



## TITLE

Tumor quantification of several fluoropyrimidines resistance gene expression with a unique quantitative RT-PCR method. Implications for pretherapeutic determination of tumor resistance phenotype.

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## **ABSTRACT**

Pretherapeutic determination of tumor resistance to chemotherapy is a main challenge, hindered by the low number of mechanisms characterized at the same time, the small size of the clinical specimens and the heterogeneity of the techniques or the lack of true quantification. The aim of the present study was to determine in real time quantitative RT-PCR, tumor cell expression of several transcripts involved in cancer cell resistance with a unique cDNA sample from a tumor biopsy. The technique had to be suitable in clinical practice for determination of several factors involved in resistance to a given drug family, for example, fluoropyrimidines resistance factors : thymidylate synthase (TS), dihydropyrimidine dehydrogenase (DPD), thymidine kinase (TK), dihydrofolate reductase (DHFR), folypolyglutamate synthetase (FPGS). A frame-shifted artificial construct was designed specifically to work within the same conditions. We validated our technique by quantifying expressions of these 5 genes starting from tissue samples of colorectal carcinoma and the surrounding normal mucosa of 33 different patients. That real time quantitative RT-PCR technique using the frame-shifted artificial construct as a standard provided a real comparison and quantification of different resistance factors. Tumor resistance phenotype determination based on that approach will be investigated in a control study.

## **KEY WORDS**

Chemotherapy, fluoropyrimidines, human colon cancer, real-time PCR

## INTRODUCTION

5-fluorouracil (5-FU)-based chemotherapy is the reference treatment in advanced tumors, especially digestive location. Other drugs, such as CPT-11 or oxaliplatin, can also be used alone or combined to 5-FU, but the development of tumor cell drug resistance remains a major obstacle to chemotherapy efficiency. Even if drug efficacy has improved mainly through drugs combination [1-3], at least about 50% of patients undergo 2 to 3 months of treatment and sometimes severe adverse side effects before a negative response rate assessment. Characterization and then quantification of the biochemical determinants to drug response before treatment could be a useful approach to improve therapy results and avoid toxic and useless treatments.

Despite several studies, little is known about factors and parameters involved in tumor cell resistance or sensitivity to 5-FU. Many resistance factors to drugs have been previously described, mainly *in vitro* but also *in vivo* and some have been associated with poor outcome and/or response to treatment in preclinical studies [4-8]. Obviously, determination of a single factor is not sufficient to predict therapeutic response. However, some difficulties remain to determine the true implication of different factors in drug resistance. For a long time, most of the studies explored only one isolated resistance factor, such as thymidylate synthase (TS), the 5-FU main target [9-13], and left aside other potential factors, ignoring or at least probably underestimating their respective importance and intrication. Used techniques such as radioenzymatic assay, immunohistochemistry, or semi quantitative RT-PCR, are very different and not easily reproducible and standardized. These methods have shown shortcomings in term of sensitivity, specificity or applicability in clinical practice: radioenzymatic assay is tedious, time consuming and needs radioisotopes; immunohistochemistry is not truly quantitative and semiquantitative RT-PCR is laborious.

Finally this analytical heterogeneity prevents the understanding of the respective predictive value of the suspected drug resistance factors in clinical practice so far.

More and more studies confirm that simultaneous quantification of several markers would permit to improve their prediction value in outcome or response to chemotherapy [14-19].

The technique of real time quantitative RT-PCR appears to be an interesting approach, with a higher sensitivity than conventional methods, and a better feasibility than enzymatic techniques with small tissue samples. Actually, it allows quantifying multiple mRNAs expression with a very low amount of material. But real time RT-PCR quantification depends firstly on the accurate quantification of the used DNA standard [20]. Multiple standard molecules are classically needed when determining the expression of several genes, and this could generate some variability due to the quantification of individual DNA standard, by either spectrophotometry or fluorescence or due to the quality of the DNA preparation itself. According to those observations, our purpose in the present study was to set up a single, simple and accurate quantitative PCR method for quantifying several resistance determinants under common amplification characteristics. This method could be a promising innovative alternative approach to achieve a real prediction of response to chemotherapy before starting a treatment.

We developed one composite standard DNA molecule for quantification of several mRNA in order to make sure we obtained accurate relative quantification of all the genes expressions. Primers sets for each studied mRNA were selected before composite standard DNA construction. Couple of primers had to work under the same PCR conditions, generate PCR products of almost the same length, and produce no amplification artefacts.

We chose to elaborate a composite standard DNA molecule for 5-FU resistance factors. This drug and oral fluoropyrimidines are widely used in treatment of numerous cancer

types, especially gastrointestinal cancer but also breast and head and neck cancers, and represent almost 50% of the chemotherapy prescription. We focused on transcripts expression of 5 genes, which are most often reported in tumor resistance to 5-FU : TS is the key enzyme of thymidine *de novo* synthesis and the main cellular target of 5-FU ; Dihydropyrimidine dehydrogenase (DPD) is the initial and rate-limiting enzyme for 5-FU catabolism and leads to dihydrofluorouracil ; Thymidine kinase (TK) catalyses the phosphorylation of deoxythymidine in the salvage pathway of dTMP synthesis ; Dihydrofolate reductase (DHFR) is the key enzyme of folates cycle, reducing folates in dihydrofolate (DHF) and tetrahydrofolate (THF) in the cell ; and the folylpolyglutamate synthetase (FPGS) is responsible for folate polyglutamylation.

We validated the protocol by evaluation of these 5 genes in tissue biopsies of colorectal carcinoma and the surrounding mucosa from 33 patients who underwent surgery for colorectal cancer.

## **MATERIALS AND METHODS**

### **Primer design**

Most of primers used in this study were designed by using primer3 freeware ([http://web.umassmed.edu/bioapps/primer3\\_www.cgi](http://web.umassmed.edu/bioapps/primer3_www.cgi)) and obtained desalted from Invitrogen. All primers had to work under the same conditions : annealing temperature of 55 °C, MgCl<sub>2</sub> final concentration of 4 mM and PCR product sizes not exceeding 500 bp (primers characteristics are depicted in Table I).

Each pair of primers was tested by capillary PCR (Lightcycler, Roche Diagnostics) using "Lightcycler FastStart DNA Master SYBR green I" (Roche Diagnostics). The absence

of amplification artefacts was confirmed by establishing a melting curve at the end of amplification and by gel electrophoresis analysis (Figure 1). Moreover, PCR product of each set of primers was cloned into pGEM-T easy vector (Promega) and sequenced (CEQ 2000 apparatus, Beckman Coulter Standard).

### **Composite standard DNA construction**

As described in Figure 2A, addition of primer sequence was done by PCR using overlapping primers, and the first overlapping primer was added by PCR on a stuffer DNA corresponding to a 259 pb sequence of the Rat V1a receptor (nucleotide 141 to 399) [21].

First sequence addition was performed in a 50  $\mu$ l volume with 10 ng of cloned Rat V1a receptor cDNA, 1  $\mu$ M of each overlapping primer, 400  $\mu$ M dNTPs, 2.5 mM MgCl<sub>2</sub> in 1X PCR buffer, overlaid with mineral oil. After a preliminary denaturing step (94 °C for 2 min 30 sec), reaction was initiated by addition of 2.5 units of Taq polymerase (Eurobio). PCR was then performed under the following conditions: 94 °C for 30 sec, 55 °C for 1 min, and 72 °C for 1 min 30 sec for 20 cycles on a Hybaid thermocycler (Coger). The PCR product was analyzed and purified after low melting agarose gel electrophoresis. Then aliquot of the purified PCR product was used for the subsequent sequence addition under the same conditions of PCR and purification. Final PCR product was purified and cloned into pGEM-T easy vector (Promega). Plasmid was extracted by "QIAprep Spin Miniprep Kit" (Qiagen), quantified by spectrophotometry at 260 nm, and sequenced. Using forward and reverse primers selected in the same conditions, we cloned the fragments of human GAPDH and  $\beta$ Actin as normalization housekeeping genes.

### ***In-vitro* transcription.**

Synthetic RNAs for the establishment of standard curves were obtained from 1µg of pGEM-T easy vector (containing composite standard DNA or fragments of GAPDH or β Actin) used as template for *in-vitro* transcription reaction with T7 RNA Polymerase ("RNA Transcription Kit", Stratagene) following the manufacturer's instructions. The obtained standard synthetic RNA were quantified by measuring absorbance at 260nm. After concentration adjustment, synthetic RNA were aliquoted and stored at -80°C.

### **Human tissue collection**

Pairs of colorectal tumors and the surrounding normal mucosa were obtained from 33 patients who underwent surgery for colon or rectal cancer at the Centre Regional de Lutte Contre le Cancer Paul Papin, Angers, during the period between December 1999 and May 2003. Patients were treated according to the diagnostic of their disease, and could receive or not a treatment containing 5-FU, before and/or after the operation.

All samples were checked by a pathologist before they were frozen and stored in liquid nitrogen until RNA isolation. Patients' characteristics are presented in Table II.

### **Total RNA extraction**

Frozen tissue fragments were included in Tissue Tek (Sakura), and 10 to 20 slices 30 µm thickness were collected using a cryostat at -27°C. Total RNA was extracted immediately using the "High Pure RNA Tissue Kit" (Roche Diagnostics) following manufacturer's procedure. Anatomopathologic exam of each sample was performed again on one interposed slice to confirm the nature of each specimen. After elution, total RNA was aliquoted and stored at -70 °C until use. RNA concentrations could not be determined since tissue samples were too small.

### **Reverse-transcription**

10  $\mu$ l of total RNA tissue sample, or known amount of standard synthetic RNA (serial dilution starting from  $10^3$  to  $10^{-2}$   $\mu$ g/ $\mu$ l synthetic RNA concentration) were mixed with 2.65  $\mu$ g of random hexamers (Amersham Pharmacia Biotech) in 15  $\mu$ l final volume, incubated at 70  $^{\circ}$ C for 5 min and chilled on ice. Then the reaction was performed at 37  $^{\circ}$ C for 1 hour with 200  $\mu$ M dNTPs, 40 units of RNase Inhibitor (RNasin, Promega) and 200 units of Moloney Murine Leukemia Virus Reverse Transcriptase (MMLV-RT) (Promega) in a 50  $\mu$ l final volume of buffer recommended by supplier. For each sample, a reaction without MMLV-RT was done as negative control. Then cDNA was purified using "Qiaquick PCR Purification Kit" (Qiagen). Purified cDNA was aliquoted and stored at  $-20^{\circ}$ C.

### **Real-time PCR**

Amplification was done in a final volume of 10  $\mu$ l containing 5  $\mu$ l of cDNA diluted at 0.1 in sterile distilled water (sample or standard), 0.5  $\mu$ M of each primer, 4 mM  $MgCl_2$ , and 1  $\mu$ l of "LightCycler FastStart DNA master SYBR green I" 10X (Roche Diagnostics). Then the following protocol was used : (i) denaturation program (95  $^{\circ}$ C for 10 min), (ii) amplification and quantification program repeated 40 cycles (95  $^{\circ}$ C for 15 sec, 55  $^{\circ}$ C for 11 sec, 72  $^{\circ}$ C for 22 sec, with a single fluorescence measurement of SYBR green I at each end of cycle), (iii) melting curve program (65-99  $^{\circ}$ C with a heating rate of 0.1  $^{\circ}$ C and continuous fluorescence measurement). In this study, the Second Derivate Maximum Method was used for Crossing Point (Cp) determination, using LightCycler Software 3.3 (Roche Diagnostics).

GAPDH, TS, TK, FPGS, DHFR and DPD mRNA were quantified starting from 2 independent RT reactions followed by PCR amplification in a one assay-run for each tissue sample.

### **Statistical analysis**

Results are presented as mean value +/- standard errors (SD). Statistical analysis was performed using SPSS 10.0 software. Correlations were performed using the Spearman rank test (rs). mRNA levels of the primary colorectal cancer and those of the corresponding normal mucosa were compared using the Wilcoxon signed rank test. Statistical differences between clinicopathological factors were evaluated using non parametric tests, Mann-Whitney U-test between 2 groups and Kruskal-Wallis test for 3 or more groups. A p value less than 0.05 was considered to be statistically significant.

## **RESULTS**

### **Selection of PCR primer sets**

Preliminary studies focused on the design of optimized primers working with good and highly comparable amplification efficiency in the standard conditions of PCR we choose SYBR green I, annealing temperature of 55 °C and MgCl<sub>2</sub> final concentration of 4 mM for the 5 genes of interest. Different couples of reverse and forward oligonucleotides for TS, DPD, TK, FPGS, DHFR and housekeeping genes GAPDH and βActin were tested using cDNA prepared from total colon tissue RNA or H<sub>2</sub>O as a template, random hexamers and MMLV-RT. Every primer had to present a good amplification curve, a single melting peak showing no artefact amplification, and a single band with the expected size after gel electrophoresis of each PCR product (Figure 1). Sequencing of cloned PCR products confirmed the specificity of amplification. Oligonucleotides sequences and physical characterization of PCR products are depicted in Table I. Since RT-PCR could provide variable quantities of cDNA products depending on the quality of RNA sample and on

efficiency of the RT reaction [22], we tested RT-PCR protocol. We got a very good reproducibility of RT reaction as Cp value standard deviation for a single RT reaction was less than 0.2 in triplicate, and for 5 different reverse transcription reactions was less than 0.25 for each gene in a one assay-run PCR (data not shown).

### **Construction of the DNA composite standard**

The construction of the DNA composite standard is outlined Figure 2. It consisted of five successive steps of PCR using overlapping primers around a 259 bp sequence from rat V1a receptor (Figure 2A). Each step of construction was checked by gel electrophoresis (Figure 2B), and the final construct was then cloned into pGEM-T easy vector. The composite standard was designed to produce amplicon size of almost the same length for the 5 primer pairs of interest: 390, 386, 385, 388, and 385 bp for TS, TK, DHFR, DPD and FPGS, respectively (Figures 2C and 2D). The quality of the standard molecule was asserted by sequencing analysis after cloning.

### **Establishment of standard curves**

Standard synthetic RNA was obtained by *in-vitro* transcription of the cloned composite standard DNA or cloned fragment of GAPDH. At first, RT-PCR amplification starting from the same concentrations of standard synthetic RNA ( $10^3 \mu\text{g}/\mu\text{l}$ ) was realized with each couple of primers and gave almost the same Cp ( $19.99 \pm 0.55$  cycles) showing a comparable amplification efficiency of each primer set. Then, standard curves were realized for each couple of primers starting from eight increasing synthetic concentrations. Slopes calculated from linear relationship between Cp and logarithm of the initial synthetic RNA amount varied from  $-3.99$  for DPD to  $-3.34$  for GAPDH and the regression was always higher than 0.99 (Figure 3). We noticed for each regression curve a slope over the 3.32 value

for an amplification efficiency of 100%, this could mean that SYBR green I present in the reaction could slightly inhibit the amplification.

### **Gene expression levels in biopsies**

At first, we evaluated the integrity of total RNA extracted from tissue sample of 35 patients (couple of colorectal carcinoma and the surrounding normal mucosa) by amplification of GAPDH mRNA, after RT-PCR performed with or without MMLV-RT. Quantification was then performed with the tissue samples of the 33 patients since 2 colorectal carcinoma samples presented significant genomic DNA contamination (data not shown).

As the purpose of our study was to truly quantify mRNA whatever the origin and/or the amount of the tissues, we had to normalize each sample in order to obtain an accurate relative quantification for every targeted gene. Quantification of the 2 housekeeping gene GAPDH and  $\beta$  Actin mRNA expression showed that they were significantly correlated ( $r_s = 0,936$ ,  $p < 0.001$ ). We choosed GAPDH because it was more sensitive to genomic contamination [23, 24], and widely used in quantification [14, 15, 25]. Then, relative mRNA levels of TS, TK, FPGS, DHFR and DPD were expressed as the ratio of targeted gene to GAPDH in order to compensate for the variation in RNA amount.

Results showed that even if expression of the 5 studied factors is widely variable, all of them were underexpressed in tumor tissue compared to normal mucosa, slightly for TK and FPGS with  $p < 0.05$ , and more significantly for TS, DHFR and DPD with  $p < 0.005$  (Wilcoxon signed-rank test, Table III).

Several clinicopathological factors, such as Gender, Location, Staging and Differentiation grading were studied. We found some correlation between few clinicopathological factors and mRNA level of every 5 genes in the colorectal carcinomas

(Table IV). DPD and FPGS expressions were significantly lower in women as compared to men. DPD mean expression was  $26.0 \pm 41.0$  (n = 16) for men and  $9.1 \pm 6.3$  (n = 10) for women with  $p = 0.023$ , and FPGS mean expression was  $41.8 \pm 20.0$  (n = 21) for men and  $24.4 \pm 8.8$  (n = 11) for women with  $p = 0.005$ . DHFR expression in tumor tissue was significantly lower for extended disease (group "B" which include metastatic disease and local relapse) when compared to more localized diseases (group "A" which include tumors stages I, II and III) (DHFR mean expression was respectively  $40.9 \pm 22.7$  (n = 11) and  $62.5 \pm 21.8$  (n = 22) with  $p = 0.010$ ). TS expression was apparently lower in well differentiated tumors as compared to poorly differentiated specimens (TS mean expression was respectively  $31.4 \pm 12.1$  (n = 17) and  $50.5 \pm 16.8$  (n = 4)) but the difference was not statistically significant ( $p = 0.081$ ). Conversely, there was no correlation between few clinicopathological factors and mRNA level of every 5 genes in normal mucosa.

We also evaluated correlations between expression of the 5 studied genes in colorectal tumor versus normal mucosa (Table V). Results showed profound differences since mRNA levels of the 5 different genes were generally more correlated between them in the normal mucosa than in the tumoral tissue (TK/DPD, TK/DHFR and FPGS/DPD mRNA levels only remained correlated in the colorectal carcinoma). These observations indicated differences and greater variability of tumoral tissue compared to normal mucosa.

## **DISCUSSION**

Since drug resistance is frequently at the origin of treatment failure, pretherapeutic characterization of the molecular resistance determinants could be of great interest. It would allow more rational decisions, on whether or not to proceed with 5-FU-based chemotherapy

as first line therapy, since other drugs, i.e. CPT-11 and oxaliplatin, are efficient in colorectal cancer in first line therapy. Many independent biological studies described numerous resistance factors, and the expression of some of them correlates with clinical outcome or chemotherapy response. Since these results are hardly comparable, sometimes conflicting and even contradictory, they could not be taken into account in clinical practice until now. Some problems stem from the fact that the used techniques are very different, most of them being semi-quantitative, tedious and time consuming, requiring significant amount of tissue, and characterizing only 1 or 2 factors. Quantitative real time PCR is a powerful method for quantifying several transcript expressions from small tissue samples. But it could be sometimes hampered because it depends highly on the accuracy of the quantification of the standard DNA used [24, 26].

The purpose of this study was to overcome these problems by setting up a real time quantitative PCR assay using a unique composite standard DNA molecule to determine transcript expression of resistance factors under the same experimental conditions. It was specifically designed to compensate variations that could occur when quantification of several factors is needed. Moreover, the technique had to be reproducible, feasible starting biopsies-sized tissue samples, easily extended in laboratories and in prospect for large scale studies.

This composite standard was set up to allow the amplification of 5 genes previously reported for their involvement in tumor resistance to 5-FU or in survival. TS, the 5-FU main target, is the most studied gene. Its overexpression is often linked to treatment failure and poor clinical outcome especially in advanced colorectal cancer [12, 15, 27, 28]. DPD is the rate-limiting enzyme of 5-FU, and has been largely studied and reported as a predictor factor. Its underexpression in primary tumor seems to be correlated with an enhanced response to 5-FU and to survival [15, 29]. For the 3 other studied factors, increase of TK [7, 19], DHFR [30, 31] or FPGS decrease [17, 32, 33] have also been related to a greater tumor cell resistance to

5-FU, or a worse survival. But these results are often conflicting and large-scale studies necessary for statistical validation and for proving their involvement and importance in practice have been limited.

In order to validate the method, we choose to quantify mRNA expression of the 5 transcripts, in biopsy-sized tissue samples of primary colorectal tumor and normal surrounding mucosa from 33 patients. In many previous studies, the true malignant nature of the tissue and the RNA quality could not be ascertained partly because tissues came from old tumor banks. In the present study, a pathologic exam of the tissue was again performed before RNA extraction to ensure its origin. Unfortunately, It was not possible to control the quality of extracted RNA and to quantify it by standard procedures since it was extracted from very small amount of material. To overcome the inherent variability in human tissue biopsies both in quality and quantity, the quantification was weighted with a control expressed gene, GAPDH. As the use of a housekeeping is necessary, we plan to improve the standard conception by including the primer sequences of this (or those) gene(s). Finally, the RT reaction, which could be limiting due to RNA quality or experimental conditions, was tested and we found that RT-PCR protocol was well reproducible.

Even if the population of interest was heterogeneous it permitted to validate our quantification technology. We observed an underexpression of the 5 studied genes in primary tumor when compared with the normal mucosa. Such difference has already been reported for DPD [34, 35]. There is less data for the other studied genes, and we have no explanation for this observation, except that mucosa is a fast renewing tissue.

Correlations were also observed between several clinicopathological parameters and the expression of the 5 genes.

Decreased FPGS and DPD mRNA expressions were found in women colorectal carcinoma as compared to men tumors whereas this difference was not observed in normal

mucosa. Tumors with low DPD expression are considered to be more sensitive to 5-FU. Female gender is often considered as a positive prognostic factor in colorectal cancer, and more recently as a positive factor for 5-FU adjuvant chemotherapy [36]. Such DPD underexpression in women primary tumor has been previously reported [37, 38] and it was proposed that low DPD expression observed in women tumor is probably associated with a better prognostic value for 5-FU chemotherapy.

Results also showed a decreased TS expression in well differentiated colorectal primary tumors. This observation is in agreement with the meaning that low TS expression is of better prognosis for survival, since well differentiated tumor are more aggressive.

Curiously, we also noticed that DHFR mRNA level was underexpressed in colorectal cancer in the case of "B" group which include metastatic diseases and local relapse. Folates present dual modulatory effects on carcinogenesis, but several points in the literature suggest that this observation might be of interest. Indeed, numerous studies suggested that low folate intake increased carcinogenesis [39, 40]. Moreover DHF and methyl-THF had an antiproliferative effect in human colon cancer cell line *in vitro* [41, 42]. At last, Levy and coll. showed that low DHFR mRNA expression correlated significantly with poor outcome in acute lymphocytic leukaemia [43]. It could mean that decrease of DHFR expression might reproduce cellular conditions close to low folate intake, conditions related to carcinogenesis and maybe to the severity of the tumoral disease.

Another question was the eventual correlation between gene expressions, because quantification of several genes that are correlated, perhaps co-regulated, would provide no additional benefit for response prediction. The use of a unique standard DNA molecule permitted that study. Our results showed that correlation between the 5 genes was quite different in the 2 types of tissues, since gene expressions were widely correlated in normal mucosa and not in primary tumor. Anterior works have also showed that TS/DPD [16, 25],

TS/TK [16], TS/FPGS [18] expressions are not correlated in tumor tissues. Those data highlight a profound difference between colorectal tumor tissue and the normal mucosa, and a high heterogeneity in tumor tissue. This could be of interest in a further study of predictive factors of chemoresistance.

In the present work, we have developed a real-time quantitative RT-PCR tool, with a unique standard DNA and homogenous PCR procedure. The assay we set up is reproducible, it allows comparison of expression levels of several genes in the same time, and is feasible starting from biopsies-sized tissue samples. Moreover, this technique could be easily used in different laboratories. Nevertheless, quantitative PCR technique show some limits if too many factors are involved in tumor resistance mechanisms. Indeed, Thymidine Phosphorylase, Uridine Kinase, Uridine Phosphorylase, or Orotate Phosphoribosyl Transferase are also linked to 5-FU resistance[5, 7, 44, 45]. Under these conditions technique microarray experiments could represent a powerful alternative. Each of the two techniques have different advantages. Since quantitative RT-PCR is sensible and could be realized with small amount of tissue, it could be quite laborious if quantification of numerous genes is needed. Conversely more tissue sample is used in microarray, but it allow more gene to be investigated. The preliminary determination of a gene implication in drug resistance is essential. the real implication of a gene is essential. For this reason, the development of two technical approachs is complementary. In the future, specific microarrays containing few genes could greatly contribute to the prediction of resistance.

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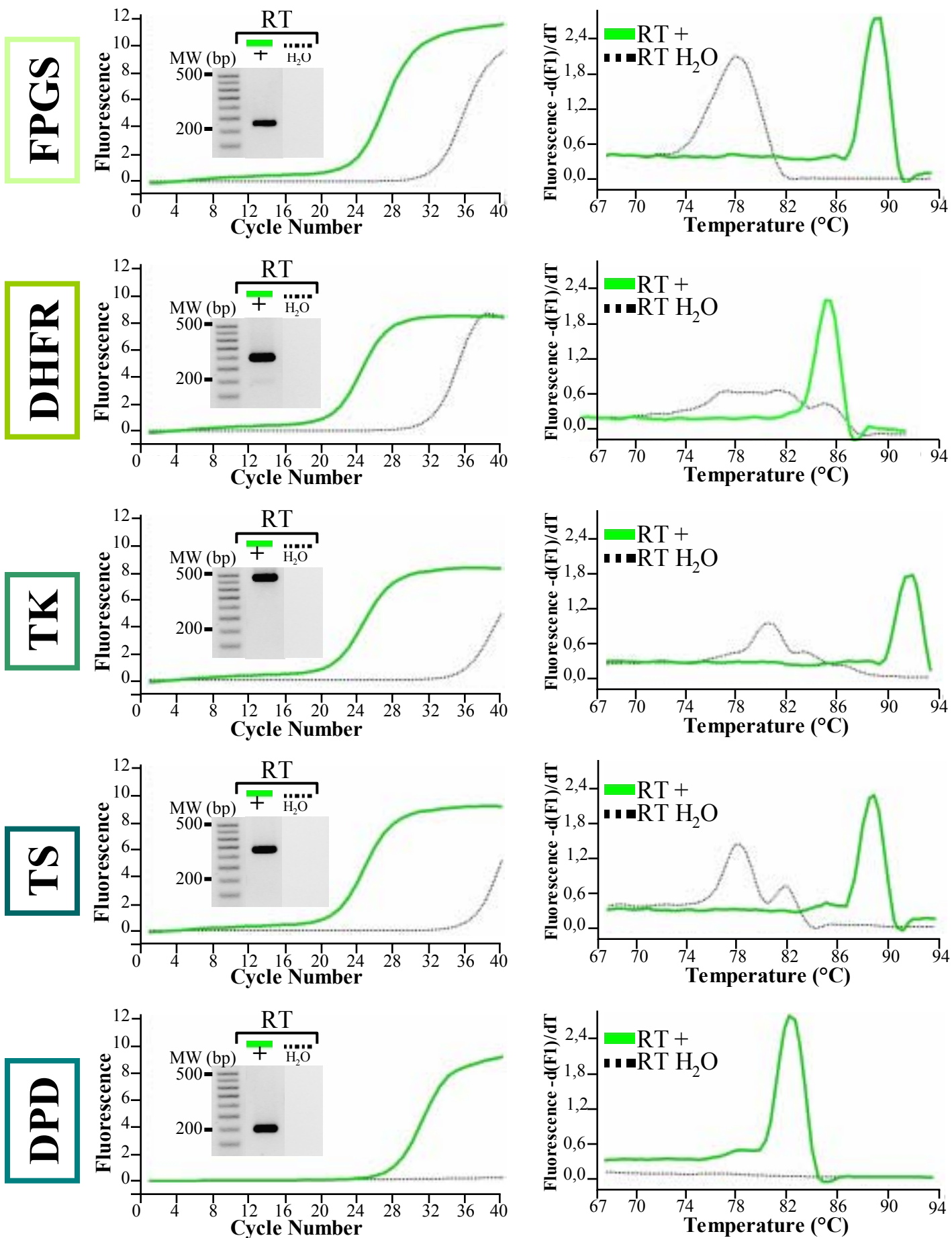
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## FIGURE LEGENDS

**Figure 1 :** Amplification curve, melting peak and gel analysis of the 5 studied genes DPD, TS, TK, DHFR, and FPGS after RT-PCR. The five couple of primers were tested using MMLV-Reverse Transcriptase, random hexamers, and cDNA made from total mucosa tissue RNA (RT+) or H<sub>2</sub>O (RT H<sub>2</sub>O) as a template. Selected primers gave a single melting peak and a single band of the expected size after agarose gel electrophoresis.

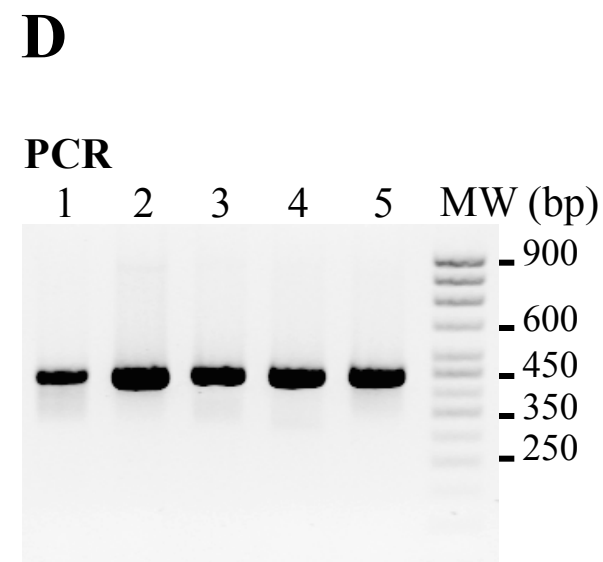
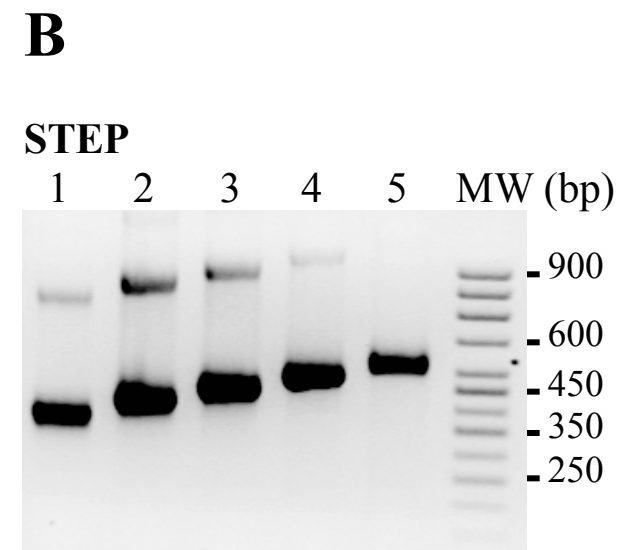
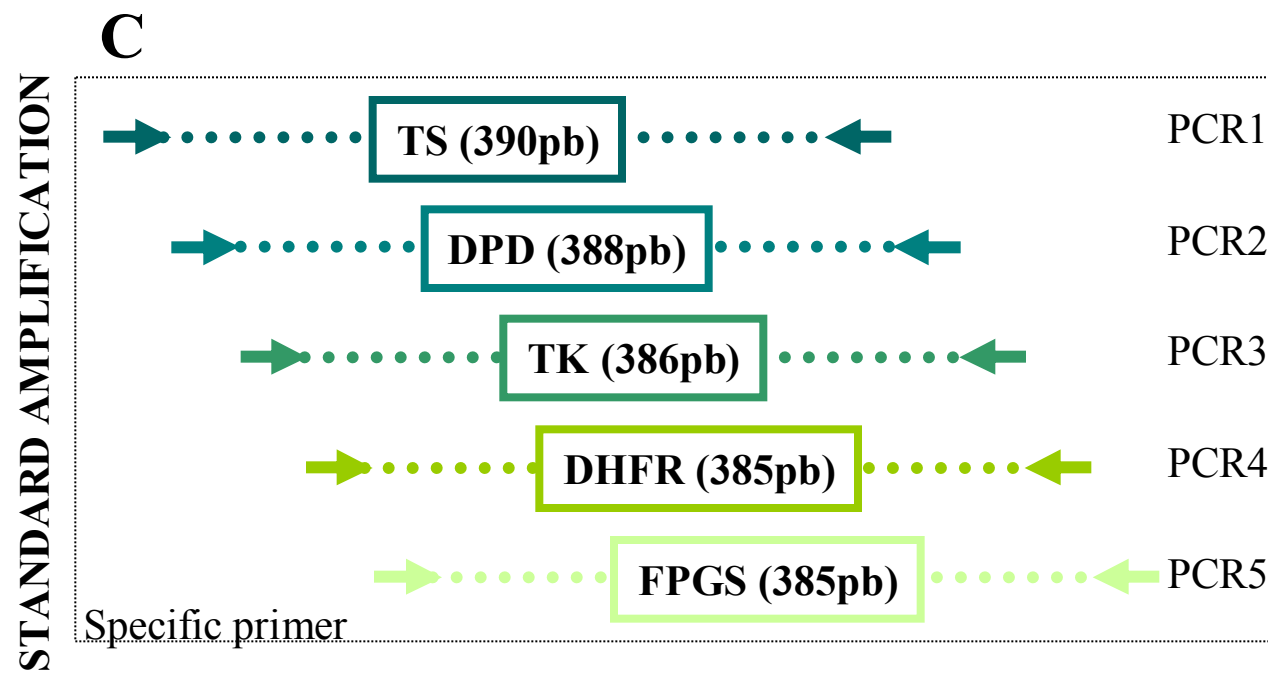
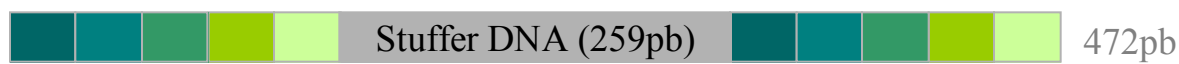
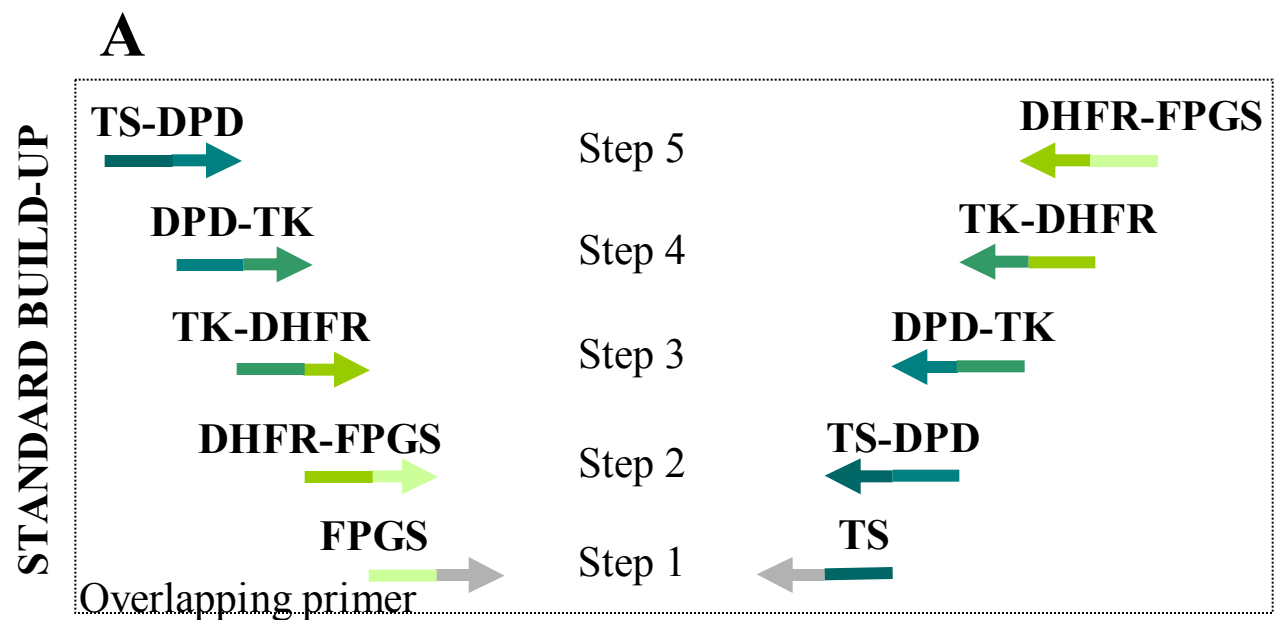
**Figure 2 :** Build-up and amplification of the composite standard DNA. The composite standard DNA was constructed by 5 successive steps of PCR around a 259 bp sequence from the V1a vasopressin Rat receptor using overlapping oligonucleotide primers (A). Each step of primers addition to the construction was checked by gel electrophoresis, and at each step the observed band was used for the next addition of primers sequence (B). After cloning into pGEM-T easy vector the final standard construct was used as a template by PCR in the standard conditions, with every 5 specific primer pairs. Every PCR product had approximately the same length (C, D).

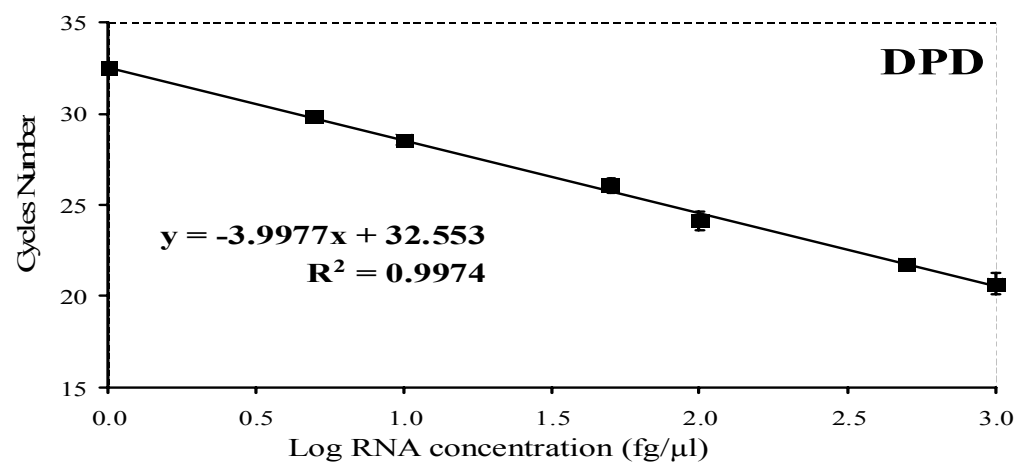
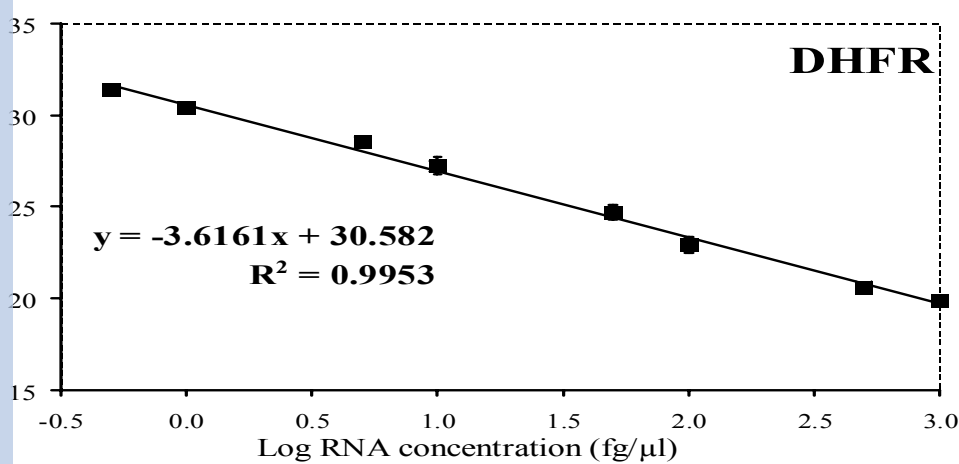
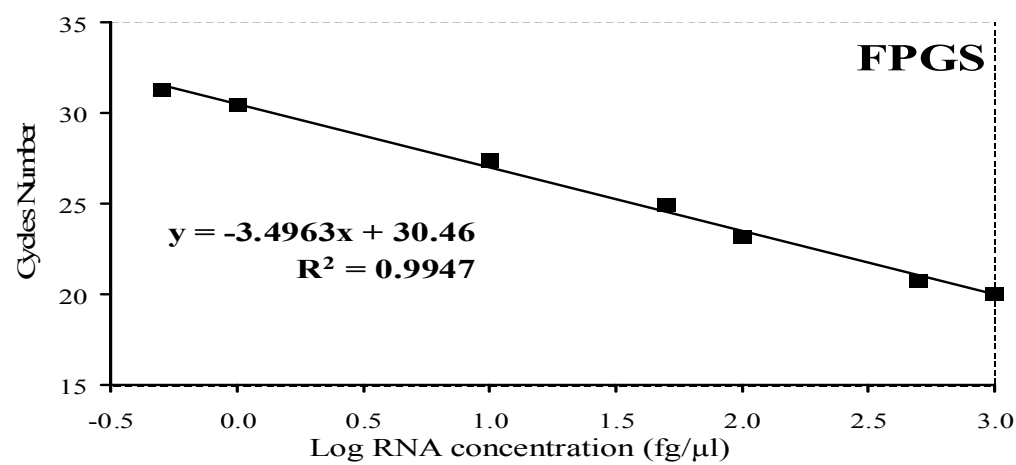
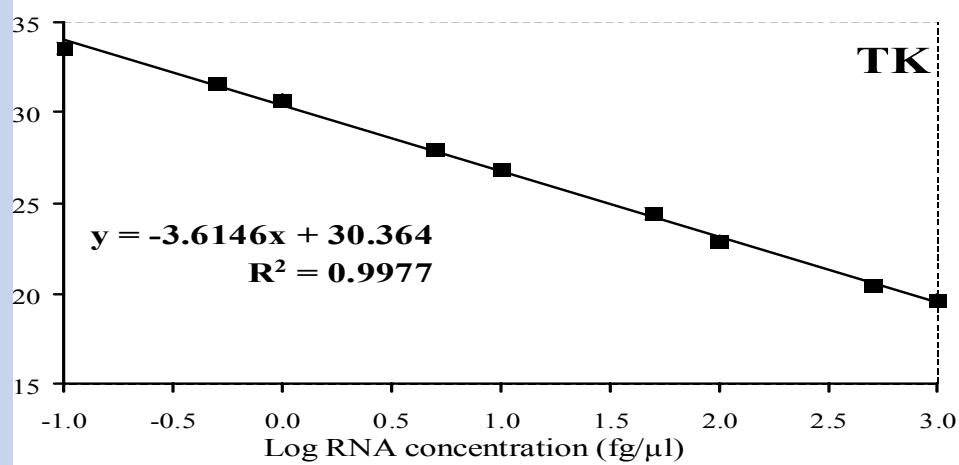
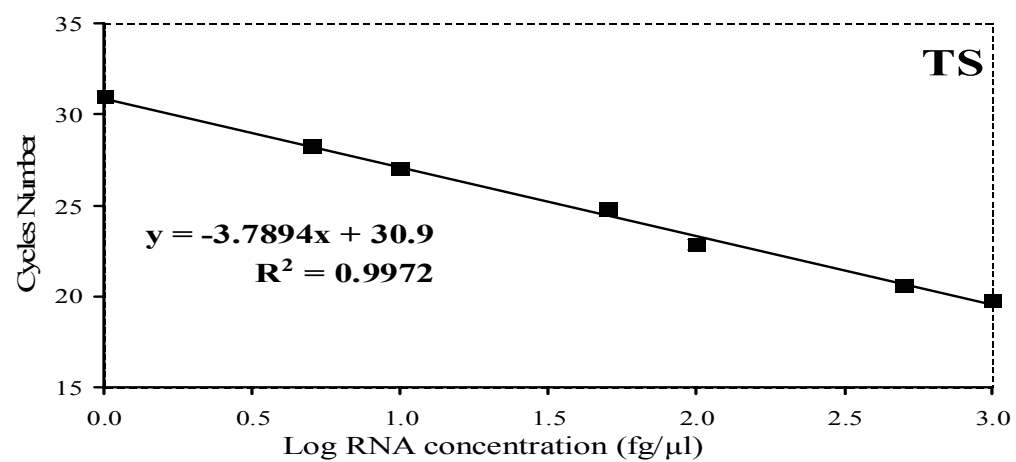
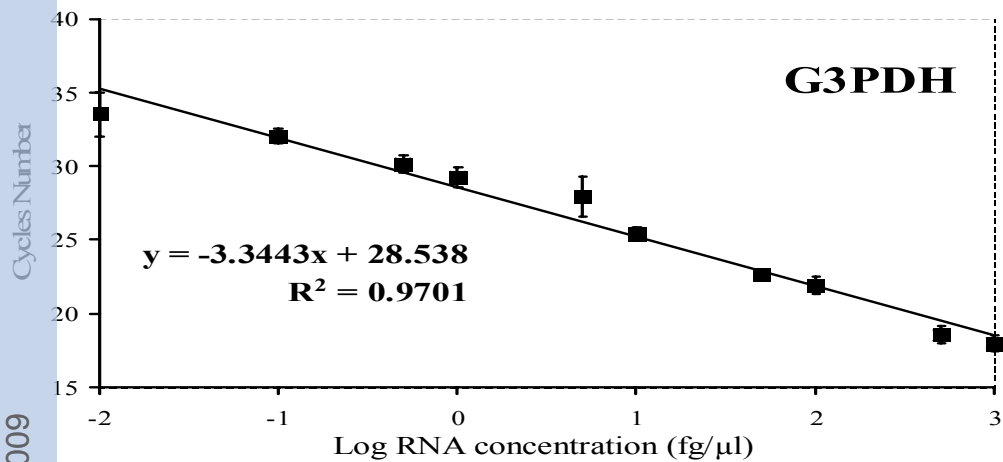
**Figure 3 :** Linear relationship between Cp and logarithm of the starting synthetic RNA concentration. The standard curves were generated with eight points (corresponding to initial amount from 10<sup>3</sup> to 10<sup>-2</sup> µg/µl of synthetic RNA), each point represents the mean ± SD of at least 4 independent experiments.



Amplification curve - Gel Analysis

Melting Peak





**Table I** : Primers couples selected for 5-FU resistance. Selection criteria were: hybridization temperature of primers (55 °C), MgCl<sub>2</sub> concentration (4 mM final), and unique amplification products with length < 500 bp.

5-FU Resistance Factors		Sequence	Length (bp)	T <sub>m</sub> (°C)	Access number
TS	Up	CATTTTGGGGCAGAATACAGAGAT	300	89	NM_001071
	Down	GATGTGCGCAATCATGTACGTGAG			
DPD	Up	TCAGAAGAGCTGTCCAATAAT	229	82.7	NM_000110
	Down	GAATACCTCAGTAGCAAATTGC			
TK	Up	AAAGCACAGAGTTGATGAGAC	475	91.7	NM_00358
	Down	AAGTAGCAGAGCCGACACAC			
DHFR	Up	AGTCAGCGAGCAGGTTCTCA	321	86.7	NM_000791
	Down	GCAGTAAGCCGAGGTCACAC			
FPGS	Up	GTCTTCCTCCAAGAGAAGGT	203	88.8	NM_004957
	Down	GACACCTTGCTTAAAGATGC			
β-Actin	Up	TCAGAAGGATTCCTATGTGG	236	90	AF261085
	Down	AGGTCTCAAACATGATCTGG			
GA3PDH	Up	GAAGGTGAAGGTCGGAGTC	226	85	XM_004814
	Down	GAAGATGGTGATGGGATTTC			

**Table II** : Clinical characteristics of the patients

<b>Variables</b>		<b>No. of patients</b>
<b>Total</b>		33
<b>Sex</b>	Male	22
	Female	11
<b>Age (year)</b>	Median	69
	Range	49-84
<b>Primary tumor site</b>	Left colon	9
	Right colon	8
	Rectum	16
<b>Stage</b>	I	5
	II	5
	III	9
	IV	10
	Local relapse	4
<b>Histology</b>	Well differentiated	18
	Moderately differentiated	11
	Poorly or undifferentiated	4

**Table III :** Comparison of mRNA expression levels of TS, TK, FPGS, DHFR, and DPD between the primary colorectal cancer and the corresponding normal mucosa (Wilcoxon signed rank test).

	<b>TS</b>	<b>TK</b>	<b>FPGS</b>	<b>DHFR</b>	<b>DPD</b>
<b>Colorectal tumor</b>					
Mean ± SD	34.7 ± 15.3	25.9 ± 10.5	35.7 ± 18.8	54.8 ± 24.1	19.5 ± 33.1
Range	14.0 - 68.4	8.4 - 53.1	7.3 - 91.8	18.7 - 98.0	2.7 - 175.7
<b>Normal mucosa</b>					
Mean ± SD	56.6 ± 22.3	32.8 ± 16.46	51.3 ± 35.2	78.3 ± 26.5	47.6 ± 72.9
Range	12.7 - 131.8	0.0 - 80.8	23.6 - 226.4	0.0 - 138.7	12.1 - 418.1
<b>P (Wilcoxon)</b>	<b>&lt;0.0001</b>	<b>0.021</b>	<b>0.017</b>	<b>0.002</b>	<b>0.001</b>

**Table IV** : Comparison of several clinicopathological factors and the mRNA expression levels of TS, TK, FPGS, DHFR and DPD in the colorectal carcinoma (Mann-Whitney *U*-test between 2 groups and the Kruskal-Wallis test for 3 or more groups. A P value less than 0.05 was considered to be statistically significant). Disease extent : "A" include stage I, II and III tumors ; "B" include stage IV and local relapse.

	<b>TS</b>	<b>TK</b>	<b>FPGS</b>	<b>DHFR</b>	<b>DPD</b>
	Mean ± SD (n)	Mean ± SD (n)	Mean ± SD (n)	Mean ± SD (n)	Mean ± SD (n)
<b><u>Disease extent</u></b>					
<b>"A"</b>	36.6 ± 16.1 (20)	26.6 ± 11.7 (20)	33.4 ± 15.8 (20)	62.5 ± 21.8 (20)	12.4 ± 8.9 (17)
<b>"B"</b>	31.7 ± 14.2 (12)	24.9 ± 8.6 (12)	39.6 ± 23.3 (12)	40.9 ± 22.7 (11)	32.9 ± 54.4 (9)
<b>P value</b>	<b>0.276</b>	<b>0.938</b>	<b>0.533</b>	<b>0.010</b>	<b>0.247</b>
<b><u>Histological Type</u></b>					
<b>Well</b>	31.4 ± 12.1 (17)	26.9 ± 12.5 (17)	38.1 ± 18 (17)	58.3 ± 23.5 (18)	14.9 ± 9.8 (13)
<b>Moderatly</b>	34.2 ± 17.2 (11)	23.2 ± 7.6 (11)	32.1 ± 22.5 (11)	49 ± 24.7 (9)	12.0 ± 9.5 (9)
<b>Poorly</b>	50.5 ± 16.8 (4)	29.6 ± 8.3 (4)	35.8 ± 13.3 (4)	52.4 ± 29.9 (4)	51.4 ± 83 (4)
<b>P value</b>	<b>0.210</b>	<b>0.619</b>	<b>0.345</b>	<b>0.509</b>	<b>0.608</b>
<b><u>Localisation</u></b>					
<b>Right colon</b>	44.1 ± 21.4 (7)	27.4 ± 11.4 (7)	34.2 ± 19.9 (7)	57.5 ± 23.7 (7)	16.1 ± 13.3 (7)
<b>Left colon</b>	35.2 ± 14 (9)	25.0 ± 13.6 (9)	37.6 ± 17.9 (9)	54.2 ± 28.9 (9)	11.0 ± 7.0 (6)
<b>Rectum</b>	30.4 ± 11.8 (16)	25.9 ± 8.7 (16)	35.4 ± 20.0 (16)	54.0 ± 23.1 (15)	25.3 ± 45.8 (13)
<b>P value</b>	<b>0.420</b>	<b>0.907</b>	<b>0.889</b>	<b>0.941</b>	<b>0.797</b>
<b><u>Sex</u></b>					
<b>Male</b>	32.2 ± 12.8 (21)	27.5 ± 11.0 (21)	41.8 ± 20.0 (21)	58.2 ± 25.7 (21)	26.0 ± 41.0 (16)
<b>Female</b>	39.5 ± 19.1 (11)	23 ± 9.2 (11)	24.2 ± 8.8 (11)	47.8 ± 20 (10)	9.1 ± 6.3 (10)
<b>P value</b>	<b>0.226</b>	<b>0.258</b>	<b>0.005</b>	<b>0.291</b>	<b>0.023</b>
<b><u>Stage</u></b>					
<b>I</b>	38.8 ± 19.4 (5)	28.4 ± 11.2 (5)	44.3 ± 16.5 (5)	59.1 ± 9.9 (5)	12.9 ± 6.7 (3)
<b>II</b>	38.6 ± 15.2 (5)	22.5 ± 10.1 (5)	23.9 ± 4.4 (5)	57.6 ± 26.7 (5)	9.2 ± 8.0 (5)
<b>III</b>	35.3 ± 17.1 (9)	27.6 ± 14.0 (9)	33.2 ± 17.7 (9)	68.3 ± 25.5 (9)	14.2 ± 11.1 (8)
<b>IV</b>	31.7 ± 16.3 (9)	26.9 ± 7.9 (9)	38.3 ± 23.6 (9)	44.6 ± 23.7 (8)	39.1 ± 67.1 (6)
<b>P value</b>	<b>0.589</b>	<b>0.731</b>	<b>0.199</b>	<b>0.161</b>	<b>0.666</b>

**Table V:** Correlations between mRNA expression levels of TS, TK, FPGS, DHFR and DPD, in the colorectal primary tumor or in the normal mucosa (Spearman rank test, \*\* statistically significant at 0.01, \* statistically significant at 0.05)

Combination	Colorectal tumor			Normal mucosa		
	Coefficient of correlation	(n)	P value	Coefficient of correlation	(n)	P value
<b>TS / DPD</b>	-0.154	(26)	0.452	0.210	(32)	0.248
<b>TS / DHFR</b>	0.103	(30)	0.586	0.090	(30)	0.638
<b>TS / FPGS</b>	-0.460	(32)	0.803	0.561**	(33)	0.001
<b>TS / TK</b>	0.085	(32)	0.642	0.506**	(33)	0.003
<b>TK / DPD</b>	0.543**	(26)	0.004	0.385*	(32)	0.029
<b>TK / DHFR</b>	0.426*	(30)	0.019	0.565**	(30)	0.001
<b>TK / FPGS</b>	0.258	(32)	0.154	0.387*	(33)	0.026
<b>FPGS / DPD</b>	0.685**	(26)	>0.001	0.609**	(32)	>0.001
<b>FPGS / DHFR</b>	0.245	(30)	0.192	0.283	(30)	0.129
<b>DHFR / DPD</b>	0.232	(25)	0.265	0.460*	(29)	0.012