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A novel point mutation in the CYBB gene promoter leading to a rare X minus chronic granulomatous disease variant – Impact on the microbicidal activity of neutrophils

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Key words: Chronic granulomatous disease, CYBB gene promoter, point mutation, cytochrome b558, NADPH oxidase, microbicidal activity.

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Abstract:

This article reports an atypical and extremely rare case of X-linked CGD in an Italian family characterized by a low expression of gp91phox (X91 CGD). A novel point mutation in the CYBB gene’s promoter (insertion of a T at position –54T to –56T) appeared to prevent the full expression of this gene in the patient’s neutrophils and correlated with a residual oxidase activity in the whole cells population. The expression and functional activity of the oxidase in eosinophils appeared to be almost normal. Gel shift assays indicated that the mutation led to decreased interactions with DNA-binding proteins. The total O$_2^-$ production in the patient’s granulocytes (5–7% of normal) supported no microbicidal power after 45 min and 60 min of contact with S. aureus and C. albicans, respectively. Despite this residual oxidase activity, the patients suffered from severe and life-threatening infections. It was concluded that in these X91 CGD neutrophils, the O$_2^-$ production per se was not sufficient to protect the patient against severe infections.
1. Introduction

Chronic granulomatous disease (CGD) is a rare congenital immunodeficiency syndrome clinically characterized by severe recurrent bacterial and fungal infections, which are relatively resistant to treatment, by conventional means [1,2]. The majority of the patients are now diagnosed before their second birthday, although some may remain undiagnosed until later childhood or even adult life [3,4]. CGD is due to a defect in the NADPH oxidase system of phagocytes [2]. This enzymatic complex is composed of a membrane-bound flavocytochrome $b_{558}$, the redox center of the oxidase, and three cytosolic components: p47phox, p67phox and p40phox. Two small G proteins, rac2 and rap1A/B, are also involved in regulating NADPH oxidase activity. Cytochrome $b_{558}$, the terminal redox membranous component of the phagocyte NADPH oxidase, is a heterodimer composed of an $\alpha$ subunit, the small p22phox, and a $\beta$ subunit, the heavy chain gp91phox or Nox2. Gp91phox is the flavinoxidase element capable of transferring electrons, while p22phox stabilizes the expression of this component [5]. The incidence of CGD is about 1 in 250,000 individuals. On the basis of the mode of inheritance, two forms of the disease are known: an autosomal form with mutations in CYBA, NCF1 and NCF2 genes encoding p22phox, p47phox and p67phox proteins, respectively, and an X-linked form with mutations in the CYBB gene encoding gp91phox, which accounts for more than 60% of CGD cases [5,7]. CYBB gene encompasses 13 exons spanning about 30 kb of human X chromosome genomic DNA [8,9]. In the so-called classical form of CGD which includes the majority of X-linked cases, cytochrome $b_{558}$ is absent and there is no detectable oxidase activity (X91°). A number of cases with “variant” forms of the disease, called X91– CGD have also been described, in which low levels of cytochrome $b_{558}$ expression are accompanied by a proportionally decreased NADPH oxidase activity [10]. Mutations associated with this phenotype are usually located in the coding region (exons) of CYBB [11-16]. These variants are of interest because
they cause a structural disorganization leading either to an incomplete loss of protein or to a partial dysfunction, or both. Mutations in the upstream promoter region of CYBB leading to the X91– CGD phenotype also have been described. To our knowledge only four point mutations were reported in this region (C-52T, C-53T, T-55C and A-57C) [17-19]. These mutations are located between the “CCAAT” and the “TATA” boxes in a consensus binding site for the ets family of transcription factors of the gp91phox promoter responsible for CYBB transcription [17]. Despite the low expression of gp91phox protein and the residual NADPH oxidase activity in phagocytes, the clinical appearance of these specific X91– CGD patients varies considerably from mild forms [18] to severe clinical phenotypes with multiple life-threatening infections during their life [17,19]. This suggests that the levels of O$_2^\cdot$ production and other derived reactive oxygen species (ROS) by activated phagocytic cells are critical for effective protection against infections. A striking point is that in most of the X91– CGD cases characterized by a mutation in the CYBB promoter, the CGD diagnosis is made in adolescents (>10 years) or in adults.

We report a novel point mutation in the CYBB gene promoter (insertion of a T at position -54/-56) leading to a rare X91– CGD case in an Italian family with two members affected by CGD. The mutation caused a reduced expression of gp91phox at the mRNA and protein level associated with a decreased ability to generate O$_2^\cdot$ and H$_2$O$_2$ (~ 7% of control neutrophils). A decrease in the binding of nuclear factors to the mutated promoter region was found in electrophoretic mobility shift assays. However, the patient’s eosinophils were NBT-positive, displayed DHR oxidation and were gp91phox-positive. Despite a weak respiratory burst activity, the patient’s granulocytes did not kill either S. aureus or C. albicans even with long incubation times. The CGD diagnosis for both patients was made in adulthood. One of them died from a bilateral pneumonia while the other suffers from sporadic respiratory and skin infections that are kept under control by conventional antibiotic therapy. These
observations raise questions about the role and the levels of ROS needed for protection against bacterial and fungal infections.

2. Materials and Methods

2.1 Materials

Phorbol 12-myristate 13-acetate (PMA) and cytochrome c (type VI, from horse heart) were from Sigma Chemical Co. (St. Louis, MO). Percoll, reagents and molecular weight markers for SDS-PAGE were obtained from Amersham Pharmacia Biotech (Little Chalfont, Buckinghamshire, UK). Nitrocellulose sheets for Western blotting were purchased from Bio-Rad Laboratories. Monoclonal antibodies (MAbs) 449 and 48 were provided by D. Roos and A.J. Verhoeven. Mab 7D5 was provided by Clinisciences (Montrouge, France). Control IgG1 mAb from a mouse myeloma was purchased from Sigma Chemical Co. Alexa Fluor 647 rat anti-human CD294 (CRTH2) was purchased from BD Biosciences (San Jose, CA, USA).

2.2 Cases report of patients II-2 and III-5 of family C (Figure 1)

Patient II-2 was born in 1947. His mother died several years ago from undefined pathology while his father passed away recently. His son (III-1) is in good health and his daughters (III-2, III-3) are carriers of the disease. The patient sisters are both CGD carriers and one of them died recently. The brother died of pneumonia at the age of 14. Diagnosis of CGD was made in 1991 immediately after his nephew (patient III-5) was diagnosed as having CGD at the age of twenty during the terminal phase of the disease. Patient II-2 was relatively well until he was 5 years old. He subsequently had a history of skin abscesses, pulmonary infections with pleuritis treated with antibiotic therapy. The clinical history of patient II-2 is characterized by episodes of bronco-pneumonia, skin ulcero-necrotic lesions and an otitis
media with tympanic membrane perforation, which responded to antibiotic treatment. At present the patient is in a relatively good health and does not receive antibiotic profilaxis.

Patient III-5, who is the nephew of patient II-2, was born in 1970 and died in 1990. He had no complaints and was healthy until the age of 19. In 1989 he developed severe multiple site infection of the skin with necrotic granuloma with involvement of the right latero cervical nodes and of the lung with pleural effusion sepsis that required hospitalization. *Serratia marcescens* was isolated from the blood and the lymph node. Lymph node and pleural lesions required surgical drainage. On that occasion, a marked defect in the respiratory burst of granulocytes was detected by a luminol-dependent chemiluminescence assay using opsonized zymosan as stimulating agent. Antibiotics including ceftazidime and netilmicin were administered and the patient recovered in five weeks. He was admitted again to hospital six months later, in March 1990, with fever, cough, dyspnoea and radiological evidence of a bilateral pneumonia. After a few days from admission the general conditions of the patient worsened and intensive care unit assistance was required. Two blood cultures were positive for *Staphylococcus sp.* (non aureus). *Pseudomonas aeruginosa* and *Candida albicans* were isolated from bronchial brushings. Antibiotic treatment, including ceftazimide, piperacilline, netilmicine and teicoplanine failed to control fever and pulmonary infection. After diagnosis was made, and before the molecular nature of the defect was known, it was hypothesized that the low activity of NADPH oxidase could be the result of enzyme instability. Accordingly, it was thought that "younger" cells could possess higher oxidase activity and kill microorganisms more efficiently. Therefore, GM-CSF was used with the aim of accelerating neutrophil release from the bone marrow and exploiting its known ability to prime neutrophils for enhanced response to agonists [20].

Human recombinant GM-CSF (kindly provided by Schering-Plough) was given, beginning with a dose of 5 µg/Kg/die escalating to a maximum of 10 µg/Kg/die and then modulating the
dose in order to maintain WBC count below $10 \times 10^9/l$. GM-CSF was given subcutaneously at 12 hrs intervals. Treatment with GM-CSF lasted for 22 days and led to a fast and sustained increase of the WBC and neutrophil count, without detectable effects on haemoglobin concentration and platelet count, and without side effects. However, in spite of the marked increase in the neutrophil count, there was no change in the percentage of NBT-positive cells and in the amount of superoxide anion produced after PMA stimulation (data not shown). After a slight and temporary clinical improvement, the patient’s conditions worsened, with high fever, unconsciousness and increasing dyspnoea, which required mechanically assisted ventilation. Left pneumo-thorax developed and death occurred 38 days after admission. During the late course of the disease *Pseudomonas* and *Staphylococcus* were again isolated from bronchial brushings and from the blood, respectively.

2.3 Isolation of polymorphonuclear and mononuclear leucocytes

Human neutrophils were isolated from citrated blood samples of patients, their parents and healthy donors after their informed consent, by sequential dextran sedimentation and Ficoll-Hypaque differential density centrifugation, followed by erythrocyte lysis by hypotonic treatment, as described elsewhere [21]. The isolated neutrophils were resuspended in Ca$^{2+}$ and Mg$^{2+}$ free HEPES – buffered saline (HBS) solution containing 140 mM NaCl, 5 mM KCl, 5 mM Glucose, 5 mM HEPES, pH 7.4 and 0.2% bovine serum albumin (BSA). Immediately before use, cell suspensions were supplemented with 1 mM CaCl$_2$ and 1 mM MgCl$_2$. Isolated mononuclear cells were transformed with the B95-8 strain of Epstein Barr virus (EBV) and were grown in RPMI-1640 supplemented with 10% fetal calf serum, 100 U/ml penicillin, 50 µg/ml streptomycin, at 37°C in a 5% CO$_2$ atmosphere [22].

2.4 Assay of superoxide anion production:
Superoxide anion was measured spectrophotometrically by the superoxide dismutase-inhibitable cytochrome c reduction at 550 nm, as previously described [19,23].

2.5 Assay of hydrogen peroxide production:

Hydrogen peroxide was measured by the dihydrorhodamine-1,2,3 (DHR) assay. In this assay, the nonfluorescent DHR is converted by hydrogen peroxide to the green fluorescent rhodamine 123 and analysed using flow cytometry [19]. Neutrophils were loaded with 2 μM DHR 123 (Molecular Probes, Eugene, OR, USA) at room temperature for 30 minutes, in the dark. After loading and addition of 1 mM sodium azide, the samples were prewarmed at 37°C in a shaking water bath and after 10 minutes the cells were stimulated with 20 ng/ml phorbol-myristate acetate (PMA) for 15 minutes; the reaction was stopped by dilution with ice cold PBS. Samples were kept on ice and cell-associated fluorescence (FL-1) was measured by flow cytometry (FACScalibur™, BD Biosciences, San Jose, CA). In experiments where expression of the eosinophil marker CRTH2 was also measured, the samples were further incubated with 2.5 μg/ml Alexa Fluor 647 rat anti-human CD294 (CRTH2) for 30 min and washed twice with cold HBS-BSA, before cytometry readings.

2.6 NBT test

Neutrophils were suspended at 0.2×10⁶/ml in HBS-BSA containing 1 mM EDTA to prevent cell aggregation and 1 mM NBT. The reaction was started by addition of 100 ng/ml PMA. After incubation for 15 min at 37°C cytospins were made, fixed with methanol and counterstained with a solution of 0.4% safranin in 25% ethanol. In some experiments the NADPH oxidase inhibitor diphenyleneiodonium (DPI) (f.c. 5μM) was included in the incubation mixture.
2.7 Cytochrome b<sub>558</sub> spectroscopy

Reduced minus oxidized optical spectra of cytochrome b<sub>558</sub> were recorded at room temperature on a DU 640 Beckman spectrophotometer, after addition of a few crystals of sodium dithionite, as previously reported [24]. A molar extinction coefficient ε<sub>426-410 nm</sub> = 200 mM<sup>-1</sup> cm<sup>-1</sup> for the Soret band was used for calculations.

2.8 Expression of gp91<sub>phox</sub> in neutrophils and eosinophils as measured by flow cytometry

Granulocytes (10<sup>6</sup> cells) were suspended in 0.1 ml HBS-BSA and incubated with 5 µg/ml of mab 7D5 (anti-gp91<sub>phox</sub>) [25] or control isotype-matched (IgG1) mab for 30 min on ice. The cells were washed twice and incubated with Alexa Fluor 488 goat-F(ab)<sub>2</sub> anti-mouse IgG1 (Invitrogen, Cergy Pontoise, France) for 30 min on ice. After two washes as above, the cells were incubated with 2.5 µg/ml Alexa Fluor 647 rat anti-human CD294 (CRTH2) for 30 min on ice and washed [26, 27]. Cell-associated fluorescence (FL1 and FL4) was measured on 100,000 events.

2.9 Assay of microbicidal activity

The microbicidal activity was tested according to a recently described method [28]. Briefly, neutrophils (final concentration: 4 × 10<sup>6</sup>/ml) were incubated at 37 °C with serum-opsonized Staphylococcus aureus (strain 502 A, ATCC 27217), Escherichia coli (ATCC 25922, a serum-resistant strain) or a clinical isolate of Candida albicans at a microorganism/neutrophil ratio of 3:1 for bacteria and 2:1 for the fungus, in HBS-BSA containing 1 mM CaCl<sub>2</sub> and MgCl<sub>2</sub>. At the indicated times of incubation aliquots were diluted 50 times in water brought to pH 11.0 with 1M NaOH. After 5 min at room temperature samples were vortexed, diluted in 0.9 % NaCl solution and plated on Petri dishes. After
overnight incubation, colony forming units (CFU) were counted and the percentage of killing was calculated with respect to the number of CFU at time 0 of incubation.

2.10 SDS-PAGE and immunoblotting

Neutrophil proteins were solubilised in 1% Triton X-100, separated by SDS-PAGE in 10% (w/v) acrylamide gel with a 5% (w/v) stacking gel [29], electrotransferred to nitrocellulose [30] and immunodetected using the monoclonal antibodies 449 and 48 directed against p22phox and gp91phox, respectively [31] and by polyclonal antibodies directed against p47phox and p67phox [32].

2.11 Electrophoretic mobility shift assays (EMSA)

Nuclear extracts. Nuclear extracts of undifferentiated PLB-985 cells were prepared with the NE-PER nuclear extraction reagents of the Pierce kit (Thermo Fisher Scientific Inc., Waltham, USA) according to the manufacturer’s instructions and stored at −80°C. The protein concentration was determined with the BioRad protein assay kit (Biorad, Marnes-La-Coquette, France).

Oligonucleotides. Oligonucleotides used as double-stranded competitors and probes are as follows: WT CYBB [−68 to −30] bp (WT, 5′-ctatgcttcttccaatgaggaatgaacagcag-3′); X91− mutated CYBB (X91− CGD, 5′-ctatgcttcttccaatgaggaatgaacagcag-3′). Oligonucleotide probes were labelled by biotin according to Biotin 3’ End DNA Pierce Labelling Kit (Thermo Fisher Scientific Inc.), followed by annealing with the complementary strand oligonucleotides.

Electrophoretic mobility shift assays were performed in a 20-µl total volume with 10 µg of nuclear protein extract in 10 mM Tris, 50 mM KCl, 1 mM DTT (pH 7.5), 50 ng/µL Poly (dI-
dC). Nuclear extracts were preincubated for 15 min on ice with or without a 20-, 50-, or 100-fold molar excess of unlabelled double-stranded oligonucleotide competitors followed by an additional 30 min on ice after the addition of 1 nM of biotin-labelled double-stranded oligonucleotides. Samples were subjected to a 5% polyacrylamide gel electrophoresis at 100 V for 2 h in a 0.5X Tris-Borate-EDTA buffer (pH 8.3) and transferred to a nylon membrane Hybond™- N+ (Amersham, GE Healthcare, Munich, Germany). After transfer the membrane was immediately cross-linked for 45–60 s at 120 mJ/cm² using an UV-light cross-linker instrument equipped with 254-nm bulbs. The biotin-labelled DNA was detected by chemiluminescence using a luminol/enhancer solution and a stable peroxide solution (Pierce labelling kit). Membranes were exposed to an x-ray film for 15 min before developing.

2.12 Preparation of RNA and DNA

Total RNA was isolated from either mononuclear cells or EBV-transformed B lymphocytes, using a modified single-step method [33, 34]. Genomic DNA was purified following classic procedures. Genomic DNA was also purified from the peripheral blood of patient II-2 and healthy donor according to the manufacturer’s instructions (Wizard kit, Promega, France).

2.13 cDNA amplification and sequencing

First-strand cDNA was synthesized from 5 µg of total RNA from Epstein Barr virus immortalized B lymphocytes or native B lymphocytes or granulocytes by reverse transcriptase (RT) reaction according to the manufacturer's instructions (QBiogen, Illkirch, France). Total gp91phox cDNA was immediately amplified by PCR in three overlapping fragments [35]. Aliquots of 10 µl PCR products in bromophenol blue solution were run together with a scale of DNA ladders (markers VI, Roche Diagnostics, Indianapolis, IN, USA) on 1.5% (wt/vol)
agarose or on 5% acrylamide gels containing 1 µg/ml ethidium bromide. The bands were photographed under UV (Vilber Lourmat, Biolabo Scientific Instruments, Archamps, France). In some cases PCR products were purified from agarose gel with the Sephaglas kit (Amersham Pharmacia, Orsay, France). All RT-PCR products were sequenced by Cogenics (Meylan, France) using the Abi Prism automatic sequencer (Perkin Elmer, Foster City, CA, USA). For each patient, the complete encoding region of CYBB was sequenced.

2.14 Semi-quantitative PCR

Semi-quantitative PCR was performed to evaluate CYBB gene expression in mononuclear cells or granulocytes. Two microliters of cDNA (obtained as described before) was used in 100 µL of total reaction mixture containing 10 µL of 10X Qbiogen incubation mix supplemented with 0.2 mM Mg++, 0.25 µM dNTP, specific primers 1 and 2* (see primer sequence in Table II), 0.25 µM β-actin primers (3 and 4*) and 25 mU/µL Taq DNA polymerase (QBiogene, Illkirch, France). The reaction mixture was thermally cycled in a Minicycler™ (MJResearch, St Bruno, Canada) once at 95°C for 5 min, 25 times at 95°C for 1 min, 57°C for 1 min, 72°C for 90 s and then once at 72°C for 7 min. Aliquots of 10-µl PCR products in bromophenol blue solution were run together with a scale of DNA ladders (marker XIV, Roche, Meylan, France) on 1.5% (wt/vol) agarose gels containing 1 µg/ml ethidium bromide and visualized on a UV transilluminator.

2.15 Genomic DNA amplification

Purified genomic DNA was amplified using a forward primer P1 CAGCAAGGCTATGAATGCTGTTC (position –427 in the CYBB promoter region and the backward one P2* GGAGAGCCCCTCATTCAACAG (position +45). PCR products were automatically sequenced by Cogenics (Meylan, France) using the Abi Prism automatic sequencer (Perkin Elmer, Courtaboeuf, France) with forward and backward primers [19].
2.16 Protein determination

Protein content was measured by using the Pierce® method [36].
3. RESULTS

Fig. 1 shows the pedigree of Family C studied in this paper. Patient II-2 was diagnosed with CGD immediately after his nephew (patient III-5) was diagnosed as having CGD at the age of 20 on the basis NBT test, PMA-induced superoxide production and cytochrome \( b \) content (Table 1). Patient II-2 was 43 at the time of diagnosis and his neutrophils in terms of respiratory burst activity (NBT and superoxide production) and cytochrome \( b \) content had characteristics similar to neutrophils of patient III-5 (Table 1). Production of superoxide anion by neutrophils from patients II-2 and III-5 after PMA stimulation was 7 and 5 % of control cells, respectively (Table 1). Neutrophils from family members II-3, II-4, II-2, III-3 and III-4 generated intermediate amounts of superoxide ranging from 34% to 60% of control (Table 1). These data are compatible with an X-linked mode of inheritance of the defect. This conclusion was confirmed by the results of the NBT test that showed that neutrophils from II-3, II-4, III-2, III-3 and III-4 were a mosaic in terms of NBT reduction, with two populations, one strongly positive, and the other negative while neutrophils from II-1 and III-1 were 100% NBT positive. In patients II-2 and III-5 a few cells were weakly NBT positive (Fig. 2 arrows and arrowheads). Such NBT positive cells were no longer apparent when DPI was included in the incubation mixture. Among the NBT positive cells, the more weakly positive (arrows) appeared to be neutrophils while the more strongly positive (arrowheads), based on the nuclear shape, the size and color of intracellular granules could be identified as eosinophils. The percentage of eosinophils as determined in Diff-Quick stained smears in patients II-2 and III-5 never exceeded 3% (data not shown).

To better establish if neutrophils from patient II-2 exhibited a NADPH oxidase activity, we measured the respiratory burst using a method much more sensitive than the NBT reduction test, i.e. the DHR oxidation assay—Fig. 3 shows that neutrophils from patient II-2 presented a homogeneous cell population with a peak response corresponding to about 5% of
control. Such a response was almost completely abolished by DPI. No response was observed with neutrophils from an X91° CGD patient. In order to establish whether a population of NADPH oxidase-positive eosinophils, as indicated by the NBT reduction test, was detectable using the DHR assay in the granulocytes of patient II-2, we conducted an experiment in which eosinophils were identified with an anti-CRTH2 mAb and with acquisition of a number of events (100,000) larger than that used in Fig. 3 (10,000). Figure 4 shows that, indeed, a small percentage of DHR and CRTH2-positive cells (eosinophils) were present in the granulocyte preparation of patient II-2. To demonstrate the presence of cytochrome $b_{558}$ in the patients’ neutrophils, reduced minus oxidized difference spectra of neutrophil homogenates and Western blot analysis were performed. As illustrated in Fig. 5A and Table 1, reduced-minus-oxidized spectra revealed a low but detectable amount of cytochrome $b_{558}$. The spectrum of X91° CGD neutrophils did not present the peaks characteristic of cytochrome $b_{558}$. Cytochrome $b_{558}$ content in patients II-2 and III-5 was 14 and 10 % of normal neutrophils, respectively, while the neutrophils of a heterozygous member of family C (patient III-4) contained 44 % of control cytochrome $b_{558}$ (Table 1).

The presence of low amounts of cytochrome $b_{558}$ in neutrophils of these patients was confirmed by Western blot analysis using an identical amount of 1% Triton X100 soluble extract in all samples (50 µg). Fig. 5B shows that in comparison with normal cells, the two bands corresponding to gp91phox and p22phox were very faint. The specificity of the bands detected by the use of monoclonal antibodies directed against the two subunits of cytochrome $b_{558}$ was confirmed by the absence of gp91phox and p22phox in the extracts from neutrophils of an X91° CGD patient. The cytosolic components of NADPH oxidase (p47phox, p67phox, p40phox and Rac2) detected by Western blot analysis were normal (data not shown). Since the spectroscopic and Western blot analyses do not distinguish eosinophils from neutrophils and since in the NBT slide test eosinophils appeared to stain more strongly than neutrophils...
(Fig. 2), a flow cytometry experiment was devised that would distinguish gp91phox expression in neutrophils and eosinophils. Figure 6 shows the dot plot profiles of granulocytes from a normal subject (top panels) and from patient II-2 (bottom panels) incubated simultaneously with the anti-eosinophil mab CRTH2 and with the anti-gp91phox 7D5 mab antibodies (gp91phox) or an isotype-matched nonspecific mAb (control mAb). The percentage of eosinophils in the two cell preparations was 0.9 for the normal subject and 2.5 for patient II-2. The right panels show that both neutrophils (99.0%) and eosinophils (0.75%) from the normal subject (top panel) express gp91phox. In the case of patient II-2 (bottom panel), despite the low but measurable NADPH oxidase activity in his granulocytes, as shown previously (Figs. 2, 3 and 4), gp91phox was not detected in the RCHT2- population (97%) and only eosinophils (1.9%) were gp91phox-positive.

RT-PCR and sequence analysis were performed to localize the mutation in the mRNA encoding gp91phox of patient II-2. mRNA was reverse-transcribed into three overlapping cDNA fragments with three pairs of oligonucleotide primers derived from the gp91phox cDNA and were sequenced as described in Materials and Methods. No mutations in the cDNA of gp91phox protein were found. A portion of the gp91phox promoter region in genomic DNA of CYBB was amplified by PCR [19]. Using sequence analysis, a thymidine insertion was found located between position –54T to –56T (Figure 7A). This mutation is located in a region between the “CCAAT” and the “TATA boxes in a consensus binding site for the ets family of transcription factors of the gp91phox promoter responsible for CYBB transcription (Fig. 7B). Genomic DNA from 50 control subjects was analyzed and no mutations were found.

Then electrophoretic mobility shift assays were performed to evaluate the impact of this new mutation of the CYBB promoter on the specific binding with regulatory transcription factors of a nuclear protein extract from PLB-985 cells. Figure 8 shows that a DNA/protein
complex (indicated by the arrow) is formed when a labelled wild-type double-strand oligonucleotide of the CYBB promoter region (−68 to −30) is used as a probe. The DNA/protein binding is specific as it is progressively inhibited by competition with a 20-, 50- and 100-fold molar excess of the unlabelled WT double-stranded oligonucleotide probe. On the contrary, adding an increasing amount of X91− CGD unlabelled double-stranded oligonucleotide probe is ineffective.

In order to study the impact of the defect of transcription factor binding on the mutated CYBB promoter region, the amount of gp91phox mRNA from patient II-2’s mononuclear cells was estimated by semi-quantitative RT-PCR using β-actin, a housekeeping gene as an internal standard. The PCR was performed with 20, 25, and 30 cycles (data not shown). Finally, 25 cycles of PCR was chosen in order to be in the exponential phase of the PCR amplification and before reaching the plateau for both tested mRNA. As shown in Fig. 9, a 624-bp fragment of comparable intensity was obtained from the control, X91+ and X910 CGD patients’ gp91phox mRNA. In contrast, patient II-2’s gp91phox mRNA amplification was very low while the signal of the housekeeping gene was similar in all the tested mRNA. We also did the same experiment with total ARN purified from the granulocyte population of patient II-2 and we obtained the same results (data not shown).

To determine the impact of the residual NADPH oxidase activity of neutrophils on microbicidal activity, in vitro killing assays were performed using S. aureus, C. albicans and E. coli as targets. Fig. 10 shows that the microbicidal activity of neutrophils from patient II-2 was superimposable to that of neutrophils from an X910 CGD patient, i.e. no killing of S. aureus and C. albicans, normal killing of E.coli. No killing of both S. aureus and C. albicans could be observed even after prolonging to 90 min the time of incubation (data not shown). However the normal killing of E. Coli for both X910 and X91− CGD highlights that it was NADPH oxidase independent.
4. Discussion

The majority of genetic lesions found in X91 CGD variants of chronic granulomatous disease are missense mutations or small deletions located in the encoding region of CYBB, whereas, up to now, only four point mutations have been described in the CYBB promoter region [17-19]. All these X91 CGD variants are characterized by a low expression of cytochrome b$_{558}$ and display some NADPH oxidase activity.

We report here for the first time the occurrence of an insertion in the promoter of CYBB gene (T insertion between position -54 and -56) leading to a rare variant of X91 CGD in an Italian family. On the basis of the amount of superoxide anion produced and the NBT test it was possible to demonstrate that neutrophils from the two members affected, II-2 and III-5, retained some oxidase activity (Table 1 and Figs. 1 and 2). Assay of hydrogen peroxide production by DHR 123 oxidation in neutrophils from patient II-2 showed a single peak of fluorescence, indicating that neutrophil population responded homogeneously to PMA with a respiratory burst. Such a response was only about 6 % of control (Fig. 3). The NADPH oxidase dependence of this response, as well as of the response observed in the NBT test was demonstrated by the complete sensitivity to inhibition by DPI (Fig 2 and Fig. 3). A homogeneous weakly responsive neutrophil population was also reported in other X91 CGD patients but no identification of the specific CYBB mutation has been done for these subjects [37-38]. The eosinophils from patient II-2 strongly reduced NBT and oxidized DHR (Figs. 2, 3, and 4). In the neutrophil population from patient II-2 (which is RCTH2), cytochrome b$_{558}$ could not be detected, probably because the amount of gp91phox expressed at the surface of each cell is low and is under the flow cytometry's sensitivity limit (Fig. 6). However, the low percentage of eosinophils in patient II-2 (about 2% of the granulocyte population) cannot explain the homogeneous weakly but DPI inhibited respiratory burst measured in the PMA-activated neutrophil population (Figs. 3 and 4).
A small population (5% to 15%) of granulocytes with completely normal respiratory burst activity has been previously found in X91-CGD patients (T-55C mutation in the promoter region of CYBB [17]. In two other X91-CGD cases caused by point mutations in the same CYBB promoter region (C-52T, C-53T), a small population of NBT-positive cells was described that turned out to be eosinophils [18]. The presence of oxidase positive eosinophils in these patients may be explained by the demonstration that gp91phox expression in eosinophils is regulated by specific transcription factors (GATA1 and GATA2) different from those operative in neutrophils monocytes and B-lymphocytic cells [39].

The new insertion mutation described here (Fig. 7) is located between the “CCAAT” and “TATA” box in a consensus binding sequence for the ets family of transcription factors of the gp91phox promoter site (5'-GAGGAAAT-3', lower strand, −57 to −50 bp), that are involved in eukaryotic gene transcription [40]. Mutations in this region are known to be related to the loss of association of DNA-binding proteins (Elf-1 and PU.1) and to prevent the full expression of the CYBB gene in neutrophils and monocytes [17, 41, 42]. Indeed, using EMSA we demonstrated a DNA–protein binding complex formation of nuclear extract from PLB-985 cells with the biotin-labelled wild-type double-strand oligonucleotide of the CYBB promoter region (−68 to −30). The specificity of the binding was demonstrated because an increasing amount of unlabelled WT oligonucleotide probe decreased the DNA–protein binding complex formation while the mutated X91− oligonucleotide probe was ineffective (Fig. 8). The consequence of this transcription factor binding defect was a reduced gp91phox mRNA expression in mononuclear cells or granulocytes (Fig. 9) correlated with a diminished cytochrome b558 expression in patient II-2’s neutrophil population of (Fig. 5). The diminished expression of gp91phox in patients II-2 and patient III-5 correlates with the low respiratory burst of their neutrophils. The finding that the eosinophils of our patients were oxidase positive is in keeping with the above mentioned observation that expression of gp91phox in
eosinophils is regulated by transcription factors different from those active on neutrophils
[39]. On the whole, the phenotypic profile of these cells is very similar to that of other X91-
CGD neutrophils with point mutations in the CYBB promoter (C-52T, C-53T, T-55C, A-57T)
[17-19].

In order to understand the role of the residual NADPH oxidase activity in the microbicidal
activity of X91- CGD neutrophils, killing of Gram-positive (S. aureus) and Gram-negative
(E. coli) bacteria and of a fungus (C. albicans) by patient II-2 neutrophils were assessed and
compared with killing by X910 CGD neutrophils (Fig. 10). As previously shown [28],
neutrophils from the X910 CGD patient were unable to kill both S. aureus and C. albicans and
ekilled normally E. coli. The results obtained with patient II-2 neutrophils indicate that the
residual NADPH oxidase activity of patient II-2 neutrophils (5-7 % of control) is not
sufficient to guarantee any killing of S. aureus and C. albicans in vitro, even after long
incubation times (90 min, data not shown). Bactericidal activity of X91- CGD neutrophils was
performed in only few studies [11,37]. The killing of S. aureus by X91- CGD neutrophils in
these studies was found only partially defective and comparable to that of X910 CGD
neutrophils. The discrepancy of these data with ours can be explained by the fact that
techniques commonly used to assess bacterial killing were affected by a methodological flaw
that undermined reliability of the results obtained [28]. In an additional study [18] a defective
killing of E. coli by X91- CGD neutrophils was demonstrated. In that study, killing of E. coli
(strain ML35) was measured by following the kinetics of perforation of the bacterial cell
envelope [43]. E. coli appear to be efficiently killed by non-oxidative mechanisms ([28,44]
and data presented in this study). Therefore we have no ready explanation for the differences
in killing between ref.18 and our results, considering that perforation assay of killing and the
classical killing assay (CFU) were shown to give comparable results [43]. It is likely that the
microbicidal defect demonstrated using the perforation technique does not reflect changes in
the number of viable bacteria detected as colony forming units or that the observed differences depend on the different strains of *E. coli* used.

The strongly defective killing of *S. aureus* and *C. albicans* which are preferential pathogens found in CGD, in spite of the low O$_2^-$ production may well explain the clinical conditions of patients II-2 and III-5, both of which suffered from recurrent and severe infections requiring hospitalization and patient III-5 died from pneumonia at the age of 20 when the diagnosis of CGD was made. It is the case for some previously described X91 CGD patients with the same type of *CYBB* promoter’s mutations [17, 45]. However the frequency and the severity of infections among the X91 CGD variant patients characterized by point mutation in the same *CYBB* promoter region, vary considerably. Reasons for discrepancies between the infectious complications and the respiratory burst capabilities in such patients are unclear. However a mild clinical expression in these X91 CGD variant patients can be explained by a high level of eosinophils (>10%) normally expressing NADPH oxidase in the blood [18]. However the compensation mechanism of eosinophil over-production is not known. Mothers who are carriers of the classical X91° CGD are generally protected against severe infections even in cases of extreme lyonization, with <15 % of neutrophils with a normal respiratory burst [46]. This indicates that even a low percentage of neutrophils with a normal respiratory burst is enough to protect against infections. In our study, neutrophils from patients II-2 and III-5 produced reactive oxygen species at a rate corresponding to 5-7 % of control. This response, however, is “diluted” among the whole cell population. Evidently, such a low level of oxidative metabolism, on a per cell basis, is not sufficient to allow microbial killing and to protect from infections. These observations raise questions about the levels of respiratory burst activity required for microbial killing and studies on this line are now underway using myeloid lines transfected with NOX mutants exhibiting various levels of oxidative activity.
Systematic characterization of gene mutations in CGD patients is essential for genetic counseling and prenatal diagnosis. It would be also of help in relating the clinical form of the disease to the type of gene affected and/or the type of mutation found. However, we believe that the number of X91 CGD variant cases with a well-documented clinical history and a defined genetic lesion described up-to-now is not sufficient to provide a clear and definitive correlation between mutation and severity of the clinical course.
Acknowledgments

The authors are grateful to Dr D. Roos and Dr A. J. Verhoeven for the generous gift of mAb449 and mAb48 antibodies raised against the two subunits of the cytochrome $b_{558}$. We extend our thanks to Dr. Marilena Granzotto for invaluable help in flow cytometry and Prof. B. Toussaint for helpful discussions concerning the EMSA experiments. Special thanks are extended to Lila Laval for her excellent secretarial work and to Linda Northrup for editing the manuscript.
References


Table 1

Superoxide anion (O$_2^-$) production, NBT reduction, and cytochrome b$_{558}$ content of neutrophils from family C members and the members of a family with a CGD patient affected by the classical X91$^0$ form of the disease. For experimental details see Materials and Methods. In parenthesis number of experiments.

<table>
<thead>
<tr>
<th>Subjects</th>
<th>O$_2^-$ production (nmol/20 min/10$^6$ cells)</th>
<th>NBT reduction (% NBT-positive neutrophils)</th>
<th>Cyt. b content (pmol/mg cell protein)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Family C</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>II-1</td>
<td>226.0</td>
<td>100 strong</td>
<td>NT</td>
</tr>
<tr>
<td>II-2</td>
<td>13.2</td>
<td>100 weak to negative</td>
<td>20.0</td>
</tr>
<tr>
<td>II-3</td>
<td>68.1</td>
<td>52 strong 48 weak to negative</td>
<td>NT</td>
</tr>
<tr>
<td>II-4</td>
<td>83.3</td>
<td>53 strong 47 weak to negative</td>
<td>NT</td>
</tr>
<tr>
<td>III-1</td>
<td>217.3</td>
<td>100 strong</td>
<td>NT</td>
</tr>
<tr>
<td>III-2</td>
<td>118.9</td>
<td>64 strong 36 weak to negative</td>
<td>NT</td>
</tr>
<tr>
<td>III-3</td>
<td>91.3</td>
<td>53 strong 47 weak to negative</td>
<td>NT</td>
</tr>
<tr>
<td>III-4</td>
<td>19.8</td>
<td>56 strong 44 weak to negative</td>
<td>72.5</td>
</tr>
<tr>
<td>III-5</td>
<td>8.9±1.8 (3)</td>
<td>100 weak to negative</td>
<td>14.8±4.3 (3)</td>
</tr>
<tr>
<td><strong>Healthy donor</strong></td>
<td>197.9±7.1 (3)</td>
<td>100 strong</td>
<td>166.0</td>
</tr>
<tr>
<td><strong>Family of patient with X91$^0$ CGD</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Father</td>
<td>181.9</td>
<td>100 strong</td>
<td>100.7</td>
</tr>
<tr>
<td>Mother</td>
<td>99.2</td>
<td>61 strong 39 negative</td>
<td>62.5</td>
</tr>
<tr>
<td>Son</td>
<td>&lt;0.5</td>
<td>100 negative</td>
<td>&lt;5</td>
</tr>
</tbody>
</table>
Table II

Sequences of oligonucleotide primers used for the semi-quantitative RT-PCR amplification of gp91\textit{phox} cDNA.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Primers</th>
<th>Sequence</th>
<th>Position</th>
</tr>
</thead>
<tbody>
<tr>
<td>β-actin</td>
<td>1</td>
<td>CCTCTGCCACCATGGGAATCTGGAGAATG</td>
<td>294</td>
</tr>
<tr>
<td></td>
<td>2*</td>
<td>GACTTCAAAGTAAGACCTCGGATG</td>
<td>1131</td>
</tr>
<tr>
<td>gp91\textit{pho}</td>
<td>3</td>
<td>ATCTGGACACCACACTCTCTCAATGAGCTGCG</td>
<td>2</td>
</tr>
<tr>
<td></td>
<td>4*</td>
<td>CGTCATACTCTGCTGTGCTGATCCACATCTGC</td>
<td>624</td>
</tr>
</tbody>
</table>

* Anti-sense strand.
Figure legends

Fig 1. Pedigree of family C. Individuals II-3, II-4, III-2, III-3, III-4 were proven to be carriers for X-linked CGD on the basis of NBT reduction and superoxide production (Table 1). In contrast, individuals II-1 and III-1 were normal in terms of NBT reduction and superoxide production (Table 1). [□], normal male; [○], normal female; [●], carrier female; [■], affected male; (†), deceased; NT, not tested.

Fig 2. NBT test in neutrophils from the patient II-2. Purified neutrophils from patient II-2 or a healthy donor were pre-incubated or not with 5 μM DPI and stimulated with 20 ng/ml PMA in presence of 1 mM NBT for 15 min at 37 °C. Arrows indicate neutrophils weakly NBT positive; arrowheads indicate eosinophils more strongly NBT positive.

Fig 3. Flow cytometric histograms of hydrogen peroxide generation from X CGD patients. Dihydrorhodamine-1,2,3 (DHR)-loaded neutrophils (10⁶ cells/ml) from a healthy donor, patient II-2 and a X91⁰ CGD patient were incubated for 15 min at 37°C without (resting) or with 20 ng/ml PMA or PMA in the presence of 5 μM DPI, as described in Materials and Methods, and analyzed by flow cytometry.

Fig 4. Flow cytometric dot plots of hydrogen peroxide generation from neutrophils and eosinophils in granulocyte preparations from a healthy subject (top panels) and patient II-2 (bottom panels) at rest (left panels) or after stimulation with PMA (right panels). Dihydrorhodamine- 1,2,3 (DHR)-loaded granulocytes (10⁶ cells/ml) were incubated for 15 min at 37°C without or with 20 ng/ml PMA. After dilution with ice-cold PBS the cells were incubated with Alexa Fluor 647 rat anti-human CD294 (CRTH2) for 30 min and washed, before cytometry readings, as described in Materials and Methods.
Fig. 5. Reduced minus oxidized spectra and Western blot analysis of cytochrome $b_{558}$ of neutrophils from the X CGD patients. The experiments were performed on 1% Triton X100 soluble extract of neutrophils. (A) Reduced minus oxidized difference spectra of cytochrome $b_{558}$ from neutrophils of a healthy donor, patient II-2 and an X910 CGD patient. (B) Immunodetection of the $\alpha$ (p22phox) and $\beta$ (gp91phox) subunits of cytochrome $b_{558}$ neutrophils of patient II-2, an X910 CGD patient and a healthy donor. All samples were subjected to SDS-PAGE and blotted onto a nitrocellulose sheet. Cytochrome $b_{558}$ $\alpha$ and $\beta$ subunits were revealed with monoclonal antibodies 449 (\(\alpha\) subunit) and 48 (\(\beta\) subunit), as described in Materials and Methods. The results of one experiment representative of the two performed, with similar results, are reported.

Fig. 6. Surface expression of gp91phox on neutrophils and eosinophils. Granulocyte preparations from a healthy subject (top panels) and from patient II-2 (bottom panels) were stained with the anti gp91phox mAb 7D5 and the eosinophil specific anti-CRTH2 mAb (right panels), or a control mAb and CRTH2 (left panels). For experimental details, see Materials and Methods.

Fig 7. Analysis of mutations in the CYBB gene from patient II-2. PCR products were gel-purified and automatically sequenced by Cogenics, Meylan, France, with forward and backward primers P1 and P2*, as described in Materials and Methods. (A) Thymidine insertion detected in the CYBB promoter region of patient II-2, compared to a healthy donor sequence. (B) Schematic representation of part of the CYBB promoter region, with localization of the “CCAAT” and “TATA” boxes, the ATG starting codon and the point mutation described in A.
Fig. 8. Electrophoretic mobility shift assay. Gel shift analysis was performed as described in Materials and Methods using the −68- to −30-bp region of the wild-type (WT) CYBB promoter as biotin labelled probe. The WT-labelled probe was incubated with 10 µg of nuclear extract from undifferentiated PLB 985 cells after preincubation with 20-, 50-, or 100-fold molar excess of unlabelled double-stranded WT or X91− CGD competitor oligonucleotide as indicated. The data are representative of three separate experiments.

Fig. 9. Semi-quantitative RT-PCR amplification of gp91phox mRNA from patient II-2. The gp91phox mRNA from patient II-2, X91+ and X910 CGD patients and from a healthy subject were amplified by RT-PCR using primers 1 and 2* (see Table II for sequences and positions). The housekeeping gene of β-actin was used as internal control. M, DNA size markers. The data are representative of three separate experiments.

Fig 10. Microbicidal activity of neutrophils from X CGD patients. Neutrophils from a control subject, patient II-2 and a patient with X910 CGD were incubated with opsonized S. aureus at MOI 3-5:1, C. albicans at MOI 1-2:1 and E.coli at MOI 3-5:1. Data are expressed as percent killing after 45 min incubation for S. aureus and E. coli, and after 60 min incubation for C. albicans. The results are means of 3-4 experiments for patient II-2 and 2-3 experiments for the X910 CGD patient. For experimental details, see Materials and Methods.
A

Patient II-2

CTGCTGTTCATTTTTTCTCATT

Healthy donor

CTGCTGTTCATTTTTTCTCATT

B

-140
gcaagcttttcagttga ccaattttag ccaatttctgataaagaa

-90
aaggaaccgttcggcccaggtgtgttgctttttca ttttcctctcatggaaga

-40
agaagcatatagttagaagaaagggacaacacacacattcaacccctctggacaacc

ATG GGG ... Exon 1