137 foxes (Vulpes vulpes) and 37 wolves (Canis lupus) in Spain 12 for 1997-2007 to detect antibodies against canine distemper virus (CDV) and against 13 canine parvovirus (CPV) by indirect ELISA. Antibodies against CDV were detected in 18.7% of the analyzed animals and antibodies against CPV in 17.2%. There was no 14 15 difference in antibody prevalence to CDV between both species, even in the same 16 region (P > 0.05), but there was a significant difference in antibody prevalence to CPV 17 between foxes (5.1%) and wolves (62.2%) (P<0.05). In fox populations there was a 18 significant difference in antibody prevalence to CDV between geographic areas (Aragón 19 26.4%, La Mancha 7.8%, P < 0.05). In wolf populations there was a significantly higher 20 antibody prevalence against CPV (P<0.05) in Castilla y León (100%) than in the 21 Cantabric region (53.3%). There was no significant sex or age related difference in the 22 antibody prevalence against CDV or CPV in foxes. These results indicate that contact 23 with CDV is widespread among wild canid populations in Spain and that CPV is 24 endemic in the Iberian wolf population. The implications of these results are briefly 25 discussed.

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26 Keywords: Canis lupus, Serosurvey, Spain, Vulpes vulpes, Wild canids.

27

#### 28 Introduction

29 Canine distemper virus (CDV) and canine parvovirus (CPV) are common pathogens of 30 domestic and wild carnivores and have a worldwide distribution. CDV is a 31 Morbillivirus (family *Paramyxoviridae*) that is very resistant to cool temperatures but 32 quickly inactivated by ultraviolet light and by heat and drying. It is transmitted by 33 aerosols or contact with oral, respiratory, and ocular fluids and exudates containing the 34 virus. Therefore, dense populations of susceptible animals are needed to sustain 35 epidemics (Williams and Barker, 2001). CDV affects species belonging to all families 36 of the order carnivora and seems to have the major impact in wild carnivores and in 37 captivity (Montali et al., 1987). It is known that CDV caused the disappearance of the 38 last wild population of black-footed ferret (*Mustela nigripes*) (Thorne and Williams, 39 1988), and has also been considered responsible of declines of endangered species such 40 as African wild dog (Lycaon pictus) during the epizootic in domestic dogs (Canis 41 *familiaris*) and wild carnivores in the Serengeti (Alexander and Appel, 1994). In Spain, 42 CDV has been identified as cause of death or disease in domestic dogs (Nieto et al., 43 1987), polecats (Mustela putorius), American mink (Mustela vison), genet (Genetta 44 genetta), fox (Vulpes vulpes) and stone marten (Martes foina) (López-Peña et al., 2001). 45 In contrast, CPV is a Parvovirus (family *Parvoviridae*) that is very hardy, able to 46 survive up to 6 months at room temperature. It is transmitted by the fecal-oral route, 47 probably mainly through ingestion of virus from the environment, rather than by direct 48 contact with infected animals (Williams and Barker, 2001). CPV has a more limited 49 host range affecting different canids (Parrish, 1990). Although CPV has been linked 50 with mortality in young wolves (Johnson et al., 1994) and coyotes (Gese et al., 1997)

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and could threaten the viability of small isolated populations (Mech and Goyal, 1993, 1995), its impact in wild canid populations is largely unknown. In Europe, the presence of the virus has been reported in wild canid populations in Italy (Martinello et al., 1997) and data on strains isolated from wolves demonstrated that the same strain of CPV can circulate among domestic and wild canids (Battilani et al., 2001). No mortality due to CPV has been reported in wild canids from Spain but CPV is common in domestic dogs (Decaro et al., 2006).

58 The Iberian wolf (Canis lupus signatus) is considered a vulnerable species and its 59 population is estimated at a minimum of 2000 individuals (Blanco, 1998). Although the 60 main factors that can affect its survival are human causes or prev availability, infectious 61 diseases can also act as a mortality source. In fact, wolf mortality due to CDV and CPV 62 has been reported (Carbyn, 1982; Mech et al., 1997). The fox (Vulpes vulpes) in 63 contrast, is an abundant species with a wide distribution in the Spanish mainland 64 (Blanco, 1998). This species is susceptible to a number of diseases including CDV and 65 CPV (Artois et al., 1996). Thus, it could be a source of infection to other less abundant 66 species that live sympatrically. Additionally, feral or free roaming domestic dogs may 67 also become a source of infections for wild canids (Alexander and Appel, 1994). 68 However, feral dog abundances in Spain are low as compared to fox abundances (The 69 authors, unpublished data).

The objective in this study was to determine the prevalence of serum antibodies to CDV and CPV in Spanish foxes and wolves, and their differences across age and sex classes and geographical regions.

73

#### 74 Materials and methods

75 Sampling

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76	In the period of 1997-2007, serum samples were collected from 37 wolf and 137 fox					
77	sses from 4 Spanish regions including the Cantabric coast, Castilla y León, Aragón					
78	La Mancha (Figure 1). All animals were legally obtained (road kills and some foxes					
79	hunted for population control).					
80	The serum samples were obtained by cardiac puncture, centrifuged, and stored at -20°C					
81	their analysis. Age class (yearling <1 year vs. adult >1 year) was determined by					
82	tooth eruption and the degree of tooth wear (Sáenz de Buruaga et al., 2001). As a					
83	consecuence of opportunistic sampling, the age or sex was not known for 26 foxes and					
84	25 wolves.					
85	Serum antibody testing					
86	podies to CDV and CPV were determined by indirect enzyme-linked inmunoassay					
87	(ELISA), using commercial kits and following the manufacturer's instructions					
88	(Ingenasa, Madrid, Spain) (Corrain et al., 2007). To test for CDV, the serum samples					
89	were diluted 1/100, and anti-dog IgG was used as conjugate. Samples were considered					
90	positive if OD (optic density) value was higher than absorbance of positive control *					
91	0.2. To test for CPV with the same dilution and conjugate, samples were considered					
92	positive if the ratio S/P (sample optic density / positive control optic density) was $\geq$					
93	0.15.					
94						

95 Statistics

96 Seroprevalence was statistically analyzed considering the variables geographical area,

- 97 sex, age and host species using the SPSS 14.0 software. We used Chi-square tests and
- 98 Fisher test, with a 95% confidence level and a P value <0.05 was considered significant.
- 99

100 **Results** 

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101 Total antibody prevalence against CDV was 18.7%, being positive 9 (24.3%) of 37 102 wolves and 23 (17.1 %) of 134 foxes. Total antibody prevalence against CPV was 103 17.2% being positive 23 (62.2%) of 37 wolves and 7 (5.1%) of 137 foxes. There was no 104 significant difference in the prevalence of antibodies against CDV between both species 105 (P>0.05), but there was a significant difference in antibodies against CPV (P<0.05). 106 The antibody prevalence distribution against CDV and CPV by sex and age is shown in 107 Table 1. Although there was no significant difference between sex and age classes in 108 anti-CDV antibody prevalence in foxes, there was a slightly higher prevalence in adults 109 than in juveniles (P=0.1). There was no significant difference by age or sex in 110 antibodies against CPV. Differences in antibody prevalence by age or sex were not 111 analyzed in wolves since these variables were unknown for most animals. 112 The antibody prevalence against CDV and CPV by regions is presented in Figure 1. In 113 foxes, there was a significant difference between Aragón (26.4%) and La Mancha 114 (7.8%) (P<0.05), but not with other regions (P>0.05). In wolves, there was a significant 115 difference in CPV prevalence between the Cantabric region (53.3%) and Castilla y León 116 (100%) (P<0.05). In the Cantabric region, where both fox and wolf sera were available,

the prevalence of antibodies against CDV and CPV was of 22.2% and 23.3% and 10.6%
and 53.3% for foxes and wolves respectively (Figure 1). There was no difference in
CDV prevalence between both species in this area (P>0.05), but there was a significant
difference between both species in antibody prevalence against CPV.

121

#### 122 Discussion

123 This is the first report of contact with CPV and CDV in the wolf in Spain. Various tests 124 are available to detect antibodies to CDV and CPV. The standard technique for CDV is 125 the virus neutralization (Williams and Barker et al., 2001), but it is expensive, time-

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consuming, requires specialised laboratory facilities and good quality sera, with little or
no haemolysis. Hence, we decided not to use this tool to analyze our serum samples
(most of them from dead-found animals and with haemolysis). Recently, the ELISA
tests have been shown to be sensitive and specific against CDV and CPV (Ohashi et al.,
2001; Phukan et al., 2005).
The prevalence to CDV in wolves in our study (24.3%) was similar to the prevalence
described for this species in North America (7-67 %, Stephenson et al., 1982; Johnson

et al. 1994, Philippa et al., 2004). In contrast, the prevalence in foxes was 17.1%, which

134 is a higher value as compared to previous results for this species in central Europe (5-

135 13%, Frölich et al. 2000, Damien et al., 2002) and in North America (11%, Amundson

and Yuill 1981). However, the different serological tools used in some cases make the

137 interpretation and comparison between studies difficult.

138 The prevalence of antibodies against CDV was not age-specific or sex-specific in foxes. 139 The regional differences in CDV antibody prevalence could be due to different fox 140 densities, different spatial aggregation, or different degree of contact with domestic 141 dogs (e.g. Gortázar et al. 2003). In regions such as La Mancha, where foxes apparently 142 had less contact with CDV, the introduction of this pathogen could cause an epidemic 143 outbreak, because most individuals would be immunologically naïve (Appel, 1987). 144 This would have conservation implications since eventually CDV could spread from the 145 abundant fox population to other sympatric carnivores and affect endangered species 146 such as the lynx. Epidemic distemper outbreaks have happened in La Mancha in 1993 147 causing a 70% decrease in fox relative abundance (dropping counts, Ramos, 1995) and 148 in North-west Spain in 1997 (Marta Muñoz, pers. comm.).

The relatively high prevalence of antibodies against CPV in the Iberian wolf (62.2%), is
within the range reported for this species in North America (13-95%, Zarnke and

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Ballard 1987; Mech and Goyal, 1993). This high prevalence suggests a high exposure to infection but does not inform about disease, because the prevalence is measured in surviving individuals (Arjo et al., 2003). The prevalence of antibodies against CPV in wolves was significantly lower in the Cantabric region (53.3%) than in Castilla y León (100%). Since no other practical way of sampling wolves was available, our sampling strategy reduced all the possible inferences to the whole population.

The anti-CPV antibody prevalence in foxes (5.1%) was within the range reported by 157 other authors in Europe (0-9%, Mulley et al., 1982, Frölich et al., 2005) although 158 159 different tools were used for serological testing. The higher prevalence against CPV in 160 wolves than in sympatric foxes is surprising and suggests that foxes are not an 161 important source of infection to wolves. An alternative explanation could be that CPV 162 does not affect foxes. In an experimental infection of CPV in foxes Barker et al. (1983) 163 demonstrated the resistance of the species to the disease. Also, Truven et al. (1998) 164 amplified DNA sequences from tissues of free-ranging foxes and compared them with 165 the prototype viruses from dogs and cats. The parvovirus sequence was indicative of a true intermediate between CPV and feline panleucopenia virus, representing a link 166 167 between those viral groups.

We conclude that foxes and wolves from Spain have contact with CDV and that CPV is endemic in Iberian wolf populations. This information is of use in the frame of carnivore conservation. Further investigations are needed to study the epidemiology of these viral agents in the wild canid populations.

172

#### 173 Acknowledgements

This is a contribution to the agreement between IREC, SDGSA-MAPA and OAPNMMA, and to the agreement between CSIC and Principado de Asturias. RS

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- 176 Acknowledges a grant from Castilla La Mancha. We thank our colleagues at SERIDA
- 177 (Gijón, Asturias), Consultora de Recursos Naturales (Vitoria), Consultores en Biología
- 178 de la Conservación (Madrid) and Kati Gerique (Valencia) for providing samples.
- 179

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- Table 1. Distribution of antibodies against CDV and CPV by sex and age in foxes and
- wolves.
- 274
- 275
- 276
- 277 Table 1.

	CDV antibody prevalence				CPV antibody prevalence				
	Age		Sex		Age		Sex		
	Adults	<1 year	Males	Females	Adults	<1 year	Males	Females	
Fox	15/66 (22.7) <sup>a</sup>	4/45 (8.9)	14/63 (22.2)	5/52 (9.6)	1/60 (1.7)	2/52 (3.8)	2/61 (3.3)	3/55 (5.5)	
Wolf	4/8 (50)	0/4 (0)	1/3 (33.3)	3/10 (30)	7/8 (87.5)	4/4 (100)	3/3 (100)	8/10 (80)	
278	a. Number of positive animals/total analyzed (%).								
270									

279

280

281

Figure 1. Geographical distribution of antibody seroprevalence against CDV and CPV in

283 foxes (black circles) and wolves (white circles).

284

