

# Isotopic evidence of distinct foraging ecology and movement pattern in two migratory predators (yellowfin tuna and swordfish) of the western Indian Ocean

Frédéric Ménard, Anne Lorrain, Michel Potier, Francis Marsac

# ▶ To cite this version:

Frédéric Ménard, Anne Lorrain, Michel Potier, Francis Marsac. Isotopic evidence of distinct foraging ecology and movement pattern in two migratory predators (yellowfin tuna and swordfish) of the western Indian Ocean. Marine Biology, 2007, 153 (2), pp.141-152. hal-00452372

HAL Id: hal-00452372

https://hal.science/hal-00452372

Submitted on 2 Feb 2010

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

# **Manuscript**

Click here to download Manuscript: MS\_Menard\_etal\_MarBiol.doc

1 Isotopic evidence of distinct foraging ecology and movement pattern in two migratory 2 predators (yellowfin tuna and swordfish) of the western Indian Ocean 3 Frédéric Ménard<sup>(1)\*</sup>, Anne Lorrain<sup>(1)</sup>, Michel Potier<sup>(2)</sup>, and Francis Marsac<sup>(1)</sup> 4 5 (1) Institut de Recherche pour le Développement, Centre de Recherche Halieutique Méditerranéenne et 6 7 Tropicale, BP 171, 34203 Sète Cedex, France <sup>(2)</sup> Institut de Recherche pour le Développement, BP 172, 97492 Ste Clotilde Cedex, La Réunion, 8 9 France 10 \* Corresponding author 11 Tel 00 33 4 99 57 32 30; E-mail address: menard@ird.fr 12 13

# Abstract

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

Ecologists primarily use  $\delta^{15}N$  values to estimate the trophic level of organisms, while  $\delta^{13}$ C, and even recently  $\delta^{15}$ N, are utilized to delineate foraging habitats. However, many factors can influence the stable isotopic composition of consumers, e.g. age, starvation or isotopic signature of primary producers. Such sources of variability make the interpretation of stable isotope data rather complex. To examine these potential sources of variability, muscle tissues of yellowfin tuna (Thunnus albacares) and swordfish (Xiphias gladius) of various sizes were sampled between 2001 and 2004 in the western Indian Ocean during different seasons and along a latitudinal gradient (23°S to 5°N). Size and latitude effects on  $\delta^{15}$ N and δ<sup>13</sup>C were investigated using linear models. Both latitude and size significantly affect the stable isotope values of the studied species but variations were much more pronounced for  $\delta^{15}$ N. We explain the latitudinal effect by differences in nitrogen dynamics existing at the base of the food web and propagating along the food chain up to top predators. This spatial pattern suggests that yellowfin and swordfish populations exhibit a relatively unexpected resident behaviour at the temporal scale of their muscle tissue turnover. The size effect is significant for both species and indicates an increase in prey size through ontogeny. However, this effect is more pronounced in swordfish as a consequence of their different foraging strategies, reflecting specific physiological abilities. Swordfish adults are able to reach very deep water and have access to a larger size range of prey than yellowfin tuna. In contrast, yellowfin juveniles and adults spend most of their time in the surface waters and large yellowfin tuna continue to prey on small organisms. Consequently, nitrogen isotopic signatures of swordfish tissues are higher than those of yellowfin tuna and provide evidence for different trophic levels between these species. Thus, in contrast to  $\delta^{13}$ C,  $\delta^{15}$ N analyses of tropical Indian Ocean marine predators allow the investigation of complex vertical and spatial segregation, both within and between species, even in the case of highly opportunistic feeding behaviours. The

- 39 linear models developed in this study allow us to make predictions of  $\delta^{15}N$  values and to
- 40 correct for any size or latitude differences in future food web studies.

### Introduction

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

Catches of tunas and billfishes have increased dramatically the past twenty years in the western Indian Ocean, very likely altering the structure and functioning of the ecosystems through trophic cascades (Essington et al. 2002; FAO 2006). Concomitantly to these topdown controls, bottom-up effects, via environmental and climatic changes, are also controlling abundance and spatial dynamics of top predators that depend on food availability (Cury et al. 2003; Franck et al. 2006; Frederiksen et al. 2006). Therefore, studies based on trophic ecology and movements of top predators are useful to assess the impact of fisheries and climate on marine resources, and to provide basic elements for an ecosystem approach to fisheries management (Sinclair and Valdimarsson 2003; FAO 2003). Unlike the Pacific and Atlantic oceans, few studies have investigated the diet of tunas and tuna-like species from stomach content analyses in the Indian Ocean (Watanabe 1960; Kornilova 1981; Roger 1994; Maldeniya 1996; Potier et al. 2004, 2007). Furthermore, stomach content analyses only reflect the composition of the most recent meal and limit our ability to address spatial and temporal variability of feeding behaviours. Small- and large-scale movements of top predators are now assessed using conventional and electronic tagging programs combined with catch statistics (e.g., Block et al. 2005). A tagging programme, the Regional Tuna Tagging Programme of the Indian Ocean Tuna Commission (RTTP-IO, http://www.rttp-io.org/en/about/) is underway in the Indian Ocean. So far, few tag returns are available and catch statistics themselves do not reflect the real movement patterns (Hilborn and Walters 1992; Walters 2003). The knowledge of the spatial dynamics of tuna in the Indian Ocean is therefore still minimal. Additional information on dietary sources, trophic levels, foraging strategies or movement patterns of migratory species can be obtained from stable isotope analyses of animal tissues (Rau et al. 1983; Fry 1988; Kelly 2000; Rubenstein and Hobson 2004; Cherel and Hobson,

2005, 2007). The stable isotope composition of an organism depends on its diet, its trophic level, but also on the isotopic signature at the base of the food web (DeNiro and Epstein, 1978, 1981; Post 2002; Fry 2006).  $\delta^{15}N$  measurements mainly serve as indicators of consumer's trophic position, while  $\delta^{13}$ C values are used to determine primary sources, inshore versus offshore or pelagic versus benthic contribution to food intake (Hobson 1999). Indeed, different oceanic processes affect isotopic baselines of  $\delta^{15}N$  and  $\delta^{13}C$  in marine pelagic food webs (Rau et al. 1982; Altabet et al. 1995; Gruber and Sarmiento 1997; Lourey et al. 2003). δ<sup>13</sup>C values of phytoplankton decrease from low to high latitudes (Lourey et al. 2003) while  $\delta^{15}N$  of particulate organic matter is driven by nutrient utilization and the nitrogen source used by primary producers (nitrate, ammonium, N<sub>2</sub> gas; Wada and Hattory 1991). The resulting spatial and temporal variability in the isotopic baseline has been shown to be incorporated and conserved through several trophic levels (up to pelagic consumers) across ocean basins (Takai et al. 2000; Wallace et al. 2006) or within a region of a single basin (Schell et al. 1989; Lesage et al. 2001; Quillfeldt et al. 2005; Cherel and Hobson 2005, 2007; Cherel et al. 2005). Hobson (1999) illustrated this approach by the new maxim "you are what you swim in" that complements the well-known dogma of stable isotopy "you are what you eat." Consequently, the stable isotope ratios of animal tissues have the potential of characterizing the isotopically distinct regions crossed by migrating fish and investigating their foraging ecology. Graham et al. (2006) successfully applied this approach to yellowfin tunas (Thunnus albacares) of the Pacific Ocean. One objective of the present study is therefore to investigate the relationships between the isotopic signature of yellowfin tuna and swordfish (Xiphias gladius) versus latitude in the western Indian Ocean, and their relative degree of residency. Indeed, for migratory species, the variability of the isotopic signature in their tissue is supposed to be low if the migration rate is quicker than isotopic tissue turnover. Conversely, for more resident species, stable isotope ratios of tissues would reflect the

66

67

68

69

70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

88

89

90

isotopic patterns at the base of the food web (Fry 2006; Graham et al. 2006; Popp et al. in press). A second objective was to document the foraging ecology of yellowfin tuna and swordfish, and potential ontogenetic effects on their trophic status. Ontogenetic shifts in tuna and swordfish foraging behaviour are also expected as larger fish usually expand their foraging habitat and exploit a larger size range of prey in the environment (Ménard et al. 2006; Young et al. 2006; Graham et al. 2007).

Inter-specific, spatial and ontogenetic differences in the stable isotope composition of muscle tissues were thus investigated for these two migratory top predators of the western Indian Ocean. Linear models and linear mixed-effects models were developed to test and disentangle potential latitudinal and size effects on the stable isotope values ( $\delta^{15}N$  and  $\delta^{13}C$ ) of each species. According to the model predictions, the trophic positions of individuals of different sizes caught in different oceanic regions can then be compared. It is indeed a prerequisite to understand these geographical and ontogenetic variations before determining the trophic position of these species. This paper is however not intended to provide a comprehensive study of the pelagic food web of the western Indian Ocean. Rather, we implement the isotope approach to gain insight into the foraging ecology and movement patterns on the studied predators, the first initiative of this nature in the Indian Ocean.

#### Materials and methods

### Sample collection

Fish were caught by industrial purse seiners with scientific observers onboard; a 20-m research longliner "Amitié" of the Seychelles Fishing Authority, and the French 24-m industrial longliner "Cap Morgane". Samples were collected from 2001 to 2004 in the western Indian Ocean along a latitudinal gradient (23°S to 5°N, Fig. 1). A total of 245 yellowfin tuna (*Thunnus albacares*) and 136 swordfish (*Xiphias gladius*) from various sizes were collected

during these cruises. Fork length (FL) ranged from 40 to 160 cm for yellowfin tuna (mean = 103 cm and median = 108 cm) and Lower Jaw Fork length (LJFL) for swordfish ranged from 68 to 225 cm (mean = 135 cm and median = 133 cm). LJFL is a reliable measure of swordfish body size that allows comparisons with tunas by reducing the bias due to the bill. Table 1 displays all the sample characteristics. Muscle tissues from the dorsal region were collected onboard from freshly caught fishes and were stored frozen at -20°C until processing.

Sample preparation and analysis

Samples were freeze dried and ground to a fine powder. Lipid extraction was performed using 20 ml of cyclohexane on powder aliquots of about 1 g, and the lipid-extracted sample was dried at 60°C before processing. One milligram sample was then placed into 8×5 mm tin cups for CF-IRMS analysis, using a Europea Scientific ANCA-NT 20-20 Stable Isotope Analyser with ANCA-NT Solid/Liquid Preparation Module (PDZ Europa Ltd., Crewz, UK). Replicate measurements of internal laboratory standards indicate measurement errors of  $\pm$  0.2% for  $\delta^{13}$ C and  $\delta^{15}$ N. Triplicate analyses performed on some samples confirmed that analytical reproducibility was very good (0.2% maximum variation). Isotopic ratios are expressed in the conventional  $\delta$  notation as parts per thousand (‰) deviation from the international standards: atmospheric nitrogen for  $\delta^{15}$ N and VPDB Belemnite for  $\delta^{13}$ C:

 $\delta X = (R_{sample} / R_{standard} - 1) \times 1000,$ 

where X is  $^{15}$ N or  $^{13}$ C and R the corresponding ratio  $^{15}$ N/ $^{14}$ N or  $^{13}$ C/ $^{12}$ C.

Lipid content in tuna and swordfish muscles may be high (greater than 50%, unpublished data). As lipids are highly depleted in  $^{13}$ C (Tieszen et al. 1983), C/N mass ratios were used to check the lipid extraction process.  $\delta^{13}$ C outliers (15 for yellowfin tuna and 33 for swordfish) were removed from the analyses according to the distributions of C/N values for each species. We estimated the corresponding thresholds to suppress any relationships between  $\delta^{13}$ C and C/N mass ratios (3.62 and 3.70 for yellowfin tuna and swordfish, respectively). The resulting

distributions of C/N were normal (mean  $\pm$  standard deviation of 3.36  $\pm$  0.11 and 3.50  $\pm$  0.11 for yellowfin tuna and swordfish, respectively) and the sampling ranges of latitude and body size were not reduced for both species.

#### Statistical analysis

141

142

143

144

145

146

147

148

149

150

151

152

153

154

155

156

157

158

159

160

161

162

163

164

165

Linear regressions were used on the  $\delta^{15}N$  and  $\delta^{13}C$  data for each species to test the covariates of interest, i.e. latitude and body size. However, all the individuals of one species are not independent and the sampling scheme is clearly unbalanced. The individuals can be grouped according to different factors (e.g. cruise, year, season, etc.; Table 1). We use the two main seasons of the monsoon system to group the individuals of each species caught during the cruises carried out from 2001 to 2004. Indeed, the ocean circulation in the West Indian Ocean is strongly related to the wind monsoon regime, which in turn strongly affects biological productivity (Tomczak and Godfrey 1994; Longhurst 1998; Schott and McCreary 2001). The Northeast (NE) monsoon becomes established in boreal winter (December to March). It is characterized by winds blowing from the Asian continent to the equatorial zone, a weak circulation north of the equator and a well marked counter-current flowing East between 2°S and 7°S. The Southwest (SW) monsoon becomes established in boreal summer (June-September), and is characterized by a reversal of the winds in the northern Indian Ocean, the development of the northward-flowing Somali Current, and an eastward flow that dominates the northern Indian Ocean. Therefore, each observation can be classified according to the season on which it was made (NE or SW monsoon, Table 1). The 5 cruises which took place during the inter-monsoons are relocated in their nearest monsoon: October and May in the SW monsoon, and November in the NE monsoon. A classification based on the 4 seasons (including the 2 additional inter-monsoon seasons) was also tested but not retained as the various models fit to the  $\delta^{15}N$  and  $\delta^{13}C$  values gave similar results to the monsoon only scenario. To test this grouping, the seasonal effect was treated as random variations around a

population mean, and the body size and the latitude were assessed as two fixed continuous covariates, using linear mixed-effects models (Ime models; Pinheiro and Bates 2000). These Ime models combine a random-effects analysis of variance model (variability amongst seasons) with a linear regression model. Lme models were tested against simple linear regression models using likelihood ratio tests. Population predicted values (obtained by setting the random effects to zero in the Ime models) were used to compare latitude and body size effects for yellowfin tuna, for swordfish, and between the two species. All the computations and tests were performed on S-Plus (Insightful 2005).

#### **Results**

Muscle  $\delta^{15}N$  and  $\delta^{13}C$  values for yellowfin tuna and swordfish plotted versus body size, latitude and season (NE and SW monsoons) are shown in Figs. 2 and 3. The  $\delta^{15}N$  values for yellowfin tuna ranged from 10.2 to 15.2‰ and from 11.8 to 16.2‰ for swordfish (Fig. 2). The  $\delta^{13}C$  values for yellowfin tuna ranged from -17.4 to -15.2‰ and from -17.4 to -15.0‰ for swordfish (Fig. 3). The range of variation for the  $\delta^{13}C$  values is therefore much more reduced than the  $\delta^{15}N$  range ( $\approx 2$ ‰ vs. 5‰).

Linear regression models with latitude and size added sequentially were significant (p = 0.026 for  $\delta^{15}N$  and p < 0.001 for  $\delta^{13}C$  for yellowfin tuna; p < 0.001 for  $\delta^{15}N$  and  $\delta^{13}C$  for swordfish). However, deviations from the models suggest that other models might be appropriate. For example, Fig. 2 reveals that intercepts of the models for  $\delta^{15}N$  may differ between seasons.

Linear mixed-effects models (Ime models) were fitted to the muscle  $\delta^{15}N$  values grouped by season. The most parsimonious model for swordfish was obtained with both latitude and size as fixed-effect covariates (p < 0.0001). For yellowfin tuna, when body size was added to a line model containing latitude only, the fit was only marginally improved (p =

0.092). This p-value evidences the low significance of body size for yellowfin tuna  $\delta^{15}N$ values, a result already exhibited in Fig. 2. According to the likelihood ratio test, lme models for both species provided a much better description of the  $\delta^{15}N$  data than the linear regression models did (p < 0.0007 and p < 0.0001 for yellowfin tuna and swordfish, respectively). Fig. 4 displays the predicted lines for each season (using the estimated random effects) and the original data for model checking. These plots exhibit the large variability of the  $\delta^{15}N$  values for each season, and confirm that latitude was the strongest linear fixed-effect for vellowfin tuna, while body size was the most significant fixed-effect for swordfish. For both species, the random effects were associated with the intercepts only. Therefore within-season intercept estimates for  $\delta^{15}N$  data were different, while slopes were identical. Interestingly, withinseason intercepts exhibited a similar pattern whatever the species: the NE monsoon intercept was always greater than the SW monsoon intercept (difference estimated at 0.37 and 0.67%) for yellowfin tuna and swordfish, respectively). The assumption of normality and independence for the random effect and the residuals were graphically assessed (not shown). On the other hand, lme models fit to the fish muscle  $\delta^{13}$ C values were not significantly better than the linear regression models (p > 0.50 for yellowfin tuna and p = 0.33 for swordfish). Table 2 lists the coefficients and standard errors estimated by the most parsimonious models fit to the  $\delta^{15}$ N and  $\delta^{13}$ C data.

209

210

211

212

213

214

215

208

191

192

193

194

195

196

197

198

199

200

201

202

203

204

205

206

207

# Discussion

Our results provide evidence for a genuine relationship between latitude and body size with  $\delta^{15}N$  and  $\delta^{13}C$  values of yellowfin tuna and swordfish. Linear mixed-effects models were used for  $\delta^{15}N$  data and provided identical slopes with different intercepts between the two seasons for both latitude and body size. Simple linear models with no seasonal effect were selected for  $\delta^{13}C$  data. Both size and latitude influence  $\delta^{15}N$  values of the two species more

strongly than  $\delta^{13}$ C values. Model predictions at the population level allow us to analyse these effects separately. Fig. 5A illustrates  $\delta^{15}$ N and  $\delta^{13}$ C predicted values of yellowfin tuna and swordfish as a function of varying sizes for fish caught at different latitudes (-10° and 0°). In the same way, Fig. 5B represents  $\delta^{15}$ N and  $\delta^{13}$ C predicted values for fish of different sizes (80 and 160 cm) as a function of a latitudinal gradient. We now attempt to examine some hypotheses supported by our results with respect to the trophic ecology of yellowfin tuna and swordfish, and to the oceanic processes affecting the isotopic baseline of  $\delta^{15}$ N and  $\delta^{13}$ C in marine food webs.

# Latitudinal effect

The range of variation for the  $\delta^{15}N$  values of yellowfin tuna and swordfish is of 2‰ along a latitudinal gradient of 30° (2.4 and 1.1‰ for yellowfin tuna and swordfish, respectively, Fig 5B), whereas those variations are less than 1‰ for  $\delta^{13}C$  (0.8 and 0.7‰ for yellowfin tuna and swordfish, respectively). Three hypotheses can be put forward to explain the  $\delta^{15}N$  and  $\delta^{13}C$  increase from the Mozambique Channel to the Somali basin: 1) dietary changes, 2) starvation and 3) a shift in  $\delta^{15}N$  baseline. Trophic level differences or starvation of the northern individuals seem highly unlikely given the regularity of the observed variations, and are not supported by any ecological data. We argue that this spatial pattern results from different oceanic processes at the base of the food web that vary by region in the western Indian Ocean, and that are conserved through different trophic levels up to top predators.

Particulate organic matter (POM)  $\delta^{15}N$  and  $\delta^{13}C$  isotopic values are not available in the western Indian Ocean to document a latitudinal pattern at the base of the food chain. However, knowledge of nitrogen dynamics in several zones of the western Indian Ocean suggests that differences in  $\delta^{15}N$  values of POM might occur. In particular, the Somali region

should have higher  $\delta^{15}N$  baseline values compared to Mozambique Channel. This is because the Arabian Sea is a major area of anoxia (Gruber and Sarmiento 1997), and is characterized by intensive denitrification that leads to an accumulation of isotopically enriched nitrate in subsurface waters (Gaye-Haake et al. 2005; Naqvi et al. 2006). Conversely, different tracers and biological indicators in the surface waters of the South Indian subtropical gyre (around 20°S, 57°E) have shown a prevailing N<sub>2</sub> fixation, known to generate lower δ<sup>15</sup>N values for phytoplankton (Capone and Carpenter 1982; Carpenter 1983; Gruber and Sarmiento 1997). Gruber and Sarmiento (1997) also found a latitudinal gradient between the Arabian Sea and 25°S in the western Indian Ocean with decreasing denitrification and increasing N<sub>2</sub> fixation from North to South. We did not sample the core of the South Indian subtropical gyre, nor the Arabian Sea; however, the northern and southern edges of our sampling zone are connected through the current system to the two most contrasted zones of denitrification (the Arabian Sea) and N<sub>2</sub> fixation (subtropical gyre, Tomczak and Godfrey 1994; Schott and McCreary 2001, Davis 2005). Therefore, the  $\delta^{15}N$  baseline values of the Somali region are likely to be strongly influenced by the Arabian Sea while those of the Mozambique Channel are under the influence of the subtropical gyre (Davis 2005).

In several studies using stable isotopes to delineate foraging locations of marine predators of the southern Ocean,  $\delta^{13}$ C displayed strong variations with latitude, whereas  $\delta^{15}$ N values responded mainly to trophic enrichment (Quillfeldt et al. 2005; Cherel and Hobson 2005, 2007). Indeed, in the southern hemisphere, the geographical  $\delta^{13}$ C gradient in POM of surface waters is well defined, and ranges from high  $\delta^{13}$ C values in warm subtropical waters in the North, to low values in cold Antarctic waters in the South (François et al. 1993; Trull and Armand 2001), with abrupt changes at fronts (Subtropical, Subantarctic and Polar fronts). Gradients in terms of sea temperature are much more reduced in our sampling zone (Tomczak and Godfrey 1994), which could explain the weak  $\delta^{13}$ C variations in the western tropical

Indian Ocean revealed by our study. Broad  $\delta^{15}N$  gradients, as observed in this study, have been found in other open ocean regions. Comparing leatherback turtle  $\delta^{15}N$  signatures in the Eastern Tropical Pacific and in the Atlantic Ocean, Wallace et al. (2006) found inter-basin differences of 5‰ between denitrification and  $N_2$  fixation zones. In the Equatorial Pacific, Graham et al. (2006) have shown basin-wide  $\delta^{15}N$  differences as high as 11‰ in tuna muscle tissue. In the Indian Ocean, we detected a much lower intra-basin difference (i.e., maximum of 2.4‰ for yellowfin tuna), most probably because our samples did not cover the core areas mentioned earlier. Consequently, even if yellowfin tuna and swordfish are migrating between the two contrasted region (the Arabian Sea versus the subtropical gyre), we are probably observing a diluted effect of this general intra-basin difference. Furthermore, muscle tissues of these species might never be at isotopic equilibrium with their recent diet because of their continuous movement, their opportunistic foraging behaviour and their muscle tissue isotopic turnover (half-life around 50 days, Graham, unpublished data). All these reasons generate variability in the data, reduce the effect of the latitudinal gradient, but do not challenge its occurrence.

The observed conservation of the  $\delta^{15}N$  baseline characteristics in these top predators has several implications. First, these species are known to be highly migratory and thus such a gradient in the data is not expected. Indeed, these data suggest that yellowfin tuna and swordfish are relatively resident species at the temporal scale of their tissue isotopic turnover, i.e., three months for yellowfin tuna (Graham, unpublished data). However, this does not preclude large basin-wide movement patterns at the temporal scale of their life time. Furthermore, the coexistence of migrating fish among more resident fish might occur and explain the rather high intra-season variability found in our study. Interestingly, the  $\delta^{15}N$  predictions along the latitudinal gradient varied two times more for yellowfin tuna than for swordfish (differences of 2.4‰ and of 1.2‰, respectively). This can be interpreted in

different ways: (i) yellowfin tuna are more resident than swordfish, (ii) swordfish have a slower turnover rate or tissue growth than yellowfin tuna, (iii) swordfish do not migrate to highly <sup>15</sup>N depleted areas such as the Arabian Sea, which the very low catch records of this species in this region suggests (Fonteneau 1997). The third hypothesis seems the most plausible given our present knowledge; however, we cannot preclude a mixed influence of the three hypotheses.

# Seasonal effect

In the mixed-effects models implemented in this paper for  $\delta^{15}N$  values, the seasonal effect is random and induced by the grouping of the data. Only intercepts differ between NE and SW monsoon predictions: compared to the SW monsoon, the NE monsoon intercepts are 0.36 and 0.67‰ higher for yellowfin tuna and swordfish, respectively. During the NE monsoon, the waters of the Arabian sea are advected to the South and invade the Somali basin (where part of our data collection was undertaken), potentially increasing the  $\delta^{15}N$  values of the baseline of this zone compared to SW monsoon (Davis 2005). Conversely, during the SW monsoon, there is a broad equatorward flow of waters from the South Equatorial Current (SEC) along the East African Coast reaching the Somali region (Tomczak and Godfrey 1994; Schott and McCreary 2001; Davis 2005). Further studies involving measurements of the  $\delta^{15}N$  of the POM over an annual cycle should be investigated to shed some light on the seasonal variations that may occur in the western Indian Ocean.

Interestingly, the seasonal effect is not significant for  $\delta^{13}C$  values. We believe that seasonal changes in the monsoon regime do not have strong consequences on the carbon isotopes ratios in the sampled areas. In addition, the  $\delta^{13}C$  ranges we observed in our data were low compared to the intra-individual variability. Our results suggest that muscle  $\delta^{13}C$  values of fish in these open sea ecosystems of the western Indian Ocean might not be useful to

document seasonal changes, to delineate foraging locations or to track fish movement. This is in contrast to studies conducted in the southern Indian Ocean where  $\delta^{13}C$  has been shown to be a useful tool (Cherel and Hobson 2005, 2007).

319

320

321

322

323

324

325

326

327

328

329

330

331

332

333

334

335

336

337

338

339

340

316

317

318

#### Size effect

Figure 5A indicates changes in the  $\delta^{15}N$  and  $\delta^{13}C$  model predictions along a gradient of body size, for fish caught at two different latitudes. In each case, the  $\delta^{15}$ N values exhibited a stronger size effect for swordfish than for yellowfin tuna. The isotopic difference between large (200 cm) and small (80 cm) swordfish was about 2‰, whereas it was less than 0.5‰ for yellowfin tuna of 40 to 160 cm. The same pattern is supported by  $\delta^{13}$ C values, but isotopic differences are much more reduced (0.8 and 0.4% for swordfish and yellowfin tuna, respectively). The lower differences for  $\delta^{13}$ C are not surprising given that  $\delta^{15}$ N is known to increase much more with trophic levels than  $\delta^{13}$ C (DeNiro and Epstein, 1981). Body size is indeed known to play a crucial role in predator-prev interactions (Sheldon et al. 1977; Cury et al. 2003). Analyses of stomach contents and nitrogen isotope ratios conducted on fish communities in different marine ecosystems have shown that prey size and trophic level generally increase with increasing predatory body size (Scharf et al. 2000; Jennings et al. 2002). In open-sea ecosystems, few studies have yet dealt specifically with size-based predation. Ménard et al. (2006) have shown that the maximum size of the prey consumed by yellowfin tunas tends to increase with tuna body length, but that large yellowfin tunas continue to consume small prey in great proportions. In addition, both adults and juveniles of yellowfin tuna generally show only minor differences in depth distributions (Brill et al. 1999, 2005). Yellowfin tuna spend most of their time in the surface layer, even if some exceptional deep dives have been evidenced by one archival tag (Dagorn et al. 2006). The diet of yellowfin tunas is then mainly composed of organisms present in the upper 100 m (Moteki et al. 2001; Bertrand et al. 2002; Potier et al. 2004, 2007), with no major ontogenetic changes (Ménard et al. 2006). An outstanding diet shift was revealed by Graham et al. (2007) but this study only concerned small juveniles of yellowfin tunas ranging around 45 cm and caught in areas surrounding Hawaiian Islands. Therefore, we conclude that the size of yellowfin tuna does not have a strong impact on its  $\delta^{15}N$  values. On the other hand, large swordfish mainly consume cephalopods (Hernandez-Garcia 1995; Markaida and Hochberg 2005; Young et al. 2006; Potier et al. 2007), while smaller swordfish have a diet focused on mesopelagic fish such as myctophids (Young et al. 2006; Potier et al. 2007). This shift in the dominant prey items has consequences on the  $\delta^{15}N$  values because mesopelagic fish such as myctophids and paralepidids have shown lower mean  $\delta^{15}$ N values than cephalopods (Young et al. 2006). In addition, swordfish can catch larger prey specimens as they grow, due to an increase of mouth-gape size, chasing predation, and diving capability (Carey and Robinson 1981). Therefore, we fully confirm that body size influences the  $\delta^{15}N$  values of swordfish, as already shown by Young et al. (2006) with much fewer data. This influence is much more pronounced than for yellowfin tuna, due to the change in the foraging ecology of swordfish through its ontogeny.

357

358

359

360

361

362

363

364

365

341

342

343

344

345

346

347

348

349

350

351

352

353

354

355

356

#### Trophic level differences

Over the body sizes and latitudes common to both species, and for a similar size or latitude, the  $\delta^{15}N$  values of swordfish were about 0.7 to 2.8% higher than those of yellowfin tuna (Fig. 5). The greatest  $\delta^{15}N$  differences were found in large fish (160 cm) sampled in the south (25°S), while the smallest differences occurred in small fish (80 cm) sampled in the north (5°N). In a recent study conducted in the same area, Potier et al. (2007) established that (i) the diet composition of swordfish was dominated by mesopelagic cephalopods (Ommastrephidae and to a lesser extent Onychoteuthidae) and by mesopelagic fish

(Nomeidae and Diretmidae), while epipelagic prey dominated the diet of yellowfin tuna, (ii) swordfish catch larger specimens of the same prey species than yellowfin tuna. This general diet pattern reflects a well-known resource partitioning between both species. Swordfish undertake large vertical migrations, allowing them to prey actively at great depth, while both adult and juvenile yellowfin tuna spend the vast majority of their time in the surface layer and prey on small organisms (Brill et al. 2005; Ménard et al. 2006; Potier et al. 2007). Consequently, swordfish have access to a larger size range of prey in the environment than yellowfin tuna, and can feed on the predators of yellowfin tuna's prey. Thus, the observed differences in  $\delta^{15}$ N values of swordfish and yellowfin tuna, once the size and latitudinal effects are removed, illustrate different trophic levels between both species due to distinct foraging strategies. Graham et al. (2007) hypothesized that mesopelagic prey might have  $\delta^{15}$ N values higher than epipelagic species. This assumption could strengthen the  $\delta^{15}$ N differences between both species, but further investigations should be carried out on the isotopic values of the forage fauna of large pelagics.

#### **Summary and conclusion**

This study revealed body size and latitudinal effects on  $\delta^{15}N$  and  $\delta^{13}C$  values of two migratory highly opportunistic predators: yellowfin tuna (*Thunnus albacares*) and swordfish (*Xiphias gladius*). However, in these open sea ecosystems of the western Indian Ocean,  $\delta^{15}N$  was much more useful than  $\delta^{13}C$  to delineate trophic relationships and to track fish movements. Linear mixed-effects models developed here will allow us to make predictions of  $\delta^{15}N$  values and to correct for any size or latitude differences in future food web studies. This study also confirmed that baseline  $\delta^{15}N$  isotopic variations can be conserved through several trophic levels, and even up to high trophic levels such as tunas and swordfish. These spatial differences together with differences in the size effects according to species illustrated the

potential of stable isotopes to investigate complex trophic ecology and foraging strategies, both within and between species, even in the case of highly opportunistic feeding behaviours.

To further investigate these spatial and size variations in the  $\delta^{15}N$  values of yellowfin tuna and swordfish, isotopic analyses of mesopelagic species together with particulate organic matter from these regions are needed. Spatial and size-based variation in the  $\delta^{15}N$  of marine pelagic fish should be considered when using  $\delta^{15}N$  to detect trophic-level variation in natural communities.

Acknowledgements - The authors gratefully thank the Seychelles Fishing Authority (SFA), the crew of the longliner "Amitié", the crew of the longliner "Cap Morgane", and the observers onboard the purse seiners for helping us to collect the samples. We also thank BS Graham for providing the unpublished data on tuna isotopic turnover and for many helpful discussions, and DP Gillikin for very thoughtful comments on the manuscript. This work, a part of the THETIS programme of the IRD (Institut de Recherche pour le Développement), is also supported by the REMIGE project funded by Agence Nationale de la Recherche (ANR 2005 Biodiv-11).

#### References

denitrification in the Arabian Sea from sediment <sup>15</sup>N/<sup>14</sup>N ratios. Nature 373:506-509

Bertrand A, Bard F-X, Josse E (2002) Tuna food habits related to the micronekton distribution in French Polynesia. Mar Biol 140:1023-1037

Block BA, Teo SLH, Boustany AB, Stokesbury MJW, Farwell CA, Weng KC, Dewar H, Williams TD (2005) Electronic tagging and population structure of Atlantic bluefin tuna. Nature 434: 1121-1127

Altabet MA, François R, Murray DW, Prell WL (1995) Climate-related variations in

416	Brill RW, Block BA, Boggs CH, Bigelow KA, Freund EV, Marcinek DJ (1999) Horizontal
417	movements and depth distribution of large adult yellowfin tuna (Thunnus albacares)
418	near the Hawaiian Islands, recorded using ultrasonic telemetry: implications for the
419	physiological ecology of pelagic fishes. Mar Biol 133:395-408
420	Brill RW, Bigelow KA, Musyl MK, Fritches KA, Warrant EJ (2005) Bigeye tuna (Thunnus
421	obesus) behavior and physiology and their relevance to stock assessments and fishery
422	biology. ICCAT Collective Volume of Scientific Papers 57:142-161
423	Capone DG, Carpenter EJ (1982) Nitrogen fixation in the marine environment. Science 217:
424	1140–1142
425	Carey FG, Robinson BH (1981) Daily patterns in the activities of swordfish, Xiphias gladius,
426	observed by acoustic telemetry. Fish Bull 79:277-292
427	Carpenter EJ (1983) Nitrogen fixation by marine Oscillatoria ( <i>Trichodesmium</i> ) in the world's
428	oceans. In: Carpenter EJ & Capone DG (Eds) Nitrogen in the Marine Environment:
429	65–103 Academic Press, New York
430	Cherel Y, Hobson KA, Weimerskirch H (2005) Using stable isotopes to study resource
431	acquisition and allocation in procellariiform seabirds. Oecologia 145:533-540
432	Cherel Y, Hobson KA (2005) Stable isotopes, beaks and predators: a new tool to study the
433	trophic ecology of cephalopods, including giant and colossal squids. Proc R Soc B,
434	272:1601-1607
435	Cherel Y, Hobson KA (2007) Geographical variation in carbon stable isotope signatures of
436	marine predators: a tool to investigate their foraging areas in the Southern Ocean. Mar
437	Ecol Prog Ser 329:281-287
438	Cury P, Shannon LJ, Shin Y-J (2003) The Functioning of Marine Ecosystems: a Fisheries
439	Perspective. In: M Sinclair and G Valdimarsson (ed) Responsible Fisheries in the
440	Marine Ecosystem FAO and CABI Publishing, Rome & Wallingford, UK, p 103-123

441	Dagorn L, Holland K, Hallier JPTM, Moreno G, Sancho G, Itano DG, Aumeeruddy R, Girard
442	C, Million J, Fonteneau A (2006) Deep diving behavior observed in yellowfin tuna
443	(Thunnus albacares). Aquat Liv Res 19:85-88
444	Davis R (2005) Intermediate-depth circulation of the Indian and South Pacific oceans
445	measured by autonomous floats. J Phys Oceanogr 35:683-707
446	DeNiro MJ, Epstein S (1978) Influence of diet on the distribution of carbon isotopes in
447	animals. Geochim Cosmochim Acta 42:495-506
448	DeNiro MJ, Epstein S (1981) Influence of diet on the distribution of nitrogen isotopes in
449	animals. Geochim Cosmochim Acta 45:341-351
450	Essington TE, Schindler DE, Olson RJ, Kitchell JF, Boggs C, Hilborn R (2002) Alternative
451	fisheries and the predation rate of yellowfin tuna in the eastern Pacific Ocean. Ecol
452	Appl 12:724-734
453	FAO (2003) Fisheries management 2. The ecosystem approach to fisheries FAO, Rome
454	FAO (2006) FAO Yearbook of Fishery Statistics Capture production 2004 FAO, Rome, Vol
455	98/1560pp
456	Fonteneau A (1997) Atlas of tropical tuna fisheries World catches and environment.
457	ORSTOM Editions, Paris, 191 p
458	François R, Altabet MA, Goericke R (1993) Changes in the $\delta^{13}$ C of surface water particulate
459	organic matter across the subtropical convergence in the SW Indian Ocean. Global
460	Biogeochem Cycles 7:627-644
461	Frank KT, Petrie B, Shackell NL, Choi JS (2006) Reconciling differences in trophic control in
462	mid-latitude marine ecosystems. Eco Lett 9:1-10
463	Frederiksen M, Edwards M, Richarson AJ, Halliday NC, Wanless S (2006) From plankton to
464	top predators: bottom-up control of a marine food web across four trophic levels. J
465	Animal Ecol 75:1259-1268

46	6 Fry B (1988) Food web structure on Georges Bank from stable C, N and S isotopic
46	7 compositions. Limnol Oceanogr 33:1182-1190
46	8 Fry B (2006) Stable Isotope Ecology. Springer, 308 pp
46	9 Gaye-Haake B, Lahajnar N, Emeis K-C, Unger D, Rixen T, Suthhof A, Ramaswamy V,
47	Schulz H, Paropkari AL, Guptha MVS, Ittekkot V (2005) Stable nitrogen isotopic
47	ratios of sinking particles and sediments from the northern Indian Ocean. Mar Chem
47	2 96:243-255
47	Graham BS, Popp B, Olson R, Allain V, Galvan F, Fry B (2006) Employing chemical tags to
47	determine trophic dynamics and movement patterns of migratory predators in the
47	equatorial Pacific Ocean. 5 <sup>th</sup> International Conference on Applications of Stable
47	Isotope Techniques to Ecological Studies Belfast-Northern Ireland 13-18 August
47	7 Graham BS, Grubbs D, Holland K, Popp BN (2007) A rapid ontogenetic shift in the diet of
47	juvenile yellowfin tuna from Hawaii. Mar Biol 150:647-658
47	9 Gruber N, Sarmiento JL (1997) Global patterns of marine nitrogen fixation and
48	denitrification. Global Biogeochem Cycles 11:235-266
48	1 Hernandez-Garcia V (1995) The diet of the swordfish Xiphias gladius Linnaeus, 1758, in the
48	central east Atlantic, with emphasis on the role of cephalopods. Fish Bull 93:403-411
48	3 Hilborn R, Walters CJ (1992) Quantitative Fisheries Stock Assessment. New York: Chapman
48	4 and Hall, 570 pp.
48	Hobson KA (1999) Tracing origins and migration of wildlife using stable isotopes: a review.
48	6 Oecologia 120:314-326
48	7 Insightful (2005) S-Plus 7 Insightful Corporation, Seattle, Washington
48	8 Jennings S, Warr KJ, Mackinson S (2002) Use of size-based production and stable isotope
48	analyses to predict trophic transfer efficiencies and predator-prey body mass ratios in
49	food webs. Mar Ecol Prog Ser 240:11-20

491	Kelly JF (2000) Stable isotopes of carbon and nitrogen in the study of avian and mammalian
492	trophic ecology. Can J Zool 78:1-27
493	Kornilova GN (1981) Feeding of yelllowfin tuna, Thunnus albacares, and bigeye tuna,
494	Thunnus obesus, in the Equatorial Zone of the Indian Ocean. J Ichthyol 20:111-119
495	Lesage V, Hammill MO, Kovacs KM (2001) Marine mammals and the community structure
496	of the Estuary and Gulf of St Lawrence, Canada: evidence from stable isotope
497	analysis. Mar Ecol Prog Ser 210:203-221
498	Longurst AR (1998) Ecological geography of the sea, Academic Press, 398 p
499	Lourey MJ, Trull TW, Sigman DM (2003) Sensitivity of $\delta^{15}N$ of nitrate, surface suspended
500	and deep sinking particulate nitrogen to seasonal nitrate depletion in the Southern
501	Ocean. Global Biogeochem Cycles 17: 1081, doi:10101029/2002GB001973
502	Maldeniya R (1996) Food consumption of yellowfin tuna, <i>Thunnus albacares</i> , in Sri Lankan
503	waters. Environ Biol Fish 47:101-107
504	Markaida U, Hochberg FG (2005) Cephalopods in the diet of swordfish (Xiphias gladius)
505	caught off the west coast of Baja California. Pacific Sci 59:25-41
506	Ménard F, Labrune C, Shin Y-J, Asine A-S, Bard F-X (2006) Opportunistic predation in tuna:
507	a size-based approach. Mar Ecol Prog Ser 323:223-231
508	Moteki M, Arai M, Tsuchiya K, Okamoto H (2001) Composition of piscine prey in the diet of
509	large pelagic fish in the eastern tropical Pacific Ocean Fisheries. Science 67:1063-
510	1074
511	Naqvi SWA, Naik H, Pratihary A, D'Souza W, Narvekar PV, Jayakumar DA, Devol AH,
512	Yoshinari T, Saino T (2006) Coastal versus open-ocean denitrification in the Arabian
513	Sea. Biogeosciences 3 621-633
514	Pinheiro JC, Bates DM (2000) Mixed-effects models in S and S-PLUS. Springer, New York,
515	528 pp

316	Popp BN, Granam BS, Olson RJ, Hannides CCS, Lott M, Lopez-Ibarra G, Galvan-Magana G
517	(In press) Insight into the trophic ecology of yellowfin tuna, Thunnus albacares, from
518	compound-specific nitrogen isotope analysis of proteinaceous amino acids. In Isotopes
519	as tracers in ecological change. T Dawson and R Seigwolf, eds Elsevier
520	Post DM (2002) Using stable isotopes to estimate trophic position: models, methods, and
521	assumptions. Ecology 83:703-718
522	Potier M, Marsac F, Lucas V, Sabatié R, Hallier J-P, Ménard F (2004) Feeding partitioning
523	among tuna taken in surface and mid-water layers: The case of yellowfin (Thunnus
524	albacares) and bigeye (Tobesus) in the western tropical Indian Ocean. Western Indian
525	Ocean J Mar Sci 3:51-62
526	Potier M, Marsac F, Cherel Y, Lucas V, Sabatié R, Maury O, Ménard F (2007) Forage fauna
527	in the diet of three large pelagic fishes (lancetfish, swordfish and yellowfin tuna) in
528	the western equatorial Indian Ocean. Fish Res 83:60-72
529	Quillfeldt P, McGill RAA, Furness RW (2005) Diet and foraging areas of Southern Ocean
530	seabirds and their prey inferred from stable isotopes: review and case study of
531	Wilson's storm-petrel. Mar Ecol Prog Ser 295:295-304
532	Rau GH, Sweeney RE, Kaplan IR (1982) Plankton <sup>13</sup> C: <sup>12</sup> C ratio changes with latitude:
533	differences between northern and southern oceans. Deep-Sea Res I 29:1035-1039
534	Rau GH, Mearns AJ, Young DR, Olson RJ, Schafer HA, Kaplan IR (1983) Animal <sup>13</sup> C/ <sup>12</sup> C
535	correlates with trophic level in pelagic food webs. Ecology 64:1314-1318
536	Roger C (1994) Relationships among yellowfin and skipjack tuna, their prey-fish and
537	plankton in the tropical western Indian Ocean. Fish Oceanogr 3:133-141
538	Rubenstein DR, Hobson KA (2004) From birds to butterflies: animal movement patterns and
539	stable isotopes. Trends Ecol Evol 19:256-263

540	Schart FS, Juanes F, Rountree RA (2000) Predator size-prey size relationships of marine fish
541	predators: interspecific variation and effects of ontogeny and body size on trophic-
542	niche breadth. Mar Ecol Prog Ser 208:229-248
543	Sheldon RW, Sutcliffe Jr, Paranjape MA (1977) Structure of pelagic food chain and
544	relationship between plankton and fish production. J Fish Res Board of Canada
545	34:2344-2353
546	Schell DM, Saupe SM, Haubenstock N (1989) Bowhead whale (Balaena mysticetus) growth
547	and feeding as estimated by $\delta^{13}C$ techniques. Mar Biol 103:433-443
548	Schott FA, McCreary JP (2001) The monsoon circulation of the Indian Ocean. Prog Oceanogr
549	51:1-123
550	Sinclair M, Valdimarsson G (2003) Responsible Fisheries in the Marine Ecosystem FAO &
551	CABI Publishing, Rome & Wallingford, UK
552	Takai N, Onaka S, Ikeda Y, Yatsu A, Kidokoro H, Sakamoto W (2000) Geographical
553	variations in carbon and nitrogen stable isotope ratios in squid. J Mar Biol Ass UK,
554	80:675-684
555	Tieszen LL, Boutton TW, Tesdahl KG, Slade NA (1983) Fractionation and turnover of stable
556	isotopes in animal tissues: Implications for $\delta^{13}C$ analyses of diet. Oecologia 57:32-37.
557	Tomczak M, Godfrey JS (1994) Regional Oceanography: an Introduction. Pergamon, Oxford,
558	442 pp
559	Trull TW, Armand L (2001) Insights into Southern Ocean carbon export from the $\delta^{13}C$ of
560	particles and dissolved inorganic carbon during the SOIREE iron release experiment.
561	Deep-Sea Res II 48:2655–2680
562	Wada E, Hattory A (1991) Nitrogen in the sea: forms, abundances, and rate processes. CRC
563	Press, Florida, USA, 208pp

564	Wallace B, Seminoff J, Kilham S, Spotila J, Dutton P (2006) Leatherback turtles as
565	oceanographic indicators: stable isotope analyses reveal a trophic dichotomy between
566	ocean basins. Mar Biol 149:953-960
567	Walters CJ (2003) Folly and fantasy in the analysis of spatial catch rate data. Can J Fish
568	Aquat Sci 60: 1433-1436
569	Watanabe H (1960) Regional differences in food composition of the tunas and marlins from
570	several oceanic areas. Report of Nankai Regional Fisheries Research Laboratory
571	12:75-84
572	Young J, Lansdell M, Riddoch S, Revill A (2006) Feeding ecology of broadbill swordfish,
573	Xiphias gladius, off eastern Australia in relation to physical and environmental
574	variables. Bull Mar Sci 79:793-809
575	

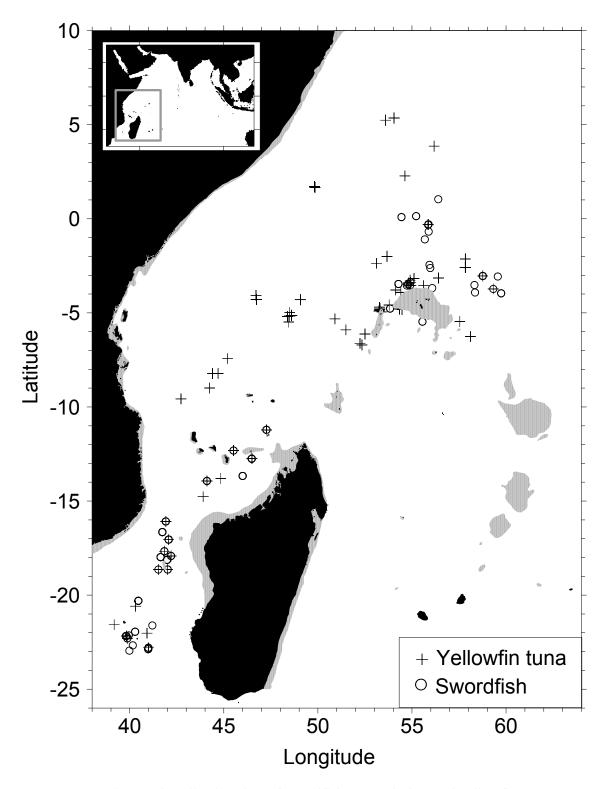
576	
577	

				Swordfish				١	Yellowfin				
				N	Si	ze	Lati	tude	N	Si	ze	Lati	tude
Cruise	Time period	Year	Group		min	max	min	max		min	max	min	max
AM1	Aug. 13-24	2001	SW monsoon	0					18	108	156	-4	2
AM2	Oct. 16-26	2001	SW monsoon	4	94	111	-4	0	5	122	140	0	0
AM6	Jul. 13-30	2002	SW monsoon	13	100	203	-4	-3	1	136	136	-3	-3
AM9	Jul. 4-9	2004	SW monsoon	5	90	226	-4	-2	1	123	123	-3	-3
ECO6	May 5-19	2004	SW monsoon	47	76	209	-17	-11	34	65	156	-17	-11
ECO9	Sept. 11-19	2003	SW monsoon	16	108	205	-23	-22	6	124	150	-22	-21
TG2	May 2 - June 21	2001	SW monsoon	0					32	42	160	-10	-6
GU1	Oct. 6 - Nov. 7	2001	SW monsoon	0					66	39	149	-4	5
AM3	Nov. 19-30	2001	NE monsoon	3	111	190	-5	-5	8	112	136	-5	-4
AM4	Jan. 28 - Feb. 8	2002	NE monsoon	3	113	162	-4	-3	10	102	153	-4	-3
AM5	Feb. 27 - Mar. 12	2002	NE monsoon	3	145	149	-1	1	0				
AM7	Dec. 10-20	2002	NE monsoon	2	114	137	-5	-5	5	88	141	-6	4
AM8	Jan. 29 - Feb. 9	2003	NE monsoon	7	76	162	-4	0	27	56	151	-5	0
ECO7	Nov. 10 - 29	2004	NE monsoon	33	68	199	-23	-18	9	96	160	-23	-18
TAL	March 28 - Apr. 29	2002	NE monsoon	0					2	58	60	-15	-14
AVAD	Feb. 2-10	2003	NE monsoon	0					21	61	164	-7	-5

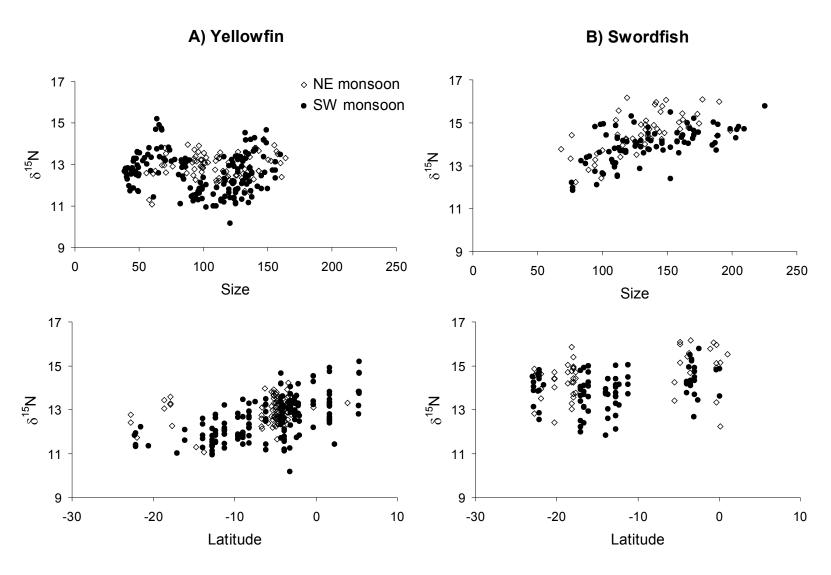
Table 2. Estimated intercepts and slopes (with standard errors) and within-season standard deviations  $\sigma_{season}$  for the random effects for linear mixed-effects models fit to the  $\delta^{15}N$  values of yellowfin tuna and swordfish, and estimated intercepts and slopes (with standard errors) for linear regression models fit to the  $\delta^{13}C$  values of yellowfin tuna and swordfish.

Со	efficients	$\delta^{15}N$	δ <sup>13</sup> C			
Intercept	Yellowfin tuna	12.995 (0.245)	-16.418 (0.0759)			
1	Swordfish	12.613 (0.414)	-16.568 (0.162)			
Latitude	Yellowfin tuna	0.0807 (0.0085)	0.0266 (0.0042)			
	Swordfish	0.0418 (0.0072)	0.0282 (0.0047)			
Size	Yellowfin tuna	0.0025 (0.0015)	0.0030 (0.0007)			
	Swordfish	0.0163 (0.0016)	0.0060 (0.0011)			
$\sigma_{\rm season}$	Yellowfin tuna	0.267	-			
22323-1	Swordfish	0.481	-			

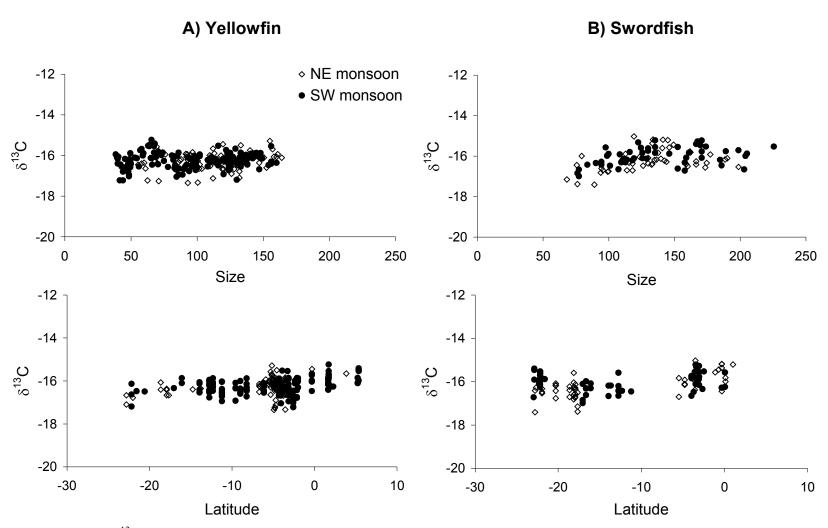
# 587 Legends of the figures 588 589 Figure 1. Muscle sample collection sites of swordfish (open circles) and yellowfin tuna 590 (crosses) in the western Indian Ocean. 591 Figure 2. Muscle $\delta^{15}$ N values from yellowfin tuna (A) and swordfish (B) plotted versus size 592 593 or latitude according to the considered season: SW monsoon (full symbols) and NE monsoon 594 (open symbols). 595 Figure 3. Muscle $\delta^{13}$ C values from yellowfin tuna (A) and swordfish (B) plotted versus size 596 597 or latitude according to the considered season: SW monsoon (full symbols) and NE monsoon 598 (open symbols). 599 Figure 4. Within-season predicted muscle $\delta^{15}N$ values (solid lines) from the linear mixed-600 601 effects models for yellowfin tuna (A) and swordfish (B) plotted versus size or latitude. The 602 original data (open circles) were superimposed on the predicted lines within each season, SW 603 monsoon (left side) and NE monsoon (right side). 604 Figure 5. Population predicted muscle $\delta^{15}N$ and $\delta^{13}C$ values from the linear models (linear 605 mixed-effects models for $\delta^{15}$ N and simple linear models for $\delta^{13}$ C) for swordfish (dashed line) 606 and yellowfin tuna (full line) plotted versus size (A) considering two different fixed latitudes 607 (0°N or 10°S); and plotted versus latitude (B) for two different sizes (80cm or 160cm). 608 609



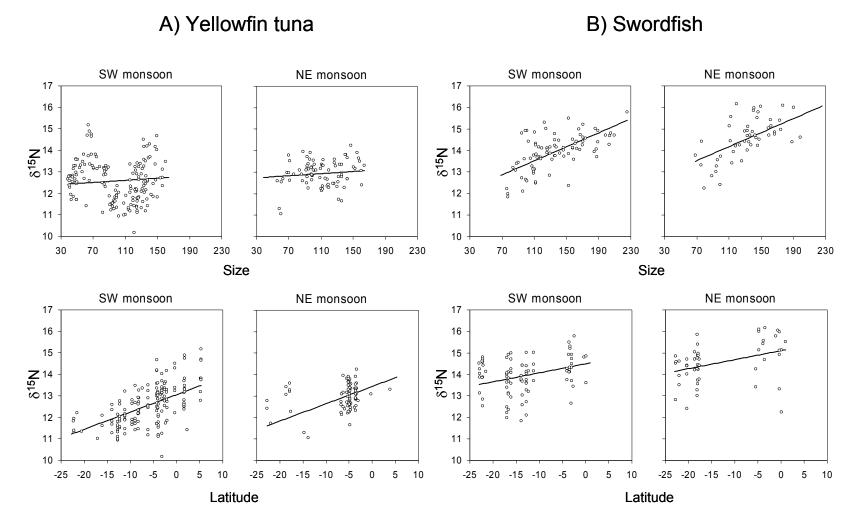
**Figure 1.** Muscle sample collection sites of swordfish (open circles) and yellowfin tuna (crosses) in the western Indian Ocean.



**Figure 2.** Muscle  $\delta^{15}$ N values from yellowfin tuna (A) and swordfish (B) plotted versus size or latitude according to the considered season: SW monsoon (full symbols) and NE monsoon (open symbols).



**Figure 3.** Muscle  $\delta^{13}$ C values from yellowfin tuna (A) and swordfish (B) plotted versus size or latitude according to the considered season: SW monsoon (full symbols) and NE monsoon (open symbols).



**Figure 4.** Within-season predicted muscle  $\delta^{15}$ N values (solid lines) from the linear mixed-effects models for yellowfin tuna (A) and swordfish (B) plotted versus size or latitude. The original data (open circles) were superimposed on the predicted lines within each season, SW monsoon (left side) and NE monsoon (right side).

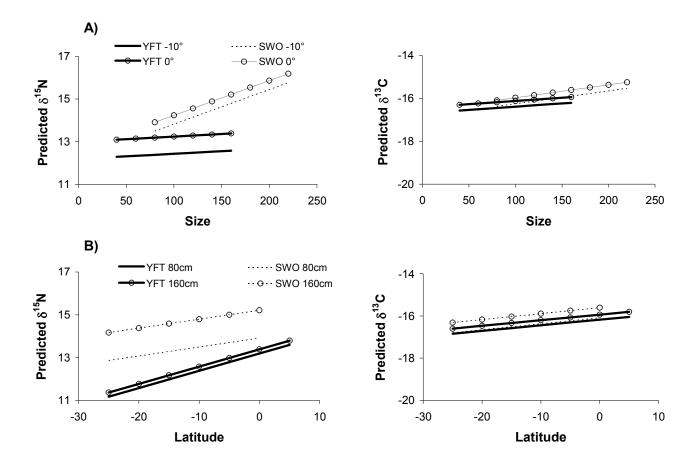


Fig 5. Population predicted muscle  $\delta^{15}N$  and  $\delta^{13}C$  values from the linear models (linear mixed-effects models for  $\delta^{15}N$  and simple linear models for  $\delta^{13}C$ ) for swordfish (dashed line) and yellowfin tuna (full line) plotted versus size (A) considering two different fixed latitudes (0°N or 10°S); and plotted versus latitude (B) for two different sizes (80cm or 160cm).