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An extremely radioresistant green eukaryote for radionuclide bio-decontamination in the nuclear industry

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Nuclear activities generate radioactive elements which require processes for their decontamination. Although biological remediation has proved to be efficient in industrial applications, no biotechnology solution is currently operational for highly radioactive media. Such a solution requires organisms that accumulate radionuclides while withstanding radioactivity. This paper describes the potentialities of an extremophile autotrophic eukaryote, Coccomyxa actinobiotis nov. sp., that we isolated from a nuclear facility and which withstands huge ionizing radiation doses, up to 20 000 Gy. Half the population survives 10 000 Gy, which is comparable to the hyper-radioresistant well-known prokaryote Deinococcus radiodurans. The cell metabolic profile investigated by nuclear magnetic resonance was hardly affected by radiation doses of up to 10 000 Gy. Cellular functioning completely recovered within a few days. This outstanding microalga also strongly accumulates radionuclides, including 238U, 137Cs, 110mAg, 54Mn, 65Zn, and 14C (decontamination above 85% in 24 h, concentration factor, 1000–450 000 mL g−1 fresh weight). In 1 h, the microalga revealed as effective as the conventional physico-chemical ion-exchangers to purify nuclear effluents. Using this organism, an efficient real-scale radionuclide bio-decontamination process was performed in a nuclear fuel storage pool with an important reduction of waste volume compared to the usual physico-chemical process. The feasibility of new decontamination solutions for the nuclear industry and for environmental clean-up operations is demonstrated.

Broader context
The nuclear industry generates radioactive toxics and requires processes for their decontamination inside the facilities themselves and of the effluents released into the environment. Radionuclide decontamination is currently performed using physico-chemical methods. Despite their robustness and efficiency, these methods are expensive, do not remove completely certain elements, especially 14C, one of the main radionuclides released in effluents, and generate large volumes of secondary wastes when applied to environmental contaminations. Biological methods have proved to be efficient and competitive in various industrial applications. However, no viable method is presently available for the bio-decontamination of highly radioactive media. Such a method would require organisms that simultaneously accumulate radionuclides while withstanding their chemical and radiological toxicity and the radioactivity of the environment. Here we report on a new autotrophic green microalga, isolated from a radioactive nuclear site, which is extremely radioresistant and strongly accumulates radionuclides, including 14C. It was used in a real-scale bio-decontamination process with considerable reduction of radioactive waste volume. This microalga provides an excellent opportunity for new decontamination technologies. It could be used in bio-processes in the nuclear industry where its performance would complement those of the conventional methods and in the environmental field for the clean-up of accidentally contaminated water where large volumes have to be processed. The characterisation of an autotrophic eukaryote with such properties may also have an important outcome for the fundamental biology of adaptation to extreme environments.

Introduction
Nuclear energy technologies generate radioactive and chemically toxic compounds during the whole nuclear fuel cycle, from mining to reprocessing or waste treatment plants. These technologies require processes for radionuclide decontamination inside the facilities themselves and of the effluents which will be released into the environment. In the event of a nuclear disaster, such as in Chernobyl or Fukushima, massive quantities of
radionuclides are released into the environment and contami-
nate water and soil for decades. In the Fukushima-Daiichi
reactors themselves, radioactivity reached 3.8 GBq L \(^{-1}\) in stag-
nant water on the basement floor of the Unit 1 turbine building, \(^{3}\)
generating dose rates of 20–40 Gy h \(^{-1}.\) \(^{4}\) The maximum radiation
dose for human survival is 10 Gy, \(^{5}\) rendering the treatment of the
contaminated water complicated.

At nuclear power plants, treatment methods for radionuclide
removal from liquid streams include evaporation, chemical
precipitation/flocculation, solid/liquid separation, reverse
osmosis or ultrafiltration, sorption, and ion exchange. \(^{6,7}\) Ion
exchange is one of the most common and effective methods
employed. \(^{8}\) Some drawbacks include high cost, incomplete
removal of certain ions and problematic disposal of spent ion-
exchangers which requires special approaches and precau-
tions. \(^{8}\) The treatment of radionuclide contamination in the
environment relies on the same physico-chemical techniques
which are generally costly and environmentally destructive,
require much reagent and energy, and generate secondary toxic
sludge or waste products. \(^{9,10}\)

Decontamination by living organisms or extracts from
organisms may constitute an alternative or a complement to the
physico-chemical processes traditionally used for radionuclide
purification. \(^{11,12}\) Such an alternative may be particularly inter-
esting for environmental applications where huge volumes of
water or soil have to be processed. In various applications,
biological remediation has proved to be competitive against
conventional methods. Bio remediation generally offers a wider
field of application, a lower consumption of energy and chem-
icals, and lower cost and impact on the environment. \(^{11,12}\) Living
organisms combine physico-chemical contaminant fixation by
biosorption with metabolism-dependent uptake and accumula-
tion. In a nuclear environment, for maximum decontamina-
tion performance, organisms must be capable of strongly
accumulating radionuclides, while withstanding their toxicity
and must at the same time be highly radioresistant. To our
knowledge, no bio-process currently exists for the decontami-
nation of highly radioactive water.

Among radiation-resistant species stand the cyanobacterium
Chroococcidiopsis sp., \(^{13}\) the archaea Pyrococcus furiosus \(^{14}\) and
Halobacterium sp., \(^{15}\) the amoeba Dictyostelium discoideum, \(^{16}\) and
the fungus Alternaria alternata \(^{17}\) which withstand ionizing
radiation doses of 2500 to 5000 Gy. The most radiation-resistant
organisms described so far are prokaryotes, \(^{18}\) which include the
bacterium Deinococcus radiodurans which grows under an
ionizing radiation dose rate of 50 Gy h \(^{-1}\) and survives doses of
up to 20 000 Gy. \(^{19,20}\) However, using D. radiodurans for the in situ
bioremediation of nuclear waste sites would require genetic
engineering for the bacterium to acquire resistance to toxic
metals and remediating capabilities. \(^{21}\) Moreover, its culture
requires a supply of carbon nutrients and is therefore sensitive
to contamination by other bacteria.

Conversely, plants and algae fix a wide range of contami-
nants, including toxic metals and radionuclides. \(^{11,13,22}\) Photo-
synthetic organisms have the advantages of requiring less
energy and mineral culture media which are less sensitive to
bacterial contamination. Microalgae possess, in addition, a
large cell wall surface, which is interesting for decontamination
by both biosorption and metabolism-dependent mechanisms.
However, the 50% lethal dose (LD \(_{50}\)) of ionizing radiation for
algae generally falls in the 30–1200 Gy range. \(^{23}\) In 2008, a
microalga of the Chlorophyceae class which tolerated ionizing
radiation with an LD \(_{50}\) of 6000 Gy was described but radioactive
isotope accumulation had not been assessed. \(^{24}\) A microalga of the
Desmidiales order, Closterium moniliferum, was recently
considered for strontium decontamination but radioactive
isotope accumulation and resistance to radioactivity have not
been evaluated. \(^{25,26}\)

This work characterizes the properties and the potential use
of a new microalga discovered in a high ionizing radiation
nuclear environment in remediation technologies. This alga
was isolated from a pool used to store spent fuel elements in
a nuclear reactor. The new species, identified at the morphol-
ogical, biochemical and genomic level, belongs to the Coccomyxa
genus and was named Coccomyxa actinabiotos, from the char-
acteristics of the place it lives in. Its radioresistance evaluated
using physiological and metabolic analyses is outstanding for a
eukaryote. Its ability to fix radionuclides and toxic metals has
also been examined. This microalga combines both properties
of extreme radioresistance and radionuclide accumulation,
being able to fix radionuclides via metabolically inactive and
active processes even in a highly radioactive environment,
which is particularly interesting for \(^{14}\)C decontamination. It is
therefore an excellent candidate for new remediation solutions
in a highly radioactive environment. Its use for the bio-decon-
tamination of radionuclides on a real-scale was validated in the
storage pool of a nuclear facility.

Materials and methods
Algae culture and identification
Algae were grown at 21 °C in different culture media in 800 mL
flasks aerated on an orbital shaker (Innova 2300, New Brunsw-
wick Scientific, Enfield, CT) at 100 rpm, under a continuous
illumination of 70 μmol photon m \(^{-2}\) s \(^{-1}\). C. actinabiotos was
grown in a modified Bold Basal Medium (BBM) (Sigma-Aldrich,
Saint-Louis, MO) diluted twice with Milli-Q water (Millipore,
Molsheim, France), C. reinhardtii in TAP medium (Gibco, Life
Technologies SAS, Saint Aubin, France), and C. chodatii in
BBM.

Scanning electron microscopy was performed on cryo-des-
icated cells using a LEO 1530 scanning electron microscope
(Zeiss, Oberkochen, Germany) operating at a voltage of 20 kV
EHT. For genomic identification, the sequence of the C.
actinabiotos nuclear genome region spanning the genes for 18S
ribosomal RNA-Intranuclear Transcribed Spacer (ITS) (ITS 1–5.8S rRNA-
ITS2-28S rRNA (500 first bases) was amplified by PCR (4065 bp)
using primers EAF3 (5′-tgacactgtcgtgatcgcagc-3′) and
ITS055R (5′-ctcctctccgctgctttcaagcggg-3′). For the phylogenetic
analysis, the same genome region was sequenced in C. chodatii
strain SAG 216-2 and C. pelletigerae strain SAG 216-5. Sequences
of the nuclear genome region spanning the genes for 18S
ribosomal RNA-ITS1-5.8S rRNA-ITS2-28S rRNA (500 first bases)
used are deposited in the EMBL/GenBank databases under
accession numbers FR850476 (C. actinabiotis strain CCAP 216-25), FN597598 (C. chodatii strain SAG 216-2) and FN597599 (C. peltigerae strain SAG 216-5).

**Radioresistance**

To assess the resistance of algae to ionizing radiation (Fig. 2 and 3), C. actinabiotis cells initially grown in BBM (10⁷ cells per mL) were concentrated to 1–2 × 10³ cells per mL and inserted inside a used nuclear fuel element (UNFE) providing a γ-radiation flux of 4000 Gy h⁻¹. Irradiated algae were allowed to recover in fresh BBM. The cell mortality and the growth were measured after acute radiation using a neutral red staining method and using a Malassez counting cell and compared to the control, as described in Farhi et al.⁴⁴ The neutral red concentration was 0.003–0.03% w/v.⁵⁷–⁵⁸ Cell observation was performed 30 min to 1 h after dye application using an Optiphot microscope (Nikon, Japan) with a magnification of 1000×. C. actinabiotis mortality was maximum 3 days after irradiation. As high survival rates were obtained for C. actinabiotis (85% mortality i.e. 15% survival at 20 000 Gy, the highest radiation dose tested), the response was plotted on a linear scale. The same protocol was used for C. chodatii and C. reinhardtii.

Changes in the algae metabolic profile after acute irradiation (Fig. 4 and 5) were analyzed using ¹H nuclear magnetic resonance (NMR). Prior to NMR analysis, cellular metabolites were extracted using a methanol–chloroform mixture.⁵⁹ The cells (0.3–0.7 g fresh weight (FW)) were ground in a mortar in liquid nitrogen in the presence of maleate (0.5 mmol g⁻¹ FW) and 1 mL of H₂O and transferred into a nitrogen-cooled vial. After adding 1.5 mL of chloroform and 4.5 mL of methanol, the mixture was vortexed and incubated for 15 min at room temperature. Following the addition of 1 mL of chloroform and 1 mL of H₂O, the mixture was vortexed and centrifuged at 10 000g at 4 °C for 10 min. The polar phase was recovered, evaporated and freeze dried. The dried extract was dissolved in 0.6 mL of D₂O containing 100 mM potassium phosphate buffer at pH 7.0, 4 mM NaN₃, (trimethylsilyl)propionate-2,2,3,3-D₄ (TSP), and 4 mM ethylenediaminetetraacetic acid. The pH was adjusted to 7.0 with KOD or DCl. The resulting solution was lyophilized again and dissolved in 0.6 mL of D₂O. NMR analysis was performed in a 5 mm o.d. glass tube, at 25 °C, using a Bruker Avance 500 spectrometer (Bruker Biospin, Wissembourg, France) equipped with a 5 mm BBI probe. Relaxed spectra were obtained from a sum of 64–128 FIDs, recorded with a resolution of 0.084 Hz per pt and a 30 s repetition period. Fast acquisition conditions were also used to increase the signal-to-noise ratio for low concentration compounds. In that case, 256 FIDs recorded with a 60° RF pulse angle, a resolution of 0.17 Hz per pt and a 4 s repetition period were added. ¹H NMR spectra, referenced to the internal TSP chemical shift, were assigned using 1D and 2D NMR. Absolute metabolite quantity was determined from the integration of the resonance line of relaxed spectra after baseline correction. The line integrals were corrected for saturation effects when fast acquisition was used. For a complex multiplet partially superimposed to another resonance, the integration was done on the resolved part of the multiplet only. Intensity of the multiplet was measured on the spectra of the pure compound.

**Radionuclide decontamination**

To determine the bio-decontamination rate of γ- and β-emitter radionuclides (Table 1), C. actinabiotis cells (250 mg fresh weight (FW)), initially grown in BBM, were washed three times with Milli-Q water and suspended in the light in 100 mL of pH 5.5 nuclear facility effluents initially containing different radionuclides. Effluent 1 contained the γ-emitters ⁶⁰Co (280 ± 8 Bq L⁻¹), ⁵⁷Co (530 ± 10 Bq L⁻¹), ¹¹⁰mAg (66 ± 7 Bq L⁻¹), ¹²⁴Sb (1460 ± 13 Bq L⁻¹), ⁵³Cr (1180 ± 12 Bq L⁻¹), ⁶⁵Zn (120 ± 7 Bq L⁻¹), and ⁵⁴Mn (230 ± 8 Bq L⁻¹), and the β-emitters ³H (200 000 Bq L⁻¹) and ¹⁴C (10 000 Bq L⁻¹). Effluent 2 contained the γ-emitters ¹³⁷Cs (67 ± 7 Bq L⁻¹) and ²³⁸U (21 ± 3 Bq L⁻¹) and the β-emitters ³H (200 000 Bq L⁻¹) and ¹⁴C (2000 Bq L⁻¹). After 24 h, the decontamination rate, calculated as (1 – Cfinal/Cinitial) × 100 where Cfinal and Cinitial represent respectively the final and the initial radionuclide concentration in water, was determined for each isotope by analyzing water and algae separately separated by centrifugation at 2000 g using γ-spectroscopy (ITECH Instruments, Châteauneuf-Les-Martigues, France). The decontamination of ¹⁴C was evaluated separately. Algae were exposed for 3 to 7 h to 2000 to 20 000 Bq L⁻¹ H⁺CO₃⁻ in 100 mL of pH 6.5 demineralized water (matrix 3), under illumination. The amount of ¹⁴C in water and algae was analyzed using liquid scintillation counting (Packard TriCarb 2900TR liquid scintillation analyzer, PerkinElmer, Waltham, MA).

Radionuclide bioconcentration factors (BCFs) (Fig. 6) were evaluated for algae directly harvested from the UNFE storage pool under illumination (200 μmol m⁻² s⁻¹). The pH 5.5 demineralized water composition changes according to the components stored; it contained typically ⁶⁰Co (400 Bq L⁻¹), ¹¹⁰mAg (1000 Bq L⁻¹), ¹²⁴Sb (1500 Bq L⁻¹), ⁵³Cr (5000 Bq L⁻¹), ⁶⁵Zn (400 Bq L⁻¹), ⁵⁴Mn (300 Bq L⁻¹), ³H (300 000 Bq L⁻¹), and ¹⁴C (20 000 Bq L⁻¹). In experiments with different microalgae, radionuclide accumulation was found to proceed rapidly, equilibrium being reached within some hours or days.⁶⁰,⁶¹ BCFs were calculated as the ratio of the radionuclide content in algae (in Bq g⁻¹ FW) to the radionuclide content in water (in Bq mL⁻¹), both concentrations being measured by γ-spectrometry. ²³⁸U BCF was assessed by incubating C. actinabiotis (60 mg FW) with 10⁻⁵ mol L⁻¹ uranyl nitrate for 24 h and measuring the ²³⁸U concentration in algae and water using an inductively coupled plasma-mass spectrometer (ICP-MS) (Hewlett-Packard 4500 Series, Agilent Technologies, Santa Clara, CA).

To determine the fixation capacities of Ag and Co, algae were suspended with an equivalent of, respectively, 6.7 × 10⁻⁷ mole of AgNO₃ g⁻¹ algae FW in BBM diluted 10 times or in deionized water for 2 days and 10⁻³ mole of CoCl₂ g⁻¹ algae FW in BBM diluted 10 times for 6 days. Experiments were performed in diluted BBM to lower the concentration of chloride which forms a precipitate with silver. The presence of this precipitate was taken into account to assess the amount of silver fixed by the algae. After exposure, phases were
separated by centrifugation at 2000g; algae were quickly washed with water, centrifuged again, and the metal concentration was assessed in algae and the liquid phase using ICP-MS.

Real-scale validation (Fig. 7) was performed in a nuclear facility storage pool as follows. Under normal conditions, the pool contains algae colonies, glued onto metallic parts, and which concentration in suspension is controlled by intermittent normal circulation and filtering of water which is operated regularly. The algae naturally grow in purified water from the dissolved CO₂ and ambient light. Water purification by resins is operated when necessary to maintain the radionuclide concentration below the regulatory level. On day 14, two nuclear mechanical components releasing ¹¹⁰ᵐAg were introduced into the pool, which also contained ⁶⁰⁹Co, ⁵⁸⁶Co, ¹²⁴⁵Sb, ⁵¹¹Cr, ³³H, and ¹⁴⁴C. Purification by resins was stopped on day 30. From day 35 to day 56, the water was purified by uptake of radionuclides by the suspended algae, which were then collected onto a micro-pore filter (diameter 50 mm, height 60 mm) installed in a mobile pool surface robot. The algae concentration in water on day 35 was roughly estimated to be 10⁴ to 10⁵ cells per mL.

Important uncertainty arises from a non-homogeneous distribution of algae according to the depth of the pool (higher concentration close to the surface). The micro-pore filter is installed in front of a 14 m² h⁻¹ pump which forces the circulation of the algae and contaminated water through the filter. The resulting highly radioactive filters containing 60–100 MBq each were changed every 2 days. The total alga dry weight collected on the filters during the experiment was 40 g. From day 60, the resins and the normal water circulation were put back into operation, while most of the suspended algae had been collected from the water.

For the direct comparison of the decontamination efficiency using algae or resins (Table 2), a nuclear effluent containing the γ-emitters ⁶⁰⁹Co (280 Bq L⁻¹), ⁵⁸⁶Co (140 Bq L⁻¹), ¹¹⁰ᵐAg (32 Bq L⁻¹), ⁵¹¹Cr (840 Bq L⁻¹), ⁶⁶Zn (100 Bq L⁻¹), and ⁵⁴Mn (150 Bq L⁻¹) and the β-emitters ³³H (260 000 Bq L⁻¹) and ¹⁴⁴C (6000 Bq L⁻¹) was contacted with the suspended alga (160 mg FW per 100 mL effluent) or with the suspended resins classically used in the nuclear effluent purification process (80 mg of Purolit NRW100 and 80 mg of Purolit NRW505 (Purolite, Paris, France) ion exchange resins per 100 mL effluent), under agitation, in the light. After 1 and 24 h, the decontamination rate was determined for each isotope by analyzing its content in water and algae or resin using γ-spectroscopy or liquid scintillation counting after phase separation by centrifugation at 2000g.

Results and discussion

*Coccomyxa actinabiots*, a new microalga species isolated from a nuclear facility

Few autotrophic eukaryotes are capable of living in radioactive nuclear sites. Most organisms previously found in such environments are bacteria such as *Kineococcus radiotolerans* or *D. radiodurans*-related strains or fungi such as *A. alternata*, which was isolated from the Chernobyl site after the nuclear disaster. Extreme environments constitute indeed a unique opportunity for new knowledge in the development of life as well as for novel biotechnologies. We have isolated from the UNFE storage pool of a nuclear site a new autotrophic microalga that is highly resistant to ionizing radiation. The slightly acidic (pH 5.3 ± 0.2), demineralized (conductivity, 1.2 ± 0.2 μS cm⁻¹), oligotrophic (10 mg L⁻¹ nitrate; <0.2 mg L⁻¹ phosphate) pool water contains radionuclides originating from the dissolution and activation of UNFE materials. It is in contact with air, continuously illuminated (200 μmol photons m⁻² s⁻¹), and maintained at 25 ± 3°C. The UNFEs generate radiologic dose rates varying between 70 μGy h⁻¹ close to the pool walls and the surface and several hundred Gy h⁻¹ close to the elements.

The alga was harvested and cultured on solid agar plates containing Bold Basal Medium (BBM), a mineral culture medium classically used for algae. After successive plating of individual alga and isolation of colonies, an axenic strain obtained from one alga cell was isolated, grown in liquid BBM under non-radioactive conditions, and identified at the morphological, biochemical, and genomic level. It is a unicellular freshwater eukaryotic green microalga measuring 6.8 ± 0.9 μm × 3.8 ± 0.6 μm, containing a parietal chloroplast with starch (Fig. 1a and b). The main pigments determined by HPLC are chlorophylls a and b, β-carotene, and lutein. It multiplies by division with an immobile vegetative stage, by flagellate zoospore production, and by auto-sporulation. The cell density doubles in 8 days when algae are grown in flasks filled with pool water, compared to about 2 days in BBM. It also grows from its internal reserves in ultra-pure water (conductivity, 0.05 μS cm⁻¹) for more than one month. Cells gather in colonies in a mucilaginous jelly in the pool but are isolated in culture.

The sequence of the nuclear genome region spanning the ribosomal RNA gene was determined. Pair-wise 18S rDNA sequence alignment yields *Coccomyxa chodatii* strain SAG 216-2, *Coccomyxa pelitigerae* strain SAG 216-5, *Coccomyxa* sp. Flensburg fjord 2 (EU127471), *Coccomyxa glaronensis* strain CCALA 306 (AM167525), and *Coccomyxa* sp. strain CPCC 508 (AM981206) as the closest species with 98%, 97%, 97%, 96%, and 96% sequence identity, respectively. From the phylogenetic cladogram (Fig. 1c), it is inferred that this microalga belongs to the *Coccomyxa* genus, which comprises to date more than 30 freshwater and marine species, including free-living, epiphytic, symbiotic with lichens, trees, or protozoans, and parasitic species. However, it exhibits two unique insertions of 545 and 436 bp in the 18S rDNA sequence and distinct ITS sequences. Both its genomic and morphological characteristics make it a new species belonging to the *Coccomyxa* genus in the *Trebouxiophyceae* class (Fig. 1c), which was named *C. actinabiots* (CCAP 216-25) meaning “lives in rays”.

*C. actinabiots* radioresistance

As *C. actinabiots* lives in a continuous ionizing radiation environment, its radioresistance properties were quantified and compared to that of *C. chodatii*, one of the taxonomically closest species (Fig. 1c), and of the reference unicellular microalga *Chlamydomonas reinhardtii*. Cells subjected to acute γ-irradiation at an intense flux of 4000 Gy h⁻¹ integrated doses of up to
20 000 Gy (2 Mrad). Maximum mortality value versus dose, determined using vital staining, reveals that C. actinabiotis exhibits an extreme radiation resistance (Fig. 2). Its LD$_{50}$ is 10 000 Gy and it survives 20 000 Gy. Irradiation of up to 6000 Gy did not affect C. actinabiotis growth (Fig. 3). A 10 000 Gy dose induced a growth lag of 2–4 days. Cells irradiated at 20 000 Gy recovered the maximum population density of the control in less than 2 weeks. The LD$_{50}$ was 1500 Gy for C. chodatii (Fig. 2), and less than 500 Gy for C. reinhardtii. As mentioned above, few organisms are capable of surviving high ionizing radiation fluxes. They are mainly prokaryotes$_{13–15,19,20}$ but they also include some eukaryotes.$^{16,17,24}$ The lethal dose corresponding to 50% mortality after $\gamma$-irradiation for other reference or radioresistant species is 250 Gy for the bacterium E. coli,$^{19}$ 2000 Gy for Dicytostelium discoideum,$^{16}$ 2500 Gy for Chroococcidiopsis sp.,$^{13}$ 5700 Gy and 10 700 Gy for D. radiodurans grown respectively on DMM and TGY media.$^{19}$ Although C. actinabiotis is an eukaryote, its survival to ionizing radiation is similar to that of the prokaryote D. radiodurans.

To assess the impact of irradiation on cellular functioning, the metabolic changes that take place in the cell upon irradiation were investigated using NMR. Metabolites, as intermediates or end-products of transcriptomic and/or proteomic changes, represent accurate indicators of the cell biochemical status.$^{44}$ Their qualitative and quantitative determination gives information on the biochemical status of the organism, cellular functioning, and pathways affected by stress or disease.$^{45–47}$ NMR is a powerful technique to obtain cell metabolic profiles which provide such information.$^{48–50}$ The C. actinabiotis metabolic content remained highly stable upon irradiation at doses of up to 10 000 Gy (Fig. 4), revealing very efficient protection and/or repair capabilities. It is unlikely that metabolites remain stable because all proteins metabolizing them would be damaged by ionizing radiation. In a less radioresistant alga species actually, a 20 000 Gy dose triggers a drastic decrease in most of the metabolites.$^{24}$ In radiosensitive species, lower ionizing radiation doses resulting in cell apoptosis induce a depletion in many metabolites.$^{31,52}$ Moreover, the fact that C. actinabiotis growth is not significantly different after irradiation at doses of up to 6000 Gy means that cells are still alive. Specifically, the sucrose pool was maintained one day after irradiation even in the 10 000 Gy irradiated sample (Fig. 5a), indicating that the sugar energy source pathways were still functional or had been rapidly repaired. Cells were still able to provide the substantial energy needed to repair the damage caused by ionizing radiations to macromolecules. Ionizing radiation actually generates damages to macromolecules (DNA and proteins), the number of DNA double-strand breaks (DSBs) being roughly proportional to dose. One gray induces about 0.002–0.005 DSBs per Mbp in many species, whether radio-resistant or not.$^{19,33,54}$ Doses of 10 000 Gy thus introduce hundreds to a few thousands of DSBs into the C. actinabiotis genome. From the metabolic perspective, a 10 000 Gy dose triggered a statistically significant increase in the pools of some main amino acids such as valine and isoleucine (Fig. 5a), also observed in other species submitted to sublethal UV- or

Fig. 2 Resistance to ionizing radiation of C. actinabiotis compared to other microalgae strains. Acute $\gamma$-irradiation was performed at a dose rate of 4000 Gy h$^{-1}$. Mortality values obtained three days after irradiation are average ± standard deviation ($n = 3$ except $n = 2$ at 20 000 Gy).

Fig. 1 C. actinabiotis cells and phylogenetic tree. (a) Microalgae harvested from the UNFE storage pool. (b) Microalgae grown in BBM observed using a scanning electron microscope. Ellipsoidal cells, surrounded by mucilage (M), contain a nucleus (N), a chloroplast (Chl), and starch granules (S). Other organelles, including vacuoles, occupy the rest of the cell. (c) Phylogenetic tree obtained after pair-wise DNA sequence alignment of the 18S rDNA sequence (BLASTn) (ref. 43) by using a maximum likelihood approach. The upper scale indicates a 1% substitution ratio.
γ-irradiation doses,24,46,54 which suggests a partial protein lysis or synthesis activity related to irradiation.24,46,54 However, 9 days after irradiation, the algae metabolite content did not significantly differ whatever the irradiation dose (Fig. 5b), suggesting the complete recovery of cellular functioning.

How can a living organism withstand such doses that damage glass and plastic, turning them brittle? In D. radiodurans, the origin of this amazing survival still remains unclear; numerous studies point to a set of resistance and repair mechanisms9,20,55–57 including multiple genome copies, extremely efficient functioning of conventional DNA repair systems, and protein protection against oxidative damage generated by irradiation via high Mn/Fe ratios. Mechanisms that protect or repair C. actinabiotis are presently unknown. Preliminary experiments using gel electrophoresis suggest that C. actinabiotis could be able to restore its genome. C. chodatii also grows after irradiation of 10 000 Gy, whereas Chlamydomonas does not; however, C. chodatii withstands lower radiation levels than C. actinabiotis (Fig. 2), indicating that C. actinabiotis most probably utilizes specific resistance and repair mechanisms.

Radionuclide and toxic metal accumulation

To demonstrate whether it is possible to take advantage of the exceptional radioresistance of C. actinabiotis to decontaminate radionuclides, the algae were incubated with synthetic and real effluents containing the main radionuclides present in nuclear effluents, namely the γ-emitters 60Co, 65Zn, 110mAg, 124Sb, 51Cr, 62Zn, 57Mn, 137Cs, and 238U and the β-emitters 3H and 14C.58 Algae and plants have been considered for radionuclide decontamination.10,59 Uptake capacities of metals or radionuclides depend among others on the organism, its growth conditions, conditioning and concentration, the radionuclide chemical speciation and concentration, the contact time, the presence of competitors within the matrix.53,59–63

C. actinabiotis in contact with nuclear effluents strongly accumulates γ-emitters. Substantial BCFs reached 450 000 for 110mAg and 35 000 for 60Co in algae directly harvested from the pool (Fig. 6). Cobalt and silver represent 77 to 94% of the γ-emitters released in liquid effluents of pressurized water nuclear reactors.64 Silver also belongs to the highest toxic class of heavy metals with cadmium, surpassed only by mercury.65,66 Values obtained in this work are among the upper values of the wide range of BCFs reported for these elements in microalgae, namely 1700–400 000 mL g−1 FW for silver24,46,57 and 300–3300 mL g−1 FW or 40 000 mL g−1 dry weight (DW) for radiocobalt.30,68,69

In 24 h, C. actinabiotis completely removed 110mAg, 65Zn, and 137Cs from nuclear effluents and fixed more than 90% of 60Co, 51Cr, 57Mn, and 238U (Table 1). Decontamination of 110mAg reached 85%. C. actinabiotis shows very efficient radionuclide uptake. In other microalgae including Scenedesmus, Cyclotella, and Chlorella sp., decontamination rates of 45–100%, 43–80%, and 45–95% have been reported for 110mAg, Co, and U, respectively.10,31,70–72

Concerning the overall resistance of C. actinabiotis to metallic toxicity, high fixation capacities were obtained for non-radioactive silver and cobalt, namely 15 and 20 mg silver g−1 FW.
in algae exposed to Ag$^+$ in water and in diluted BBM, respectively, and 1.5 mg cobalt g$^{-1}$ FW in algae exposed to Co$^{2+}$ in diluted BBM, i.e. about 150–200 mg Ag and 15 mg Co g$^{-1}$ DW, at the same level as the reported extremes.\textsuperscript{65,69,73–75} Maximal values of 300 mg silver g$^{-1}$ FW were reported in a Pseudomonas containing bacterial community\textsuperscript{76} and up to 6 mg cobalt g$^{-1}$ DW in the zinc hyperaccumulator Thlaspi caerulescens.\textsuperscript{77}

\textit{C. actinabiotis} fixes various radionuclides and toxic metals, both concentration factors and fixation capacities revealing a great affinity for silver and cobalt. Algae and plants are able to chelate and immobilize metallic contaminants on their surface, as well as to incorporate and sequester them within their cytosol or vacuoles, changing their speciation into less-toxic forms.\textsuperscript{11,22} Mechanisms for metallic ion fixation in \textit{C. actinabiotis} might be related to the mucilage shell surrounding the cells (Fig. 1b) and to intracellular concentration.

Biological radionuclide decontamination has mainly been implemented in low dose rate environments. Moreover, most studies have addressed uranium whose main isotope activity is low, typically 1.3 Bq L$^{-1}$ for a 100 $\mu$g L$^{-1}$ solution of $^{238}$U. The originality of \textit{C. actinabiotis} lies in the combination of its extreme resistance to ionizing radiation and its ability to uptake very efficiently toxic metals and radionuclides, enabling its use in highly radioactive environments.

**Real-scale radionuclide biodecontamination using \textit{C. actinabiotis}**

Despite significant research efforts on biodecontamination, very few industrial set-ups for radionuclide biodecontamination are operational.\textsuperscript{9,10} Real-scale radionuclide bio-decontamination was then tested \textit{in situ} in a 360 m$^3$ storage pool of radioactive components using \textit{C. actinabiotis} in suspension in water and compared to conventional methods. The storage pool water is usually purified by ion-exchange resins that fix radionuclides. An extremely active nuclear component releasing $^{110m}$Ag was introduced in the pool on day 14 (Fig. 7), leading to an increase in radioactivity in water despite the resin-based purification. Resin-based purification was stopped on day 30 and the level of radionuclides strongly increased thereafter. The classical purification process was then replaced by algae-based decontamination for 21 days. The decrease in the concentration of the main radionuclide, $^{110m}$Ag, observed between days 35 and 56, originates from its uptake by algae in suspension, which were then collected onto filters. Fig. 7 shows that exponential growth of algae was observed in the storage pool (Fig. 7).

Fig. 5  Changes in the cellular metabolic content in response to irradiation. Metabolite concentration, in $\mu$mol g$^{-1}$ FW, as a function of the radiation dose, in kGy, (a) 1 day and (b) 9 days after acute irradiation. Abbreviations as in Fig. 4.

Fig. 6  Bio-concentration factors, in mL g$^{-1}$ FW, of the radionuclides $^{110m}$Ag, $^{60}$Co, $^{51}$Cr, $^{65}$Zn, $^{54}$Mn, $^{124}$Sb, and $^{238}$U by the microalga \textit{C. actinabiotis}.
determined in algae and water using 310 e was similar to that obtained with 80 mg resin per 100 mL standard deviation. With the physico-chemical decontamination method using ion-exchange resins. The volume of radioactive waste generated was then at least 100 decrease in the activity profile and hence in the purification efficiency by microalgae is comparable to that by resins. On the whole, the algae removed 740 ± 7 MBq γ-emitters, including 310 ± 5 MBq 110mAg, 270 ± 5 MBq 51Cr, and 30 ± 5 MBq 60Co from the pool in 21 days. The mean activity collected by C. actinabiota was 20 ± 5 MBq g−1 DW. A challenge in the nuclear industry is the reduction of the ultimate radioactive waste volume that requires careful and safe storage and disposal. Once dried, the algae volume was reduced by 90%. The volume of radioactive waste generated was then at least 100 times lower than that of resins. The decontamination efficiency of algae was also directly compared to that of the resins. When using 160 mg algae FW i.e. 16 mg DW per 100 mL nuclear effluent (which corresponded to 107 cells per mL), the decontamination efficiency was similar to that obtained with 80 mg resin per 100 mL effluent for the γ-emitter radionuclides 110mAg, 58Co, 60Co, and 54Mn after a 1 h contact time (Table 2). 51Cr decontamination by resins was higher. Conversely, 65Zn decontamination by algae was twice greater, probably because zinc is a physiological metal taken up by the cells via active mechanisms. Decontamination efficiency using algae increased after 24 h, though less or identical to that obtained using resins, except for 14C. The removal of 14C is usually problematic using ion-exchange resins and only reached 27% in this experiment. Whatever the contact time, the decontamination efficiency was far superior using illuminated algae (Table 2) owing to 14C incorporation inside the cells through metabolically mediated processes, particularly through photosynthesis. In this kind of application, living organisms are advantageous for overall maximal decontamination performance. Algae also have the advantage of smaller waste volumes compared to resins. Once dried, the algae retained the totality of the γ-emitters and 97% of the 14C fixed during the decontamination step, yielding for this experiment a ultimate waste volume reduction of 5 compared to resins.

Conclusions
The newly discovered microalgae C. actinabiota not only exhibits an exceptional radioresistance but also possesses several other assets. From its photosynthetic activity, it can produce the organic materials it needs for its metabolism and growth. It only needs light, water, CO2, and a few dissolved minerals to grow. It can thrive in a radioactive environment and is also capable of capturing and concentrating, rapidly and efficiently, radionuclides in nuclear facility effluents. The feasibility of the bio-decontamination of radionuclides on a real-scale has been demonstrated. This alga is an excellent candidate for new methods of remediation. The algae-based methods could be used inside nuclear facilities, where they would complement or replace conventional methods and reduce the volume of radioactive waste, at the exit point from nuclear facilities to reduce radioactive emissions into the environment, or for the decontamination of accidentally
polluted water. An industrial pilot of the bio-process is currently under development at the French Atomic Energy Commission (CEA) and at the Laue Langevin Institut.

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