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Note

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Summary

The ELISA assay was used to measure the complex plasminogen-plasmin as well as chymosin in milk products. The results showed that plasmin concentration was slightly higher than the corresponding enzymatic activity, which could be explained by the blockage of several active sites by inhibitors or caseins. Measurement of chymosin in a St Paulin type cheese and a hard cheese « Comté » showed evidence for residual activity of chymosin in both curds. However, active enzyme was found only in the « St Paulin » whey. Therefore, the heat stability of chymosin is enhanced when bound to caseins and some activity may subsist in hard cheeses.

Résumé

Dosage de la plasmine et de la chymosine dans les produits laitiers par la méthode ELISA

La méthode immunologique ELISA a été appliquée au dosage du complexe plasminogène-plasmine ainsi que de la chymosine dans les produits laitiers. Les résultats montrent que la concentration en plasmine est légèrement supérieure à l'activité enzymatique correspondante, ce qui peut s'expliquer par un blocage d'une partie des sites actifs de l'enzyme soit par des inhibiteurs soit par les caséines. Le dosage de la chymosine dans un fromage de type Saint-Paulin et un fromage à pâte cuite pressée « Comté » a mis en évidence une activité résiduelle de la chymosine dans les deux caillés. Par contre, seul le lactosérum de fromage Saint-Paulin possédait encore de la chymosine active. Cela montre que la stabilité de l'enzyme liée aux caséines est accrue et qu'une certaine activité peut subsister dans les fromages à pâte cuite.

Introduction

Quantitative determination of proteolytic enzymes in milk products is important for a better understanding of the biochemical processes which occur during cheese ripening and conservation. Coagulating enzymes as well as heat
resistant plasmin (alkaline milk proteinase) may play an important role in the milk industry. As the enzymes encountered in milk products are partially bound to their substrate, it is difficult to determine their activity without performing an extraction, as it is usually done, for example, to measure chymosin activity in cheese.

In this short communication we describe an immunological method (ELISA) to measure quantitatively and specifically free and bound plasmin and its zymogen, plasminogen as well as chymosin. The results are compared with the enzymatic determination of plasmin activity.

I. Material and methods

A. Purification of bovine plasminogen

Bovine plasminogen was purified by affinity chromatography as described by DeutsCH and MERTZ (1970), using fresh blood plasma. The protein thus obtained was freed from immunoglobulin contamination by gel filtration on Bio-Gel P-100. Plasminogen was activated to plasmin with urokinase according to CASTELLINO and SODETZ (1976).

B. Samples preparation

Milk samples were prepared from fresh milk or lyophilized milk powder for the determination of the enzymatic activity of plasmin, as described by RICHARDSON and PEARCE (1981) (previous experiments showed that lyophilization of raw milk did not alter the enzymatic activity). When necessary, caseins were separated by isoelectric precipitation. After centrifugation, they were diluted in a solution of 0.4 M sodium-citrate (SC).

C. ELISA assay

Antibodies, obtained in rabbits, were diluted 1/200 in 0.1 M phosphate buffer, pH 7.2. One hundred microliter of this solution were added to each well of a Nunc plastic plate and incubated at 37 °C for 90 minutes. The wells were then washed three times with PBS containing 0.05 % of Tween-20 (PBS-Tween). Antigen solution diluted 1/25 to 1/400 with SC-Tween were added to the wells (100 μl each) and incubated as described above. After washing three times, an antigen-alkaline phosphatase conjugate, diluted 1/40 with PBS-Tween, was added. A further incubation-washing cycle was performed. A solution of 1 mg/ml of p-nitrophenylphosphate disodium salt solubilized in 1.0 M diethanolamin-HCl buffer, pH 9.8, containing 0.01 % of MgCl₂, 0.05 % AcZn and 0.05 % thimerosal (w/v) was added to each well. After 30 minutes the enzymatic reaction was stopped by adding 20 μl of 6.0 M NaOH and the intensity of the reaction determined by measuring the absorbance in each well at 405 nm.

D. Determination of plasmin activity

Plasmin activity was determined using the fluorimetric method described by RICHARDSON and PEARCE (1981). The aminomethyl coumarin peptide (Suc-
Ala-Phe-Lys-AMC) was purchased from Bachem (Bubendorf, Switzerland). The specific activity of the enzyme was determined by active site titration according to the method described by Chase and Shaw (1970).

II. Results and discussion

A. Immunological quantification and enzymatic activity of plasmin and plasminogen in raw milk

Table 1 shows the results obtained with 9 individual cow milk samples. The results of the enzymatic determinations of plasmin and plasminogen are in good agreement with the values published by Richardson and Pearce (1981): 140 to 730 ng/ml of plasmin and 550 to 2750 ng/ml of plasminogen. The values obtained by the ELISA assay are about 2.5 times higher than the enzymatic values. These discrepancies could be due to the fact that several active sites of the enzyme are occupied by inhibitors, or are still bound to casein whereas the rest of the molecule of the antigen can still be recognized by the antibody. It is also possible that the sample preparation may alter the enzymatic activity without modifying the antigenic structure. The activation of plasminogen to plasmin by urokinase may also be partially hindered by the presence of caseins. If the titration of the purified antigen has not been somewhat overestimated, it seems that the immunological method can determine the total amount of plasminogen and plasmin in milk.

<table>
<thead>
<tr>
<th>Sample Nr.</th>
<th>Enzymatic activity</th>
<th></th>
<th>ELISA</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Plasmin</td>
<td>Plasmin + Plasminogen</td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>192</td>
<td>1650</td>
<td>4277</td>
</tr>
<tr>
<td>2</td>
<td>469</td>
<td>1400</td>
<td>4756</td>
</tr>
<tr>
<td>3</td>
<td>231</td>
<td>953</td>
<td>2679</td>
</tr>
<tr>
<td>4</td>
<td>110</td>
<td>1200</td>
<td>2644</td>
</tr>
<tr>
<td>5</td>
<td>475</td>
<td>1240</td>
<td>3250</td>
</tr>
<tr>
<td>6</td>
<td>203</td>
<td>1300</td>
<td>4370</td>
</tr>
<tr>
<td>7</td>
<td>300</td>
<td>1140</td>
<td>3522</td>
</tr>
<tr>
<td>8</td>
<td>155</td>
<td>1150</td>
<td>2739</td>
</tr>
<tr>
<td>9</td>
<td>257</td>
<td>1850</td>
<td>4739</td>
</tr>
<tr>
<td>Mean Val.</td>
<td>266</td>
<td>1320</td>
<td>3664</td>
</tr>
</tbody>
</table>
B. Quantification of plasmin + plasminogen and chymosin in cheese

The amount of plasmin and plasminogen bound to the curd or present in the whey was determined in two types of cheeses: one hard cheese «Comté» heated to 56 °C for 40 minutes and one St Paulin type cheese heated to 33 °C. Table 2 shows the results obtained in raw milk, whey and in the curd, 20 hours after manufacture. The distribution of plasmin and plasminogen is nearly the same in both cheeses since 85 to 90 % of the enzyme are bound to the curd, as previously noted by CASEY et al. (1987). These results are in agreement with the analyses of the enzymatic activity determined in various types of Swiss hard cheeses (BAER, unpublished data). On the contrary they do not confirm the data obtained by BENSILMANE (1986), who detected almost no activity in «Comté» cheese. Titration of chymosin shows that the ratio of the concentration of the enzyme found in the curd compared to the amount added to the milk is almost the same in both types of cheeses. However the concentration of chymosin determined in the whey St Paulin cheese is much higher than in «Comté» cheese which was heated at higher temperature.

<table>
<thead>
<tr>
<th></th>
<th>Plasmin + plasminogen</th>
<th>Chymosin</th>
</tr>
</thead>
<tbody>
<tr>
<td>Making of hard cheese (Comté)</td>
<td>curd 90 %</td>
<td>18 %</td>
</tr>
<tr>
<td></td>
<td>whey 10 %</td>
<td>&lt; 10 %</td>
</tr>
<tr>
<td>Making of St Paulin type cheese</td>
<td>curd 88 %</td>
<td>20 %</td>
</tr>
<tr>
<td></td>
<td>whey 12 %</td>
<td>80 %</td>
</tr>
</tbody>
</table>

These results may indicate that the enzyme is immunologically heat resistant when bound to its substrate, as previously described by ANDRÉN et al. (1983) who demonstrated a close relationship between the enzymatic activity of chymosin and its immunological response. Thus it seems that the titration by the ELISA assay of chymosin in cheese reflects its enzymatic activity. These results indicate that a non negligible chymosin activity still remains in hard cheeses, as suggested by COLLIN et al. (1987) who proposed that the proteolysis of casein αs1 in «Comté» cheese may be due to chymosin activity. However these results should be confirmed.
Conclusions

The results presented in this short communication indicate that the ELISA assay could be adapted to the measurement of other milk enzymes. However, the data presented here must first be confirmed by further analyses. It may also be possible to differentiate plasmin from plasminogen using monoclonal antibodies.

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BENSLIMANE S., 1986. Variations de l'activité de la plasmine et du plasminogène dans les laits de vaches montbéliardes et dans les fromages de type Comté. Thèse de doctorat 3<sup>e</sup> cycle, Université de Besançon.


