



A simple and highly sensitive radioenzymatic assay for lysophosphatidic acid quantification.

Jean Sébastien Saulnier-Blache, Alexia Girard, Marie-Françoise Simon, Max Lafontan, Philippe Valet

► To cite this version:

Jean Sébastien Saulnier-Blache, Alexia Girard, Marie-Françoise Simon, Max Lafontan, Philippe Valet. A simple and highly sensitive radioenzymatic assay for lysophosphatidic acid quantification.. J Lipid Res, 2000, 41 (12), pp.1947-51. inserm-00110161

HAL Id: inserm-00110161

<https://inserm.hal.science/inserm-00110161>

Submitted on 2 Nov 2006

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

A Simple and Highly Sensitive Radioenzymatic Assay for Lysophosphatidic acid Quantification.

Jean Sébastien Saulnier-Blache*, Alexia Girard, Marie-Françoise Simon, Max Lafontan and Philippe Valet.

I.N.S.E.R.M U317, Institut Louis Bugnard, Université Paul Sabatier, CHU Rangueil, Batiment L3, 31403, Toulouse cedex 04, France.

* To whom correspondence should be addressed

Phone: (33)0562172956

Fax: (33)0561331721

Email: saulnier@rangueil.inserm.fr

Abbreviations: LPA: lysophosphatidic acid; LPAAT: lysophosphatidic acid acyl-transferase; PA:phosphatidic acid; AGPAT: acyl glycerol phosphate acyl transferase; LB: Lubroth ; GC: gas chromatography; PAF: platelet activating factor; PLD: phospholipase D; PLA2: phospholipase A2 ; LDL:low density lipoprotein;.

ABSTRACT

The objective of the present work was to develop a simple and sensitive radioenzymatic assay to quantify lysophosphatidic acid (LPA). For that, a recombinant rat lysophosphatidic acid acyl transferase (LPAAT) produced in *Escherichia coli* was used. In the presence of [^{14}C]oleoyl CoA, LPAAT selectively catalyzes the transformation of LPA and alkyl-LPA into [^{14}C]phosphatidic acid. Acylation of LPA was complete and linear from 0 to 200 pmoles with a minimal detection of 0.2 pmoles. This method was used to quantify LPA in butanol-extracted lipids from bovine sera, as well as from human and mouse plasma. This radioenzymatic assay represents a new, simple, and highly sensitive method to quantify LPA in various biological fluids.

Key words: radioenzymatic detection, lysophosphatidic acid, lysophosphatidic acid acyl-transferase, phosphatidic acid, serum, plasma, [^{14}C]oleoyl-CoA, butanol, recombinant protein, *Echerichia coli*, bioactive phospholipid, thin layer chromatography.

Introduction

Lysophosphatidic acid (LPA: 1-acyl-2-hydroxy-*sn*-glycero-3-phosphate), is a key intermediate in glycerolipid synthesis, and a bioactive phospholipid (1, 2). It controls a wide variety of cellular responses (mitogenesis, cytoskeletal rearrangements, cell adhesion, ion transport, apoptosis) through the activation of specific G-protein coupled receptors (3, 4).

LPA has been detected in various biological fluids including blood (5-9), aqueous humors (10), or adipose tissue microdialysates (11). Abnormally high concentrations of LPA have been found in ascites and plasma from patients bearing ovarian cancer (12, 13). Several cell types have been shown to produce LPA in vitro: platelets (14, 15), adipocytes (11), ovarian cancer cells 16.. LPA has also been shown to be produced from oxidized LDL (17). Several possible metabolic pathways could be involved in cellular production of LPA: phospholipase A2-dependent deacylation of phosphatidic acid (18), lysophospholipase D-dependent hydrolysis of lysophosphatidylcholine (19, 20), acyl transferase-dependent acylation of monoacyl glycerol-phosphate (21), or acyl dihydroxyacetone-phosphate reductase-dependent reduction of acyl dihydroxyacetone-phosphate(22).

In order to understand the precise role of LPA in the development of particular pathologies such as cancer or obesity, and to clarify the metabolic origin of LPA produced by the cells, a precise and sensitive detection of LPA is necessary. In the present work, a recombinant lysophosphatidic acid acyltransferase (LPAAT) was used to develop a simple and highly sensitive radioenzymatic assay to quantify LPA in various biological fluids.

Material and Methods

Preparation of recombinant LPAAT:

The rat 1-acyl-*sn*-glycerol-3-phosphate acyltransferase cDNA encoding for an enzyme behaving as a LPAAT was previously cloned in a bacterial expression vector pTrcHis (23). Fifty μ l of DH5 α -competent-cell suspension (Life Technologies) were transformed with 0.5 μ g of pTrcHis-AGPAT plasmide (generous gift from Dr. Kazuhiko Kume) and grown in LB media containing 50 μ g/ml ampicilin. When A600 reached approximately 0.6, bacteria were cultured for 3 hours in the presence of 1 mM IPTG (isopropyl- β -D-thiogalactopyranoside). Bacteria were collected by centrifugation at 3000 g for 10 min, and the pellet was disrupted by sonication in 0.2 M Tris-HCl (pH 7.4). After centrifugation at 10,000 x g for 20 min, the supernatant was recovered and further centrifuged at 100,000 x g (microsomes) for 90 min. The yield of a typical preparation is 6 to 7 mg microsomal protein per liter of culture. The resulting pellet was resuspended in the same buffer to a final protein concentration of 1 μ g/ μ l and was used for radioenzymatic detection of LPA. Kinetic analysis allowed to determined that the activity of transformation of LPA into PA of a typical microsome preparation varied from 0.8 to 1 nmol/min/mg protein.

Preparation of acyl- and alkyl-LPA stock solutions

Na⁺ salt 1-oleoyl-*sn*-glycero-3-phosphate (LPA) (from SIGMA) was solubilized in PBS buffer containing 1% lipid-free albumin (from ICN). 1-O-hexadecyl-*sn*-glycero-3-phosphate (alkyl-LPA) was obtained from 1-O-hexadecyl-2-acetyl-*sn*-glycerol-3-phosphocholine (PAF) (from SIGMA). PAF was dispersed by sonication in a 50 mM Tris buffer (pH 7.4) containing 5 mM Ca⁺⁺, and incubated first with pig pancreas PLA₂ (from Sigma) for 2h followed by 30 min treatment with *Streptomyces chromofuscus* PLD (from SIGMA). Lipids were extracted using Bligh and Dyer procedure (24) in acid conditions. After evaporation alkyl-LPA was solubilized in ethanol. The real concentration of the stock solutions of LPA and alkyl-LPA was determined by phosphorus measurement (25).

Lipid extraction from serum and plasma

1-oleoyl-LPA or lipids contained in serum or plasma were extracted with 1 volume of 1-butanol. After vigorous shaking and centrifugation (5 min at 3000 g), the upper butanol phase was collected and evaporated under nitrogen at 50°C. This procedure allowed 85 to 90% extraction of LPA.

In vitro acylation of LPA

Dry lipid extract was resuspended in 200 μ l of reaction medium (1 μ l [¹⁴C]oleoyl CoA (RAS 55 mCi/mmol, NEN), 20 μ l Tris (pH 7.5) 200 mM, 10 μ l of recombinant LPAAT, 8 μ l of sodium orthovanadate 500 μ M, and 161 μ l H₂O containing 1mg/ml Tween 20), and

incubated for 120 min at 20°C. The mixture was vortexed every 15 min. The reaction was stopped by addition of 400 µl of CHCl₃/MeOH/HCl 12M (40/40/0.26) followed by a vigorous shaking and a 10 min centrifugation at 3000 g. The lower CHCl₃ phase was evaporated under nitrogen, resuspended in 20 µl of CHCl₃/MeOH (1/1), and spotted on a silica gel 60 TLC glass plate (Merck), then separated using CHCl₃/MeOH/NH₄OH/H₂O (65/25/0.9/3) as a solvent. Alternatively, a two dimensional TLC using CHCl₃/MeOH/NH₄OH/H₂O (65/25/0.9/3) as the solvent of the first migration, and CHCl₃/MeOH/acetic acid/H₂O (40/20/5/0.5) as the solvent of the second migration, was performed. The plate was autoradiographed overnight to localize [¹⁴C]-labeled lipids. Identification of radioactive spots was performed by co-migration with cold lipids visualized under iodine vapors. [¹⁴C]phosphatidic acid spots were then scraped and counted with 3 ml of scintillation cocktail. The following formula was used to convert radioactivity in pmoles: $dpm = 1/(2.22 \times RAS)$ pmoles, RAS being the specific radioactivity (of [¹⁴C]oleoyl CoA).

Origin of bovine sera

Fetal calf sera were from BioWhittaker Europe (Lot #6SB0011), Gibco Life Technologies (Lot #4001585K) and Boehringer Mannheim France (Lot #210463). Donor calf sera were from BioWhittaker Europe (Lot #6M1955) and Gibco Life Technologies (Lot #3016082D).

Blood collection

Human plasma was collected on EDTA with the consent of the subjects and was approved by the Ethical Committee of the Hospital. Mouse plasma was collected from FVB mice following carotid section. Collection was performed in tubes containing 50 µl of a 2% solution of EDTA, disodium salt. The tubes were centrifuged for 5 min at 1500 g and LPA was measured in the platelet poor plasma. Animals were handled in accordance with the principles and guidelines established by the National Institutes of Medical Research (INSERM).

Results

The ability of the LPAAT to acylate LPA into PA was tested after semi-purification of the enzyme from rat LPAAT cDNA-transformed *E. coli*. For that, 1-oleoyl-LPA or butanol-extracted lipids from serum (known to contain significant amount of LPA (6-9)) were incubated in the presence of LPAAT-transformed *E. coli* microsomes and [^{14}C]oleoylCoA. In order to identify the products of the reaction, they were separated by two dimensional thin layer chromatography (Figure 1).

In the absence of LPA (cont), no [^{14}C]PA was detected. A radioactive spot was found at the depot (start). It corresponded to traces of [^{14}C]oleoylCoA extracted with 1-butanol. Two additional spots (up right corner) were also observed. The lower spot comigrated with cold oleic acid (not shown) and was therefore identified as [^{14}C]oleate. Since [^{14}C]oleate was also observed in the absence of microsomes (not shown), and resulted from a partial degradation of [^{14}C]oleoylCoA. The upper spot comigrated with diacylglycerol and triacylglycerol and was therefore identified as [^{14}C]neutral lipids likely resulting from the presence of monoacyl- and/or diacyl-glycerol acyltransferases endogenously expressed in *E. coli* and co-purified with LPAAT in microsomes.

In the presence of LPA (standard or from serum) a major spot comigrating with cold PA was observed. [^{14}C]PA was interpreted as the result of the transfert of [^{14}C]oleate onto LPA. This confirming the presence of LPAAT in *E. coli* microsomes. A minor but nonidentified spot was also observed. Because it represented less than 2% of [^{14}C]PA this minor spot was further neglected.

Whereas the vast majority (about 90 %) of LPA species found in biological fluids and tissues are acyl-LPA, a small proportion (about 10%) are alkyl-LPA (26). LPAAT-dependent acylation of alkyl-LPA was tested. Alkyl-LPA was synthesized from PAF (see Methods) and exposed to semi-purified LPAAT and [^{14}C]oleoylCoA. As shown in figure 2, both acyl-LPA or alkyl-LPA led to the formation of [^{14}C]PA. No [^{14}C]PA was formed from PAF or lyso-PAF.

The sensitivity of radioenzymatic assay was then tested. For that, increased quantities of 1-oleoyl-LPA were incubated in the presence of semi-purified LPAAT and [^{14}C]oleoylCoA. This led to the formation of increased amount of [^{14}C]PA (Figure 3A). Conversion of the radioactivity into pmoles revealed that LPA acylation into PA was complete and linear (slope close to 1) from zero to 200 pmoles, with a detection set point of 0.2 pmole (Figure 3-B).

The radioenzymatic assay was finally used to quantify LPA in bovine sera from different origin. As shown in Table 1, the concentration of LPA measured in 20 μl of serum varied from 0.28 to 2.56 μM . Concentration of LPA in donor calf sera was significantly higher (2.4 to 4.7 fold) than the mean concentration (0.54 μM) of LPA found in fetal calf sera. Although present at much lower concentrations than in serum, significant concentrations of LPA (from 51 to 283 nM) were also found in human and mouse platelet -poor plasma (Table 2).

Discussion

Several procedures have previously been used to detect LPA. Gas chromatography (GC) allows quantification of methylated fatty acids derived from TLC-purified LPA (15). This is an indirect assay allowing determination of the relative fatty acid composition of LPA. However, this method is not very sensitive and requires relatively high amounts of purified LPA. In addition, classical GC does not allow to determine the presence alkyl-LPA. For that it is necessary to employ both mild alkaline hydrolysis and gas chromatography coupled to mass spectrometry (26).

The presence of LPA in a biological fluid can also be demonstrated by using bioassays. This type of assay consists in measuring the influence of a LPA-containing fluid on a cellular response known to be controlled by LPA, such as platelet aggregation (17), calcium mobilization in *xenope* oocytes (10), or cell spreading (11). Since bioactive phospholipids other than LPA are also able to generate those biological responses, bioassays cannot be considered specific enough for LPA quantification. LPA can also be detected following labeling cellular phospholipid with [³²P]ortho-phosphate, and separation of [³²P]LPA by TLC (7, 11, 18). Although very sensitive, this method is only applicable on cultured cells, and is not quantitative.

The LPAAT, also known as 1-acyl-*sn*-glycerol-3-phosphate acyltransferase (EC 2.3.1.51) catalyses LPA conversion into phosphatidic acid (PA), a key step in glycerolipid synthesis. LPAAT has been demonstrated to be highly specific for LPA and not influenced by LPA fatty acid composition (27). Therefore the high specificity of LPAAT made this enzyme an interesting partner to develop a new assay for LPA quantification.

By using a rat recombinant LPAAT produced in *E. coli* (23) we have developed a simple, rapid, and highly sensitive radioenzymatic assay to quantify LPA in various biological fluids. By using this assay it is possible to quantify LPA in crude butanol-lipid extracts without prior time consuming and poorly efficient TLC-purification of LPA. It is also important to notice that, this assay allows to quantify both acyl- and alkyl-LPA. Although present to a much lower concentration than acyl-LPA, alkyl-LPA has been reported to be much more active as acyl-LPA on platelet aggregation(28). Although our assay does not discriminate between acyl-LPA and alkyl-LPA, the amount of alkyl-LPA present in a biological fluid can nevertheless be determined. For that the biological fluid can be treated with a lysophospholipase, such as the phospholipase B, known to specifically hydrolyze acyl-LPA without altering alkyl-LPA. Phospholipase B-resistant LPA will correspond to alkyl-LPA.

Our radioenzymatic assay is also highly sensitive. It is possible to detect less than 0.5 pmole LPA. This high sensitivity allows determination of LPA concentration from low volumes of biological samples. It was for example possible to quantify LPA in 200 µl of plasma or 25 µl of serum. Those volumes are entirely compatible with current blood sampling in both human and animals. This high sensitivity allowed us to easily quantify as low as 51

nM LPA in human and mouse plasma. It is important to notice that, by using gas chromatography, Xu et al. considered that the lower detectable concentration of LPA in human plasma was 100 nM (5).

Finally, the method is reliable since the concentration values determined in bovine sera are in agreement with those previously published (6-9). However, it is noticeable that donor calf sera contain a higher concentration of LPA than fetal calf sera. Although the precise reasons of this difference remain to be determined, this information needs to be taken into consideration to choose cell culture serum.

In conclusion, this radioenzymatic assay represents a new, and highly sensitive method for LPA quantification. Because of its simplicity and rapidity of execution, this method could easily be introduced in any research or hospital laboratory interested in fundamental or pathological issues concerning LPA.

Acknowledgements: We thank Dr. Kazuhiko Kume (University of Tokyo) for having generously provided us with pTrcHis-AGPAT vector. We also thank David Sibrac for his skillful technical assistance and Stéphane Gesta for fruitful discussion. This work was supported by grants from the “Institut National de la Santé et de la Recherche Médicale” (APEX #4X405D), the “Association pour la Recherche sur le Cancer” (#5381), the “Institut de Recherche Servier”, and the “Laboratoires Clarins”.

References

1. Gaits, F., Fourcade, O., F, L. B., Gueguen, G., Gaigé, B., Gassama-Diagne, A., Fauvel, J., Salles, J.-P., Mauco, G., Simon, M.-F. and Chap, H. 1997. Lysophosphatidic acid as a phospholipid mediator: pathways of synthesis. *FEBS Lett.* 410: 54-58.
2. Jalink, K., Hordijk, P. L. and Moolenaar, W. H. 1994. Growth factor-like effects of lysophosphatidic acid, a novel lipid mediator. *Biochim. Biophys. Acta.* 1198: 185-196.
3. Goetzl, E. and An, S. 1998. Diversity of cellular receptors and functions for the lysophospholipid growth factors lysophosphatidic acid and sphingosine 1-phosphate. *FASEB J.* 12: 1589-1598.
4. Chun, J., Contos, J. J. and Munroe, D. 1999. A growing family of receptor genes for lysophosphatidic acid (LPA) and other lysophospholipids (LPs). *Cell Biochemistry and Biophysics* 30: 213-242.
5. Xu, Y., Shen, Z., Wiper, D. W., Wu, M., Morton, R., Elson, P., Kennedy, A., Belinson, J., Markman, M. and Casey, G. 1998. Lysophosphatidic acid as a potential biomarker for ovarian and other gynecologic cancers. *JAMA* 280: 719-723.
6. Tigyi, G. and Miledi, R. 1992. Lysophosphatidates bound to serum albumin activate membrane currents in *Xenopus* oocytes and neurite retraction in PC12 pheochromocytoma cells. *J. Biol. Chem.* 267: 21360-21367.
7. Eichholtz, T., Jalink, K., Fahrenfort, I. and Moolenaar, W. H. 1993. The bioactive phospholipid lysophosphatidic acid is released from activated platelets. *Biochem. J.* 291: 677-680.
8. Tokumura, A., Iimori, M., Nishioka, Y., Kitahara, M., Sakashita, M. and Tanaka, S. 1994. Lysophosphatidic acids induce proliferation of cultured vascular smooth muscle cells from rat aorta. *Am. J. Physiol.* 267: C204-C210.
9. Sasagawa, T., Suzuki, K., Shiota, T., Kondo, T. and Okita, M. 1998. The significance of plasma lysophospholipids in patients with renal failure on hemodialysis. *J Nutr Sci Vitaminol* 44: 809-18.
10. Liliom, K., Guan, Z., Tseng, J.-L., Desiderio, D. M., Tigyi, G. and Watsky, M. A. 1998. Growth factor-like phospholipids generated after corneal injury. *Am. J. Physiol.* 274: C1065-C1074.
11. Valet, P., Pagès, C., Jeanneton, O., Daviaud, D., Barbe, P., Record, M., Saulnier-Blache, J. and Lafontan, M. 1998. Alpha2-adrenergic receptor-mediated release of lysophosphatidic acid by adipocytes. *J. Clin. Invest.* 101: 1431-1438.
12. Xu, Y., Gaudette, D., Boynton, J., Frankel, A., Fang, X., Sharma, A., Hurteau, J., Casey, G., Goodbody, A., Mellors, A., Holub, B. and Mills, G. 1995. Characterization of an ovarian cancer activating factor in ascites from ovarian cancer patients. *Clin. Cancer Res.* 1: 1223-1232.

13. Westermann, A.M., Havik, E., Postma, F.R., Beijnen, J.H., Dalesio, O., Moolenaar, M.W. and Rodenhuis, S. 1998. Malignant effusions contain lysophosphatidic acid (LPA)-like activity. *Ann. Oncol.* 9: 437-442.
14. Mauco, G., Chap, H., Simon, M.-F. and Douste-Blazy, L. 1978. Phosphatidic and lysophosphatidic acid production in phospholipase C- and thrombin-treated platelets. Possible involvement of a platelet lipase. *Biochimie* 60: 653-661.
15. Gerrard, J. M. and Robinson, P. 1989. Identification of the molecular species of lysophosphatidic acid produced when platelets are stimulated by thrombin. *Biochim. Biophys. Acta.* 1001: 282-285.
16. Shen, Z., Belinson, J., Morton, R. and Yan, X. 1998. Phorbol 12-myristate 13-acetate stimulates lysophosphatidic acid secretion from ovarian and cervical cancer cells but not from breast or leukemia cells. *Gynecol Oncol* 71: 364-368.
17. Siess, W., Zangl, K., Essler, M., Bauer, M., Brandl, R., Corrinth, C., Bittman, R., Tigyi, G. and Aepfelbacher, M. 1999. Lysophosphatidic acid mediates the rapid activation of platelets and endothelial cells by mildly oxidized low density lipoprotein and accumulates in human atherosclerotic lesions. *Proc Natl Acad Sci U S A* 96: 6931-6936.
18. Fourcade, O., Simon, M. F., Viodé, C., Rugani, N., Leballe, F., Ragab, A., Fournié, B., Sarda, L. and Chap, H. 1995. Secretory phospholipase A2 generates the novel lipid mediator lysophosphatidic acid in membrane microvesicles shed from activated cells. *Cell* 80: 919-927.
19. Tokumura, A., Harada, K., Fukuzawa, K. and Tsukatani, H. 1986. Involvement of lysophospholipase D in the production of lysophosphatidic acid in rat plasma. *Biochim Biophys Acta* 875: 31-38.
20. Tokomura, A., Miyake, M., Nishioka, Y., Yamano, S., Aono, T. and Fukuzawa, K. 1999. Production of lysophosphatidic acids by lysophospholipase D in human follicular fluids of in vitro fertilization patients. *Biol Reprod* 61: 195-199.
21. Halder, D. and Vancura, A. 1992. Glycerophosphate acyltransferase from liver. *Methods Enzymol* 209: 64-72.
22. Dunlop, M. E. and Larkins, R. G. 1985. Pancreatic islets synthesize phospholipids de novo from glucose via acyl-dihydroxyacetone phosphate. *Biochem. Biophys. Res. Commun.* 132: 467-473.
23. Kume, K. and Shimizu, T. 1997. cDNA cloning and expression of murine 1-acyl-sn-glycerol-3-phosphate acyltransferase. *Biochem. Biophys. Res. Com.* 237: 663-666.
24. Bligh, E. and Dyer, W. 1959. A rapid method of total lipid extraction and purification. *Can. J. Biochem. Physiol.* 37: 911-917.
25. Böttcher, C., Gent, C. V. and Priest, C. 1961. A rapid and sensitive submicrophosphorus determination. *Anal. Chim. Acta* 24: 203-204.
26. Sugiura, T., Nakane, S., Kishimoto, S., Waku, K., Yoshioka, Y., Tokomura, A. and Hanahan, D. 1999. Occurrence of lysophosphatidic acid and its alkyl ether-linked analog in rat

brain and comparison of their biological activities toward culture neural cells. *Biochim Biophys Acta* 1440: 194-204.

27. Yamashita, S., Hosaka, K., Miki, Y. and Numa, S. 1981. Glycerolipid acyltransferases from rat liver: 1-acylglycerophosphate acyltransferase, 1-acylglycerophosphorylcholine acyltransferase and diacylglycerol acyltransferase. *Methods Enzymol.* 71: 528-536.

28. Simon, M. F., Chap, H. and Douste-Blazy, L. 1982. Human platelet aggregation induced by 1-alkyl-lysophosphatidic acid and its analogs: a new group of phospholipid mediators? *Biochem Biophys Res Commun* 108: 1743-50.

Legend of the Figures

Figure 1: LPAAT-dependent acylation of LPA into [^{14}C]PA

Lipids were butanol-extracted from 2ml PBS buffer containing either 200 pmoles of 1-oleoyl-LPA (LPA), or 300 μl of fetal calf serum (serum), or nothing (cont). After evaporation, lipids were incubated in the presence of semi-purified LPAAT and [^{14}C]oleoylCoA as described in Material and Methods. The products of the reaction were separated by two dimensional TLC and autoradiographed as described in Material and Methods.

Figure 2: LPAAT-dependent acylation of alkyl-LPA in [^{14}C]PA

Alkyl-LPA was synthesized from PAF as described in Material and Methods. 200 pmoles of LPA, PAF, lyso-PAF and alkyl-LPA were incubated in the presence of semi-purified LPAAT and [^{14}C]oleoylCoA as described in Material and Methods. The products of the reaction were separated by one dimensional TLC and autoradiographed as described in Material and Methods.

Figure 3: Sensitivity and linearity of LPAAT-dependent acylation of LPA into [^{14}C]PA.

Butanol-extracted 1-oleoyl-LPA (0 to 200 pmoles) was evaporated and incubated in the presence of semi-purified LPAAT and [^{14}C]oleoylCoA as described in Material and Methods. The products of the reaction were separated by one dimensional TLC and autoradiographed (A) as described in Material and Methods. Each spot of [^{14}C]PA was scraped, counted, and the radioactivity was converted into pmoles (B). The insert in (B) represents the lowest amounts of LPA.

Tables

Table 1 : Quantification of LPA in different bovine sera.

Calf Sera	Company	Concentration of LPA (μ M)
Fetal	BioWhittaker	0.65 ± 0.21
	GIBCO	0.28 ± 0.04
	Boehringer	0.71 ± 0.07
Donor	BioWhittaker	2.56 ± 0.43
	GIBCO	1.31 ± 0.11

Lipids present in 25 μ l of fetal or donor calf serum were extracted with butanol, evaporated, and incubated in the presence of semi-purified LPAAT and [14 C]oleoylCoA as described in Material and Methods. The products of the reaction were separated by one dimensional TLC and autoradiographed as described in Material and Methods. Each spot of [14 C]PA was scraped and counted. The radioactivity was converted into pmoles and the concentration of LPA was calculated. Values corresponded to mean \pm SEM from 3 separate determinations.

Table 2 : Quantification of LPA in human and mice plasma

	Concentration of LPA (nM)	n
Human	83 ± 8 (51-136)	14
Mouse	170 ± 50 (80 - 283)	12

Lipids present in 200 μ l of human or mouse platelet-poor plasma were extracted with butanol, evaporated, and incubated in the presence of semi-purified LPAAT and [14 C]oleoylCoA as described in Material and Methods. The products of the reaction were separated by one-dimensional TLC and autoradiographed as described in Material and Methods. Each spot of [14 C]PA was scraped, counted. The radioactivity was converted into pmoles and the concentration of LPA was calculated. Values correspond to mean \pm SEM from n separate determinations. Extreme values are reported in parenthesis.