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Characterization of the organic composition of aerosols from Rondônia, Brazil, during the LBA-SMOCC 2002 experiment and its representation through model compounds

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Abstract

The chemical composition of carbonaceous aerosols collected during the LBA-SMOCC field experiment, conducted in Rondônia, Brazil, in 2002 during the transition from the dry to the wet season, was investigated by a suite of advanced analytical techniques.

5 The period of most intense biomass burning was characterized by high concentrations of submicron particles rich in carbonaceous material and water-soluble organic compounds (WSOC). At the onset of the rainy period, submicron total carbon (TC) concentrations had decreased by about 20 times. In contrast, the concentration of supermicron TC was fairly constant throughout the experiment, pointing to a constant
10 emission of coarse particles from the natural background. About 6–8% of TC (9–11% of WSOC) was speciated at the molecular level by GC-MS and liquid chromatography. Poly-hydroxylated compounds, aliphatic and aromatic acids were the main classes of compounds accounted for by individual compound analysis. Functional group analysis by proton NMR and chromatographic separation on ion-exchange columns allowed characterization of ca. 50–90% of WSOC into broad chemical classes (neutral species/light acids/humic-like substances). In spite of the significant change in the chemical composition of tracer compounds from the dry to the wet period, the functional groups and the general chemical classes of WSOC changed only to a lesser extent. Model compounds representing size-resolved WSOC chemical composition for
20 the different periods of the campaign are then proposed in this paper, based on the chemical characterization by both individual compound analysis and functional group analysis deployed during the LBA-SMOCC experiment. Model compounds reproduce quantitatively the average chemical structure of WSOC and can be used as best-guess surrogates in microphysical models involving organic aerosol particles over tropical areas affected by biomass burning.
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1. Introduction

The Large Scale Biosphere-Atmosphere Experiment in Amazonia-Smoke, Aerosols, Clouds, Rainfall and Climate (LBA-SMOCC) experiment was conducted in the Amazon Basin in the period Sept.–mid-Nov. 2002, with the principal purpose of investigating how and to what extent aerosol particles produced by biomass burning alter cloud formation. Previous studies have highlighted that smoke particles emitted by biomass burning are enriched in organic carbon (OC) and also contain a variable amount of elemental carbon (EC), operationally defined as the fraction of carbon that is refractory at high temperature in an inert atmosphere (Chow et al., 2001; Mayol-Bracero et al., 2002a, b). Elemental carbon is often used as a synonym for “soot carbon” defined by IPCC(2001) as “Particles formed during the quenching of gases at the outer edge of flames of organic vapors, consisting predominantly of carbon, with lesser amounts of oxygen and hydrogen present as carboxyl and phenolic groups and exhibiting an imperfect graphitic structure”.

Although carbonaceous aerosols are less hygroscopic than particles consisting of sulfate or sodium chloride, the polar organic compounds known to occur in biomass burning aerosol may absorb water from the gas phase, thus enhancing the ability to nucleate cloud droplets (“CCN ability”) of the particles (Svenningsson et al., 2005). The organic compounds that have an affinity to water are generally isolated by extracting aerosol samples with water and measuring their total carbon concentration by liquid total organic carbon (TOC) analysis. These water-soluble organic compounds (WSOC) constitute a variable fraction of the aerosol TC. They can be internally or externally mixed with other aerosol constituents (e.g., inorganic soluble and insoluble components) and, most importantly, they contain a wide range of chemical species that are expected to show very different water solubilities (Mochida and Kawamura, 2004). All these factors are of primary importance in determining the CCN ability of biomass burning particles.

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In order to evaluate the net effect of carbonaceous particles from biomass burning on the formation of clouds, it is indicated to compare their characteristics with those formed during unpolluted episodes in the same region. The SMOCC aircraft campaign revealed that forested areas distant from the biomass burning sources show a background of CCN with low concentrations and a rather constant vertical profile in the troposphere (Andreae et al., 2004). Previous studies highlighted the occurrence of natural sources of aerosol particles from the biota in the same area (Graham et al., 2003). The processes responsible for particle formation in the Amazon basin – biomass burning and biogenic emission – are geographically distinct and follow very different temporal trends. In particular, the dramatic increase in the aerosol load observed during the dry period in the rural areas of Rondônia and Mato Grosso can be mainly attributed to biomass burning. In the same period, biomass burning products strongly impact the TC concentrations and composition in the neighboring forested areas. Biogenic organic aerosols have mainly been characterized at forest sites located in a large distance from the main pollution sources (Kubátová et al., 2000; Graham et al., 2003; Claeys et al., 2004a). However, primary biological particles (e.g., pollen and fungal spores) were found to increase also in rural areas at the end of the dry period, when the increase in precipitation impedes extensive burning activities (Graham et al., 2002).

The chemical composition of carbonaceous particles produced by biomass burning in Rondônia was characterized during the 1999 LBA-EUSTACH-2 campaign (Mayol-Bracero et al., 2002a). It was found that WSOC account for 45–75% of TC and that a substantial fraction of the thermally refractory carbon determined by evolved gas analysis (EGA) analysis is also water-soluble. This soluble refractory carbonaceous material was linked to complex polycarboxylic acids, which are denoted by the generic term “HULIS” (humic-like substances) and were determined by analysis of water extracts. Another important fraction of WSOC was identified as neutral compounds mainly consisting of sugar-like compounds such as levoglucosan, which is the most abundant product of the pyrolysis of cellulose at temperatures higher than 300°C (Shafizadeh,

1984).

Except for levoglucosan, linking the organic composition of the aerosol to the chemical reactions occurring during the complex and varying combustion processes remains an issue. The relatively low concentrations of sugars in biomass burning smoke, compared to the abundance of their degradation products, likely results from the fact that biomolecules undergo pronounced chemical transformations, even at relatively low temperatures (<200°C), owing to heterogeneous reactions with oxygen and reactions between amines and sugars (Moens et al., 2004). Combustion studies on Gramineaceae (Knicker et al., 1996) have shown that at 350°C cellulose and hemicelluloses are completely degraded to volatile products (with levoglucosan as a major compound) in a time span of 1 min. The remaining char products contain mainly complex aliphatic compounds and newly formed aromatic compounds, and have a very low content of oxygenated substances. The composition of these char residues is completely different from that of biomass burning aerosol which instead shows a large fraction of oxygenated compounds (Graham et al., 2002). The oxygenated compounds may form during the low-temperature stages of combustion. In addition, a substantial fraction of compounds volatilized at the high temperatures of the combustion process may condense onto particles, when the smoke plume cools down. Finally, the photochemical production of newly formed condensable compounds within the plume and in the regional haze will increase the fraction of oxygenated species relative to soot (Gao et al., 2003; Reid et al., 2004). Therefore, there are a number of processes that are responsible for the high content of oxygenated water-soluble organic compounds in biomass burning particles, ranging from combustion itself, through the early aging stages to actual in-situ secondary processes. Chemical transformation within the aerosol particles, e.g., polymerization of low-molecular weight compounds, is not a likely mechanism for altering significantly the oxygen to carbon ratio of the organic matter. However, it may alter its solubility through the modification of functional groups of specific classes of compounds, for example, benzoic acids may be converted to high-molecular weight

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aromatic acids (Hoffer et al., submitted, 2005¹) and unsaturated acids may be further oxidized to dihydroxy acid derivatives (Claeys et al., 2004b).

The biogenic sources of organic aerosols are still only partly understood and cover a wide range of different sources and source processes. The identification of biomolecules (mainly sugars and lipids) in the coarse fraction of aerosols collected at forest sites has highlighted the importance of the direct emission of primary biological particles, like spores, pollen, plant debris, soil detritus and insect body parts, to the atmosphere (Graham et al., 2003; Simoneit et al., 2004). Recently, another source process has been characterized for the formation of secondary organic aerosols, i.e., photo-oxidation of isoprene which is emitted in large amounts by the tropical forest vegetation (Claeys et al., 2004a; Wang et al., 2005).

Biomass burning and biogenic sources show a very different seasonal dependence. In a simplified scheme since the Amazon forest ecosystem is productive throughout the year, while biomass burning mainly occurs in the dry period, the latter gives rise to episodes of very high aerosol loads, which are superimposed on a relatively constant natural background of primary biological particles and photo-oxidation products of biogenic volatile organic compounds (secondary organic aerosol). Consequently, the atmospheric concentrations of biomass burning aerosols, along with their composition, will vary according to the strength and type of the combustion sources, as well as the ambient conditions, with strong diel and day-to-day variations (Falkovich et al., 2005; Schkolnik et al., 2005; Fuzzi et al., to be submitted, 2005²). The aim of this

¹Hoffer, A., Gelencsér, A., Blazsó, M., Guyon, P., Artaxo, P., and Andreae, M. O.: Chemical transformation in organic aerosol from biomass burning, *Atmos. Chem. Phys. Discuss.*, submitted, 2005.

²Fuzzi, S., Decesari, S., Facchini, M. C., Cavalli, F., Emblico, L., Mircea, M., Andreae, M. O., Trebs, I., Hoffer, A., Guyon, P., Artaxo, P., Rizzo, L.V., Lara, L. L., Pauliquevis, T., Maenhaut, W., Raes, N., Chi, X., Mayol-Bracero, O. L., Soto, L., Claeys, M., Kourchev, I., Rissler, J., Swietlicki, E., Tagliavini, E., Schkolnik, G., Falkovich, A. H., Rudich, Y., Fisch, G., and Gatti, L. V.: Overview of the inorganic and organic composition of size-segregated aerosol in Rondônia,

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study is to trace the changes in the organic aerosol composition during the transition from the biomass burning period to the rainy period, with particular focus on WSOC. A combination of state-of-the-art techniques for speciation of polar organic compounds has been exploited for the chemical characterization, together with functional group analysis by ¹HNMR. The results and implications of some specific analytical methods have been published in separate papers (Falkovich et al., 2005; Schkolnik et al., 2005). Here, they are included to provide comprehensive compositions for the main periods of the LBA-SMOCC field campaign. In past studies on tropical areas affected by biomass burning, only limited analyses (e.g., ion chromatography or GC-MS) on selected samples have been performed, providing sets of measurements that are difficult to inter-compare (Reid et al., 2004). Here, we provide what is probably the most complete set of analyses of biomass burning aerosols, for a full assessment of the organic chemical composition. Finally, we propose a model representation of the water-soluble fraction of OC, derived from both individual compounds and functional group composition for the different periods of the campaign. These model compositions should be useful in microphysical models of aerosol hygroscopic growth, as well as in laboratory studies for the determination of the hygroscopic behavior of mixed organic/inorganic systems (Svenningsson et al., 2005).

2. Experimental

2.1. Sampling

The SMOCC field campaign was conducted at a ground-based station on the Fazenda Nossa Senhora Aparecida (FNS) (10°45'44" S, 62°21'27" W, 315 m a.s.l.), which is located approximately 8 km southwest of the town Ouro Preto do Oeste in the state of Rondônia, Brazil. The place was deforested by fire about 20 years ago and the area

Brazil, from the biomass burning period to the onset of the wet season, to be submitted to J. Geophys. Res., 2005.

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is now a pasture site with *Brachiaria brizantha* as a dominant grass species. Sampling of aerosol particles was conducted from 9 September to 14 November 2002, by deploying a series of impactors and filter-based techniques. A full description of the sampling platform is provided by Fuzzi et al. (2005²). According to meteorological conditions, the sampling period was subdivided into dry (intense burning; 11 September to 7 October), transition (8 October to 29 October) and wet (30 October to 14 November) periods. With respect to the determination of TC and organic and elemental carbon (OC and EC), and of the organic chemical composition, sampling was conducted by:

5 a) a stacked filter unit (SFU) sampler that separates coarse ($10 > d > 2.0 \mu\text{m}$, where d is the aerodynamic diameter) and fine particles ($d < 2.0 \mu\text{m}$) by sequential filtration on 8.0 and $0.4 \mu\text{m}$ pore-size Nuclepore® polycarbonate filters, with a flow rate of 10–15 l/min.;

10 b) three high-volume dichotomous impactors (HVDS) with front and back quartz fiber filters, segregating fine (PM_{2.5}) and coarse ($> 2.5 \mu\text{m}$) particles, operating at a flow rate of ca. 300 l/min; c) two microorifice uniform deposit impactor (MOUDI) devices,

15 one (from Institute of Physics of the University of Sao Paulo (IFUSP)) with Nuclepore® polycarbonate filters as impaction foils, the other (from Ghent University (UGent)) with aluminum foils on the impaction stages; d) a 5-stage Berner impactor with aluminum and Tedlar foils on the impaction stages. Sampling time varied from 12 h in the dry period (when the highest aerosol concentrations were encountered) to 24 and 48 h at

20 the end of the campaign. Positive artifacts during sampling were either avoided by using inert substrates (polycarbonate, Tedlar, Aluminum) or corrected by the analysis of back-filters (in the case of quartz filters). In contrast, presumably semi-volatile organic compounds (Eatough et al., 2003) could not be efficiently collected by our sampling apparatus.

25 The comparison of aerosol measurements performed with analogous sampling systems (e.g., HVDS and SFU) indicates that the uncertainties in the flow measurement and potential other minor sampling biases, such as differences in the size cutoff between samples, led to differences of up to 25% in the concentration data reported from the various samplers and groups. Where such biases could be quantified and correc-

tions applied, this is indicated in the text and tables. Otherwise, values are reported as obtained from the species mass measured on the particular samples and the sample volumes determined with the flow meters attached to the individual samplers.

2.2. TC/OC/EC analysis

5 Ghent University (UGent) analyzed the fine and coarse filter samples (both front and back filters) of two HiVol samplers (HVDS1 and HVDS3) for OC, EC and TC (TC=OC+EC) with a thermal-optical transmission (TOT) technique (Birch and Cary, 1996; Schmid et al., 2001), using a thermal-optical carbon analyzer from Sunset Laboratory Inc. (Tigard, OR, USA). The analysis was done in a two-stage procedure; one
10 or two 1.5-cm² rectangular punches of each quartz filter were heated stepwise (up to 900°C) in a non-oxidizing helium (He) atmosphere, and then (again up to 900°C) in an oxidizing atmosphere of 2% oxygen and 98% He. The carbon that evolves at each temperature is oxidized to carbon dioxide (CO₂), and then reduced to methane (CH₄) for quantification with a flame ionization detector (FID). The transmittance of light from
15 a He-Ne laser through the filter punches is continuously monitored and used for setting the OC/EC “split” point, thereby correcting for pyrolysis/charring during the first stage of the analysis.

Max Planck Institute for Chemistry (MPIC) measured total carbon (TC) and the elemental carbon after water extraction (EC_w) on HiVol samples (HVDS2) by evolved gas analysis (EGA) (Mayol-Bracero et al., 2002; Hoffer et al., 20051). The combustion
20 was performed in an oxygen atmosphere, the temperature was increased linearly from 50°C to 780°C at a rate of 20°C/min, and the conversion was completed over a MnO₂ catalyst at 800°C. The EC concentration was determined integrating the last peaks of the thermograms obtained after water extraction (Mayol-Bracero et al., 2002).

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2.3. TOC analysis

UGent analyzed the fine ($<2.5\ \mu\text{m}$) filter samples (both front and back filters) from HVDS1 for total organic carbon (TOC). Filter punches of 1 or $1.5\ \text{cm}^2$ were placed in a 15 ml tube, 5 or 10 ml Millipore Simplicity water was added, and the tube was hand-shaken during 5 min, after which it was allowed to stand for 30 min. The sample extract was then filtered through a PVDF syringe filter (pore size $0.2\ \mu\text{m}$) and analyzed for TOC, thereby correcting for the inorganic carbon, with a Shimadzu TOC-V CPH analyzer. The TOC data were used as water-soluble OC.

ISAC subjected a sub-set of coarse HiVol (HVDS3) filter samples, having collected supermicron particles ($>2.5\ \mu\text{m}$), to TOC analysis. Upon extraction of 1/4 of filter with 30 ml of deionized water, samples were filtered to remove the filter debris and analyzed by a Shimadzu TOC5000A analyzer. Blank levels were 1 ppm C and 0.25 ppm C, when filtering with hydrophilic cellulose filters and PTFE hydrophobic filters, respectively. Only coarse ($>2.5\ \mu\text{m}$) filters from the transition and the wet periods could be analyzed, because of the relatively low interference from fine particles (Graham et al., 2002).

Pre-cleaned Tedlar substrates mounted on a 5-stage Berner impactor were extracted in 6 ml of deionized water in an ultrasonic bath, and analyzed by means of a Shimadzu TOC5000A analyzer. Blank levels were around 0.2 ppm C and a total of 37, 12, 5 size-segregated samples were analyzed for the dry, transition and wet periods, respectively.

2.4. GC-MS_{MPIC}

The fine fraction of the aerosols collected by the HVDS2 sampler was analyzed for individual polar compounds by gas chromatography-mass spectrometry (Hewlett Packard 6890 GC-MSD) after derivatization. The method was adapted from Graham et al. (2002). The samples ($3.5\text{--}5.3\ \text{cm}^2$ of the filter) were extracted in 4–6 ml of acetonitrile for 1 h, agitated once every 15 min. The extract was then filtered through a $0.45\ \mu\text{m}$ pore size PTFE syringe filter (Pall). An internal stan-

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5 dard (3,3-dimethylglutaric acid) was added to 3 ml of filtered sample solution, which was then brought to dryness under a gentle stream of N₂. After this process, 50 μl pyridine and 50 μl bis(trimethylsilyl)trifluoroacetamide (BSTFA), containing 1% trimethylchlorosilane (TMCS) as a catalyst (Supelco), were added to the samples, which were then put into an oven at 70°C for 30 min. The sample was injected onto a HP5-MS column (30 m × 250 μm × 0.25 μm) equipped with a Supelco guard column (deactivated methylsiloxane, 1 m × 0.32 mm) in a splitless mode at 280°C. The temperature of the oven was held at 65°C for 10 min and ramped at 10°C/min to 310°C and held for 10 min. The detector was operated both in SIM and SCAN mode, and it was calibrated with aliquots of a stock solution of authentic standards. Repeated analysis of the samples showed that the precision of the method was about 20%.

2.5. GC-MS_{UA}

15 A series of polar organic compounds in aerosol samples, i.e., the front fine filters of the HiVol quartz filter samples (HVDS1) and aluminum foils from the MOUDI collections, were quantified using GC-MS techniques that incorporated derivatisation into trimethylsilyl derivatives. Two methods were employed: the first method was targeted to the quantitation of sugar-like compounds (i.e., anhydrosugars (levoglucosan, mannosan, galactosan, 1,6-anhydro-β-D-glucofuranose), the 2-methyltetrols (2-methylthreitol, 2-methylerythritol), C₅ alkene triol derivatives of isoprene (2-methyl-1,3,4-trihydroxy-1-butene (cis and trans) and 3-methyl-2,3,4-trihydroxy-1-butene), the monosaccharides (fructose, glucose), and the sugar polyols (arabitol, mannitol)), while the second method was targeted to the quantitation of acidic compounds (i.e., hydroxy monocarboxylic acids (glyceric acid, threonic acid + isomer), dicarboxylic acids (succinic acid, fumaric acid, glutaric acid), hydroxy dicarboxylic acids (malic acid, α-hydroxy glutaric acid, β-hydroxy glutaric acid, tartaric acid) and aromatic acids (hydroxy benzoic acids, vanillic acid, isovanillic acid)).

25 Method 1: The first method was adapted from one previously described for the determination of levoglucosan in urban aerosols (Pashynska et al., 2002). A part

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of the quartz filter sample (1/16 or 1/32) of the whole filter area was used for extraction. Before extraction, the recovery standard, methyl β -L-xylanopyranoside was added. In the case of analysis of aluminum foils, 1/2 or 1/4 of the whole foil was used and the foil was also spiked with a second internal recovery standard, deuterated (d_3)-malic acid (2,2,3- d_3 -malic acid; Cambridge Isotope Laboratories, Andover, MA, USA), for measurement of malic acid. Sample workup consisted of extraction with dichloromethane:methanol (4:1, v/v) and trimethylsilylation. GC-MS analysis was performed with a TRACE GC2000 gas chromatograph and a Polaris Q ion trap mass spectrometer equipped with an external electron ionization source (ThermoFinnigan, San Jose, CA, USA) using an electron energy of 70 eV. For data acquisition and processing, Xcalibur version 1.2 software was used. The gas chromatograph was equipped with a deactivated silica precolumn (2 m \times 0.25 mm i.d.) and a CP Sil 8CB low-bleed capillary column (95% dimethyl-, 5% phenylpolysiloxane, 0.25 mm film thickness, 30 m \times 0.25 mm i.d.; Chrompack, Middelburg, The Netherlands). The following temperature program was applied: the initial temperature was 50°C and kept for 5 min, the temperature was then increased to 200°C at the rate of 3°C/min and kept at that temperature for a further 2 min and then raised to 310°C at the rate of 30°C/min. The total analysis time was 62 min. The GC-MS instrument was operated in the full scan mode in the mass range m/z 45-500, and quantification was based on mass chromatographic data (i.e., extracted ion chromatograms). For derivatisation of standard solutions of all saccharidic compounds, the same procedure was applied. The quantification of the monosaccharide anhydrides (levoglucosan, mannosan, and galactosan), the polyols (erythritol, arabitol, and mannitol), the monosaccharides (fructose and glucose), and the 2-methyltetrols (2-methylthreitol and 2-methylerythritol) was based on an internal standard calibration procedure employing methyl β -L-xylanopyranoside (and d_3 -malic acid in case of the Al foils) as internal recovery standard and pure reference compounds, if available. For assessing the amount of the 2-methyltetrols, 2-methylthreitol and 2-methylerythritol, and the C_5 alkene triol derivatives of isoprene (Wang et al., submitted, 2005²), for which no pure reference compounds were available, the response

factor of erythritol was used. In the case C₅ alkene triol derivatives of isoprene were observed, which are related to 2-methyltetrols, they were lumped with the 2-methyltetrols. Duplicate analyses show that the precision of the determinations was about 10%. All reported concentrations are corrected for procedural blanks.

Method 2: The second method employs the same analytical principles as method 1 and is also based on the use of internal recovery standards. A part of the quartz filter sample (1/4) of the whole filter area was used for extraction. All glassware used for sample workup was deactivated with 5% dimethyldichlorosilane in toluene in order to minimize adsorption and loss of polar acidic compounds. Before extraction, three recovery standards (3.75 μg of each) were added to the filter sample: (a) deuterated glutaric acid (2,2,4,4-d₄-pentanedioic acid; Cambridge Isotope Laboratories), (b) deuterated malic acid (2,2,3-d₃-malic acid; Cambridge Isotope Laboratories) and (c) tropic acid (3-hydroxy-2-phenylpropionic acid; Fluka, Buchs, Switzerland) were added. Deuterated glutaric acid served as internal recovery standard for the dicarboxylic acids, i.e., glutaric acid, succinic acid and fumaric acid. Deuterated malic acid served as internal recovery standard for the hydroxy mono- and dicarboxylic acids, i.e., malic acid, glyceric acid, α- and β-hydroxy glutaric acid, threonic acid, an isomer of threonic acid, and tartaric acid. Tropic acid served as internal recovery standard for the aromatic acids, i.e., 2-, 3- and 4-hydroxybenzoic acid, vanillic acid and isovanillic acid. Extraction was performed in 25 ml Pyrex flasks with methanol (3 times with 20 ml) under ultrasonic agitation for 15 min. The subsequent steps were the same as for method 1. For assessing the amounts of the analytes, the response factor of pure reference compounds was used. Duplicate analyses show that the precision of the determinations was about 10%. All reported concentrations are corrected for procedural blanks.

2.6. IC

A 1/4 of each HiVol sample was extracted twice into 5.0 ml of water, by short vortex agitation followed by 15 min of gentle shaking. The combined extract was centrifuged for 5 min and filtered through a GHP Acrodisk® syringe filter (25 mm, 0.45 μm pore size;

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Gelman, Pall Corporation, NY, USA), which had been previously washed with 10 ml water. Each MOUDI and half of each SFU sample were extracted in the same manner into 4.0 ml of water. These were filtered using a GHP Acrodisk® syringe filter (13 mm, 0.45 μm pore size). It has been validated that further extraction was not needed.

5 IC analysis was carried out using a Varian ProStar HPLC system equipped with a Dionex ED50 electrochemical detector. Anions were determined using a Dionex AS11 analytical column and ASRS-Ultra suppressor in autosuppression mode. For simultaneous separation of inorganic and short-chain ($\text{C}_1\text{--C}_9$) organic anions, gradient elution by 0.4–25 mM NaOH (2 mL/min) was employed (Table 1). Cations were determined using a Dionex CS12 column and CSRS-Ultra suppressor in autosuppression mode with 10 20 mM methanesulfonic acid (MSA) as an eluent (1 ml min^{-1}). All ions were quantified against standard calibration curves. A thorough validation of extraction and analytical method can be found in Falkovich et al. (2004).

2.7. IEC

15 Sample extracts obtained as described for IC analysis were purified using AccellTM QMA solid-phase extraction cartridges (Waters, MA, USA) in order to eliminate HULIS, which interfered with detection. The samples were separated using ion exclusion liquid chromatography using a Dionex ICE-AS1 column and a Varian ProStar 230I HPLC pump, and polyhydroxy compounds were detected by photodiode array (Varian ProStar 330) at 194 nm (for a detailed description of the method, please see Schkolnik 20 et al., 2005). The method uncertainty is 15% for concentrations $>0.2\ \mu\text{g/m}^3$, and 23% for concentrations $<0.2\ \mu\text{g/m}^3$.

2.8. IC–UV

25 Ion-exchange chromatography on a tertiary amine stationary phase (DEAE) was used to fractionate WSOC according to their ionic nature at pH 8. The whole set of 5-stage Berner impactor samples collected during the SMOCC field campaign was analyzed by

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the IC-UV technique, implemented on a Hitachi L-7100 HPLC system equipped with a Gilson autosampler, a Toso-Haas DEAE-TSK gel column (7.5 mm i.d. × 7.5 cm l.), and an UV detector (260 nm). The aliquots of samples for HPLC analysis were dried under vacuum and re-dissolved with 300 μ l of mobile phase A, necessary to fill the 100 μ l loop of the HPLC system. The injection of samples dissolved in the first mobile phase instead of water allowed the suppression of the injection peak in the chromatogram, in order to perform a more accurate integration of the peak arising from non-retained analytes.

The mobile phase was an aqueous solution of 20% acetonitrile and NaClO_4 at the following concentrations: A) 0 M; B) 0.02 M; C) 0.4 M. The pH of the mobile phases B and C was held constant at 8.0 with a 0.01 M TRIS:HCl buffer. A first isocratic elution was followed by a gradient (from 12 to 15 min) changing the solvent composition from A to B; after 6 min of isocratic conditions, a second gradient from 21 to 26 min allowed the system to reach the 100% C composition. A last five minute gradient changed the composition of the mobile phase to 100% A. Flow rate was 0.7 ml min^{-1} . The compounds eluted A) with the first eluent (from 3 to 17 min), B) after the increase of the NaClO_4 concentration to 0.02 M (from 17 to 26 min), or C) after the second increase of the ionic strength (from 26 to 32 min) were classified as neutral compounds (NC), mono-/di-acids (MDA) and polyacids (PA), respectively. Compared to the original analytical procedure proposed by Decesari et al. (2000), the elution method used in the present study improves the separation between NC and the acidic fractions, allowing to overcome the elution problems with the phenolic compounds (Chang et al., 2005). The attribution of the separated fractions to chemical classes has been verified by injection of standard compounds (their retention time in minutes is indicated in parentheses in the following listing) selected to represent all the classes of polar organic compounds known to occur in biomass burning aerosol (Graham et al., 2002). NC: D-galactal (4.5), D-glucal (4.5), furfuryl alcohol (5.0), benzyl alcohol (5.6), vanillin (9.8), phenol (10.2), m-cresol (11.9); MDA: glyoxylic acid (19.2), sodium formate (19.7), sodium oxalate (23.4), malic acid (23.9), potassium hydrogen-phthalate (25.1); PA: 1,3,5-benzene-tricarboxylic acid

(28.3), citric acid (28.5), Suwannee River fulvic acid (ca. 29). Clearly, the retention times of all the standards match with those expected according to the classification into the three main chromatographic fractions.

Calibration factors to convert the peak areas of NC, MDA and PA into their specific carbon concentrations were provided by chromatographic fractions isolated on preparative glass columns of DEAE-cellulose gel. Buffer solutions of ammonium bicarbonate were used for selective elution of NC, MDA and PA. In order to improve the resolution while keeping low the volume of mobile phase necessary to elute the strongly retained compounds, two columns, A and B (lengths: 1 cm + 4 cm), were initially used in series to separate NC from the acidic compounds retained on the column. MDA were subsequently eluted with the 0.02 M buffer, while in a second step PA were eluted with 1 M buffer directly from column A by-passing B. This procedure allows the elution of each fraction with an amount of ammonium bicarbonate low enough to be easily removed by rotary evaporation. The isolated fractions were then analyzed for TOC and by HPLC, providing an external calibration for the HPLC technique. The precision of the procedure for providing calibration factors is 8% for NC and PA and 20% for MDA, while the variability between samples belonging to homogenous sets (e.g., the samples of the dry, transition or wet periods) introduces an uncertainty of 10–20% up to 30% in the case of NC in the coarse aerosol samples.

3. Results

3.1. Concentrations and trends of TC, EC and total WSOC

Figure 1 shows the temporal trends of TC from the TOT analysis of HVDS1 filters in the fine and coarse size fractions throughout the experiment. EC data provided by EGA analysis of water-extracted HVDS2 filters (fine fraction) are also reported after correction for sampling biases between the two HiVol systems. Data are already corrected for the back-up filter contribution. EC accounts for 5 to 20% of TC in the fine fraction of the

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aerosol (Table 1). These data are in agreement with results reported in the literature on biomass burning in tropical forests (Reid et al., 2004). It should be noted that the EC values provided by EGA analysis of water-extracted filters are about six-fold higher than those measured by TOT analysis (not shown in the figure), while they are substantially in agreement with those determined by means of a Ruprecht and Patashnik (R&P) 5400 carbon analyzer (Fuzzi et al., 2005²). The partitioning between OC and EC in TOT analysis depends strongly on the temperature program used (Schmid et al., 2001), especially for biomass smoke aerosols, and suffers from artifacts when filters are heavily loaded (Kubatova et al., 1999).

The figure also shows the concentrations of total WSOC determined on the same HVDS filters used for TOT analysis, plus the total WSOC determined on the Berner impactor samples obtained by lumping the impactor stages corresponding to an aerosol diameter $<1.2\mu\text{m}$ (fine fraction), and those sampling between 1.2 and $10\mu\text{m}$ (coarse fraction). Generally, the Berner impactor was less efficient than the HVDS in sampling WSOC in the fine fraction, providing concentrations 20% lower on average during the first part of the campaign, when the aerosol concentrations were highest and the discrepancy between the samplers largest. The air concentrations of WSOC are always well correlated with those of TC and PM (the latter are not shown in the figure), with temporal trends that reflect the main factors controlling the aerosol loads in the boundary layer as discussed in a parallel paper (Fuzzi et al., 2005²). Clearly, the high aerosol concentrations of submicrometer carbonaceous particles observed in September and in the beginning of October (i.e., the dry period) must be attributed to the intense biomass burning activities in Rondônia and Mato Grosso, and other upwind Brazilian states at that time. Under high pressure meteorological conditions, the height of the tropical continental boundary layer undergoes a diurnal cycle, which causes fluctuations in the aerosol loads with clear maxima during the night.

After the intense precipitation event on 8 October, the concentrations of biomass burning particles started to increase again but without reaching the peaks observed in the dry period. This period was denoted as the “transition period”. Finally, “wet”

conditions with frequent rain episodes became established after 1 November, leading to a substantial decrease in the concentrations of the fine particles, down to approx. 4% relative to the average concentrations observed in the dry period.

Conversely, the TC in the coarse fraction did not undergo a significant decrease from the dry to the wet period (Fig. 1b), suggesting that its concentrations were only marginally affected by the biomass burning emissions and were controlled by other sources that also hold for periods of intense precipitation. In contrast to the strong dominance of submicrometer WSOC mass during dry conditions, the concentrations of the WSOC in the fine and coarse fractions were comparable (0.72 and $0.50 \mu\text{g C/m}^3$ on average, respectively) during the wet period. The trends in WSOC concentrations in the fine and coarse fractions closely follow those of TC. However, the average WSOC/TC ratios are different, indicating that coarse carbonaceous particles have a lower WSOC content (Table 1b). The analysis of the chemical tracers indicates that biological particles contributed to the coarse fraction of the aerosol (see the following discussion). Large biological particles emitted by vegetation typically contain substances that are essentially water-unextractable, such as cell membranes and walls, epi-cuticular materials, as well as water-soluble compounds trapped inside intact cells (Graham et al., 2003). Therefore, the occurrence of large biological particles is a plausible explanation for the less soluble character of coarse carbonaceous particles collected at FNS. Conversely, WSOC dominate the composition of TC in fine particles from biomass burning sources. During the dry period, a limited but significant increase in the WSOC percentage was observed during day-time compared to the night. This difference can be attributed either to the different combustion stages occurring during the day compared to night (i.e., more frequent flaming than smoldering fires), or to the photochemical production of secondary organic compounds during day (Hoffer et al., 2005¹). Therefore, different sources of biomass burning aerosols and secondary processes are likely to affect the soluble/insoluble character of the fine particles in the dry period.

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3.2. OC speciation in the fine and coarse aerosol

Using the diverse analytical techniques for individual compound analysis, as described in the experimental section, we identified and quantified a series of polar organic compounds with 1 to 12 carbon atoms per molecule, comprising different chemical classes: mono-carboxylic, dicarboxylic and tricarboxylic acids, aromatic acids and aldehydes, sugars, sugar-alcohols and anhydrosugars. A summary of the results of WSOC speciation from the analysis of filter samples is given in Table 2, while a pie chart (Fig. 2) shows the relative concentrations of the different identified chemical classes determined in fine PM for the various periods of the campaign. The four sets of data available for the composition of submicron particles exhibit discrepancies in the concentrations of specific classes of compounds. However, since the statistics for each series are not based on the same number of samples, a general conclusion on the recovery of the different sampling systems and analytical methods cannot be provided here.

In general, the IC analysis provided higher concentrations of monocarboxylic, tricarboxylic and aromatic acids compared to the GC–MS methods. The GC–MS analysis following the UA methods (denoted by GC-MS_{UA}) provided higher values for the dicarboxylic acids and lower for the sugar-derivatives compared to the analysis performed with the MPIC method (denoted by GC-MS_{MPIC}). The recovery of low-molecular weight compounds, such as C₃–C₆ dicarboxylic acids (DCA) by GC-MS can be affected by the different extraction techniques employed. The levoglucosan concentrations obtained with the GC-MS_{UA} method are in agreement with the results of the IEC-UV method which does not require a derivatization step (Schkolnik et al., 2005). Therefore, the deviation between the two methods shown in the Table is merely due to the different sets of samples analyzed. Further, it is noted that the levoglucosan concentrations obtained with the GC-MS_{UA} method are in good agreement with those for the LBA-EUSTACH-2 campaign, which had been conducted at the same pasture site (Graham et al., 2002). A possible reason for the large discrepancies obtained for measurement of polar carboxylic acids between the two GC-MS methods is the different polarity of

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the extraction solvents used: the GC-MS_{UA} method uses methanol as an extraction solvent, and internal recovery standards that allow to correct for losses during sample workup, while the GC-MS_{MPI}C method uses acetonitrile (less polar than methanol) as extraction solvent and assumes a 100% recovery.

The data in Tables 2a and b can be directly compared with the GC-MS results from the 1999 LBA-EUSTACH-2 campaign (Graham et al., 2002). The coarse filters collected during the SMOCC campaign were subjected to a less comprehensive set of analyses than used in LBA-EUSTACH, and sugar-derivatives were not measured. In the case of the submicron particles, on the other hand, the combination of speciation methods in the present study, comprising both GC and LC techniques provided a more detailed picture of the organic composition. In particular, in addition to the di- and tri-carboxylic acids, benzoic and vanillic acids, and sugar-derivatives already identified and measured by Graham et al. (2002), the analysis of the LBA-SMOCC samples indicated higher concentrations of monocarboxylic acids and significant concentrations of dicarboxylic acids, hydroxy mono- and dicarboxylic acids, and 2-methyltetrols. The identified chemical compounds are classified as follows:

- C₁-C₃ monocarboxylic acids: acetic, formic, methanesulfonic, lactic, glyoxylic and glyceric acids. They represent 4 to 7% of total aliphatic acids. Monocarboxylic acids have multiple sources, including both biomass burning and biogenic emissions (Souza et al., 1999);
- C₂-C₉ aliphatic dicarboxylic acids: oxalic acid was found to be by far the most abundant carboxylic acid identified, followed by malonic, succinic and malic acid, and with a clear increase in the relative contribution of the latter species towards the end of the campaign. Oxalic acid can originate from multiple sources, both primary and secondary. It was recently found as the most abundant carboxylic acid in biomass burning smoke in South Africa (Gao et al., 2003). Conversely, malic acid shows a more pronounced biogenic origin and was found to accompany photo-oxidation products of isoprene (i.e., 2-methyltetrols) in the Amazon basin

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(Claeys et al., 2004a);

- Aliphatic tricarboxylic acids: they include citric and tricarballilic acids, and account for only 1 to 2% of the mass of total speciated organic compounds. They have been found previously (Graham et al., 2002);
- One-ring aromatic acids and aldehydes: the aromatic compounds include methoxylated species (vanillin, syringaldehyde and the corresponding acids) and hydroxy-benzoic acids. The methoxylated compounds are more abundant compared to the hydroxy-benzoic acids in the dry season, whereas the contrary holds starting from the transition period. The concentration of the aromatic compounds clearly decreases relatively to aliphatic acids from 1:5 in the dry period to 1:25 in the wet period. One-ring aromatic acids and aldehydes are primary compounds known to be produced by the combustion of lignins (Simoneit, 2002) and are, therefore, enriched in fresh biomass burning smoke;
- Sugars and sugar-derivatives: they comprise both pyrogenic (anhydrosugars; Graham et al., 2002) and biogenic (sugars and sugar-alcohols; Simoneit et al., 2004) compounds. Levoglucosan is the most abundant single compound identified in submicron aerosols during the whole campaign, including the wet period. However, the concentration of levoglucosan and the other anhydrosugars clearly decreases relative to the biogenic compounds (sugars and sugar-alcohols), from 10:1 in the dry to 9:10 in the wet period;
- 2-Methyltetrols: they were identified as major biogenic compounds in the Amazon basin, and were attributed to photo-oxidation of isoprene (Claeys et al., 2004a). Principal component analysis confirmed that the concentrations of 2-methyltetrols in the SMOCC filter samples do not correlate with those of pyrogenic compounds, such as levoglucosan and potassium sulfate Maenhaut et al., 2005, in preparation). Figure 2 also shows that 2-methyltetrols account for a very small fraction of the identified WSOC mass during the dry period, whereas they add up to 10% of

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the speciated mass in the wet period, following the decrease of the concentrations of the pyrogenic organic compounds.

The tracer analysis clearly indicates a more pronounced signature of the biogenic sources in the wet period compared to the previous periods of the campaign. Organic compounds of clearly pyrogenic origin account for 52, 35 and 21% of the total speciated compounds in the dry, transition and wet periods, respectively, while the corresponding fractions for the biogenic species are 8, 11 and 31% (Fig. 2). The relatively high concentrations of levoglucosan and other pyrogenic compounds in the samples from November suggest that the biomass burning activity was not totally suppressed at the beginning of the wet season. In that period, persistent fire activity could be detected in Rondônia and Mato Grosso in areas temporarily free from precipitations (www.master.iag.usp.br/queimadas/). Finally, the increase in the concentrations of aliphatic carboxylic acids compared to anhydrosugars and aromatic compounds towards the end of the campaign may reflect more pronounced secondary sources due to oxidation of either pyrogenic and biogenic VOCs. The enrichment of the aliphatic carboxylic acids compared to anhydrosugars is known to occur during the transport of biomass burning products (Gao et al., 2003). After the onset of wet conditions, the longer transport from distant sparse fires, as well as more pronounced in-cloud processing, would favor the production of secondary organic compounds, rendering a more chemically aged character to the aerosol OC. The same conclusion can be derived from the change in the size-segregated inorganic chemical composition from the dry to the wet period (Fuzzi et al., 2005²).

Table 2 also reports the sum of concentrations of identified organic compounds and the aerosol TC, OC and WSOC for the same periods. The recovery of the speciation methods is higher in the case of the GC-MS_{UA} analysis on the HVDS1 samples, where the identified compounds account for by 6 to 8% of TC, and 9 to 11% of WSOC on a carbon basis. It is clear that a large part of the organic matter was not amenable to GC-MS analysis because it could not be eluted or derivatised into stable products, and eventually eluded identification at the molecular level.

3.3. OC speciation in the size-segregated samples

Selected samples of 12-stage and 10-stage MOUDI impactors underwent OC speciation by GC-MS, IC and IEC-UV techniques, providing the size-distributions of the most important identified organic compounds in the three periods of the campaign (Falkovich et al., 2004; Claeys et al., in preparation, 2005³; Schkolnik et al., 2005). The resulting size-segregated organic composition in terms of identified chemical classes is plotted for samples representative from the dry, transition and wet periods in Fig. 3a and 3b. The concentrations of the speciated OC classes are expressed as percentages of the aerosol mass in each size bin. Figure 3a reports the size-segregated composition of the carboxylic acids determined by IC analysis. Mono-carboxylic acids, oxalic acid and C₃–C₉ dicarboxylic acids are the main contributors to the acidic organic fraction in all the size intervals, although aromatic compounds are also important in the dry and transition periods, but only for particles with a diameter lower than 1 μm. Conversely, aromatic acids were not detected in the sample from the wet period in all size intervals. In all periods, the carboxylic acids are less abundant in the finest size range, as well as in the coarse fraction. Their contribution to aerosol mass is also higher in the dry compared to the other two periods. The contribution of the 2-methyltetrols determined by GC-MS to PM (Fig. 3b) also shows a decrease toward the wet period, but only in the submicron fraction, whereas the opposite is true for the coarse fraction. Anhydrosugars are the most abundant compounds in all seasons, with levoglucosan as the main species in the submicron and supermicron fractions, respectively. Sugar-alcohols (arabitol and mannitol) contribute up to almost two thirds of the speciated polyols in the size intervals 1.8–10 μm in the dry period, but this fraction increases to 60–90% in the transition phase and to 80–100% in the wet period. Finally, the contribution of the

³Claeys, M., Kourtchev, I., Pashynska, V., Vas, G., Vermeylen, R., Cafmeyer, J., Chi, X., Artaxo, P., and Maenhaut, W.: Polar organic marker compounds in boundary layer aerosols during the LBA-SMOCC 2002 biomass burning experiment in Rondnia, Brazil: time trends, diurnal variations and size distributions, in preparation, 2005.

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2-methyltetrols to OC increases significantly in the wet period, but only in the submicron size intervals.

The data reported in Figures 3a and 3b show that the contribution of the identified classes of organic compounds to aerosol PM changes with the aerosol diameter, and is generally highest in the 0.4–4 μm range. Since TC was not determined systematically on MOUDI samples, a size-segregated carbon balance was not attempted. However, the available TC size-distributions indicate a relatively constant TC/PM ratio for submicrometer aerosols, which are essentially carbonaceous particles (Fuzzi et al., 2005²). Therefore, the fraction of TC speciated at the molecular level is also expected to be higher for particles with diameter larger than 0.4 μm than for those in the finest size range. Finally, it should be noted that, since submicron particles with diameter lower than 0.4 μm account for 30 to 40% of PM₂ (Fuzzi et al., 2005²), the recovery values relative to OC obtained on PM₂ and PM_{2.5} filter samples (see previous section) must be considered as averages between the low percentage of speciated OC in the finest particles and the high percentage of identified organic compounds in larger (>0.4 μm) submicron particles.

3.4. Apportionment of WSOC into main chemical classes

Compared to the methods for WSOC speciation discussed so far, the IC-UV technique presented in the experimental section is a chromatographic method for the separation of broad chemical classes of WSOC, which is not targeted to speciation at the molecular level. The neutral compounds (NC), mono-/di-acids (MDA) and the polyacids (PA) separated by IC-UV were identified mainly on the basis of their chromatographic behavior (i.e., the retention time). The quantitative analysis is based on the determination of the TOC on isolated fractions used to calibrate the UV detector. Therefore, the concentrations of NC, MDA and PA could be derived only as μg of carbon per cubic meter, as for total WSOC. Other than some improvements in the chromatographic conditions, the method is essentially the same as the one used for the analysis of PM_{2.5} samples during the LBA-EUSTACH campaign (Mayol-Bracero et al., 2002a).

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The NC, MDA and PA were detected by UV absorption at 260 nm in all samples analyzed, provided that the injected WSOC amount exceeded 500 ng C. Using calibration following the procedure described in the experimental section, the HPLC peak areas of the three main regions of the chromatogram provided the concentrations of NC, MDA and PA in the size range 0.05–3.5 μm (stages I to IV of the Berner impactor) for 37 samples from the dry period, 12 samples from the transition period and 5 samples from the wet period, whereas the concentrations in larger particles (3.5–10 μm) were often below the detection limit. Table 3 reports the statistics for the concentrations of NC, MDA and PA in size-segregated samples from the three periods and also separately for nocturnal and diurnal samples. The mean relative contributions of the three IC-UV classes to the total water-soluble carbon are also included. Average size-segregated distributions for the three chemical classes and for the WSOC fraction unresolved by the IC-UV method are shown in Fig. 4 for the three main periods of the campaign. A constant feature for all samples is the substantially lower relative concentration of PA in the coarse size range compared to the submicron size fraction. PA show a particularly low abundance in coarse particles from the wet period. MDA exhibit rather constant contributions to WSOC in the submicron particles, whereas their contribution to WSOC in the coarse particles is quite variable. Finally, the NC fraction of WSOC decreased steadily from the finest size range (0.05–0.14 μm) to the coarsest, although a mode in the range 1.2–3.5 μm was also observed in some samples from the transition and wet periods. Table 3 also highlights some systematic differences in the composition of the aerosol samples collected at night-time compared to day-time. In particular, the NC fraction is significantly higher during the night (often by a factor of two compared to the day) in all periods. Conversely, PA are relatively more abundant during day in the dry period, but differences in the other two seasons are less evident. MDA also show limited diel variations, tending towards enrichment during night (similar to NC), especially in the finest size fraction during the transition period. The high content of neutral compounds of nocturnal samples is clearly correlated with the higher nocturnal concentrations observed for levoglucosan (Schkolnik et al., 2004). Therefore, the diel

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variations in the IC–UV composition can be interpreted in terms of different combustion conditions (e.g., smoldering vs. flaming processes) active at different times of the day. The production of PA during the day can also be explained by heterogeneous chemical processes promoted by sunlight (Hoffer et al., 2005¹). Although the chemical classes separated by the IC–UV method are not unambiguous tracers of the WSOC sources, the variations in their relative concentrations can be interpreted on the basis of the correlation with other chemical constituents of the aerosol (e.g., K), which can be more readily associated with a particular source or source processes. A full treatment of this subject is given in a parallel paper (Fuzzi et al., 2005²).

The fraction of WSOC not accounted for by the sum of NC, MDA and PA is size-dependent, being around 0–15% for the smallest particles, and increasing to 70% in the coarsest size range. The proportion is lower in the dry period, compared to the transition and especially the wet period, during which 50% of WSOC could not be accounted for even in the accumulation mode size range. The organic materials that can contribute to WSOC, especially in the wet period, and not accounted for by the IC–UV analysis could be either semi-volatile compounds that were lost during the calibration procedure, or hydrophilic colloids (very high molecular weight humic-like particles, fragments of biological membranes and other cell constituents) that are extracted from the impactor foils but are not elutable from the HPLC column.

4. Discussion

4.1. Speciated WSOC and chemical classes

The analytical techniques described in the experimental section can be grouped into methods for individual compound analysis (GC–MS, IC and IEC–UV), and methods targeted to the separation of main chemical classes (IC–UV). The former allow the identification and measurement of six categories of polar organic compounds, but leave uncharacterized a substantial fraction of OC. Conversely, the IC–UV technique provides

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a high recovery in the measurement of the water-soluble fraction of OC by fractionation into three main categories, whose specific chemical compositions, however, rests poorly defined. Therefore, in general, the IC–UV chromatographic classes do not correspond directly to any of the categories of polyols and carboxylic acids accounted for by individual compound analyses, but are more likely to include them.

To better understand the chemical nature of the “neutral compounds”, “mono-/di-acids” and “polyacids”, the WSOC speciated by GC–MS, IC and IEC in size-segregated samples can be classified according to their number of carboxylic groups per molecule; the estimation of their contribution to each of the three IC–UV classes is then straightforward. However, the concentrations of the identified WSOC were obtained on MOUDI samples, while those of total WSOC and of NC, MDA and PA were available only for the Berner impactor, making the comparison difficult owing to the different size resolutions and sampling efficiencies of the two impactors. To make the two sets of data comparable, we first defined a sub-set of four MOUDI samples analyzed by IC and IEC for their organic and inorganic composition, and collected approximately in parallel with Berner impactor samples. Then, the MOUDI size-distributions for the identified WSOC and inorganic compounds were converted to continuous size-distributions according to an inversion algorithm, using the MICRON inversion program (Wolfenbarger and Seinfeld, 1990), which takes into account the collection efficiency and species concentration in each MOUDI stage. A more detailed description of the use of the program is given by Havránek et al. (1996). The concentrations of the chemical species measured in MOUDI samples could then be retrieved for the size intervals of the Berner impactor by integrating the continuous size distributions between the Berner cut-offs. To take into account the different sampling efficiencies of the two impactors, the concentrations of organic species were normalized to those of sulfate, which had been measured on both sets of impactor samples. The identified WSOC from MOUDI were then expressed as $\mu\text{gC m}^{-3}$ and compared to the concentrations of total WSOC and of the IC-UV classes from the Berner impactor.

Table 4 shows the resulting average carbon budget for the dry period ($n=2$) and for

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the transition phase ($n=2$). Levoglucosan accounts for 1 to 12% of NC in the fine fraction, and for an even higher percentage in the size range 1.2–3.5 μm . Similarly, the identified mono- and di-carboxylic acids account for 6 to 16% of total MDA in the fine fraction, and 9–18% in coarse particles. In contrast, the contribution of the identified tricarboxylic acids (citric and tricarballylic) to total PA is very low, usually less than 1%. In conclusion, the speciated WSOC represent around 1.5% of the total water-soluble carbon in the 0.05–0.14 μm size interval, and up to 10% in the 0.42–3.5 μm size range. The identified WSOC are mostly NC and MDA, while PA remains essentially uncharacterized at the molecular level. This finding is in agreement with the hypothesis that PA consist mostly of medium-to-high molecular weight compounds (HULIS). These results for the identified fractions of the three IC–UV classes analyzed in the impactor samples are in agreement with analogous data obtained on comparable PM_{2.5} samples collected during the 1999 LBA-EUSTACH-2 experiment (Mayol-Bracero et al., 2002a).

4.2. A synthetic representation of the WSOC composition by model compounds

In the previous section, chemical characterization of the IC–UV chromatographic classes was attempted in terms of organic compounds identified at the molecular level, showing that large fractions of NC, MDA and especially PA could not be speciated using this approach. An alternative characterization method is the functional group analysis performed by ¹HNMR spectroscopy on the water-extracts of filter and impactor samples, as well as on chromatographic fractions separated on IC preparative columns (Tagliavini et al., submitted, 2005⁴). ¹HNMR analysis provides the functional group composition of the mixture, but little information on the speciation into individual compounds. The functional groups containing hydrogens directly bound to carbon atoms are determined directly, while COOH groups are assessed after derivatization

⁴Tagliavini, E., Moretti, F., Decesari, S., Fuzzi, S., and Maenhaut, W.: Organic functional groups approach to the characterization of WSOC from biomass burning aerosol samples: H NMR and chemical derivatization outcomes, submitted to Atmos. Chem. Phys. Discuss., 2005.

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into methyl esters. A fully detailed procedure is reported in a companion paper, together with a discussion of the results found (Tagliavini et al., 2005⁴). As a brief survey, we outline the following points: 1) significant differences were found among the three periods with respect of the composition of fine particles, with samples from the wet period exhibiting a lower content of aromatic groups and more unsaturated and hydroxylated aliphatic moieties; 2) the coarse fraction of the aerosol from the transition and wet periods, on the other hand, is dominated by oxygenated carbon atoms, indicating the occurrence of poly-hydroxy-compounds; 3) carboxylic groups (COOH) account for 13.3%, 19.7% and 18.4% of the water-soluble carbon accounted for by NMR analysis in fine aerosol samples collected during the dry, transition and wet periods, respectively; 4) when applied to size-segregated samples, the functional group approach provides significantly different compositions in the size range 0.05–0.14 μm compared to the 0.42–1.2 μm range, the formers containing much more alkylic and less oxygenated moieties; 5) the picture derived from IC separated fractions is consistent with what previously observed, i.e., that the neutral fraction is dominated by polyols (levoglucosan, mannitol, and many others), but still contains polyhydroxy and polyalkoxy benzenes, the mono-/di-acids fraction can be seen as a mixture of hydroxylated alkyl and benzoic monocarboxylic and dicarboxylic acids, the polyacidic fraction exhibits a humic-like character.

Since the functional group composition provided by ¹HNMR refers to the mixture of WSOC in the sample, the functional groups of the individual compounds identified by GC–MS, IC and IEC are part of that composition. The contribution of the speciated compounds to the ¹HNMR composition can be computed on the basis of their concentrations and molecular structure. Figure 5 shows the functional group composition of WSOC determined on two Berner impactor samples collected under extremely polluted conditions during the dry period, and on HVDS fine aerosol samples representative of all three periods of the campaign. The functional group compositions of the IC–UV classes isolated from the PM_{2.5} HVDS sample collected on 25–26 September is also reported. The concentrations of each functionality accounted for by the speciated or-

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ganic compounds was calculated on the basis of Table 4 for size-segregated data and GC–MS analysis of HVDS1 filters for the PM_{2.5} samples. Levoglucosan, malic acid and vanillic acid were used as proxies for the functional group composition of polyhydroxylated compounds, aliphatic carboxylic acids and aromatic acids, respectively.

5 The difference between the functional group composition of the samples analyzed by ¹HNMR and that attributable to the mixture of speciated compounds provides the gross composition of the organic fraction which the WSOC speciation methods failed to characterize. It turns out that COOH and H-C-O groups are the functional groups for which the individual compound analysis provides the best recoveries. Conversely, there is a
10 large amount of water-soluble carbon occurring as aliphatic and aromatic C-H groups that cannot be explained by the available set of speciated organic compounds.

The general chemical characteristics observed by ¹HNMR for the WSOC fraction that escaped the GC–MS and IC methods can be helpful in addressing further improvements in analytical techniques for WSOC speciation. However, the available functional
15 group compositions for the unidentified WSOC can already be utilized to provide at least a set of “model compounds”. Following the definition given by Fuzzi et al. (2001), the model compounds are hypothetical compounds that reproduce the observed functional group composition and other chemical properties of a mixture (or a fraction) of WSOC. The molecular formulae of the model compounds do not follow unequivocally
20 from the functional group composition, but they are constrained by it. Using this approach the chemical composition of a mixture or a fraction of WSOC is represented in terms of a limited number of compounds, when speciation of organic compounds is not feasible, but a consistent pool of structural data and other information on the chemical properties of the mixture is available instead.

25 The molecular structure of the model compounds and their relative concentrations were chosen in order to match as much as possible the chemical composition of the samples in Fig. 5 with respect to the functional groups (from ¹HNMR characterization) and main chemical classes (NC, MDA and PA from IC–UV analysis). The model compounds used in this study are shown in Fig. 6, and their contributions (as mass

percentage) to WSOC in the different samples representative for the three periods of the campaign are reported in Table 5.

The model compositions in the table describe the WSOC fraction determined by the IC–UV method, with an explicit representation of the WSOC classes speciated at the molecular level.

In order to differentiate the contributions of the WSOC identified by the GC-MS, IC and IEC methods, they were lumped into three classes, which were in turn represented by three model compounds:

- “Levogluconan”, accounting for all the speciated polyhydroxylated compounds (anhydrosugars, sugar-alcohols, saccharides, 2-methyltetrols);
- “Malic acid”, for the identified C₁–C₉ aliphatic mono- and di-carboxylic acids.
- “Vanillic acid” for the identified aromatic acids.

No specific class for the identified tricarboxylic acids was introduced because of their very low contribution to PA.

The WSOC fraction represented by the model compounds “levogluconan”, “malic acid” and “vanillic acid” was derived from the average carbon balances for the dry and the transition periods reported in Table 4. In contrast, the WSOC fractions accounted for by “levogluconan” and “malic acid” in the wet period could only be extrapolated from the average data for the transition period, while the contribution of “vanillic acid” was set to zero. This is qualitatively in agreement with Fig. 3 showing that low-molecular weight aromatic compounds were not detected in the wet period, and that, despite of the decrease of anhydrosugars, the content of polyhydroxylated compounds in the fine fraction of PM decreased only slightly from the transition to the wet period, due to the simultaneous increase of the 2-methyltetrol concentrations.

The fractions of NC, MDA and PA that could not be attributed to the chemical classes speciated at the molecular level were represented in our chemical model by the smallest number of model compounds necessary to explain the observed variability in the functional group composition of the samples.

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The unidentified NC fraction was apportioned to two model compounds: “cellobiosan” (Gao et al., 2003) and compound (1), a compound close to matairesinol, a plant phenolic compound that had also been found in biomass smoke (Simoneit et al., 2003). The former was introduced to account for the unidentified polyhydroxylated compounds, while the latter reproduces the aromatic and H-C-C=groups of the NC. It is noted that the resulting model NC composition is lacking the saturated aliphatic moieties observed in the samples, for which we could not find a suitable candidate molecule.

The closure of the functional group balance of MDA after subtracting the contribution of the speciated compounds was attempted by introducing: 1) a C₆ aliphatic di-carboxylic acid, namely adipic acid, to account for the excess COOH groups determined by the derivatization-NMR technique; 2) a methoxylated C₁₁ aromatic acid (syringyl-propionic acid; Nolte et al., 2001) representing the unidentified aromatic species; 3) a hypothetical C₁₉ aliphatic mono-carboxylic acid, named simply “compound (2)”, which reproduces the abundance of saturated aliphatic moieties determined by ¹HNMR, especially in the 0.05–0.14 and 3.5–10 μm size intervals of the nocturnal sample from the dry season. Acyclic compounds like compound (2) have already been reported for biomass smoke particles (Simoneit et al., 2003), but prevalently in the water-insoluble fraction. Compound (2) must exhibit a limited solubility in water, probably higher than that of cortisone, a compound with similar structure but with no carboxylic groups and a water solubility of 0.28 g/kg H₂O. This solubility can be sufficient for the extraction of compound (2)-like WSOC, for which a minimum amount of water of 15 g per 0.1 mg C of sample was always used. Therefore, we propose that a fraction of organic compounds of medium polarity were efficiently extracted and quantified in the WSOC mixture. For simplicity and because of the higher availability of standards, such organic compounds of medium polarity are represented to contribute only to the MDA fraction, although this fraction had to be artificially augmented relative to NC and PA in the Aitken mode, and in the coarse fraction of the nocturnal sample collected in the dry period.

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The polyacidic fraction is represented in all samples by the same single species, compound (3), a fulvic-like substance. This is a hypothetical compound, whose structure resembles that of the Suwannee River fulvic acid (Averett et al., 1989), but it is less aromatic while showing more saturated aliphatic moieties. Therefore, the choice of the Suwannee River fulvic reference material to reproduce the chemical properties of the PA fraction in laboratory experiments cannot be considered as optimal. Nonetheless this standard is still preferable to the other commercial standards of humic substances or of other high-molecular weight water-soluble substances.

The estimation of an average molecular weight for the model compounds representing the (large) fraction of WSOC not speciated at the molecular level is probably the largest source of uncertainty in our chemical model. Unfortunately, the functional group analysis by ¹HNMR does not provide any information on the molecular weight. We can assume that the derivatization-GC-MS and ion chromatography methods were rather efficient in the determination of the low-molecular weight fraction of WSOC. Consequently, the model compounds introduced to represent the uncharacterized fraction must include medium-high molecular weight species. In our model, the compounds having a molecular weight higher than 300 Da account for 60–70% of WSOC in the fine fraction, consistent with the results of EGA analysis which indicates a fraction of 60–80% on the basis of the thermal evolution behavior. The maximum molecular weight attributable to the model compounds reproducing the more refractory fraction of WSOC cannot be determined accurately. However, it cannot be very high for the less polar compounds, like compounds (1) and (2), whose homologues with 30 carbon atoms would probably be insoluble even in the great excess of water used for extraction of the samples. Conversely, the polyhydroxylated species with more than six carbon atoms, represented in the model by cellobiosan, can be perfectly soluble even when occurring in very large polymeric forms. Although polyhydroxylated compounds larger than cellobiosan have not been found in samples of ambient aerosol yet, such polymers may form in wood smoke (Kawamoto et al., 2003), and therefore the molecular weight of the WSOC fraction represented by the model compound “cellobiosan” re-

mains largely uncertain.

Figure 5 shows that a good fit between the observed functional group composition and that of the model was obtained for fine aerosol samples from the dry and the transition period, for the size-segregated composition of sample 25–26 September in the 0.42–1.42 μm range (accounting for most of the PM), and for the internal composition of the MDA and PA fractions. In contrast, the representation of the composition of the NC fraction, of the coarse and fine aerosol samples from the wet period is not as good. In some cases, as for the wet period sample, the WSOC fraction accounted for by the IC–UV method could not account for the total functional groups estimated by $^1\text{HNMR}$. Therefore, only a relative functional group composition could be reproduced.

The list of model compounds representative of the WSOC composition in the aerosol samples collected during the SMOCC campaign is more complex than that presented in the previous work by Fuzzi et al. (2001), based on samples from the Po Valley. In contrast to this first study, a more detailed picture of the low-molecular weight WSOC emerged from the GC-MS and IC analyses performed during the LBA-SMOCC experiment. The identified species were represented explicitly in the chemical model, i.e., by introducing specific model compounds to account for them (levoglucosan, malic acid and vanillic acid). They constitute a subset of model compounds known with a higher confidence, whereas the remaining unidentified compounds are more susceptible to modifications following different approaches in defining the chemical classes, and improvements in the characterization of un-specified WSOC. For this reason, we believe that the current representation is more flexible and provides a better integration of the information from the individual compound analysis with the results from the functional group characterization.

5. Conclusions

During the LBA-SMOCC field campaign, biomass burning emissions frequently caused episodes of very high concentrations of submicron carbonaceous aerosol (submicron

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TC concentrations $>50 \mu\text{gC m}^{-3}$) in the period 9 September–8 October 2002, while at the beginning of November, after the end of the intense burning period, the measured submicron TC concentrations were 20 times lower. At the same time, biogenic sources produced a constant background of aerosol throughout the campaign, which was found predominantly in the coarse mass fraction. Fine and coarse aerosol TC differ also with respect to their water-soluble fraction, which is 64% on average in the first case and only 34% in the latter. The EC accounted for 12% on average of TC in the fine fraction of the aerosol. A unique combination of analytical techniques for individual compound analysis was employed to speciate the aerosol organic compounds, resulting in the apportionment of up to 8% of the submicron TC (ca. 11% of WSOC). Carboxylic acids and poly-hydroxylated compounds, comprising both pyrogenic and biogenic species, were the main classes of compounds speciated. Pyrogenic compounds include anhydrosugars, aromatic acids and aldehydes, while biogenic species include sugars, sugar-alcohols, 2-methyltetrols and malic acid. The ratio between total identified pyrogenic and biogenic compounds changed from 6:1 in the dry period to 2:3 in the wet period, showing that biomass burning was still active at the end of the field campaign.

Three main chemical classes of WSOC isolated by the IC-UV method (i.e., neutral compounds, mono-/di-acids and polyacids) were detected in size-segregated aerosol samples throughout the campaign, although the concentrations of polyacids were low in the coarse fraction, especially at the onset of the wet period. The speciated polyhydroxylated compounds and low-molecular weight carboxylic acids contributed 1–20% of both, neutral compounds and mono-/di-acids. Conversely, the polyacidic fraction remained almost entirely uncharacterized at the molecular level. An insight into the unresolved fraction of WSOC was provided by ^1H NMR functional group analysis, showing that most of the less polar compounds, characterized by extended saturated aliphatic and aromatic moieties, escaped molecular speciation.

The size-segregated composition of WSOC was summarized by a set of model compounds, which reproduce both the composition of the identified organic compounds and the functional groups of the whole WSOC mixture. The model compounds retain

the average structural information on WSOC derived by the chemical characterization. It is remarkable that despite the significant change in the relative strength of pyrogenic and biogenic sources from the dry to the wet period, the main chemical classes and functional groups of WSOC (and consequently the sets of model compounds) change only to a limited extent. This must be attributed to the fact that the decrease in the hydroxylated compounds and carboxylic acids of pyrogenic origin toward the end of the campaign was compensated by the increased contribution of analogous classes of compounds of biogenic nature. In this way, the pyrogenic and biogenic emissions provided a rather constant pool of very polar organic compounds in the submicron fraction of the aerosol throughout the dry-to-wet season transition. One consequence of this fact is the surprisingly similar cloud droplet nucleating ability (CCN ability) of the pyrogenic and biogenic aerosols over Amazonia (Andreae et al., 2004).

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Table 1. (a) EC/TC for the fine fraction of the aerosol and (b) WSOC/TC ratios for the fine and coarse fractions in the different periods of the LBA-SMOCC campaign.

a) EC/TC

period	fine		
	N/D	N	D
dry	0.14 (0.04)	0.15 (0.04)	0.11 (0.03)
trans.	0.12 (0.03)	0.13 (0.02)	0.09 (0.03)
wet	0.05 (0.03)	0.06 (0.03)	0.06 (0.01)
all samples	0.12 (0.04)		

b) WSOC/TC

period	fine			coarse		
	N/D	N	D	N/D	N	D
dry	0.64 (0.08)	0.61 (0.06)	0.67 (0.08)	0.37 (0.13)	0.35 (0.13)	0.37 (0.13)
trans.	0.69 (0.09)	0.66 (0.03)	0.68 (0.12)	0.31 (0.07)	0.31 (0.02)	0.28 (0.05)
wet	0.53 (0.11)	0.54 (0.16)	0.46 (0.06)	0.28 (0.12)	0.33 (0.14)	0.21 (0.04)
all samples	0.64 (0.09)			0.34 (0.12)		

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Table 2. Concentrations of the identified organic compounds (in ng m^{-3}) in fine and coarse filter samples, following GC–MS, IC and IEC analysis. Average values and range of variation are reported for the dry (Table 2a), transition (Table 2b) and wet (Table 2c) periods. Total identified organic compounds, WSOC, OC and TC data are reported as $\mu\text{g m}^{-3}$. MCA: aliphatic mono-carboxylic acids; DCA: aliphatic dicarboxylic acids; TCA aliphatic tricarboxylic acids. TC determinations on HVDS2 samples were performed by evolved gas analysis (Hoffer et al., submitted, 2005¹).

(a) Dry period	Fine HVDS1 GC-MS _{UA} (N=51)	HVDS2 GC-MS _{MPIC} (N=17)	HVDS3 IC+IEC (N=16)	SFU IC (N=19)	Coarse SFU IC (N=19)
MCA			529 (106–1310)	139 (22.9–413)	80.3 (6.1–264)
glyceric acid	60.0 (18.6–142)	64.5 (25.5–109)			
formic acid			122 (50–262)	88.5 (12.1–370)	17.2 (2.5–43.8)
MSA			11.3 (4.4–16.4)	25.2 (0.0–213)	32.0 (0.0–465)
glyoxylic acid			71.0 (17.3–161)	40.5 (0.0–115)	0.80 (0.0–15.1)
Oxalic acid			1097 (364–2192)	516 (92.2–932)	162 (19.2–305)
DCA C₃–C₉			778 (150–1332)	566 (139–1026)	19.7 (0.0–106)
malonic acid		170 (25.9–389)	113 (49.4–255)	53.7 (0.0–210)	
methylmalonic acid		6.4 (3.3–10.3)			
succinic acid	554 (105–1860)	166 (68.8–290)			
methylsuccinic acid		20.4 (8.0–38.7)			
maleic acid		23.9 (7.8–83.5)	22.9 (9.2–46.8)	9.8 (0.0–29.3)	
fumaric acid	59.8 (18.9–232)	11.9 (5.5–19.5)	7.9 (2.0–19.3)	5.2 (0.0–20.3)	
malic acid	334 (128–721)	188 (107–254)			
glutaric acid	48.3 (7.6–157)	26.8 (12.2–44.8)			
α -hydroxyglutaric acid	156 (51.9–377)				
β -hydroxyglutaric acid	39.3 (13.6–93.0)				
2-ketoglutaric acid		35.3 (0.0–56.6)			
threonic acid (+ isomer)	70.2 (20.4–173)				
tartaric acid	42.8 (15.6–87.1)				
adipic acid		8.8 (4.2–18.7)			
pimelic acid		5.5 (3.1–10.5)			
azelaic acid			9.0 (0.0–27.5)		

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Table 2. Continued.

TCA				
citric acid			48.1 (18.1–84.7)	48.4 (0.0–114)
tricarballic acid		25.6 (16.7–40.6)	50.7 (5.7–12.8)	42.8 (0.0–89.3)
Aromatic acids				
phtalic acid		33.9 (18.0–63.6)	19.6 (0.0–49.2)	
isophtalic acid		4.1 (1.4–8.2)		
2-hydroxybenzoic acid	4.7 (0.4–17.1)			
3-hydroxybenzoic acid	2.5 (0.2–12.2)	11.8 (3.1–25.1)	74.0 (23.5–163)	78.2 (0.0–183)
4-hydroxybenzoic acid	28.0 (1.9–98.5)	54.4 (16.7–138)		
3,4-dihydroxy benzoic acid		57.0 (25.8–121)		
vanillic acids	68.2 (4.1–214)	60.2 (13.8–151)	83.4 (23.1–155)	16.8 (0.0–67.3)
syringic acid		83.0 (7.6–216)	97.0 (27.8–304)	86.4 (0.0–225)
Aromatic aldehydes				
4-hydroxybenzaldehyde		5.1 (0.7–17.8)		
vanillin		11.4 (1.4–30.7)		
syringaldehyde		74.3 (6.9–240)		
Other aromatics				
4-methylbenzocatechin		5.8 (1.2–18.5)		
Anhydrosugars				
galactosan	49.3 (6.5–219)	89.4 (18.3–215)		
mannosan	128 (19.9–456)	168 (37.8–384)		
levoglucosan	1868 (239–6291)	4115 (849–8795)	1163 (82–5797)	
anhydroglucofuranose	82.2 (13.7–366)			
Sugar-alcohols				
glycerol		15.9 (7.3–39.8)		
erythritol		57.5 (20.6–172.7)		
threitol		13.2 (1.6–26.7)		
arabitol	14.0 (0.0–34.8)	34.4 (10.0–63.9)		
mannitol	18.4 (3.2–44.0)	27.8 (8.3–49.6)		
sorbitol		1.1 (0.4–2.1)		
inositol		1.1 (0.0–2.3)		

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Table 2. Continued.

Sugars					
fructose		23.0 (12.5–49.3)			
sucrose		5.4 (1.4–12.2)			
Methyl-tetrols					
2-methyl-threitol	45.1 (7.3–82.6)				
2-methyl-erythritol	124 (22.4–267)				
Total identified	3.8 (0.94–11.14)	6.93 (1.92–14.10)	3.01 (0.86–5.66)	1.68 (0.36–3.69)	0.26 (0.08–0.55)
WSOC	17.2 (3.7–44.2)	37.1 (21.1–61.4)	20.7 (8.4–44.2)		
OC	26.6 (5.1–64.1)		31.3 (12.8–62.1)		
TC	27.5 (5.4–65.7)	57.5 (27.7–101.2)	32.4 (13.3–64.1)		

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Table 2. Continued.

(b) Transition	Fine HVDS1 GC-MS _{UA} (N=20)	HVDS2 GC-MS _{MPIC} (N=13)	HVDS3 IC + IEC (N= 5)	Coarse SFU IC (N= 8)	SFU IC (N= 8)
MCA			146.1 (100–191)	27.8 (8.4–46.0)	10.6 (0.3–25.7)
glyceric acid	27.2 (13.7–40.0)	15.3 (7.7–21.5)			
formic acid			28.9 (16.9–40.8)	17.0 (3.7–31.5)	2.0 (0.1–4.3)
MSA			6.1 (2.9–10.4)	3.4 (0.0–11.1)	
glyoxylic acid			22.3 (17.4–28.2)	10.9 (0.0–24.4)	
Oxalic acid			386.5 (309–436)	177 (65.9–265)	55.5 (8.4–123)
DCA C₃-C₉			306.7 (229–412)	295 (63.6–514)	
malonic acid		35.1 (3.3–64.7)	42.9 (26.8–53.0)	14.4 (0.0–43.8)	
methylmalonic acid		1.2 (0.5–2.1)			
succinic acid	232 (84.0–512)	24.8 (9.4–52.6)			
methylsuccinic acid		2.6 (1.0–5.9)			
maleic acid		5.2 (1.1–21.4)	16.3 (8.3–25.4)	7.3 (0.0–15.8)	
fumaric acid	30.7 (11.2–53.0)	3.0 (1.3–5.6)	4.7 (3.3–7.6)	3.5 (0.0–7.7)	
malic acid	225 (137–347)	77.6 (43.7–116)			
glutaric acid	18.2 (7.0–36.5)	3.9 (1.4–8.1)			
α -hydroxyglutaric acid	65.3 (31.5–112)				
β -hydroxyglutaric acid	22.4 (11.0–38.4)				
2-ketoglutaric acid		9.3 (3.2–26.0)			
threonic acid (+ isomer)	35.2 (19.4–50.9)				
tartaric acid	40.3 (21.4–65.8)				
adipic acid		1.5 (0.6–3.2)			
pimelic acid		0.8 (0.5–1.4)			
azelaic acid					
TCA					
citric acid			18.8 (13.2–30.1)	22.9 (7.3–44.9)	
tricarballic acid		12.6 (6.6–20.8)	14.9 (12.8–18.6)	16.0 (4.9–31.7)	

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Table 2. Continued.

Aromatic acids				
phtalic acid			3.6 (0.0–6.0)	
isophtalic acid		1.0 (0.4–1.7)		
2-hydroxybenzoic acid	0.6 (0.1–1.9)			
3-hydroxybenzoic acid	0.3 (0.0–1.3)	0.9 (0.2–2.6)	49.3 (36.3–76.9)	38.2 (6.1–79.5)
4-hydroxybenzoic acid	4.1 (0.6–12.1)	5.1 (0.9–15.0)		
3,4-dihydroxy benzoic acid		8.9 (1.7–19.8)		
vanillic acids	10.8 (1.6–32.9)	3.6 (0.6–10.6)	8.8 (0.0–23.2)	12.6 (0.0–24.0)
syringic acid		7.0 (0.4–18.5)	27.4 (22.5–38.2)	39.0 (0.0–77.1)
Aromatic aldehydes				
4-hydroxybenzaldehyde		0.5 (0.1–1.8)		
vanillin		1.1 (0.2–3.2)		
syringaldehyde		4.1 (0.8–13.3)		
Other aromatics				
4-methylbenzocatechin		0.4 (0.1–0.9)		
Anhydrosugars				
galactosan	9.6 (2.3–27.1)	10.0 (1.6–25.3)		
mannosan	25.7 (6.1–58.5)	18.7 (3.7–43.5)		
levoglucosan	335 (88.7–721)	473 (100–994)	228 (29–542)	
anhydroglucofuranose	18.9 (5.7–35.2)			
Sugar-alcohols				
glycerol		3.7 (1.5–6.2)		
erythritol		8.4 (3.5–17.0)		
threitol		1.6 (0.3–3.0)		
arabitol	8.3 (3.8–18.6)	13.3 (5.8–27.2)		
mannitol	17.0 (8.0–39.3)	20.1 (4.2–51.5)		
sorbitol		0.9 (0.2–2.3)		
inositol		0.4 (0.0–0.8)		
Sugars				
fructose		6.5 (2.1–10.1)		
sucrose		1.8 (0.9–5.1)		

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Table 2. Continued.

Methyl-tetrols					
2-methyl-threitol	21.9 (6.4–74.9)				
2-methyl-erythritol	78.9 (18.8–262)				
Total identified	1.2 (0.63–1.95)	0.96 (0.37–1.79)	1.01 (0.77–1.20)	0.63(0.06–1.25)	0.06 (0.01–0.15)
WSOC	5.2 (1.3–8.5)	8.0 (5.7–11.3)	6.5 (4.9–8.2)		
OC	7.2 (2.4–12.8)		9.3 (6.7–12.1)		
TC	7.6 (2.6–13.3)	10.5 (6.0–16.6)	9.7 (7.1–12.5)		

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Table 2. Continued.

(c) Wet period	Fine HVDS1 GC-MS _{UA} (N=7)	HVDS2 GC-MS _{MPIC} (N=7)	HVDS3 IC+IEC (N=0)	Coarse SFU IC (N=7)	SFU IC (N=7)
MCA				5.2 (2.5–6.7)	11.2 (4.6–20.8)
glyceric acid	3.4 (1.1–5.3)	1.7 (0.6–2.8)			
formic acid				9.4 (6.6–18.0)	0.4 (0.0–1.3)
MSA					
glyoxylic acid					
Oxalic acid				36.7 (23–51.6)	16.0 (7.7–28.3)
DCA C₃–C₉				33.9 (14.7–61.0)	4.0 (0.0–11.4)
malonic acid		5.6 (2.0–7.9)		5.2 (0.0–8.1)	
methylmalonic acid		0.2 (0.1–0.5)			
succinic acid	24.0 (7.1–41.0)	2.8 (1.3–4.2)			
methylsuccinic acid		0.3 (0.1–0.6)			
maleic acid		1.0 (0.3–3.0)			
fumaric acid	6.0 (3.3–10.3)	0.6 (0.3–1.0)			
malic acid	64.5 (33.2–94.6)	17.2 (9.2–27.4)			
glutaric acid	3.7 (0.0–9.8)	0.5 (0.2–0.9)			
α -hydroxyglutaric acid	7.3 (3.2–15.1)				
β -hydroxyglutaric acid	3.9 (1.7–6.0)				
2-ketoglutaric acid		0.9 (0.0–1.5)			
threonic acid (+ isomer)	5.6 (3.0–9.0)				
tartaric acid	12.3 (5.0–20.3)				
adipic acid		0.4 (0.2–0.6)			
pimelic acid		0.2 (0.1–0.4)			
azelaic acid					
TCA					
citric acid					
tricarballic acid		3.0 (1.4–5.9)			

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Table 2. Continued.

Aromatic acids			
phtalic acid		1.7 (0.5–2.6)	
isophthalic acid		0.1 (0.0–0.5)	
2-hydroxybenzoic acid	0.05 (0.03–0.12)		
3-hydroxybenzoic acid	0.02 (0.00–0.08)	0.1 (0.0–0.3)	
4-hydroxybenzoic acid	0.22 (0.05–0.54)	0.0 (0.0–0.1)	
3,4-dihydroxy benzoic acid		0.5 (0.1–1.1)	
vanillic acid	0.9 (0.2–2.9)	0.3 (0.1–0.9)	
isovanillic acid			
syringic acid		0.4 (0.0–1.2)	
Aromatic aldehydes			
4-hydroxybenzaldehyde		0.0 (0.0–0.1)	
vanillin		0.2 (0.0–0.9)	
syringaldehyde		0.1 (0.0–0.4)	
Other aromatics			
4-methylbenzcatechin		0.1 (0.0–0.2)	
Anhydrosugars			
galactosan	0.9 (0.0–2.0)	0.7 (0.2–1.2)	
mannosan	3.5 (1.6–6.6)	1.5 (0.4–2.8)	
levoglucosan	50.2 (13.2–134)	39.8 (10.7–82.7)	39.6 (0–79.3)
anhydroglucofuranose	2.0 (0.9–4.1)		
Sugar-alcohols			
glycerol		1.5 (0.9–2.7)	
erythritol		2.5 (0.6–3.7)	
threitol		0.2 (0.1–0.4)	
arabitol	7.5 (4.5–10.0)	11.6 (6.5–15.1)	
mannitol	15.1 (9.4–17.8)	26.2 (13.5–35.1)	
sorbitol		1.2 (0.8–1.6)	
inositol		0.1 (0.1–0.2)	
Sugars			
fructose		4.4 (2.8–6.6)	
sucrose		1.0 (0.4–2.7)	

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Table 2. Continued.

Methyl-tetrols				
2-methyl-threitol	7.8 (2.6–17.1)			
2-methyl-erythritol	29.2 (11.1–55.1)			
Total identified	0.25 (0.15–0.41)	0.16 (0.08–0.22)	0.09 (0.05–0.13)	0.03 (0.01–0.04)
WSOC	0.72 (0.55–1.71)	1.6 (0.95–2.0)		
OC	1.2 (0.73–2.1)			
TC	1.3 (0.76–2.3)	1.9 (1.0–2.4)		

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Table 3. Concentrations of WSOC, neutral compounds (NC), mono-/di-acids (MDA) and polyacids (PA) determined by the IC–UV method for the Berner impactor samples from the dry (Table 3a), transition (Table 3b) and wet (Table 3c) periods. Mean values, medians, standard deviations and range of variations are reported for the whole set of samples for each period (N/D), and specifically for the samples collected at night (N) and during day-time (D). The concentrations are expressed as $\mu\text{gC m}^{-3}$. The cut-offs of the five size intervals of the Berner impactor are expressed in μm .

a) Dry period

	WSOC				NC					MDA					PA					
	mean	median	5 perc	95 perc	mean	median	5 perc	95 perc	%WSOC	mean	median	5 perc	95 perc	%WSOC	mean	median	5 perc	95 perc	%WSOC	
N/D (n = 37)																				
0.05 - 0.14	1.35	1.07	0.31	2.68	0.40	0.23	0.05	1.16	28%	0.46	0.37	0.09	0.93	35%	0.38	0.33	0.09	0.76	31%	
0.14 - 0.42	3.59	3.18	1.09	7.62	0.99	0.63	0.19	2.94	26%	1.08	0.84	0.30	2.58	30%	1.08	0.78	0.30	2.74	30%	
0.42 - 1.2	8.96	6.27	2.19	19.49	2.18	1.11	0.28	6.51	21%	2.97	2.09	0.61	6.83	32%	2.72	2.13	0.61	5.30	31%	
1.2 - 3.5	0.64	0.51	0.21	1.22	0.12	0.06	0.01	0.33	15%	0.18	0.13	0.06	0.34	28%	0.10	0.06	0.01	0.25	14%	
3.5 - 10	0.29	0.23	0.10	0.59	0.04	0.03	0.01	0.08	9%	0.09	0.07	0.05	0.16	18%	0.02	0.02	0.01	0.04	5%	
N (n = 18)																				
0.05 - 0.14	1.75	1.44	0.63	3.91	0.65	0.59	0.19	1.32	43%	0.60	0.54	0.22	1.04	37%	0.43	0.38	0.22	0.80	25%	
0.14 - 0.42	3.73	3.31	1.30	7.53	1.32	1.19	0.32	3.23	28%	1.18	1.00	0.41	2.58	31%	0.97	0.76	0.30	1.90	25%	
0.42 - 1.2	12.76	13.29	3.10	22.01	3.43	2.75	0.58	6.74	24%	4.17	3.34	0.76	7.68	34%	3.65	2.65	0.73	8.13	26%	
1.2 - 3.5	0.91	0.68	0.31	1.65	0.20	0.13	0.04	0.43	18%	0.24	0.19	0.07	0.44	25%	0.15	0.09	0.03	0.36	12%	
3.5 - 10	0.40	0.36	0.18	0.61	0.04	0.03	0.02	0.08	8%	0.06	0.07	0.00	0.10	17%	0.02	0.02	0.00	0.04	4%	
D (n = 18)																				
0.05 - 0.14	1.12	1.02	0.45	2.19	0.20	0.16	0.07	0.42	17%	0.35	0.31	0.09	0.84	34%	0.39	0.32	0.17	0.75	35%	
0.14 - 0.42	3.61	3.18	1.58	7.28	0.70	0.57	0.24	1.55	17%	1.04	0.79	0.42	2.59	27%	1.23	1.04	0.55	2.73	31%	
0.42 - 1.2	5.58	4.66	2.21	10.44	1.05	0.71	0.29	2.39	16%	1.91	1.42	0.63	4.44	31%	1.93	1.56	0.69	4.58	32%	
1.2 - 3.5	0.38	0.36	0.17	0.65	0.05	0.03	0.01	0.14	9%	0.11	0.10	0.00	0.23	31%	0.07	0.04	0.01	0.18	13%	
3.5 - 10	0.16	0.13	0.10	0.32	0.03	0.02	0.00	0.06	11%	0.03	0.00	0.00	0.16	32%	0.01	0.00	0.00	0.03	11%	

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Table 3. Continued.

b) Transition											
	WSOC		NC			MDA			PA		
	mean	st. dev.	mean	st. dev.	%WSOC	mean	st. dev.	%WSOC	mean	st. dev.	%WSOC
N/D (n = 12)											
0.05 - 0.14	0.47	0.19	0.13	0.07	24%	0.18	0.08	37%	0.12	0.05	26%
0.14 - 0.42	1.33	0.56	0.32	0.17	21%	0.37	0.15	28%	0.32	0.14	24%
0.42 - 1.2	2.87	1.44	0.46	0.37	14%	0.74	0.44	25%	0.61	0.34	21%
1.2 - 3.5	0.37	0.14	0.11	0.04	28%	0.13	0.06	37%	0.04	0.02	12%
3.5 - 10	0.25	0.15	0.07	0.02	24%	0.07	0.01	29%	0.01	0.00	4%
N (n = 4)											
0.05 - 0.14	0.47	0.24	0.15	0.10	29%	0.20	0.12	42%	0.11	0.05	25%
0.14 - 0.42	1.48	0.47	0.38	0.18	25%	0.45	0.16	30%	0.32	0.12	21%
0.42 - 1.2	3.62	1.55	0.62	0.45	16%	0.84	0.29	24%	0.69	0.26	19%
1.2 - 3.5	0.49	0.03	0.08	0.02	16%	0.16	0.03	33%	0.04	0.01	9%
3.5 - 10	0.40	0.15	0.05	0.05	14%	0.08	0.01	21%	0.01	0.00	3%
D (n = 3)											
0.05 - 0.14	0.32	0.12	0.02	0.02	12%	0.11	0.05	34%	0.09	0.04	27%
0.14 - 0.42	0.84	0.18	0.06	0.07	14%	0.24	0.06	29%	0.22	0.06	26%
0.42 - 1.2	1.58	0.34	0.12	0.04	7%	0.32	0.08	20%	0.31	0.08	19%
1.2 - 3.5	0.22	0.04	0.00	0.00		0.06	0.02	26%	0.02	0.01	8%
3.5 - 10	0.09	0.02	0.00	0.00		0.01	0.03	51%	0.00	0.00	

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Table 3. Continued.

c) Wet period

	WSOC		NC			MDA			PA		
	mean	st. dev.	mean	st. dev.	%WSOC	mean	st. dev.	%WSOC	mean	st. dev.	%WSOC
N/D (n = 5)											
0.05 - 0.14	0.12	0.09	0.04	0.01	38%	0.05	0.02	47%	0.03	0.02	24%
0.14 - 0.42	0.37	0.13	0.05	0.03	10%	0.08	0.03	23%	0.06	0.02	16%
0.42 - 1.2	0.62	0.48	0.07	0.04	8%	0.12	0.07	22%	0.10	0.05	18%
1.2 - 3.5	0.21	0.14	0.03	0.01	25%	0.04	0.02	25%	0.01	0.00	4%
3.5 - 10	0.28	0.28	0.05	0.04	12%	0.04	0.02	26%	0.00	0.00	2%
N (n = 3)											
0.05 - 0.14	0.18	0.10	0.05	0.01	29%	0.06	0.02	35%	0.04	0.02	21%
0.14 - 0.42	0.46	0.07	0.06	0.02	13%	0.10	0.01	21%	0.07	0.02	16%
0.42 - 1.2	0.91	0.40	0.09	0.03	10%	0.16	0.04	19%	0.13	0.02	16%
1.2 - 3.5	0.29	0.11	0.01	0.02	13%	0.05	0.01	19%	0.01	0.00	4%
3.5 - 10	0.43	0.28	0.05	0.04	11%	0.06	0.02	15%	0.00	0.00	2%
D (n = 2)											
0.05 - 0.14	0.05	0.00	0.02	0.02	56%	0.03	0.02	59%	0.01	0.00	26%
0.14 - 0.42	0.23	0.04	0.01	0.01	7%	0.05	0.02	23%	0.04	0.02	16%
0.42 - 1.2	0.19	0.00	0.01	0.01	8%	0.05	0.02	28%	0.04	0.02	22%
1.2 - 3.5	0.09	0.02	0.01	0.02	37%	0.03	0.00	31%	0.00	0.00	
3.5 - 10	0.06	0.01	0.00	0.00		0.02	0.00	37%	0.00	0.00	

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Table 4. Contribution of levoglucosan and carboxylic acids identified by IEC and IC, respectively, to total WSOC and neutral compounds (NC), mono-/di-acids (MDA) and polyacids (PA) measured by the IC-UV method. The speciated carboxylic acids include C₁–C₃ aliphatic monocarboxylic acids (MCA), oxalic acid, C₃–C₉ aliphatic dicarboxylic acids (DCA), C₇–C₈ aromatic acids and C₆ aliphatic tricarboxylic acids (TCA). Average mass balances were calculated on a carbon basis for the dry and transition periods, on the basis of a limited number of samples for which the IFUSP MOUDI and the five-stage Berner impactor were operated approximately in parallel. The concentrations of the polar organic compounds determined on the MOUDI samples by IEC and IC were converted into five-stage size-distributions for comparison with the total WSOC and NC, MDA and PA data, according to the procedure described in the text.

Size intervals (µm)	%WSOC						%NC	%MDA				%PA	
	Levoglucosan	Aliphatic carboxylic acids				Aromatic acids		Aliphatic carboxylic acids			Aromatic acids		
		MCA	Oxalic	C ₃ –C ₉				MCA	Oxalic	C ₃ –C ₉			
Dry period													
0.05–0.14	0.3%	0.3%	0.5%	0.5%		0.1%	1.0%	0.8%	1.4%	1.2%	0.3%		
0.14–0.42	1.3%	0.6%	0.8%	1.1%	0.1%	1.9%	7.5%	1.9%	2.6%	3.6%	6.3%	0.2%	
0.42–1.2	2.7%	1.1%	1.3%	1.6%	0.1%	1.6%	12.1%	3.1%	3.5%	4.4%	4.3%	0.3%	
1.2–3.5	4.7%	1.2%	1.9%	1.4%	0.1%	1.2%	27.9%	3.7%	5.7%	4.1%	4.1%	0.3%	
3.5–10	0.0%	1.0%	1.8%	0.9%	0.1%	2.6%	0.1%	0.6%	1.0%	0.7%	2.1%	1.6%	
Transition period													
0.05–0.14	0.2%	0.4%	0.2%	0.4%	0.1%	0.2%	0.6%	1.0%	0.9%	1.0%	0.6%	0.3%	
0.14–0.42	0.1%	0.2%	0.2%	0.2%	0.1%	0.5%	0.3%	0.6%	1.4%	0.9%	2.1%	0.2%	
0.42–1.2	2.2%	0.9%	0.7%	1.4%	0.2%	1.1%	12.5%	3.0%	4.5%	4.6%	3.7%	0.7%	
1.2–3.5	4.5%	3.1%	1.0%	0.9%	0.1%	0.7%	10.7%	7.0%	4.7%	1.8%	1.9%	0.4%	
3.5–10		0.6%	0.3%	0.7%	0.5%	1.7%		2.1%	2.5%	2.8%	7.1%	10.0%	

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Table 5. Model composition of water-soluble carbon in size-segregated samples from the different periods of the SMOCC campaign. Model compounds with underscored names were evaluated on the basis of WSOC speciation, while the others were based only on functional group analysis and the apportionment of WSOC into the three main chemical classes from the IC–UV method: neutral compounds (NC), mono-/di-acids (MDA) and polyacids (PA). Model compounds corresponding to organic species never detected in aerosol samples are hypothetical compounds labelled with numbers 1 to 3 in the table. Their structure is shown in Fig. 6, and recommendations for suitable surrogates among the commercial standards are given in the footnotes.

			Dry (day)			Dry (night)			Dry (day/night)		Transition			Wet			Transition/ Wet
Size range (µm)			0.05- 0.14	0.14- 0.42	0.42- 1.2	0.05- 0.14	0.14- 0.42	0.42- 1.2	1.2-3.5	3.5-10	0.05- 0.14	0.14- 0.42	0.42- 1.2	0.05- 0.14	0.14- 0.42	0.42- 1.2	1.2-10
NC	<u>levoglucosan</u>	C ₆ H ₁₀ O ₅	0.5%	2.1%	3.4%	0.4%	1.9%	3.2%	10.7%	0.2%	0.2%	0.1%	3.9%	0.3%	0.1%	4.4%	18.5%
	compound 1 ^a	C ₁₈ H ₂₂ O ₆	2.3%	3.4%	3.2%	2.5%	3.0%	3.1%	5.7%	4.1%	6.0%	6.8%	4.8%	5.1%	5.4%	4.5%	0.0%
	cellobiosan	C ₁₂ H ₂₀ O ₁₀	12.0%	16.4%	13.8%	13.1%	14.5%	13.8%	2.9%	9.7%	27.3%	30.9%	18.2%	23.1%	24.6%	16.4%	46.5%
MDA	<u>malic acid</u>	C ₄ H ₆ O ₅	2.7%	4.9%	6.3%	2.2%	4.4%	5.9%	12.8%	18.1%	1.5%	1.0%	6.4%	2.2%	1.6%	7.3%	4.2%
	<u>vanillic acid</u>	C ₈ H ₈ O ₄	0.1%	2.3%	1.6%	0.1%	2.1%	1.5%	2.1%	8.0%	0.2%	0.5%	1.5%	0.0%	0.0%	0.0%	0.0%
	adipic acid	C ₆ H ₁₀ O ₄	17.7%	14.4%	14.3%	2.4%	10.2%	10.7%	0.0%	0.0%	33.6%	32.7%	30.3%	35.3%	34.3%	32.5%	30.8%
	syringyl- propionic acid	C ₁₁ H ₁₄ O ₅	3.7%	3.0%	3.0%	1.7%	10.1%	7.2%	0.0%	0.0%	2.4%	2.2%	2.1%	0.0%	0.0%	0.0%	0.0%
	compound 2 ^b	C ₁₉ H ₂₄ O ₅	16.7%	11.9%	13.0%	45.7%	16.6%	8.9%	46.4%	43.0%	3.5%	3.2%	3.0%	2.8%	2.4%	2.4%	0.0%
PA	compound 3 ^c	C ₃₆ H ₄₂ O ₁₆	44.3%	41.6%	41.4%	32.0%	37.2%	45.8%	19.2%	16.9%	25.2%	22.7%	29.8%	31.2%	31.6%	32.5%	0.0%

^a Can be approximated to compound with CAS 580-72-3;

^b Can be approximated to compound with *Sigma-Aldrich* Compound Number R221511;

^c Can be approximated to the Suwannee River fulvic acid (reference standard from IHSS).

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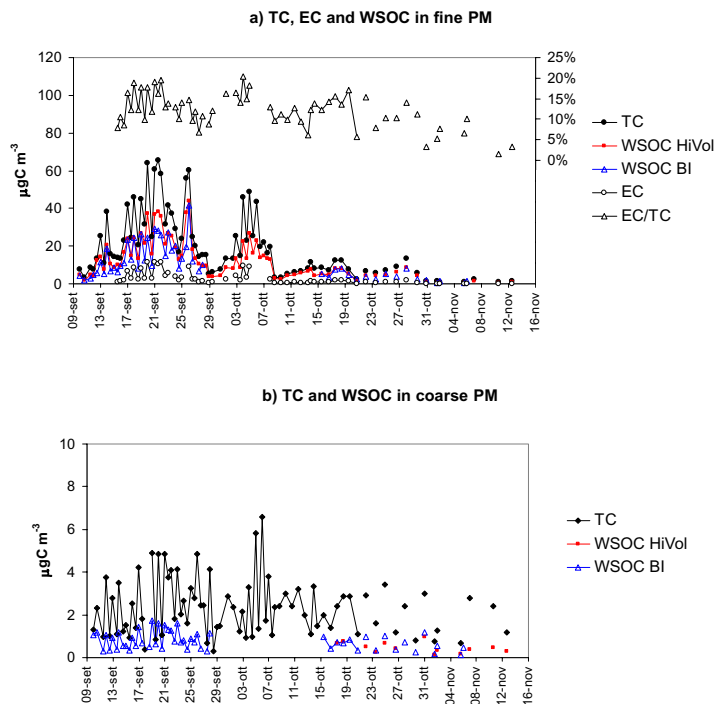


Fig. 1. Trends of TC, EC and WSOC concentrations ($\mu\text{gC m}^{-3}$) in the fine (a) and coarse (b) fractions of the aerosol during the LBA-SMOCC experiment. The profile of the EC/TC (%) ratio is also reported for the fine fraction. The concentrations of the carbon classes were obtained by the analysis of HiVol (HVDS) fine ($\text{PM}_{2.5}$) and coarse ($>2.5 \mu\text{m}$) filter samples. The WSOC concentrations determined on the Berner impactor (BI) samples are also reported for the fine fraction ($0.05\text{--}1.2 \mu\text{m}$) and for the coarse one ($1.2\text{--}10 \mu\text{m}$).

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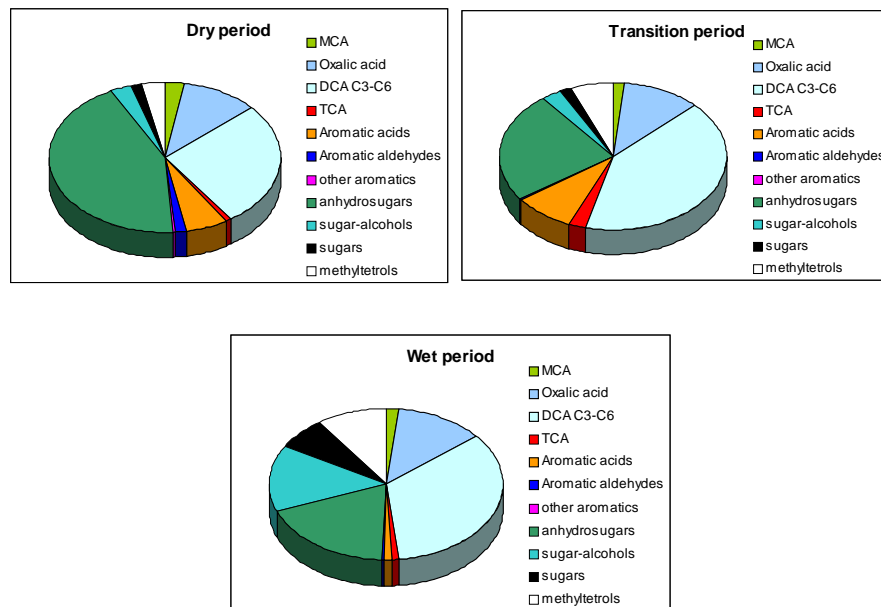


Fig. 2. Mean composition of the WSOC fraction characterized at the molecular level by GC–MS, IC and IEC methods in the different periods of the campaign. Data are obtained from Table 2: oxalic acid, mono- and tri-carboxylic acids and aromatic acids from IC analysis on SFU samples; sugars, sugar-alcohols, aromatic aldehydes and other aromatic compounds from GC–MS_{MPI}C analysis on HVDS2 filters; C₃–C₆ dicarboxylic acids (DCA), anhydrosugars and methyl-tetrols from GC–MS_{UA} of the HVDS1 samples.

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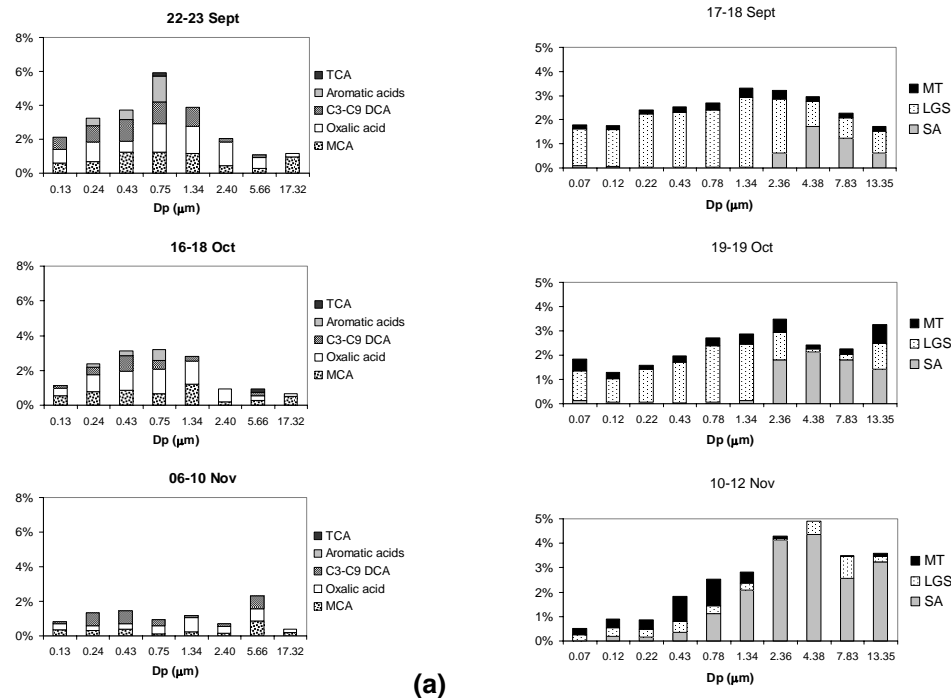
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(a)

(b)

Fig. 3. Fraction of PM accounted for by the organic compounds speciated in size-segregated aerosol samples. **(a)** Carboxylic acids from IC analysis of IFUSP MOUDI samples: aliphatic monocarboxylic acids (MCA), oxalic acid, C₃–C₉ dicarboxylic acids (DCA), C₇–C₈ aromatic acids and aliphatic tricarboxylic acids (TCA). **(b)** 2-methyltetrols (MT), levoglucosan (LGS) and sugar-alcohols (SA) (arabitol and mannitol). Both figures report the results for three different samples representative of the three main periods of the campaign. The horizontal axis gives the geometric mean (μm) of the lower and upper cut-off diameters of the MOUDI stages.

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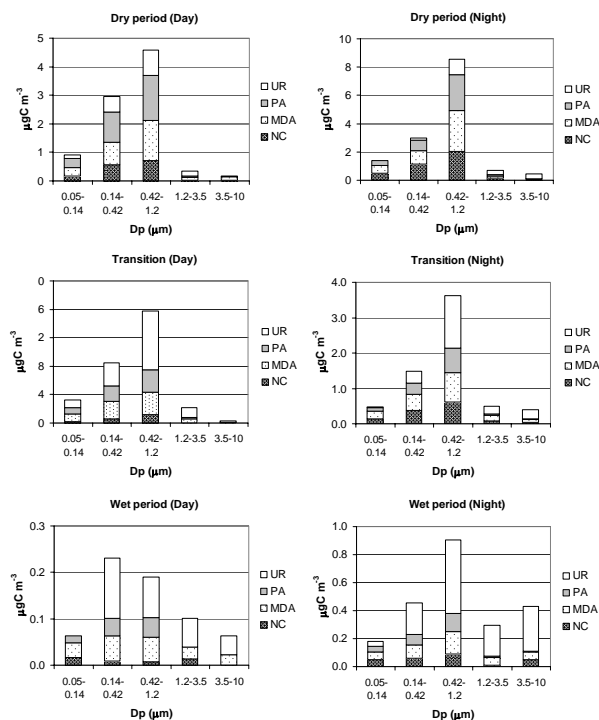


Fig. 4. Average concentrations of the WSOC chemical classes determined by the IC-UV method on the Berner impactor samples. NC: neutral compounds; MDA: mono-/di-acids; PA: polyacids; UR: unresolved, i.e. WSOC fraction not accounted for by IC-UV analysis. Mean concentrations are reported for the three main periods of the campaign and separately for the samples collected during night-time and those collected during day-time.

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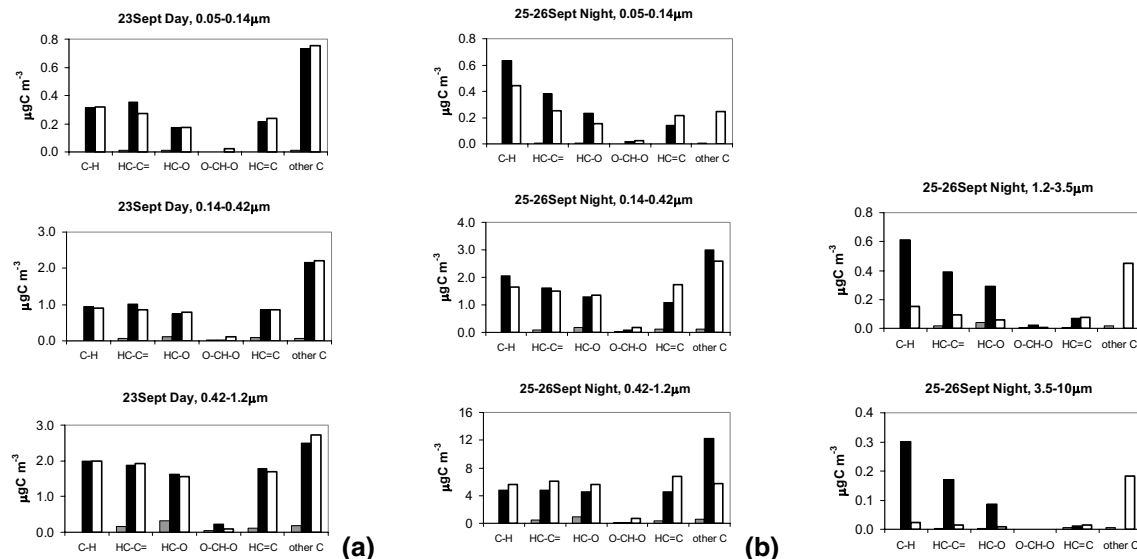


Fig. 5. Comparison between the functional group composition (black bars) determined by 1HNMR analysis of size-segregated samples and HVDS filters and that reconstructed on the basis of the individual compounds identified by GC-MS, IC and IEC techniques (gray bars). WSOC functionalities derived from the chemical model are also reported (white bars). The comparison is shown for the selected samples: **(a, b)** Two size-segregated samples from the dry period; **(c)** three PM_{2.5} HVDS samples from the dry, transition and wet periods, respectively; **(d)** chromatographic fractions of the PM_{2.5} HVDS sample collected on 25–26 September; **(e)** A coarse (PM_{<2.5}) HVDS sample from the transition season. NC (neutral compounds); MDA (mono-/di-acids); PA (polyacids); (d) one coarse HVDS sample from the transition period.

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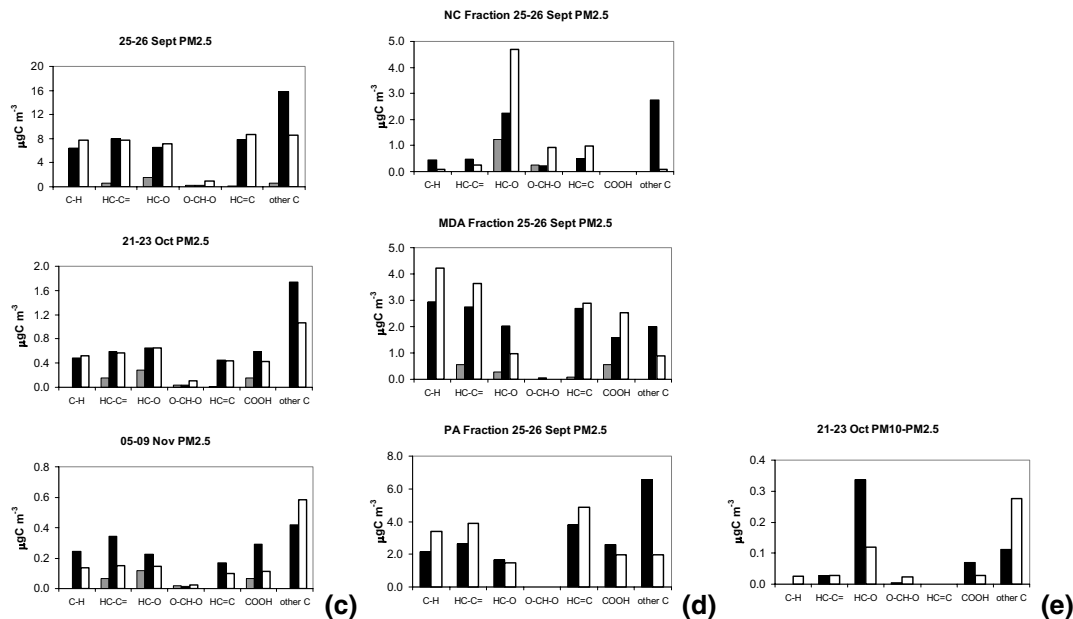


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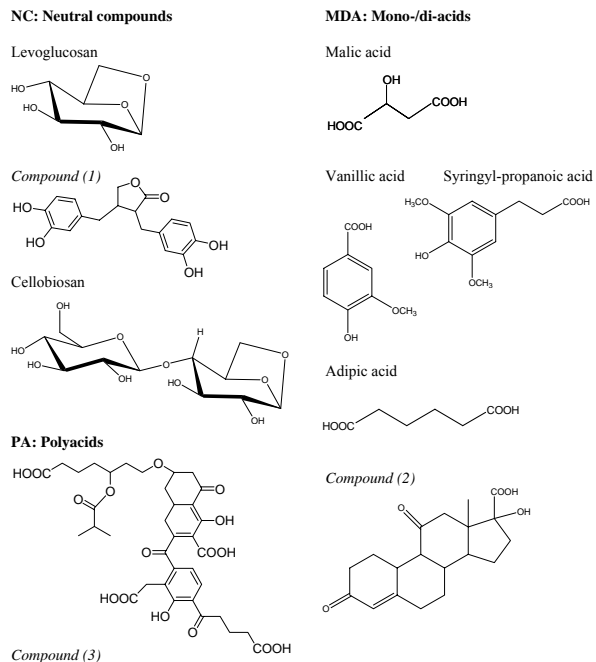


Fig. 6. Model compounds proposed to represent the chemical composition of WSOC on the basis of both speciation methods and functional group analysis. The three compounds labeled with numbers are hypothetical and not available commercially. On the basis of the chemical structure, they can be approximated to: the organic compound with CAS number 580-72-3 (Compound 1); the organic compound with Sigma-Aldrich Compound Number R221511 (Compound 2); the Suwannee River fulvic acid (reference standard from IHSS).

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